

Review Article

An Over View of Dermatophytosis in Camels

Abstract:

Dermatophytosis is a fungal infection of the skin caused by *dermatophytes* spp.-filamentous fungi which have ability to invade the epidermis and keratinized tissues such as hair, skin or nails. *Trichophyton verrucosum* is the most common dermatophytes species isolated from camel. The disease is characterized by circumscribed crusty hairless lesion, (1-2 cm) distributed over the head, neck, shoulder, limbs and flanks. Dermatophytosis can be diagnosed by direct examination, fungal culture, skin biopsy and molecular diagnosis methods. This overview forecast more light of the different aspects of this disease.

Key words:

Dermatophytosis,Camel,Clinical feature, Diagnosis.Treatment

Introduction:

Camels in their natural habitat are exposed to severe stress conditions which make them susceptible to many diseases [1, 2]. In last decades camels were reported to be resistant to many disease causing agents [3, 4], now it has been realized that they are susceptible like other livestock or even more, to the common disease causing pathogens [5-7].

Dermatophytes are among the most frequent causes of superficial skin infections in man and animals, known as Dermatophytosis (ringworm). It is caused by fungi in the genera *Microsporum*, *Trichophyton* and *Epidermophyton*. Ecologically, dermatophytes are classified to three groups anthropophilic (mostly associated with humans), zoophilic (associated with animals) and geophilic (found in the soil). Dermatophytosis in camels is the most frequent mycosis worldwide it has public health and economic importance. There are two forms of the disease sporadic as well as epidemic form [8-10]. Ringworm occurs in camels less than 3 year age and is characterized by circumscribed

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31 crusty hairless lesion, 1-2 cm in diameter distributed over the head, neck, shoulder,
32 limbs and flanks [1]. *T. verrucosum* is the most common cause of dermatophytosis in camels
33 [11].

34 **Epidemiology:**

35 Dermatophytosis was reported to be a common disease of camels worldwide [10,12]. Different
36 prevalence rate of the disease of 48 % [13] and 43.5% [14] were reported in camels, while [15]
37 reported lower prevalence of 8.58% in camels suffering from dermatophytosis. Camels less
38 than 3 year age were more susceptible to the disease than older animals.

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39 **Predisposing factors:**

- 40 1- Age
- 41 2- Breed
- 42 3- Production system
- 43 4- Close confinement
- 44 5- Immunosuppression (including immunosuppressive treatment)[16,17]

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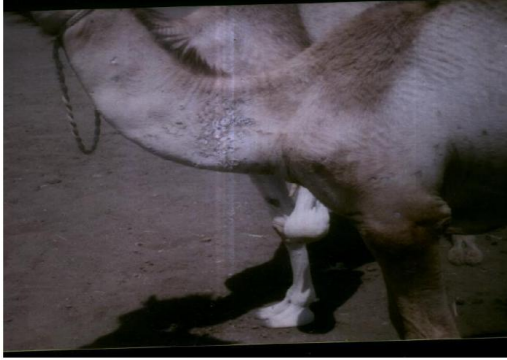
46 **Transmission**

47 The transmission of dermatophytosis is usually occurs by direct contact with infected host
48 (animals or humans) or asymptomatic carriers indirect contact with contaminated fomites besides
49 contact with soil [18,19].

52 **Clinical features:**

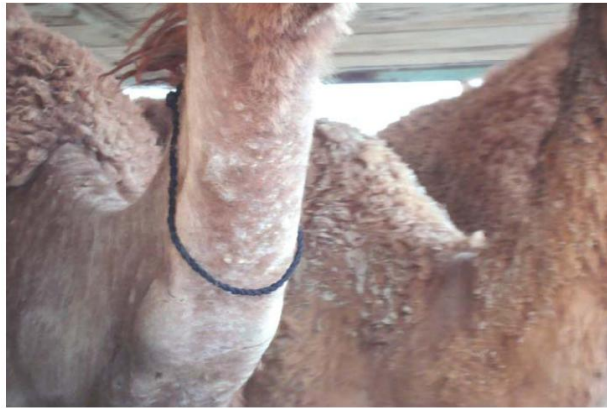
53
54 Ringworm in camels is characterized by circumscribed crusty hairless lesion, 1-2 cm in
55 diameter [1] on the head, the neck and shoulders with a possible extension to the flanks and
56 legs, leading sometimes to emaciation [20] (fig.1-8).

57



58

59 **Fig.1: Localized lesions of ringworm on camel neck [11]**



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61 **Fig.2: Circular lesions on the neck of camel[56]**

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63

64 **Fig.3: Alopecic ringworm lesions [57]**

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66

67 **Fig.4: A young camel calf has Crusty and hairless lesions on the shoulder.**[44]

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68

69 **Fig.5: Acamel from Elobied areas showing generalized lesions of ringworm**
70 **giving moth-eaten appearance of wool.**[11]



71

72 **Fig.6: Affected camel showed white hairless patches on different parts of the body. The**
73 **lesions typically consisted of an area of alopecia.**[44]

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74

75 **Fig.7: Epidermophyton fast spreading lesions with circular patches**[29]

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76

77 **Fig.8: Epidermophyton lesions giving just burning appearance [29]**

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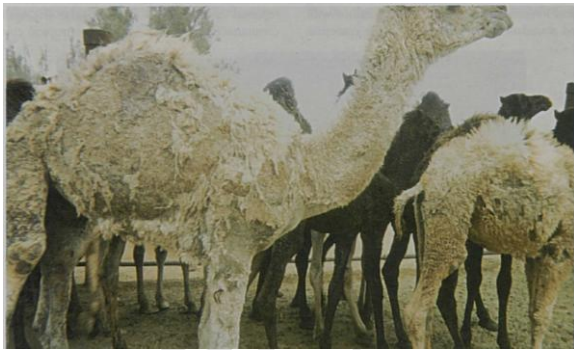
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80 **Mixed infection of dermatophytosis and other skin diseases:**

81 Mixed infection of dermatophytosis and *Sarcoptic scabiei* has been reported by [21, 22]. Mixed
82 infection of dermatophytosis caused by *M.gypseum* and *Dermatophilus congolensis* in dairy farm
83 camel was recorded by [15] fig. 9.

84

85



86

87 **Fig.9: Hair matting and crusty, hairless lesions on the flanks of a camel calf**

88

89 **Etiology:**

90 The disease in camel is mainly caused by *Trichophyton verrucosum* [12, 14, 23, 24, 25]. *T.*
91 *mentagrophytes* has been isolated by [11]. [26-28] were able to isolate *T. schoenleinii*. *T.*
92 *dankaliense* was isolated by [4]. [29] has been isolated *T. equinum* *T. concentricum* , *T.*
93 *tonsurans* , *T. violaceum* , *T. soudanense* , *T. rubrum* , *M. canis* , *M. nanum* and *M. ferrugineum*.
94 *M.gypseum* has been isolated by [15, 30, 31] *Epidermophyton floccosum* has been reported by
95 [32]

96

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references???

98 **Diagnosis:**

99 Dermatophytosis diagnosis is based on the clinical signs however in order to confirm the
100 diagnosis culturing and direct microscopic examination of skin scrapings from the periphery of
101 the lesions should be indicated [33].

102 **Collection of samples:**

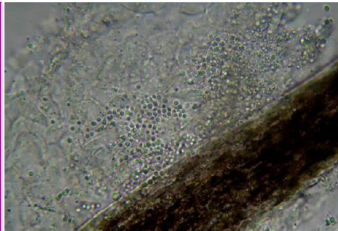
103 Skin scraping samples from the cattle that were suspected to be infected with dermatophytes will
104 be collected **on the basis of** gross lesion on their body after cleaning with ethyl alcohol 70%. Hair
105 and scrapings samples should be collected with forceps or scalpel just behind the extending
106 margin in the infected area. Samples can be kept in polyethylene bags [25].

107 **Direct examination:**

109 Each Sample from infected camel should be divided into two portions, one portion for direct
110 microscopic examination and the other for culture. Fungal hyphae and/or ectothrix spores are
111 determined to be seen in the direct examination when they appear to make hairs or hair
112 fragments thicker and rough with irregular surface.

113 Potassium Hydroxide (KOH) 10 or 20% is used as a clearing agent because it has keratinolytic
114 activity [34-36]. Infected hairs appear pale, wide and filamentous compared with normal hairs
115 when examined at x4 or x10 magnification, appearing. Arthrospores can be visible on high
116 magnification (x40).(fig.10)

117
118



119 **Fig.10: KOH preparation showing hair surrounded with chain of large ectothrix spores**
120 **X400[58]**

122 **Fungal culture:**

124 Fungal culture is considered the ‘gold standard’ for diagnosis [37]. Sabouraud’s dextrose agar
125 (SDA) containing cycloheximide, penicillin and streptomycin were used in most diagnostic
126

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127 laboratories. Plates should be incubated at 25°C for 5 weeks. Dermatophytes test media (DTM) is
128 recommended as the best media for isolation of dermatophytes because the presence of the red
129 color indicated positive result, this can help in early identification of highly suspected cultures
130 [38]. The isolates should be examined macroscopically and microscopically after staining with
131 lactophenol cotton blue using wet mount technique [39].(fig.11-20)

132 In addition to technique steps mentioned above, pigment production on corn meal agar, urease
133 activity on urea agar base, growth at 37°C on SDA.(fig.21)

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136 **Fig.11: Colony of *T. verrucosum* on the modified SDA. [55]**

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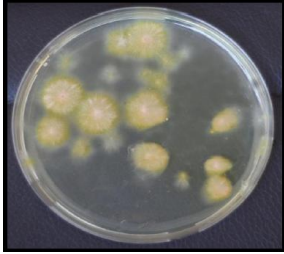
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139 **Fig.12: Colony *T. mentagrophytes*: surface of colony show powder-like shape, white, loose
140 irregular mycelium on the edge. [58]**

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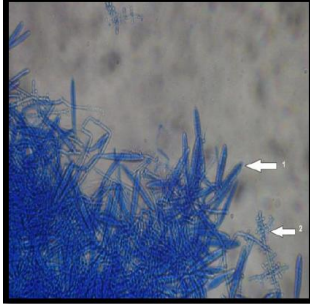
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145 **Fig.13: Colony surface of *M.canis*. [59]**
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148
149 **Fig.14: Colony of *Microsporum gypseum*. [60]**
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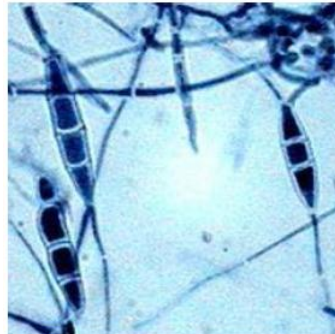
151
152 **Fig.15: Colony surface of *E.floccosum*[59]**
153



154
155 **Fig.16: 1)Macroconidia . 2) Microconidia in *T.mentagrophytes* . [59]**
156



159
160 **Fig.17: Macroconidia in *E.floccosum*. [59]**
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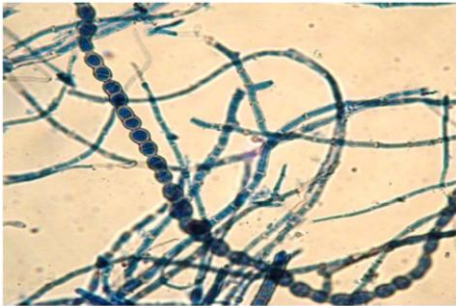
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165 **Fig.18: *Microsporum canis* microscopic observation in lactophenol cotton blue [61]**
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Fig.19: *Microsporium gypseum* microscopic observation in lactophenol cotton blue [60]



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Fig.20: Microscopic appearance of *T. verrucosum*. [52]



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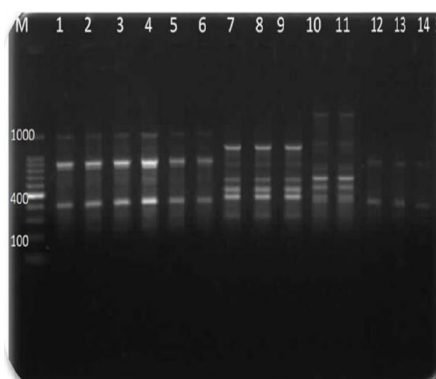
Fig.21: Growth of *T.mentogrophytes* on urea agar after 4 days showing hydrolysis of the urea.[62]

178 **Molecular diagnosis:**

179
180 Diagnosis with conventional methods is time-consuming because it might take up to 4 weeks or
181 longer to give the final results [40]. Furthermore, morphological identification may be confusing
182 due to polymorphism of dermatophytes [41]. During the last decade, a wide variety of molecular
183 techniques has become available as possible alternatives for routine identification of fungi in
184 clinical microbiology laboratories [42, 43].

185 Molecular identification for *Trichophyton* species isolated from camel skin lesions was done
186 using (GACA)₄ all the strains were amplified simply resulting PCR bands ranged from 2-5.
187 Three profiles of *Trichophyton mentagrophytes* have been detected so *T.mentagrophytes* is
188 known to be a species complex [44].(fig.22)

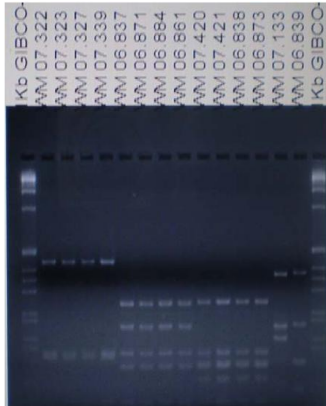
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191 **Fig.22: Agarose gel electrophoresis for PCR using (GACA)₄. M, molecular weight marker.**
192 **Lanes (1-6) first profile of *T.mentagrophytes*Lanes (7-9) second profile of *T.mentagrophytes*.**
193 **Lanes (10&11) third profile of *T. mentagrophytes*Lanes (12-14) *T.verrucosum*. [44]**

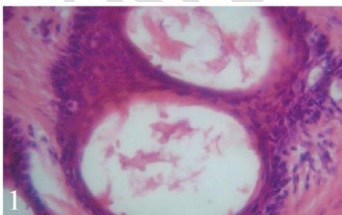
194 *Trichophyton* species isolated from camel and human were identified using restriction fragment
195 length polymorphism (RFLP), *Mva*1 was used as restriction enzyme. Five different patterns
196 of two to four bands were obtained. None of these different species gave the same profile pattern
197 [45]. (fig.23)



198
 199 **Fig.23: RFLP profiles of *Trichophyton* spp isolates. WM 07.322, WM 07.323, WM 07.327,**
 200 **WM 07.339 were *T.mentagrophytes var mentagrophytes*, WM06.837, WM 06.871, WM**
 201 **06.884, WM 06.861 were *T.mentagrophytes var interdigitale*, WM 07.420, WM07.421, WM**
 202 **06.838, WM 06.873 were *T.tonsurans*, WM 07.133 *T.mentagrophytes var erinacei* and WM**
 203 **06.839 was *T.rubrum*.[45]**

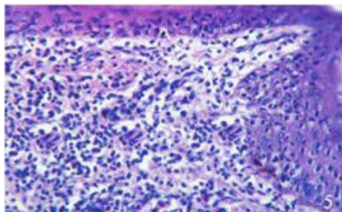
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 205 **Skin biopsy:**

206 Specimens from infected skin should be taken and fixed in 10% formaline solution then
 207 dehydrated, cleared and embedded in paraffin wax, sectioned at 4 µm thickness should be stained
 208 by haematoxylin and eosin for microscopical examination [46]. haematoxylin and eosin staining
 209 (H&E) may or may not identify dermatophytes and special stains such as periodic acid Schiff
 210 (PAS) and Grocott methenamine silver (GMS) are needed.
 211 Microscopically, the hair follicles and sweat glands exhibited cystic dilatation (fig. 24) and were
 212 lined by atrophied epithelium.



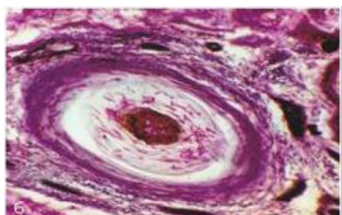
213
 214 **Fig.24: Ring worm in camels, noticed destructed hair shaft, perifolliculitis and cystic**
 215 **dilatation of the hair follicles (H&E X650). [46]**

216 Occasionally, perivascular dermatitis, and intra-epidermal pustules characterized by focal
217 aggregation of neutrophils mixed with eosinophil and karyorrhectic debris were reported (fig. 25).



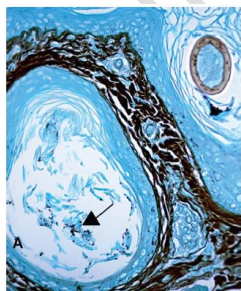
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219 **Fig.25: Ring worm in camels, noticed intra epidermal pustules and acanthosis (H&E**
220 **X400). [46]**
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223 The branched fungal hyphae were seen when sections stained with PAS (fig.26)
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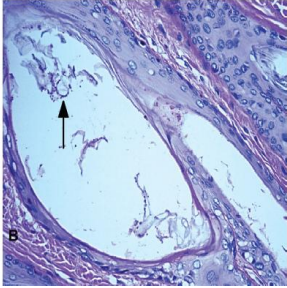


226
227 **Fig.26: Ring worm in camels, noticed trichophyton hyphae positive for PAS-reaction (H&E**
228 **X400). [46]**
229

230 The surface as well as intrafollicular hairs was colonised by large numbers of refractile or
231 slightly basophilic arthrospores and hyphae. These were coloured bright magenta with periodic
232 acid-Schiff stain and black with Gomori's methenamine silver stain. The Keratin-filled follicles
233 ruptured leading to prominent furunculosis [22]. (fig.27,28)



234
235 **Fig.27: Refractile arthrospores and hyphae appear: bright magenta with Periodic Acid-**
236 **Schiff stain [22].**



237 **Fig.28: Refractile arthrospores and hyphae appear black with Gomori's Methenamine**
238 **Silver stain [22].**
239

240
241 **Treatment:**

242 Optimal therapy of dermatophytosis requires a combination of topical antifungal therapy,
243 concurrent systemic antifungal therapy and environmental decontamination. The treatment
244 should be continued until two consecutive negative cultures (at weekly or bi-weekly intervals)
245 are obtained [47]. Topical treatments speed resolution of clinical lesions and may help prevent
246 zoonotic contagion. Systemic therapies that have prolonged residual activity in the skin and hair
247 provide the most effective treatments.

248 **Topical Therapy:**

- 249 1- 2% solution of tincture iodine [23].
250 2- 10% iodine ointment daily for three weeks [48].
251 3- Enilconazole Wash or spray with diluted emulsion (2000 ppm) four times at 3–4-day
252 intervals [49].

253
254
255 **Systemic Therapy:**

- 256 1- Griseofulvin 10 mg/kg body weight for 7 days in mild infections; in severe cases 2–3
257 weeks [49].

258 **Environmental decontamination:**

259 Dermatophytes can remain viable in infected soil for many years [50-52], so 10% hypochlorite
260 solution can be used as disinfectant [53].
261
262
263

264 **Vaccination:**

265 Live attenuated vaccine is used for prophylaxis and therapy for dermatophytosis caused by
266 *T.verrucosum* and *T.mentagrophytes* every five years [54].

267
268 **Conclusion:**

269 Dermatophytoses are the most common fungal infections in camels. Many studies were done
270 considering different aspects of the disease (eg. epidemiology, clinical presentation and
271 diagnosis, treatment, prevention, and control). Infected camel with dermatophytes can be a source
272 of infection to human this can lead to public health problem.

273
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