

Morpho-Physiological and Yield Responses of Sweet Potato (*Ipomoea batatas* (L.) Lam.) Genotypes to Frequency of Irrigation under Greenhouse Condition

ABSTRACT

Introduction: The sweet potato (*Ipomoea batatas* Lam.), is one of the root and tuber crops grown from low land to high land region of Ethiopia. However, its productivity depends on adaptability and tolerance to different environmental stresses and the capacity of the crop to enhance water use efficiency under moisture stress conditions. The objective of this study was to evaluate impact of irrigation interval on morpho-physiological characteristics of sweet potato varieties. **Methodology:** The trail was a 3 x 2 factorial arrangement in CRD design consisting: three irrigation intervals (daily-control), four days and seven days interval) combined with two sweet potato genotypes (Awassa-83 and Kulfo) with three replications. **Result:** The morpho-physiological indicators, morphological traits, water use efficiency (WUE), Relative leaf water content (RLWC), leaf gas exchange, stomata density, and tuber yield were evaluated. The result indicated that morphological traits were significantly ($P \leq 0.05$) responded to genotype and irrigation frequencies. As compared to daily irrigation, an extended watering interval to seven days irrigation interval significantly reduced leaf number, vine length, branch number and internode length by 34%, 20%, 27% and 19%, respectively. Stomata density was strongly responded to genotypes than effect of irrigation frequency. Genotype Awassa-83 had approximately 2.0 more stomata per mm^2 than genotype Kulfo regardless to irrigation frequency. The interaction effect between genotype and irrigation frequency revealed significant influence on photosynthesis and transpiration rate. The rate of assimilate accumulation was significantly reduced (by 62%) in Awassa-83 irrigated due to extended irrigation interval to seven days than variety irrigated daily. Delay irrigation for four and seven days reduced transpiration rate in genotype Awassa-83 by 22% and 25%, respectively. Result on WUE indicated that Kulfo was found better in efficiently utilizing water under extended irrigation interval than Awassa 83. The leaf water content was significantly ($P \leq 0.001$) responded to irrigation frequency than genotypes. The higher leaf relative water content was obtained from daily irrigation than extended irrigation interval. **Conclusion:** Finally it was observed that tuber yield under daily and four days irrigation interval was not statistically different in both varieties, This is therefore, the four days irrigation interval is recommended for sweet potato production from farmers economic point of view

Key words: Photosynthesis, Stomata, genotype, WUE, sweet potato, tuber yield

INTRODUCTION

Sweet potato (*Ipomoea batatas* L. Lam) is a dicotyledonous and tuberous root crop which belongs to the genus *Ipomoea* of the family Convolvulaceae that believed to be originated in the Central America (Norman *et al.*, 1995). Among these approximately 50 genera and more than 1,000 species of Convolvulaceae, Some members of the family are weeds (e.g. hedge bindweed, *Convolvulus seppephum* L.) and ornamentals (e.g. morning glory, *Ipomoea*

45 *purpurea* (L) Roth) (Okereke et al., 2015) but *Ipomoea batatas* is the only crop plants of
46 major importance as food (Bovell-Benjamin, 2007; Onwueme and Charles, 1994) .
47 Production of sweet potato in the world is about 106.5 million tons of tubers with a
48 productivity of 4–6 MT/ha. In Ethiopia Sweet potato is the third most important root and
49 tuber crops next to Enset (*Ensete ventricosum*) and potato in terms of area and total
50 production. Even if it grows in most parts of the country at elevation from 1000 -2500 m.a.s.l
51 altitude and (between 3-15°N and 33-48°E) latitude, 96 % of the production area is covered
52 by the Southern Nations Nationalities People’s Region state (SNNPRS) and Oromia region of
53 Ethiopia. Sweet potato is used as human food, animal feed and human health and raw
54 material for industrial production of starch, sugar and alcohol (Woolfe, 1992). The yellow
55 fleshed variety is a good source of beta-carotene, sources of vitamin A which are used to
56 alleviate problem of night blindness of millions of children in sub-Saharan Africa including
57 Ethiopia (Taboge et al., 1994). The wide range of variation in productivity can be related to
58 difference in climatic factors including; UV- radiation, water stress, temperature, relative
59 humidity, altitude as well as, crop genotype variation (Zelege, 2010). Sweet potatoes are
60 often cultivated on non-irrigated lands and have been considered drought tolerant if some
61 drought happen near the end of its life cycle (Cattivelli et al., 2008; ZHANG et al., 2001).
62 However, soil moisture stress particularly at early growth stage is a crucial factor that limits
63 its growth and development through affecting storage root production and yield (Pardales and
64 Yamauchi, 2003). In addition, water stress also causes a reduction in growth rate, stem
65 elongation, leaf expansion and stomatal movements and changes in a number of
66 physiological and biochemical processes governing plant growth and productivity
67 (Fernández, 2014). Moreover, physiological and morphological process like water-use
68 efficiency, growth performance and above-ground biomass of sweet potato are very sensitive
69 to water stress and generally leads to loss of storage root’s productivity(Daryanto et al.,
70 2017). In sweet potato, the function of stomatal closure and reduce CO₂ assimilation, under
71 water deficit stress has been well studied, especially in the sensitive genotypes (Kubota,
72 2003) and the stresses may cause a variety of plant responses which can be additive,
73 synergistic or antagonistic(Fernández, 2014).
74 Therefore, the purpose of this study was to investigate the effect of genotypes and irrigation
75 interval and compare their effect and interaction on growth, physiology, yield and adaptive
76 mechanism of two sweet potato varieties.

77 MATERIALS AND METHODS

78 Description of the Study Areas

79 The study was conducted at Hawassa, main campus of Hawassa University, under
80 greenhouse condition, during September 2016 to March 2017. Hawassa is located at 7° 04’N ,
81 and 38° 31’ E on the escarpment of the Great Rift valley with an elevation of 1700 meters
82 above sea level, which is located about 275 km south of Addis Ababa, the capital city of
83 Ethiopia. The mean annual rainfall and temperature of Hawassa are 900-1100mm and 27 °C,
84 respectively. The yearly average maximum and minimum temperature of the area was 26 °C
85 and 12.4 °C, respectively. In general, the area receives short rainy season (March-May),
86 “Belge” and long rainy season (July-October), “Meher”.

87 **Planting Materials and Description of the Genotypes**

88 Two sweet potato genotypes known as Awassa-83 and Kulfo were collected from Southern
89 Agricultural research institute. They are well performing sweet potato genotype in terms of
90 yield, nutritional value and under wide range of agro ecological conditions.

91

92 Table 1. Description of varieties used for the experiment

Variety	Altitude (m.a.s.l)	Maturity (days)	Flesh color	Yield tone/ha	Years of release
Awassa-83	1500-2500	150-180	White	36.6	1998
Kulfo	1200-2000	150	Orange	31.5	2005

93 Source: (MoARD, 2009)

94 **Experimental Design and Treatments**

95 A factorial experiment with completely randomized design (CRD) with three levels of
96 irrigation frequency (daily watering, four days interval and seven days interval) and two
97 sweet potato genotypes (Awassa-83 and Kulfo) was used to run the pot(pan) experiment
98 under partially automated greenhouse condition. The pan was field with soil collected from
99 field and air dried for three weeks so as to have a constant weight. Then after, a total of 90
100 experimental pots of 16.9 L volume, which accommodate 17.2 kg of soil per pans was filled
101 with soil, which was calculated based on the bulk density of the soil. Tip cutting of each
102 genotypes, 30 cm long, were planted directly in each pan.

103 **Greenhouse Climate Condition**

104 The greenhouse was partially automated to regulate temperature through side and roof
105 ventilation system. During the experimental period (150 days) ambient air humidity was
106 maintained through regulation of vents and manual irrigation system. Temperature and
107 relative humidity data were recorded on randomly selected 25 days using mini data loggers
108 (Testo 174, Version 5.0.2564.18771, Lenzkirch, Germany) (Fig 1) during the experimental
109 period from September to March, 2017. Data logger was hanged closer to the plant canopy
110 (30cm above the ground) and covered from the top with flat carton to avoid direct sun and
111 moisture The vapor pressure deficit of the greenhouse was calculated based on the
112 temperature and relative humidity recorded using VPD-Auto grow software
113 (www.autogrow.com/wp-content/uploads/2016/03/VPD_HDCALC.xls). Data were measured
114 every hour for 25 days. Each point represents the average value of 25 days measurements.

115 Table 2. Greenhouse daily climatic variables recorded during the experiment period on
116 randomly selected days (average of 25 days)

Hour	Temperature (°C)	Relative humidity (%)	VPD (KPa)
13:00 pm	36.6	22.8	4.74
14:00 pm	35.7	23.2	4.49
15:00 pm	33.5	23.4	3.96
16:00 pm	31.8	24.7	3.54
17:00 pm	27.5	30.3	2.56
18:00 pm	24.3	38.1	1.88

19:00 pm	22.9	44.0	1.56
20:00 pm	21.9	47.5	1.38
21:00 pm	21.0	49.1	1.27
22:00 pm	20.1	51.4	1.14
23:00 pm	19.3	53.6	1.04
24:00 pm	18.6	56.7	0.93
1:00 am	18.0	59.2	0.84
2:00 am	17.3	61.8	0.75
3:00 am	16.6	62.7	0.70
4:00 am	16.1	64.6	0.65
5:00 am	15.6	66.3	0.60
6:00 am	15.9	66.0	0.61
7:00 am	21.7	52.6	1.23
8:00 am	27.4	40.3	2.18
9:00 am	31.3	32.5	3.08
10:00 am	33.8	27.7	3.80
11:00 am	35.0	26.5	4.13
12:00 am	36.5	24.2	4.65

117 Note: VPD = Vapor pressure difference and KPa = Kilo Pascal

118

119 From the result it was observed that extremely higher (36.6°C) and lower (15.6°C)
 120 temperature was recorded during middle of the day (12:00am-1:00pm) and before dawn (5:00
 121 am to 6:00am), respectively (Table 2). However, the recorded average daily temperature of
 122 24.9 °C is the optimal temperature for vegetative and tuber production for most of sweet
 123 potato genotypes (Ramirez, 1992a, b).

124 Regarding to relative humidity, greenhouse daily maximum relative humidity (66.3%) was
 125 recorded at 5:00 am which was coincided with greenhouse minimum temperature (15.6%)
 126 and minimum vapor pressure difference (0.60KPa). Likewise, greenhouse daily minimum
 127 relative humidity (22.8%) was recorded at 1:00 pm which coincided with maximum daily
 128 temperature (36.6%) and maximum daily vapor pressure deficit (4.74KPa).

129

130 **Soil Sampling, Preparation and Analysis**

131

132 Composite soil sample, made from twelve sub-samples, was collected from Hawassa
 133 University research field in a diagonal pattern from 0-20 cm soil depth. The samples were air-
 134 dried, ground to pass through a 2 mm sieve, except for analysis of organic carbon, where the
 135 samples were passed through 0.5 mm sieve. Working samples were obtained from each
 136 submitted samples and analyzed for selected Physico-chemical properties such as texture, soil
 137 pH, and organic carbon, using standard laboratory procedures at Hawassa University, College
 138 of Agriculture, Plant and Soil Analysis Laboratory. Organic carbon content of the soil was
 139 determined by reduction of potassium dichromate and oxidation reduction titration with
 140 ferrous ammonium (Walkley and Black, 1934). Soil particle size distribution was determined
 141 by hydrometer method (differential settling within a water column) using particles less than 2
 142 mm diameter. The pH of the soil was measured in 1:2.5 (weight/volume) soil samples to
 143 CaCl₂ solution ratio using a glass electrode attached to digital pH meter. Organic matter and
 144 total nitrogen was obtained by derivation from soil organic carbon content. Moreover, in

145 order to determine the bulk density of the soil, actual moisture content, and moisture content
146 at field capacity, twelve soil samples were taken from experimental soil by using soil core
147 sampler and determined using gravimetric method at Melka Werer Agricultural Research
148 Center.

149

150 Table 3. Selected physical and chemical properties of the experimental soil collected from the
151 study area.

Physical and chemical properties	Values
pH	7.6
Soil texture	Loam
Bulk density (g/cm ³)	1.018
Organic matter (OM %)	5.4
Organic carbon (OC %)	3.1
Total nitrogen (%)	0.11
Moisture content at FC (v/v %)	35.5
Soil moisture content (v/v %)	29.7

152

153 The results of the physical and chemical properties of the soil of the study site were presented
154 in Table 3. The analysis indicated that soil texture, level of organic carbon, total nitrogen
155 and soil moisture were found to the recommended growing media quality (Jones Jr, 2002;
156 Tadesse et al., 1991) and the actual soil moisture content of the soil and moisture content at
157 field capacity were 29.7% and 35.5%, respectively (Table 3).

158 **Plant growth parameters**

159 During the experimental periods (60 days after the start of the treatments) nondestructive
160 sampling for vine length, number of leaves, branch number (>2 cm), internode length, were
161 recorded from two plants in each treatment. At 60 days after the start of the treatments
162 destructive sampling were carried out to measure total leaf area, specific leaf area (SLA), and
163 Leaf Area Ratio (LAR) per plant. A LI-3100 leaf area meter (LI-COR, Inc., Lincoln,
164 Nebraska, USA) was used to measure total leaf area. Moreover, leaf dry weight was
165 determined after drying the leaves at 70°C for 48 hours and specific leaf area was calculated
166 (SLA= leaf area/leaf dry mass (cm²g⁻¹). At the age of 60 days after the start of the treatment,
167 the leafiness of the plant was determined by calculating the leaf area ratio (LAR) which is
168 expressed in cm²g⁻¹ of plant dry weight.

169

170 **Stomata density**

171 Two Sweet potato plants with intact root from each treatment were used for the measurement
172 of stomata density at 60 days after the start of the treatment. Epidermal impressions were
173 made on fresh intact lower leaves of the two genotypes following the procedure of (Torre *et*
174 *al.*, 2003). Stomata number was counted using Automated Upright Leica Microscope
175 DM5000 B, fixed with digital Leica DFC425/DFC425C image processing camera.

176

177 **Photosynthesis and Gas exchange parameters**

178 Photosynthesis(A), Transpiration rate (E) and Stomata conductance (g_s) were measured
179 during the vegetative stage at 60 days after the start of the treatment on fully developed
180 intact leaves at the 5th node using an open system LCA-4 ADC portable infrared gas analyzer
181 (Analytical Development Company, Hoddeson, England). These measurements were done
182 between 12:00 and 15:00 h with the following specifications/adjustments: Leaf surface area
183 was 6.25 cm², ambient carbon dioxide concentration 340 $\mu\text{mol mol}^{-1}$, temperature of the leaf
184 chamber varied from 34 to 47°C, leaf chamber molar gas flow rate was 410 $\mu\text{mol s}^{-1}$, ambient
185 pressure 828 mbar and photosynthetic active radiation (PAR) at the leaf surface was
186 maximum up to 1500 $\mu\text{mol m}^{-2} \text{s}^{-1}$. Data was collected every five min for 15 min using three
187 leaves in each of 3 plants per treatment.

188 **Instantaneous water use efficiency (IWUE)**

189 The ratio of carbon gain in photosynthesis and loss of water in transpiration was calculated
190 based on the data generated by open system LCA-4 (LCA-4 Software Version 1.04) ADC
191 portable infrared gas analyzer used at the growth stage of 60 days after the start of the
192 treatments. The ratio of leaf photosynthesis (A) to leaf transpiration rate (E) indicates the
193 efficiency of the genotype to produce dry matter per water loss through the leaves.

194 **Leaf relative water content (LRWC)**

195 Leaf relative water content was measured using the method of (Kamara et al., 2003). Leaf
196 discs (10 mm in diameter) were taken from young fully expanded leaves at 60 days after the
197 start of the treatment in the field sealed in tubes. The tubes containing leaf samples were
198 immediately placed on ice box which was not frozen, and immediately brought to the
199 laboratory. Leaf discs that were cut from the leaves were directly weighed to determine fresh
200 weight (FW). Samples were then floated in 100ml of distilled water in a closed Petri dish
201 under low light (50 $\mu\text{mol m}^{-2} \text{s}^{-1}$) for 24 hours. Leaf samples were taken out of water and
202 were surface dried with tissue paper, and their turgid weights (TW) were recorded. The leaf
203 relative water content takes into account the turgid mass of leaves, and so it is the proportion
204 of the leaf water content related to the maximum water content that can potentially be
205 achieved by the leaf. The samples were packed in paper bags, and oven dried at 65 °C for 48
206 hours for dry weight (DW) determination. The leaf discs were weighed using an analytical
207 balance with precision of 0.00001 g. Then calculation of leaf relative water content was
208 computed as following the methodology of (Turner, 1981):

209
$$\text{LRWC (\%)} = \frac{\text{FW}-\text{DW}}{\text{TW}-\text{DW}} \times 100$$

210 **Dry matter accumulation and tuber yield**

211

212 At harvesting time; leaf, vine plus petiole, root and tuber components were taken from three
213 plants and weighed separately. The tubers were washed to remove soil and allowed surface
214 air dried for approximately 30 minutes, and weighed to obtained fresh weight. Each plant part
215 was allowed to dry for 48 hours in an oven at 70°C.

216

217 **Harvest index (HI)**

218

219 At harvest , 152 and 168 days after planting for genotype Kulfo and Awassa-83 respectively,
220 a pan area of within each treatment of sweet potato genotypes was harvested (0.1125 m²), and
221 whole plant part with in the pan was oven dried up to a constant weight, weighed and then
222 converted into biological yield (biomass) (g/m²). The harvested bottom part (tuber) is
223 considered as economic yield (tuber yield in g / m²). Harvest index was calculated according
224 to the following the methodology of (Ludlow and Muchow, 1990):
225 Harvest index (%) = (tuber yield / Biological yield) × 100.

226 **Statistical Analysis**

227

228 Analysis of variance (ANOVA) was carried out using SAS statistical software version 9.00
229 (SAS Institute, 2002). Mean separation was done by using Tukey's procedure (P ≤ 0.05).
230 When there was a statistically significant interaction between the factors, the interaction was
231 considered, rather than the main effects, otherwise, only the main effects of treatments was
232 presented. Pearson's simple correlation coefficient was used to analyze correlation between
233 selected parameters.

234

235 **RESULT AND DISCUSSION**

236 **Morphological Characters**

237 **Vine length, leaf number, branch number and internode length**

238 The result indicated that leaf number, branch number and vine length were significantly
239 (P≤0.05) responded to genotype and irrigation frequencies. Result in table 4 indicated that,
240 vegetative growth was more enhanced in Kulfo than Awassa-83. Although genotype has been
241 contributing to the differences in growth performance of the plant, prolonged irrigation
242 interval (more than a day) showed stronger effect on vegetative growth. The longer the
243 irrigation interval (lower irrigation frequency), the more the reduction was observed in
244 vegetative growth in both genotypes.

245 Analysis of variance revealed that as compared to daily irrigation an extended watering
246 interval to seven days significantly reduced leaf number, vine length, branch number and
247 internode length by 34%, 20%, 27% and 19%, respectively (Table 4).

248 The result is in agreement with the finding of (Sokoto and Gaya, 2016) who reported
249 significantly less number of leaves under lower irrigation frequency on sweet potato. On the
250 other hand, vine length reduction under lower irrigation frequency has also been reported in
251 many other crop species. Previous research (Katsoulas et al., 2006) found that the main
252 length of harvested shoot of rose during the period of measurements irrigated with high
253 frequency produced slightly longer stems than those irrigated with low frequency. Moreover,
254 (Laurie and Magoro, 2008) also reported that reduction in vine length of sweet potato has
255 been positively correlated to the decline in irrigation rates from 100% full irrigation to 30%
256 irrigation. Similar to the present study, (Ebel et al., 1995) found that an extended irrigation
257 interval led to decrease in percentage of vine length in sweet potato. Branch number was also
258 found to be significantly reduced when extended irrigation interval was considered. Report

259 from (Nair and Nair, 1995; Prabawardani et al., 2007) noted that number of branches per
 260 plant were significantly influenced under water stress condition.

261 Table 4. Main effects of genotype and irrigation frequency on leaf number (LN), vine-length
 262 (VL), branch number (BR) and internode length (INL)

Treatments	Leaf Number	Vine Length(cm)	Branch Number	Internode length (cm)
Genotype				
Awassa-83	85.61 ^b	73.42 ^b	5.92 ^{b*}	1.66
Kulfo	203.83 ^a	106.89 ^a	7.98 ^a	1.53
Tukey's HSD _(0.05)	33.231	7.167	0.3681	Ns
Irrigation frequency(I)				
Daily watering	164.25 ^a	101.21 ^a	8.13 ^a	1.80 ^a
Four days interval	161.08 ^a	87.88 ^b	6.75 ^b	1.53 ^b
Seven days interval	108.83 ^b	81.38 ^b	5.96 ^c	1.45 ^b
Tukey's HSD _(0.05)	49.833	10.748	0.5521	0.2186
F test values				
Genotype (G)	60.08 ^{***}	103.55 ^{***}	148.00 ^{***}	3.78 ^{ns}
Irrigation frequency (I)	5.55 [*]	12.60 ^{**}	56.14 ^{***}	10.18 ^{**}
G x I	3.81 ^{ns}	1.06 ^{ns}	3.76 ^{ns}	0.84 ^{ns}
SEM ±	26.42	5.70	0.45	0.12
CV (%)	22.36	7.74	5.16	8.9

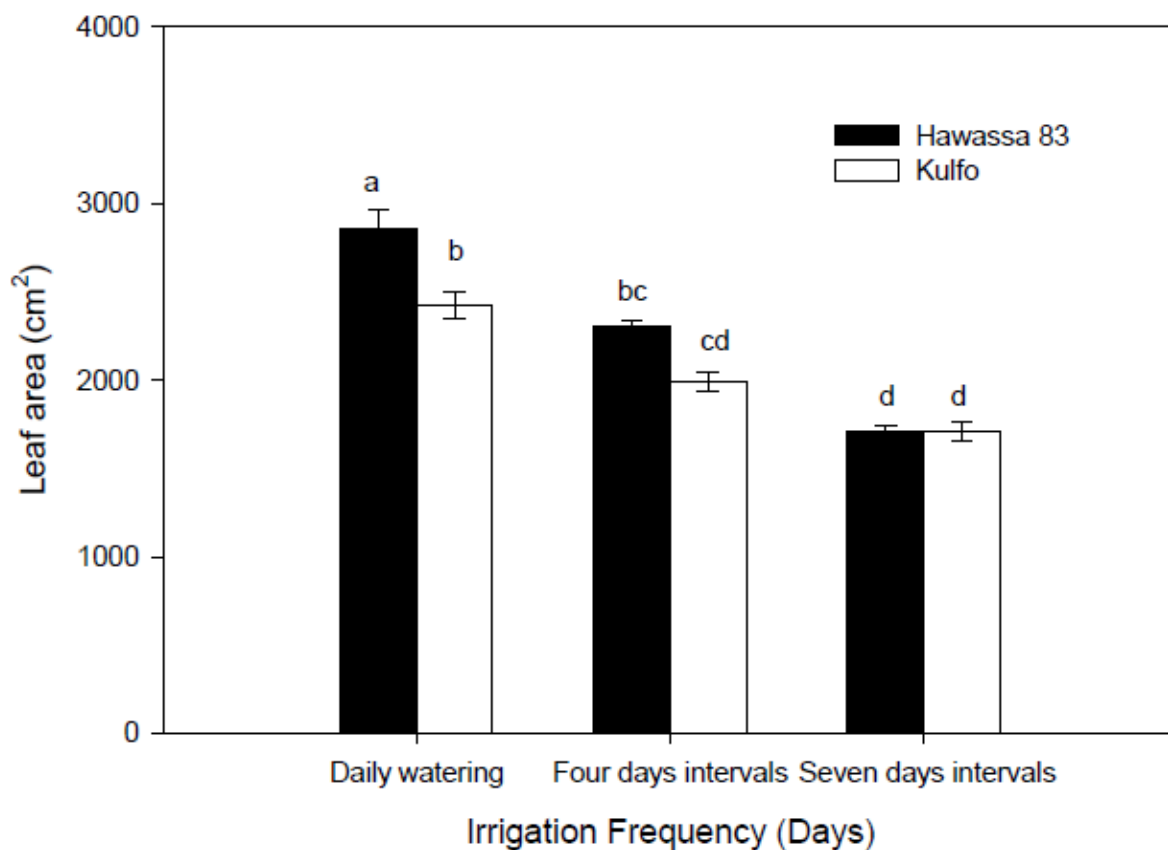
263 *Means in the same column followed by the same letter are not significantly different at the 5% probability
 264 level.

265 The former author reported that when water was withheld, the inter node length of sweet
 266 potato cultivars found to be significantly declined. This is mainly due to decrease in turgor
 267 pressure within cells during cell growth and development forcing the inhibition of cell
 268 expansion which could in turn reflected in decrease in internode length, leaf number and vine
 269 length. This probably could be one of the adaptation strategies in plants against moisture
 270 stress to minimize potential water loss from the surface of the plant.

271 **Total leaf area**

272 Total leaf area production per plant was significantly affected by the interaction effect of
 273 genotype and irrigation frequency. Result on figure 2 showed that, total leaf area was
 274 significantly ($P \leq 0.05$) influenced by interaction between genotype and irrigation frequency.
 275 Maximum leaf area was obtained when Awassa-83 was irrigated daily than Kulfo. As
 276 compared to daily irrigation, extending irrigation interval to four days and seven day
 277 significantly reduced leaf area 20% (Hawassa 83) and 18% (Kulfo) and 36 % (Hawassa 83)
 278 and 28% (Kulfo), respectively (Fig 2). This indicated that Awassa-83 was more sensitive to
 279 moisture stress than Kulfo genotype and adaptation to moisture stress was largely observed in
 280 kulfo than Awassa-83 genotype.

281



282
 283 *Figure 1. Illustration of genotypes response to irrigation frequency on leaf area (cm²). Error*
 284 *bars represent standard errors of means with three replications. Means with same letter (s)*
 285 *are not significantly different at p ≤ 0.05.*
 286

287 As (Meyer and Boyer, 1972) stated that the occurrence of water deficits in young growing
 288 plants would also be expected to cause a reduction in cell turgor which would slow leaf
 289 expansion and growth. These observations are supported by previous findings reporting
 290 reduction in leaf area under decreasing soil water regimes to 40 % and 20 % of the field
 291 capacity significantly reduce leaf production compared to growth under well-watered
 292 conditions (Saraswati, 2007). Our results show that, specific leaf area and leaf area may
 293 have a higher plasticity in response to a large range of water status, and these parameters are
 294 clearly associated with photosynthesis and water use efficiency.

295 **Physiological characteristics**
 296 **Leaf anatomy and Stomata density**

297 Result indicated that irrigation frequency did not show significant differences (P > 0.05) on
 298 stomata density per mm². However, stomata density was significantly influenced due to
 299 genotype effect. From the analysis it was observed that genotype Awassa-83 had
 300 approximately 2.0 more stomata number per mm² than genotype Kulfo (Table 5). Although
 301 irrigation did not have significant effect on density of stomata, result on table 5 indicated that
 302 delaying irrigation by seven days reduced density of stomata per mm² than plant irrigated
 303 daily or every four days interval.

304

305 Table 5. Effect of genotype and irrigation frequency on stomata density (SD), specific leaf
 306 area (SLA) and Leaf area ratio (LAR) of sweet potato grown under greenhouse

Treatments	SD(mm ²)	SLA(cm ² g ⁻¹)	LAR (cm ² g ⁻¹)
Genotype			
Awassa -83	16.06 ^a	245.87 ^{a*}	46.22 ^a
Kulfo	14.54 ^b	222.71 ^b	39.37 ^b
Tukey's HSD _(0.05)	0.7744	12.244	5.0898
Irrigation Frequency			
Daily watering (control)	15.42	246.60 ^a	44.50
Four days interval	15.78	225.22 ^b	44.79
Seven days interval	14.70	231.05 ^{ab}	39.09
Tukey's HSD _(0.05)	Ns	18.361	Ns
F-test values			
Genotype (G)	18.29 ^{***}	16.98 ^{**}	8.58 [*]
Irrigation frequency (I)	3.21 ^{ns}	5.16 [*]	2.52 ^{ns}
G x I	0.05 ^{ns}	3.44 ^{ns}	1.67 ^{ns}
SEM ±	0.62	9.73	4.05
CV (%)	4.93	5.09	11.58

307 *Means in the same column followed by the same letter are not significantly different at the 5%
 308 probability level.

309 The result verified with findings of (Saraswati, 2007) who noted that stomatal density of
 310 sweet potato cultivars was unaffected by soil water stress conditions. However, in this study
 311 there was significant variation between genotypes considered. This might be related to the
 312 variability in genetic make-up of the genotypes. Previous report indicated that, an increase in
 313 stomata density under water deficit, indicated that an adaptation to moisture stress vary from
 314 genotype to genotype (Martínez et al., 2007; Xu and Zhou, 2008).

315
 316 **Specific leaf area and leaf area ratio**

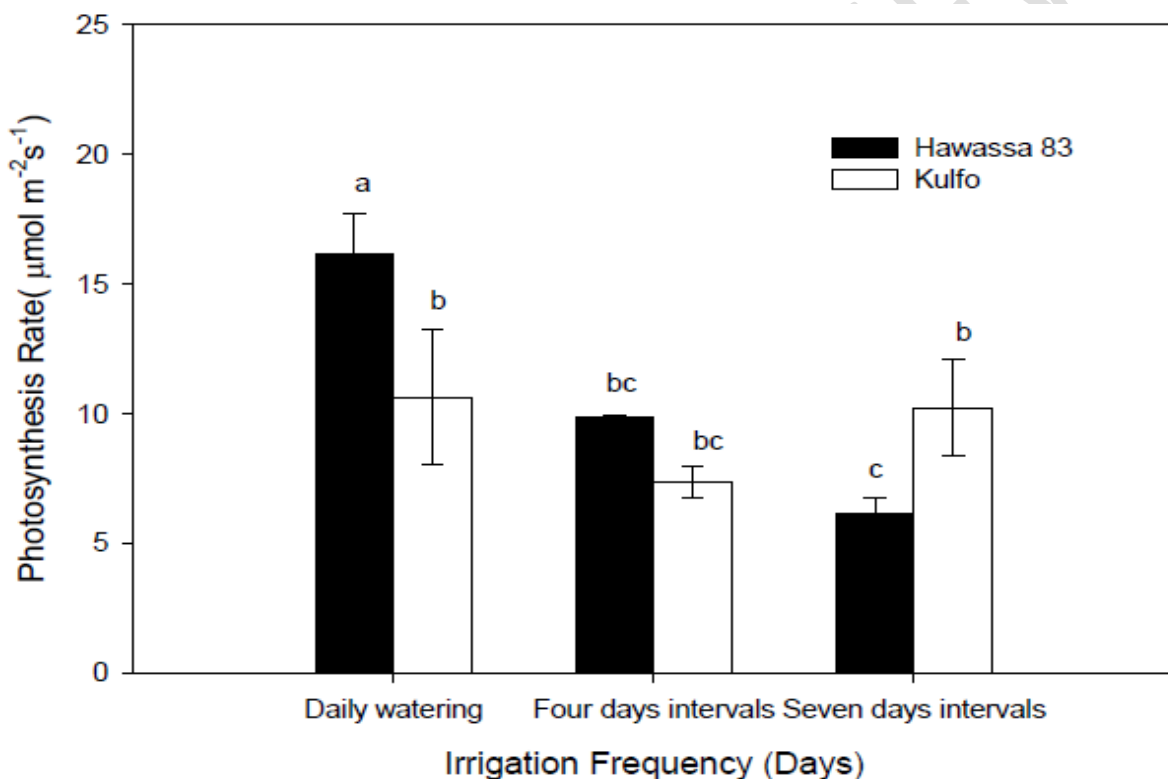
317 The main effects of genotype and irrigation frequency was significant ($P \leq 0.05$) on specific
 318 leaf area (SLA). Irrigation at four days interval significantly ($P \leq 0.05$) reduced SLA by 9%
 319 compared to the daily irrigation in both genotypes. Regarding genotypes, Kulfo had
 320 significantly ($P \leq 0.01$) superior performance over genotype Hawassa 83, implying that
 321 genotype Kulfo possibly had thicker leaves than genotype Hawassa 83 (Table 5). Unlike,
 322 LAR was not significantly ($P > 0.05$) affected by different irrigation frequencies. The highest
 323 and least LAR was observed from daily irrigation and followed by seven days interval
 324 respectively, although it was statistically at par with (Table 5). However, different genotypes
 325 showed significant ($P \leq 0.05$) difference in LAR. Genotype Awassa-83 had better
 326 performance in leaf area ratio than Kulfo, this implies that genotype Awassa-83 was leafy
 327 (Table 5).

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332

333 **Photosynthesis (A)**

334 The highest assimilation rate was produced from the interaction between genotype Awassa-
335 83 and daily irrigation followed by Kulfo by seven days interval, while the least assimilation
336 rate was observed from Awassa-83 by seven days interval. There was significant difference
337 in the rate of assimilate due to the interaction between genotype and irrigation intervals. The
338 highest amount of assimilation rate ($16.16 \mu\text{mol m}^{-2}\text{s}^{-1}$) was produced from genotype
339 Awassa-83 treated with daily irrigation (Figure 4). However, there was no significance
340 difference in the rate of assimilation between Hawassa genotype irrigated every day and four
341 day interval. In this study it was observed that water extended water holding for seven days
342 significantly reduced assimilation rate by 62% compared to Awassa-83 treated with daily
343 irrigation. Genotype Kulfo had produced statistically similar assimilation rate over the entire
344 irrigation frequency considered in this trial (Figure 4).



345

346 *Figure 2. The interaction effects of genotype and irrigation frequency of sweet potato on*
347 *photosynthesis ($\mu\text{mol m}^{-2}\text{s}^{-1}$). Means with same letter (s) are not significantly different at $p \leq 0.05$.*

348 Result indicated that genotype Awassa-83 has shown strong reduction in assimilation rate as
349 irrigation interval prolonged. Quite in opposite, genotype Kulfo had stable performance
350 across irrigation frequencies. This might imply genotype Awassa-83 was more sensitive to
351 moisture stress than genotype Kulfo. In line with this study, (Shao et al., 2008) noted that, as
352 the soil water availability declines, leaf cells lose their turgor; this affects the leaf
353 photosynthesis due to stomatal closure and physical disruption of the leaf cells. Moreover,
354 report indicated that, higher irrigation frequency increased g_s and with high g_s values favored
355 CO_2 assimilation and plants showed higher daily carbon gain on tomato (Pires et al., 2011).

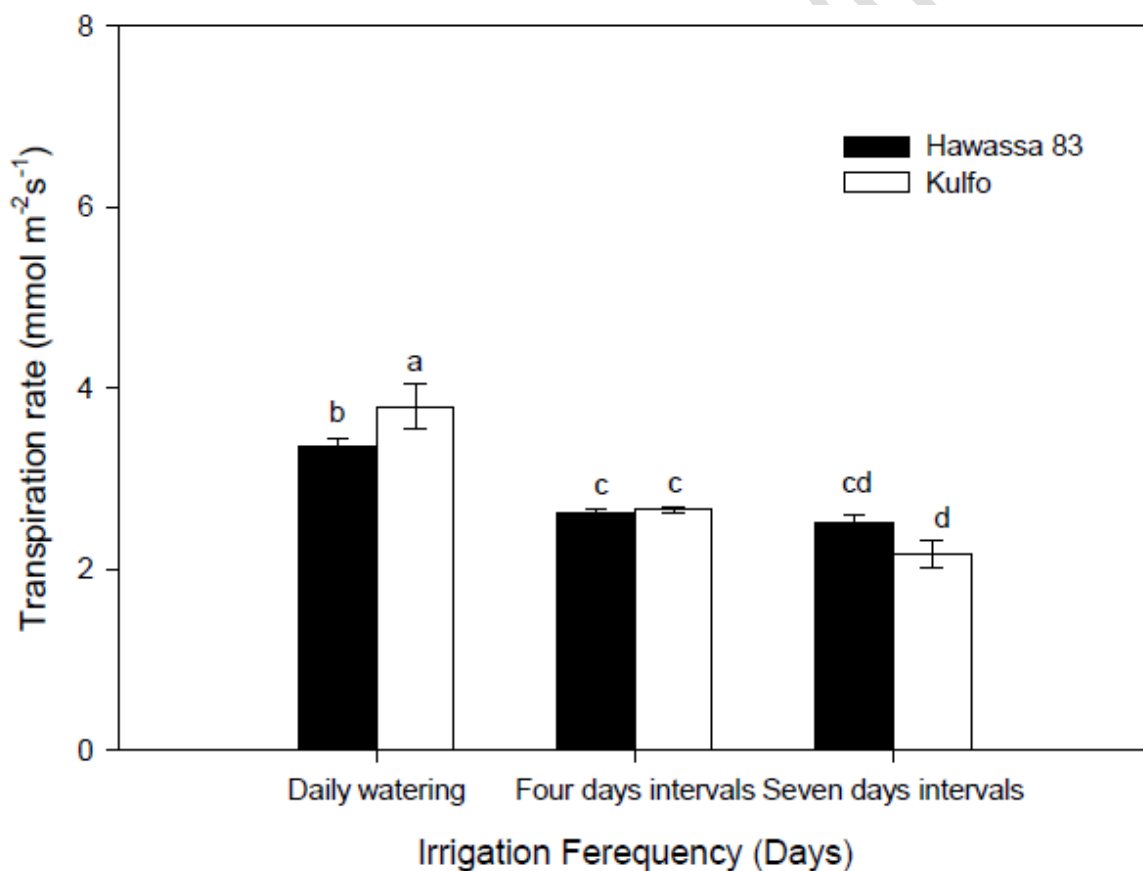
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359 **Transpiration rate (E)**

360 The results of interaction effect between genotype and irrigation frequency showed
361 significant ($P \leq 0.05$) effect on transpiration rate, though it was not large enough to be
362 extremely different from main effects. Nonetheless, the highest rate of transpiration (3.81
363 $\text{mmol m}^{-2}\text{s}^{-1}$) was recorded when genotype Kulfo was irrigated daily whereas the least (2.17
364 $\text{mmol m}^{-2}\text{s}^{-1}$) was observed from genotype Kulfo and seven days irrigation interval (Figure
365 5). In contrast, withholding irrigation for four days or seven days significantly reduced
366 transpiration rate in both genotypes (Awassa-83 and Kulfo) as compared to treating both
367 genotype with daily irrigation. Consequently, four and seven days delay in irrigation
368 significantly reduced transpiration rate in genotype Awassa-83 by 22% and 25%,
369 respectively. And stronger decline in transpiration rate were observed when genotype Kulfo
370 was irrigated with four and seven days irrigation intervals with 30% and 43% reductions,
371 respectively (Figure 5).



372

373 *Figure 3. Illustration of response of sweet potato genotypes as influenced by irrigation efficiencies on*
374 *transpiration rate ($\text{mmol}^{-2}\text{s}^{-1}$). Means with same letter (s) are not significantly different at $p \leq 0.05$.*

375 Overall, genotype Kulfo combined with daily irrigation gave significantly higher
376 transpiration rate. The reduction in the rate of transpiration with decrease in the rate of
377 irrigation might be associated with lower number of stomata density in genotype Kulfo,
378 which finally attributed to have relatively lower transpiration rate under extended watering
379 interval. Parallel with the result report from (Garnier et al., 2001) indicated that tolerance in
380 drought in different plant species associated with lower number of stomata and reduction in

381 the rate of water lost which attributed to its capability to maintain cellular integrity by
 382 conserving water under drought conditions. (Saraswati, 2007) also reported that water
 383 stressed plants transpired less water compared to the well-watered plants in sweet potato
 384 cultivars. In addition, one of the adaptive features of plants growing in drought condition is
 385 reduction in the size of stomata opening and leaf size to reduce loss of moisture through
 386 transpiration.

387

388 **Stomatal conductance (g_s)**

389 The main effects of irrigation frequency showed significant ($P \leq 0.001$) effect on stomatal
 390 conductance (g_s). The highest stomatal conductance was obtained in response to daily
 391 watering followed by four days interval while the least was observed from seven days
 392 interval. As compared to the effect of daily irrigation, genotypes treated to four and seven
 393 days water holding significantly reduced Stomatal conductance by 36% and 44% ,
 394 respectively (Table 7). Unlike, there was no significant ($P > 0.05$) difference between
 395 genotype in relation to stomatal conductance.

396 Table 7. Main effects of genotype and irrigation frequency on stomatal conductance (g_s) in
 397 mmol m⁻²s⁻¹

Treatments	g_s (mmolm ⁻² s ⁻¹)
Genotype	
Awassa -83	110.0 ^{a*}
Kulfo	100.0 ^a
Tukey's HSD _(0.05)	Ns
Irrigation Frequency	
Daily watering (control)	143.3 ^a
Four days interval	91.7 ^b
Seven days interval	80.0 ^b
Tukey's HSD _(0.05)	0.0254
F-test values	
Genotype (G)	1.65 ^{ns}
Irrigation frequency (I)	25.04 ^{***}
G x I	2.39 ^{ns}
SEM ±	0.01
CV (%)	15.71

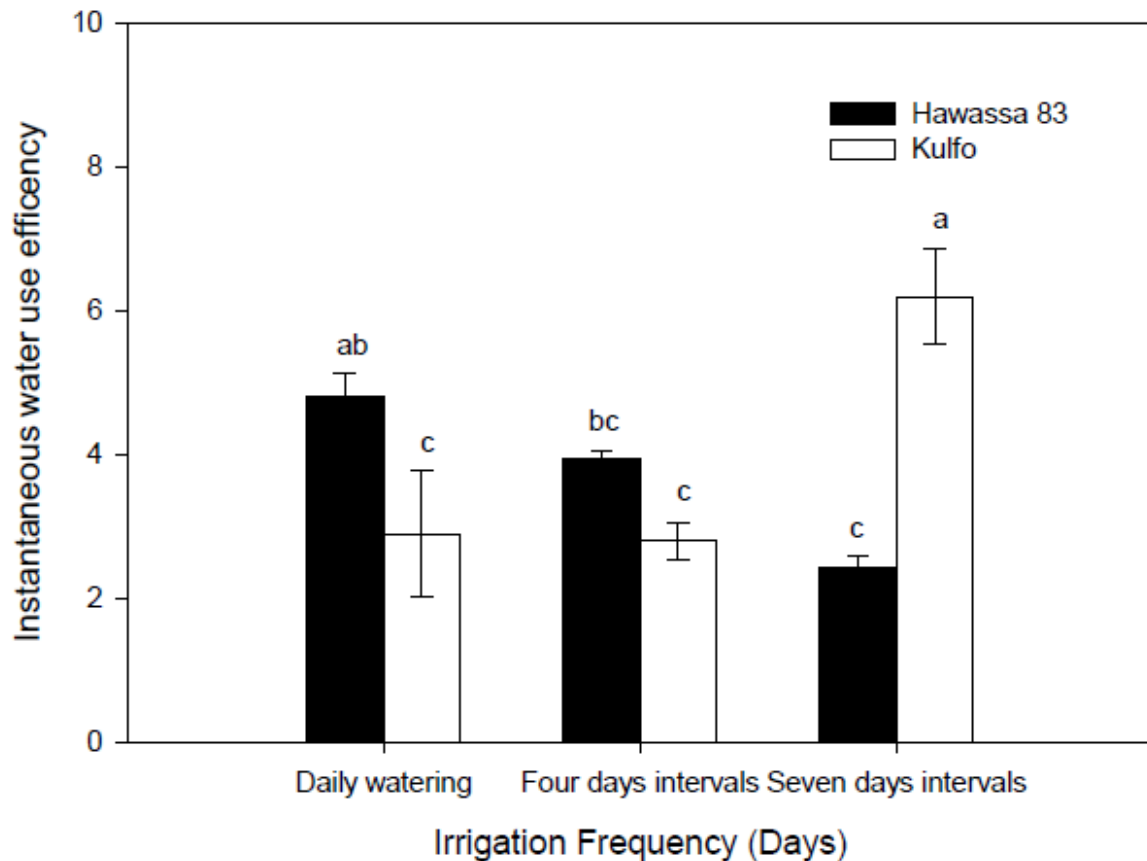
398 **Means in the same column followed by the same letter are not significantly different at the 5%*
 399 *probability level.*

400 The difference in stomata conductance might be associated to dry out of soil and the leaf
 401 water potential that play significant role in influencing the stomatal conductance (Liang et
 402 al., 2002).Previous report indicated that a severe decline in stomatal conductance values for
 403 sweet potato plants subjected to drought stress. (Yooyongwech et al., 2014) also noted that
 404 stomatal conductance (g_s) in sweet potato genotypes declined significantly when plants were
 405 subjected to mild and extreme water deficit stress.

406

407 **Instantaneous water use efficiency (IWUE)**

408 The analysis of variance revealed that there was statistically significant ($P \leq 0.001$)
 409 differences in IWUE due to the interaction effect between genotype and irrigation interval.
 410 The interaction of genotype Kulfo and seven days interval resulted with the highest
 411 instantaneous water use efficiency as compared to genotype Awassa-83 (Figure 6). In
 412 response to genotype by irrigation frequency, extended watering interval for seven days with
 413 genotype Awassa-83 had significant reduction (i.e., 49%) compared to the combination of
 414 genotype Awassa-83 and daily irrigation (Figure 6). On the other hand, higher irrigation
 415 frequency of daily and four days watering intervals resulted in significant reduction on
 416 genotype Kulfo in instantaneous water use efficiency with 55% and 53%, respectively over
 417 genotype Kulfo which was irrigated with seven days irrigation interval (Figure 6).



418

419 *Figure 4. The interaction effect of genotype and irrigation frequency on instantaneous water use*
 420 *efficiency ($\mu\text{mol mmol}^{-1}$). Means with same letter (s) are not significantly different at $p \leq 0.05$.*

421 Result indicated that, under seven days irrigation interval, genotype Kulfo was able to
 422 conserve and utilize water efficiently than Hawassa 83. This was attributed to low
 423 transpiration rate as a result of small leaf surface area, and few stomata density. Quite in
 424 opposite, genotype Awassa-83 responded differently to irrigation frequency suggesting that
 425 sweet potato genotypes had different response to irrigation interval. Nevertheless, in this
 426 study the highest IWUE was observed from the interaction between Kulfo and seven days
 427 interval. This result supported with the finding of (Kang and Wan, 2005) who noted that
 428 water use efficiencies of radish was significantly increased by decreasing irrigation level.
 429 Moreover, (Pires et al., 2011) reported that the highest IWUE values were noticed in plants
 430 subjected to high irrigation frequency than to low irrigation frequency on tomato.

431

432

433 **Leaf relative water content**

434 Sweet potato significantly ($P \leq 0.001$) responded to different irrigation frequency on leaf
 435 relative water content. The higher leaf relative water content was obtained from daily
 436 irrigation whereas the lowest was observed from seven days interval. Unlike, seven days
 437 interval had significant deviation on leaf relative water content from daily irrigation. In
 438 quantitative term seven days interval recorded 15% reduction compared to the daily irrigation
 439 (Table 8). Regarding on genotype difference, there was no significant variation in leaf
 440 relative water content between Awassa-83 and Kulfo (Table 8).

441

442 Table 8. The main effects of genotype and irrigation frequency on leaf relative water
 443 content(LRWC)

Treatments	LRWC (%)
Genotype	
Awassa -83	61.73 ^{a*}
Kulfo	64.20 ^a
Tukey's HSD_(0.05)	Ns
Irrigation Frequency	
Daily watering (control)	67.67 ^a
Four days interval	63.55 ^a
Seven days interval	57.68 ^b
Tukey's HSD_(0.05)	4.1771
F-test values	
Genotype (G)	4.9 ^{ns}
Irrigation frequency (I)	27.17 ^{***}
G x I	0.03 ^{ns}
SEM ±	1.57
CV (%)	3.06

444 *Means in the same column followed by the same letter are not significantly different at the 5%
 445 probability level.

446 Leaf relative water content was substantially diminished when sweet potato genotypes were
 447 subjected to prolonged irrigation frequency (seven days interval). Under extended irrigation
 448 interval tissues and cells were not well hydrated enough (lower LRWC %) which might have
 449 an impact on normal physiological activities. In line with this, (Saraswati, 2007) indicated
 450 that water stress caused significance decrease in the relative water content in sweet potato.
 451 Under lower soil field capacity, the leaf relative water content declined compared to that of
 452 the same cultivars grown at higher soil field capacity.

453 **Yield and yield components**

454 **Dry Mass Production, Biomass Yield, Tuber Yield and Harvest Index**

455 The interaction effect of genotype and irrigation frequency showed non-significant ($P > 0.05$)
 456 effect on leaf dry mass and root dry mass production. Quite in reverse, storage root dry mass
 457 was significantly influenced by the interaction effect of genotype and irrigation frequency.

458 The main effect of irrigation frequency and genotype were found to be significant on root dry
 459 mass and leaf dry mass of sweet potato except for main effect of genotype on leaf dry mass.
 460 Significantly ($P \leq 0.01$) maximum leaf dry mass accumulation was observed from daily
 461 irrigation. Comparatively, irrigating in seven days interval had recorded significantly
 462 reduced performance by 36% over the daily irrigation in leaf dry mass (Table 9). In contrast,
 463 concerning the genotype difference for leaf dry mass accumulation, there was no significant
 464 variation between genotype Awassa-83 and genotype Kulfo (Table 9).

465 Root dry mass was also significantly ($P \leq 0.01$) affected by irrigation frequency (Table 9). It
 466 was observed that irrigating the genotype in every seven days interval gave the highest root
 467 dry mass and found to be significantly different from daily irrigation with 59% (Table 10).
 468 On the other hand, irrigation frequency treatment, which was irrigated once in every four
 469 days interval, was statistically at par with daily irrigation for root dry mass accumulation.
 470 Regarding on root dry weight accumulation genotype Awassa-83 had accumulated
 471 significantly ($P \leq 0.001$) higher (90 %) dry mass than sweet potato genotype Kulfo (Table 9).

472 Table 9. Main effects of root dry weight (RDM) and leaf dry weight (LDM) of sweet potato
 473 as influenced by genotype and irrigation frequency

Treatments	RDM(g)	LDM(g)
Genotype		
Awassa-83	4.83 ^{a*}	7.26 ^a
Kulfo	2.54 ^b	6.13 ^a
Tukey's HSD _(0.05)	0.6063	Ns
Irrigation frequency		
Daily watering (control)	2.83 ^b	8.20 ^a
Four days interval	3.73 ^{ab}	6.60 ^{ab}
Seven days interval	4.50 ^a	5.29 ^b
Tukey's HSD _(0.05)	0.9092	1.7732
F test values		
Genotype (G)	67.98 ^{***}	4.30 ^{ns}
Irrigation frequency (I)	11.98 ^{**}	9.58 ^{**}
G x I	2.72 ^{ns}	0.41 ^{ns}
SEM ±	0.48	0.94
CV (%)	16.01	17.19

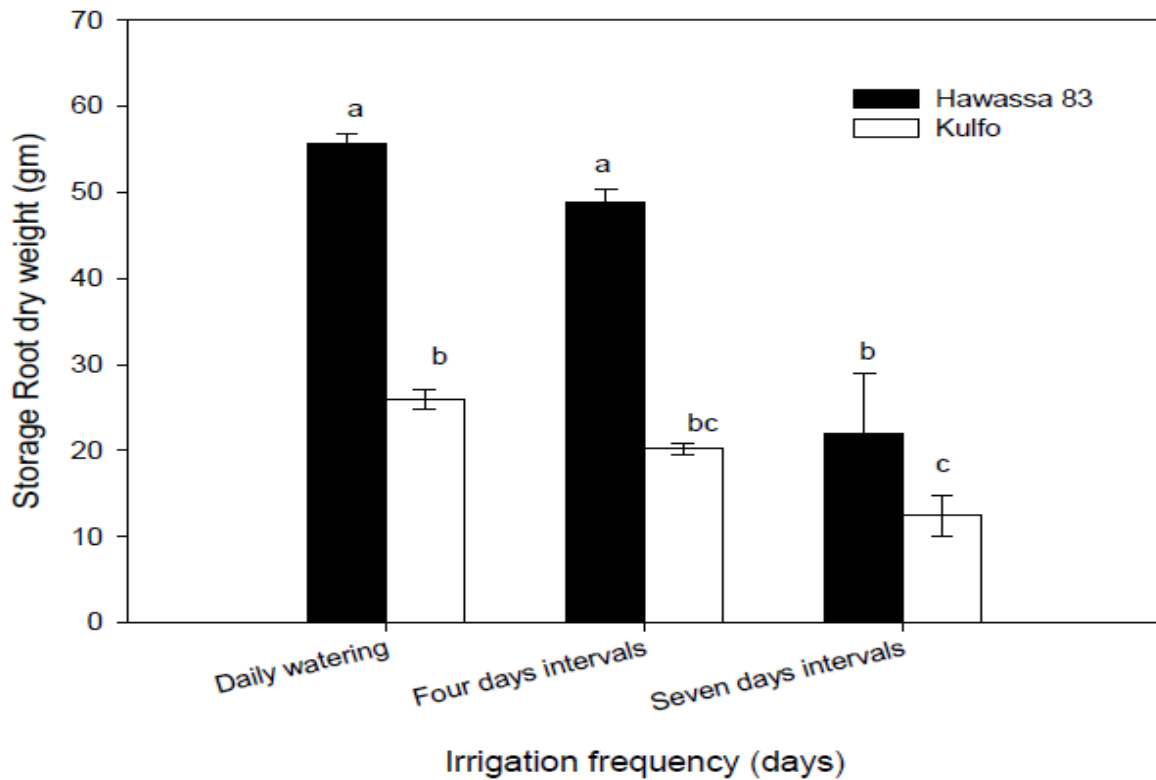
474 *Means in the same column followed by the same letter are not significantly different at the 5%
 475 probability level.

476
 477 Report from (Saraswati, 2007) indicated that water stress significantly reduced dry leaf
 478 masses of different sweet potato cultivars. However, in this study we investigated that
 479 reduction in irrigation frequency increased root dry mass than frequently irrigated genotypes.

480 Storage root dry mass

481 ANOVA analysis indicated that the interaction between genotype and irrigation frequency
 482 showed significant ($P \leq 0.05$) influence on storage root dry mass. Higher storage root dry
 483 mass accumulation was found from Awassa-83 and daily irrigation than kulfo under similar
 484 growth condition. It was observed that reduction in irrigation frequency significantly reduced

485 storage root dry mass by 61% (Figure 8) and the effect was stronger genotype Kulfo than
486 Awassa-83. Genotype Kulfo, has shown similar performances in all irrigation frequencies.



487

488 *Figure 5. The interaction effect of genotype and irrigation frequency of sweet potato on*
489 *storage root dry mass. Error bars represent standard errors of means with three replications.*
490 *Means with same letter (s) are not significantly different at $p \leq 0.05$.*

491 Similar observation also reported by (Masango, 2014) where storage root dry mass with
492 lower irrigation frequencies was lower compared to with higher irrigation frequencies. The
493 result is in agreement with the findings of (Tshisola, 2014) who indicated lower tuber dry
494 weight at the low irrigation frequency compared to the high irrigation frequency in Irish
495 potato.

496 **Biomass yield, tuber yield and harvest index**

497 Total dry biomass was significantly ($P \leq 0.001$) affected by the main effects of genotype and
498 irrigation frequency. From daily irrigation the highest total dry biomass was obtained from
499 genotype treated with daily irrigation and the least was from genotype treated with seven
500 days interval. Total dry biomass for daily irrigation found to be increased by 77% and 20%
501 compared with seven days interval and four days interval respectively (Table 10). Moreover,
502 the main effect of genotype was also significant on total dry biomass and hence, significantly
503 greater production of total dry biomass was obtained from genotype Awassa-83 (Table 10).

504 In this study, both genotypes produced maximum total dry biomass under daily irrigation.
505 With respect to genotype difference for total plant dry biomass genotype Awassa-83 had
506 produced significantly superior total dry biomass. As (Tshisola, 2014) indicated, in line
507 with this finding, reported higher biomass accumulation at the high irrigation frequency.

508

509

510 **Tuber yield**

511 In this study, tuber yield was significantly influenced by main effects of genotype and
512 irrigation frequency. Although remarkably higher tuber yield was recorded from genotype
513 irrigated daily and every four days intervals, the difference was not statistically significant at
514 ($P>0.05$). Genotype irrigated every seven days gave the lowest tuber yield and significantly
515 different from daily irrigation. Daily irrigation produced more than two fold tuber yield over
516 seven days interval (Table 10). Furthermore, genotype Awassa-83 produced significantly
517 more (26%) tuber yield over Kulfo (Table 10).

518 This finding was consistent with the finding of (Sokoto and Gaya, 2016) who reported that
519 high tuber yield at higher irrigation interval because the rate of tuber yield increased with
520 progressive increase in irrigation frequency, this perhaps due to improved root system which
521 enables the plant to utilize more moisture from the soil. This finding aligned correctly with
522 previous findings of several other investigations (Masango, 2014; Tshisola, 2014).

523

524 **Harvest index**

525 Anova analysis result indicated that, maximum harvest index was observed from daily
526 irrigation whereas minimum was recorded from seven days interval. In comparison to daily
527 irrigation, seven days interval deviates significantly from daily irrigation whereas four days
528 interval was found to be insignificant. As to the magnitude of reduction, seven days interval
529 irrigation frequency treatment was diminished by 27% compared to the daily irrigation (Table
530 10). In addition to this, there was also genotype difference for harvest index, genotype
531 Awassa-83 had significantly higher (40%) harvest index than genotype Kulfo (Table 10).

532 Under non-limiting condition (control), both genotypes found to have significantly higher
533 harvest index. Furthermore, in this study genotype Awassa-83 had higher harvest index than
534 genotype Kulfo. As (Bhagsari and Ashley, 1990) noted that frequently irrigated treatment
535 produced relatively higher HI values on sweet potato, demonstrating that more assimilates
536 were translocated efficiently to the main sink, compared to the other plant parts. The study of
537 (Masango, 2014) also agreed with the current result, sweet potato crop under higher irrigation
538 frequency had better harvest index, thus enabling photosynthesis efficiently translocate to the
539 main sink (storage root).

540

541 Table 10. Main effects of genotype and irrigation frequency on total dry biomass (TDBM),
542 tuber yield (TY) and harvest index (HI)

Treatments	TDBM (g m^{-2})	TY (g m^{-2})	HI (%)
Genotype			
Awassa -83	100.27 ^a	216.27 ^{a*}	56.86 ^a
Kulfo	66.78 ^b	172.01 ^b	40.64 ^b
Tukey's HSD _(0.05)	9.2172	31.758	6.4956
Irrigation frequency			

Daily watering (control)	104.47 ^a	261.79 ^a	53.59 ^a
Four days interval	87.21 ^b	228.24 ^a	53.51 ^a
Seven days interval	58.91 ^c	92.39 ^b	39.15 ^b
Tukey's HSD _(0.05)	13.822	47.623	9.7407
F-test values			
Genotype (G)	62.66 ^{***}	9.22 [*]	29.61 ^{***}
Irrigation frequency(I)	39.42 ^{***}	50.50 ^{***}	10.36 ^{**}
G x I	3.46 ^{ns}	1.12 ^{ns}	1.98 ^{ns}
SEM ±	7.33	25.25	5.16
CV (%)	10.74	15.93	12.97

543 *Means in the same column followed by the same letter are not significantly different at the 5%
544 probability level.

545

546

547 **Conclusion**

548 Morphological parameters of sweet potato genotypes were significantly influenced depending
549 on irrigation interval, genotypes and their interaction. Extension of irrigation interval to seven
550 days significantly reduced leaf area, leaf number, vine length, branch number and internode
551 length of the sweet potato genotypes. Growth reduction was stronger with Hawassa 83 and
552 when irrigation frequency with holds for longer period of time (seven days) than daily or
553 every four day irrigation intervals. Physiological parameters such as Stomata density,
554 specific leaf area and leaf area ratio were remained constant under different irrigation
555 intervals. Similarly, photosynthetic rate, transpiration rate and stomata conductance were
556 reduced as irrigation intervals extended to seven day intervals. Extension of an irrigation
557 interval to seven days strongly reduced instantaneous water use efficiency in Hawassa 83 but
558 increased in Kulfo genotype suggesting that kulfo had better water utilization efficiency than
559 Hawassa 83.

560 Finally, although yield and yield component did not respond to the interaction effect between
561 irrigation interval and genotype, extension of irrigation interval to seven days significantly
562 reduced tuber dry matter, total tuber yield and harvest index and the effect was stronger in
563 kulfo than Hawassa 83 genotype.

564

565

566

567

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