

**COMPARATIVE PROXIMATE, ANTIOXIDANT VITAMINS AND MINERAL COMPOSITION OF LEAVES OF FOUR SELECTED TROPICAL NUTRITIONAL PLANTS NAMELY: *Ocimum gratissimum*, *Piper guineense*, *Gongronema latifolium* and *Vernonia amygdalina*.**

**ABSTRACT**

**Aim:** The aim of the study was to carry out a comparative analysis of the proximate, antioxidant vitamins and mineral composition of the leaves of four selected tropical nutritional plant namely: *Ocimum gratissimum*, *Piper guineense*, *Gongronema latifolium* and *Vernonia amygdalina*. **Methodology:** The macro and micronutrients in the plant leaves were extracted by cold maceration in ethanol and subjected to quantitative proximate, antioxidant vitamins and minerals analysis. **Results:** For all four plants, carbohydrates was the major macronutrient constituents followed by fats, proteins, ash and fiber in that order. *G. latifolium* had the highest carbohydrate (64.09% w/w) and protein (12.53%) composition while *V. amygdalina* had the highest fat (29.43%) composition. Results of Ash analysis of the four leaves showed *P.guineense* to have the highest total mineral content (14.82%) followed by *V. amygdalina* (10.75%), *O. gratissimum* (4.60%) and *G. latifolium* (1.87%) in that order. *O. gratissimum* and *P.guineense* had the highest composition of fiber (7.53% and 7.22% respectively) closely followed by *G. latifolium* (6.03%) and *V. amygdalina* (2.92%). Vitamin analysis revealed that leaves of the four vegetable plants contained high levels of vitamin C and appreciable quantities of vitamins A and E. *V. amygdalina* leaf contained the highest concentration of vitamin C (4.54%) and A (0.12%) while vitamin E was pretty much the same for the four plants (0.07-0.09%). The mineral assay indicated that the leaves of the plants contain high levels of Magnesium (Mg), Phosphorus (P) and Calcium (Ca) relative to their copper (Cu), Zinc (Zn), Potassium (K) and Sodium (Na) contents. **Conclusion:** In conclusion, these plants were shown to be rich in carbohydrates, proteins and fats, vitamins and minerals justifying their use in diets. The plants were particularly rich in antioxidant vitamins and mineral justifying the therapeutic uses of various preparations of these leafy vegetables, in traditional medicine, for the treatment and management of diseases that have their etiology and pathophysiology in free radical generation and oxidative stress.

Key words: Proximate, Vitamins, Minerals, nutritional plants, *Ocimum gratissimum*, *Piper guineense*, *Gongronema latifolium* and *Vernonia amygdalina*.

**1. INTRODUCTION**

Plants since prehistoric times, have been used as spices, food and medicine in all cultures [1-3]. More than 70% of people in developing countries depend on plants (vegetables and fruits) for regular dietary needs [4]. It is well known that apart from energy needs, many plants and foods are ingested because of their perceived medicinal and health benefit. Indeed a significant amount of research has shown a correlation between a healthy diet and lifestyle and significant reductions in diseases and associated chronic conditions [5]. Plants are also a rich source of vitamins and minerals. Studies have identified a vast majority of antioxidant vitamins from vegetable plants like vitamins A, C and E [6]. Selective intake of

54 food containing these antioxidant vitamins, minerals and phytochemicals can prevent the  
55 onset of degenerative diseases like cardiovascular diseases, cancer and diabetes.

56

57 Given the plethora of vegetable plants available it becomes difficult to identify which plant  
58 should be added to our diet to address particular nutrient deficiency or ameliorate particular  
59 ailments. Moreover, some plants may contain appreciable levels of anti-nutrients rendering  
60 them unsafe for human consumption. It is with a view to establishing the relative proximate,  
61 antioxidant vitamins and mineral composition in some commonly used vegetable leafs in the  
62 southern region of Nigeria namely *Ocimum gratissimum*, *Piper guineense*, *Gongronema*  
63 *latifolium* and *Vernonia amygdalina*, that the current study is being carried out. The focus on  
64 the four plants stems from their common use as vegetables and spices in soups in the  
65 southern part of Nigeria. The plants have also been employed in ethnobotany for the  
66 treatment of various diseases. A comparative analysis of the proximate, antioxidant vitamins  
67 and minerals composition of the four plants will provide a bio-rational basis for the choice of  
68 the plants for addressing some nutrient deficiency. Earlier work in our laboratory had carried  
69 out a comparative analysis of the phytochemical composition of the four plants [7].

70

71 *O. gratissimum* commonly called African basil and belonging to the family *Lamiaceae*, is a  
72 herbaceous perennial flowering plant which is woody at its base. The leaf is called scent leaf  
73 because it possesses a pleasant aroma which is responsible for its use as spice and  
74 condiments in cooking. It is widely distributed in tropical Africa and Asia, especially India.  
75 The plant is economically important for its food flavoring (as spice and condiments) [8] and  
76 essential oil which has been widely used in food industries [9,10].

77

78 *P. guineense* (family *Piperaceae*) is a climbing perennial plant native to the tropical regions  
79 of Central and Western Africa. It is commonly referred to as Ashanti pepper, West African  
80 pepper or African black pepper. *P. guineense* is economically important for its culinary uses  
81 as well as medicinal, cosmetic and insecticidal uses [11]. It is a highly spicy plant and the  
82 leaves have pungent taste and a pleasant aroma when crushed. It thus imparts "heat",  
83 "pungency" and a spicy aroma to classic West African soups (stews). The plant oils is used  
84 as aromatics in the drink industry [12].

85

86 *G. latifolium*, commonly called "utazi," "aroeke" in the South Eastern and South Western  
87 parts of Nigeria respectively, belongs to the family *Asclepiadaceae*. It is primarily used as  
88 spice and vegetable for cooking and in traditional medicine [13]. A non-wood forest plant, it  
89 is native to West Africa and widely distributed elsewhere in tropical Africa and subtropical  
90 Asia.

91

92 *V. amygdalina*, popularly called bitter leaf, belongs to the family *Asteraceae*. It is widely used  
93 in the West African sub-region for a number of medicinal and nutritional purposes [14,15]. It  
94 has also been employed as a digestive tonic and appetizer [16].

95

96

97

## 98 2. MATERIALS AND METHOD

99

### 100 2.1 Plant Materials

101

102 Mature leaf samples of *O. gratissimum*, *P. guineense*, *G. latifolium* and *V. amygdalina* were  
103 harvested from local farms in Cross River State, South-south Nigeria.

104

### 105 2.2 Methods

106

#### 106 2.2.1 Extraction Procedure

107

108 Fresh leaves of each plant were washed and air dried at room temperature (25°C) for two  
109 weeks. The dried leaves were pulverized using a mechanical grinder. A weighed quantity,  
110 200g, of each plant material was extracted by cold maceration in absolute ethanol for 48  
111 hours. The extracts were double filtered, first with a white muslin cloth then with Whatman  
112 no.1 filter paper. The resulting ethanol leaf extracts were concentrated in vacuum using a  
113 rotary evaporator (at temperatures between 40°C and 45°C) to obtain a semi-solid mass.  
114 Weighed quantities of each extract were used in the macro and micro nutrient analysis  
115 according to experimental protocol.

## 117 2.2.2 Proximate Analysis

118  
119 Proximate composition of the leaf extracts was determined using methods prescribed by the  
120 Association of Official Analytical Chemists (AOAC) [17] and the Food and Agriculture  
121 organization (FAO) [18].

## 123 2.2.3 Determination of Mineral Composition

124  
125 Potassium and sodium were determined by the Flame photometric method while iron,  
126 copper, zinc, calcium and magnesium were determined by atomic absorption  
127 spectrophotometric method as described by James [19] and the Association of Official  
128 Analytical Chemists, AOAC [20]. Phosphorus was determined spectrophotometrically by the  
129 vanadomolybdate yellow method.

## 131 2.2.4 Determination of Some Antioxidant Vitamins

132  
133 Vitamin A and E concentration was determined by the spectrophotometric method as  
134 described by Pearson [21]. Vitamin C was determined by the method of AOAC [22].

# 137 3. RESULTS & DISCUSSIONS

## 139 3.1 Proximate Analysis

140  
141 The result of the proximate composition of the fresh leaves of the four plants is shown in  
142 Figure 1. For all four plants, carbohydrates was the major macronutrient constituents  
143 followed by fats, proteins, ash and fiber in that order. *G. latifolium* had the highest  
144 carbohydrate composition followed by *O. gratissimum*, and then *P.guineense* and finally *V.*  
145 *amygdalina*. Dietary carbohydrate is a major macronutrient for both humans and omnivorous  
146 animals; human adults in the Western countries obtain approximately half their daily caloric  
147 requirements from dietary carbohydrate while it is the major source of energy in other  
148 countries [23]. Carbohydrate is stored as glycogen, and although it is important for short-  
149 term energy needs, it is of very limited capacity for providing for energy needs beyond a few  
150 hours.

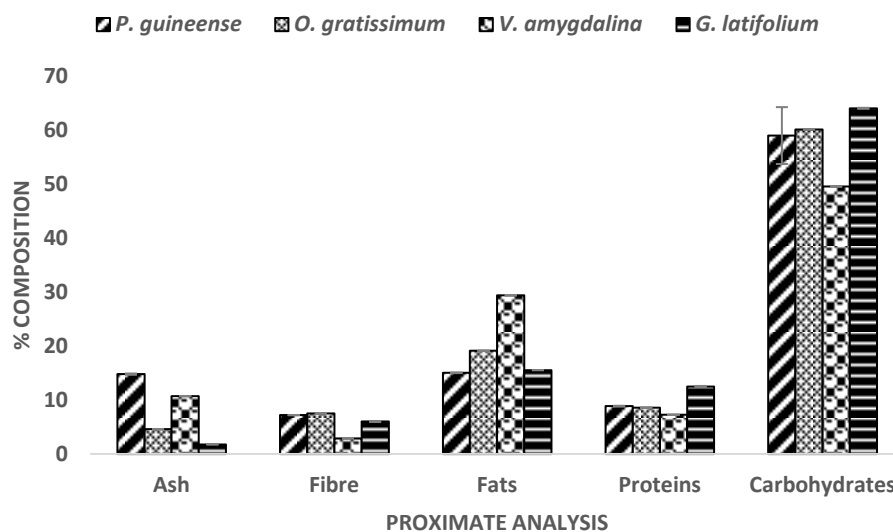
151  
152 Fats, the second highest macro nutrient in the four plants, constitute the highest energy in  
153 humans. *V. amygdalina* had the highest fat composition followed by *O. gratissimum*, *G.*  
154 *latifolium* and *P.guineense*.

155  
156 Protein is the second largest store of energy in the body after adipose tissue fat stores [24].  
157 The result of macronutrient analysis revealed that all the four plants were a fairly rich source  
158 of protein and may be used as a protein supplement for patients with protein deficiency  
159 diseases. *G. latifolium* had the highest protein composition followed by *P.guineense*, *O.*  
160 *gratissimum* and *V. amygdalina* in that order.

161

162 Ash, which refers to the inorganic residue remaining after ignition or complete oxidation of  
 163 organic matter in a food sample, is a measure of the total amount of minerals present within  
 164 the food [25]. Results of Ash analysis of the four leaves shows *P.guineense* to have the  
 165 highest total mineral content followed by *V. amygdalina*, *O. gratissimum* and *G. latifolium* in  
 166 that order.

167  
 168 **Fibre** is a measure of the quantity of indigestible cellulose, pentosans, lignin and other like  
 169 components in foods. Insoluble fibers can help promote bowel health and regularity. It also  
 170 support insulin sensitivity and may help reduce the risk of diabetes. *O. gratissimum* and  
 171 *P.guineense* had the highest composition of crude fiber closely followed by *G. latifolium* and  
 172 *V. amygdalina* (Figure 1).  
 173

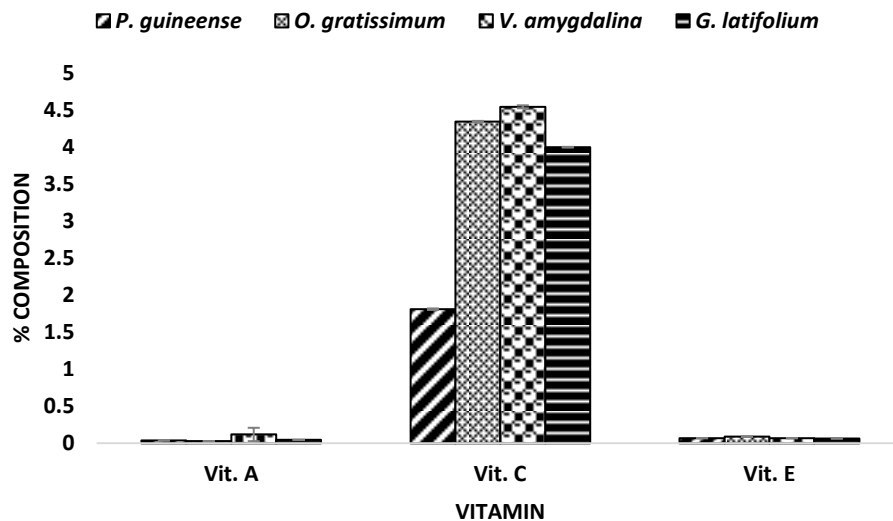


174  
 175  
 176 Fig 1: Proximate Analysis of crude leaf extracts of *P.guineense*, *O.gratissimum*, and *V. amygdalina* and *G.latifolium*. Values (%w/w) are expressed as mean  $\pm$  SEM.  
 177  
 178

### 180 3.2 Vitamins

181  
 182 The protective action of fruit and vegetables has been attributed to the presence of  
 183 antioxidants, especially antioxidants vitamins including ascorbic acid,  $\alpha$ -tocopherol and beta-  
 184 carotene [26-28]. The results of this study (Figure 2) revealed that leaves of the four  
 185 vegetable plants contain appreciable concentration of vitamin C, vitamin E and beta-  
 186 carotene (vitamin A). *V. amygdalina* leaf contained the highest concentration of vitamin C  
 187 and vitamin A. Other reports have also shown the plant to be rich in Vitamin C and A [28,29].  
 188 The level of Vitamin E was pretty much the same for the four plants with *O.gratissimum*  
 189 having a slightly higher amount. These results seem to suggest that fresh leaves of the  
 190 plants are good sources of antioxidant vitamins. Vegetable leaves/ Spices provide a variety  
 191 of vitamins and minerals as well as macronutrients to the diet [30]. These antioxidant  
 192 vitamins appear to be partly responsible for the antioxidant properties of the leaves. Vitamin  
 193 C is an antioxidant which helps to protect the body against cancer and other degenerative  
 194 diseases such as arthritis and type 2 diabetes mellitus and also strengthens the immune  
 195 system [31]. Vitamin C has also been shown to facilitate iron absorption by its ability to  
 196 reduce inorganic ferric ion to the ferrous form [32]. This suggests that the vegetable leaves  
 197 may be beneficial to people suffering from iron-deficiency anemia. Vitamin E ( $\alpha$ -tocopherol)  
 198 appears to be the most important lipid soluble antioxidant protecting membranes from lipid  
 199 peroxidation by acting as a chain-breaking antioxidant [33]. It also limits the oxidation of LDL

200 cholesterol and may help prevent or delay the development of atherosclerosis and/or  
 201 coronary heart disease (CHD) [34]. This probably explains why high vitamin E intake is  
 202 associated with lower rates of heart diseases. Beta-carotene is a lipid-soluble antioxidant. It  
 203 is the precursor of vitamin A, so it is necessary for the production and re-synthesis of  
 204 rhodopsin. High levels of beta-carotene intake have been correlated with lower risk of lung  
 205 cancer, coronary heart disease, stroke and age-related eye disease [35].  
 206



207  
 208 Fig 2: Quantitative Analysis of some Antioxidant Vitamins in *P.guineense*, *O.gratissimum*,  
 209 and *V. amygdalina* and *G.latifolium*. Values (%w/w) are expressed as mean  $\pm$  SEM.  
 210

### 211 3.3 Minerals

212  
 213 The results of the quantitative analysis of mineral elements (Figure 3) indicate that the  
 214 leaves of the plants contain high levels of Magnesium (Mg), Phosphorus (P) and Calcium  
 215 (Ca) relative to their copper (Cu), Zinc (Zn), Potassium (K) and Sodium (Na) contents. In  
 216 addition to the numerous biological roles these minerals play, they also serve as co-factor in  
 217 certain biochemical reactions including those involving antioxidant enzymes. Magnesium  
 218 serves as a co-factor for the enzyme catalase, a primary antioxidant that detoxifies hydrogen  
 219 peroxide by dismutation to water and oxygen. Similarly Copper and Zinc, are vital co-factor  
 220 of the different forms of SOD found in plants and animals [36]. Superoxide dismutase (SOD)  
 221 is a primary antioxidant enzyme that catalyses the dismutation or disproportion of superoxide  
 222 anion radicals ( $O_2^-$ ) to hydrogen peroxide and molecular oxygen [37]. It is therefore  
 223 suggested that these minerals contribute to the antioxidant properties of the plants probably  
 224 by boosting the levels of antioxidant enzymes such as SOD and catalase.  
 225

226 Except for *P.guineense*, Phosphorus (P) was the major constituents of the mineral elements  
 227 assayed. *O.gratissimum* had the highest phosphorus content closely followed by *V.*  
 228 *amygdalina* and *G.latifolium* in that order. Phosphorus is an ubiquitous mineral in the human  
 229 body and has diverse functions ranging from the transfer of genetic information to energy  
 230 utilization [38]. It forms the backbone of DNA and RNA, it is an essential component of  
 231 phospholipids that form all membrane bilayers and is an integral component of the body's  
 232 key energy source, adenosine triphosphate (ATP). Phosphorus also plays a vital role in the  
 233 dissociation of oxygen from hemoglobin, it is the main intracellular buffer and therefore is  
 234 essential for pH regulation of the human body and is a key component of the second  
 235 messenger molecules such as cyclic adenosine monophosphate (cAMP), cyclic guanine  
 236 monophosphate (cGMP) and inositol polyphosphates. Taken together with the equally high  
 237 level of carbohydrates, the four plants are a very good source of energy.  
 238

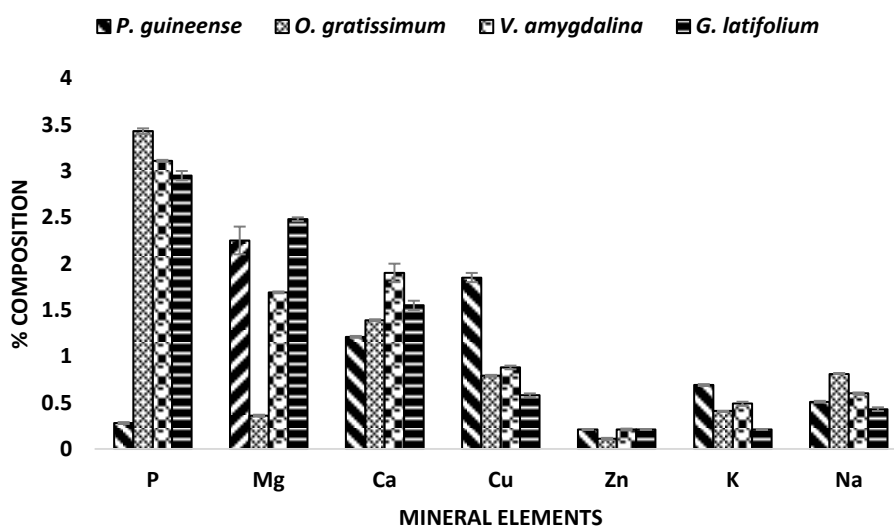
239 **Comparatively**, *G.latifolium* had the highest Mg content with *O.gratissimum* having the  
 240 lowest. *G.latifolium* is thus the plant of choice to address Mg deficiency. Mg plays an  
 241 essential role in a wide range of fundamental biological reactions. Apart from its cofactor  
 242 role, it is involved in bone mineralization, the building of proteins, muscle contraction, nerve  
 243 transmission and immune system health [4,39]  
 244

245 Calcium (Ca) is the most tightly regulated ion in the extracellular fluid (ECF). In higher  
 246 mammals, the most obvious role of calcium is structural or mechanical being responsible for  
 247 the mass, hardness, and strength of the bones and teeth [40]. Calcium is also involved in  
 248 cell movement and muscle contraction to nerve transmission, glandular secretion, and even  
 249 cell division where it acts as both a signal transmitter from the outside of the cell to the inside  
 250 and as an activator or stabilizer of the functional proteins involved. Calcium also plays a role  
 251 in the regulatory activities of parathyroid hormone [PTH], calcitonin [CT], and a key activity of  
 252 vitamin D. Ca was more predominant in *V. amygdalina* followed by *G.latifolium*,  
 253 *O.gratissimum* and *P.guineense* in that order.  
 254

255 Copper is a constituent of many enzymes including superoxide dismutase. It is also required  
 256 for iron metabolism [4,41]. It was more prevalent in *P.guineense*.  
 257

258 Zinc plays a catalytic, structural, and regulatory role in the body [42]. Zinc is essential for  
 259 general growth and proper development of the reproductive organs and for normal  
 260 functioning of the prostate gland. Apart from SOD, Zinc is a co-factor of over 300 enzymes  
 261 including carbonic anhydrase, which is crucial to maintenance of acid-base balance in the  
 262 blood, and alcohol dehydrogenase that break down alcohol. It is also a component of insulin  
 263 and plays a role in its processing, storage, secretion and action [43]. The Zinc content of *P.*  
 264 *guineense* may be responsible for the observed stimulated sexual behaviors of mature male  
 265 rats fed with extract of *P. guineense* [44]. The level of the mineral was pretty much the same  
 266 in *V. amygdalina*, *G.latifolium* and *P. guineense*. *O.gratissimum* had the lowest level of the  
 267 mineral.  
 268

269 Sodium (Na) and potassium (K) (and chloride ions Cl<sup>-</sup>) are the major electrolytes located in  
 270 all body fluids. While sodium is extracellular, potassium is intracellular. They are responsible  
 271 for the maintenance of acid/base balance, nerve transmission and muscle contraction and  
 272 regulation of fluid movement in and out of cells [45]. *P. guineense* had the highest amount  
 273 of potassium while *O.gratissimum* had the highest level of sodium.



274  
 275  
 276 Fig 3: Quantitative Analysis of some Minerals in *P.guineense*, *O.gratissimum*, *V. amygdalina*  
 277 and *G.latifolium*. Values (%w/w) are expressed as mean  $\pm$  SEM.

278  
279  
280  
281  
282  
283  
284  
285  
286  
287  
288  
289  
290  
291  
292  
293  
294  
295  
296  
297  
298  
299  
300  
301  
302  
303  
304  
305  
306  
307  
308  
309  
310  
311  
312  
313  
314  
315  
316  
317  
318  
319  
320  
321  
322  
323  
324  
325  
326  
327  
328  
329  
330  
331

#### 4. Conclusion

In summary, the four plants, *P.guineense*, *O.gratissimum*, *V. amygdalina* and *G.latifolium*, have been shown to be rich in carbohydrates, proteins and fats, vitamins and minerals justifying their use in diets. The plants are particularly rich in antioxidant vitamins and mineral. Taken together with earlier work on the comparative phytochemical analysis of these plants [7], the findings have good correlation with the therapeutic uses of the various preparations of these leafy vegetables in traditional medicine for the treatment and management of diseases that have their etiology and pathophysiology in free radical generation and oxidative stress like diabetes, arthritis, rheumatism, eye problems and infectious diseases such as AIDS. Increased consumption of the leaves of these plants is therefore recommended, especially as they have been shown to contain low levels of anti-nutrients [7], for optimized health and wellness, and to boost the endogenous antioxidant system and in so doing, help prevent the development of certain free radical related diseases.

#### COMPETING INTERESTS

The Authors declare that no competing interests exist.

#### References

1. Farnsworth NR, Akerele O, Bingel AS, Soejarto DD, Guo Z. World Health Organ. 1985; 63: 965.
2. Bandaranayake, WM. Quality Control, Screening, Toxicity, and Regulation of Herbal Drugs. In: Ahmad I, Aqil F, and Owais M, editors. Modern Phytomedicine. Turning Medicinal Plants into Drugs. WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim; 2006; pp25-57.
3. Odugbemi TA. A textbook of Medicinal Plants from Nigeria. Lagos: University of Lagos Press; 2008.
4. Nahak G, Suar M, Sahu RK. Antioxidant Potential and Nutritional values of vegetables: A Review. Research Journal of Medicinal Plant. 2014; 8(2): 50-81.
5. Poe K. Plant-Based diets and phytonutrients: Potential health benefits and disease prevention. Arch Med. 2017; 9(6): 7.
6. Godber JS. Nutrient bioavailability in humans and experimental animals. J Food quality. 1990; 13: 21-36.
7. Mgbeje BIA, Umoh EU and Emmanuel-Ikpeme C. Comparative Analysis of Phytochemical Composition of Four Selected Tropical Medicinal Plants Namely: *Ocimum gratissimum*, *Piper guineense*, *Gongronema latifolium* and *Vernonia amygdalina*. Journal of Complementary and Alternative Medical Research. 2019: 7(3): 1-11.
8. Akinmoladun AC, Ibukun EO, Afor ED, Obuotor EM & Farombi, EO. Phytochemical constituent and antioxidant activity of extract from the leaves of *Ocimum gratissimum*. Science Research Essay. 2007; 2(5): 163-166.
9. Lachowicz KJ, Jones GP, Briggs DR, Bienuenu FE, Palmer MV, Tings SST & Hunter MO. Characteristics of essential oils from basil (*Ocimum basilicum*) grown in Australia. J. Agric. Food Chem. 1996; 144: 877-881.
10. Machale KW, Niranjana KU & Pangarkar VG. Recovery of dissolved essential oils from condensate waters of basil and *Mentha arvensis* distillation. J. Chem. Tech. Biotech. 1997; 69: 362-366.
11. Okwute SK. Plants derived pesticidal and antimicrobial agents for use in Agriculture. A review of phytochemical and biological studies on some Nigeria plants. J of Agric. Science and Technology. 1992; 2(1): 62-70.

- 332 12. Rehm SS & Espig GT. The cultivated plants of the tropics and subtropics cultivation,  
333 economic value, utilization. Germany: Verlay Josef Margraf; 1991, 552 pps. ISBN 3-  
334 8236-1169-0.
- 335 13. Ugochuwku NH, Babady NF, Cobourne MN & Gasset SR. The effect of *Gongronema*  
336 *latifolium* extracts on serum lipid profile and oxidative stress in hepatocytes of diabetic  
337 rats. J Biosci. 2003; 28(1): 1-5.
- 338 14. Igile GO, Fafunsho M, Fasanmade A, Burda S, Jurzysta M, & Oleszek W. Toxicity of  
339 *Vernonia amygdalina* leaves, extracts and purified saponins in mice. Proc. Eurp. Food  
340 Tox. 1994; 2: 394-399.
- 341 15. Okafor JC. Conservation and use of traditional vegetable from woody forest species in  
342 south eastern Nigeria. Fame Agriculture Centre, Enugu, Nigeria; 2005. Pp 55-59.  
343 Available online at <http://www.biodiversityinternational.org>.
- 344 16. Iwu MM. Handbook of African medicinal plants. 1<sup>st</sup> ed. Florida: CRC Press; 1993; pp.  
345 221-222.
- 346 17. AOAC. *Official Methods of Analysis* 14th ed. Association of Official Analytical Chemists,  
347 Washington DC; 1984.
- 348 18. FAO. Manuals of food quality control. Food analysis: general techniques, additives,  
349 contaminants and composition. FAO Food and Nutrition Paper. 1986; 14(7): 203-232.
- 350 19. James CS. *Analytical Chemistry of Food*. New York: Chapman Hall; 1995. Available at  
351 <http://dx.doi.org/10.1007/978-1-4615-2165-5>.
- 352 20. AOAC. Official methods of analysis. 15th ed. Association of Official Analytical Chemists  
353 Washington D.C; 1990 <http://dx.doi.org/10.3923/pjn.2009.1204.1208>.
- 354 21. Pearson, D. Chemical analysis of food. 7th ed. Edinburgh, New York: Churchill  
355 Livingstone; 1976, Pp. 7-9.
- 356 22. AOAC. Official Methods of Analysis. 13th ed. Association of Official Analytical Chemists,  
357 Washington D.C; 1980.
- 358 23. Keim NL, Levin RJ, and Havel PJ. Carbohydrates. In: Ross AC, Caballero B, Cousins  
359 RJ, Tucker KL, Ziegler TR, editors. Modern Nutrition in health and disease. 11th edition.  
360 Philadelphia: Lippincott Williams & Wilkins; 2014; pp 36-57.
- 361 24. Cahill GF. Starvation in man. N Engl J Med. 1970; 282:668-75.
- 362 25. Ismail BP. Ash content determination. In: Nielsen S, ed. Food Analysis Laboratory  
363 Manual. Food science Text series. Cham: Springer; 2017; Pp 117-119.
- 364 26. Cao GH, Sofic EE & Prior RL. Antioxidant capacity of tea and common vegetable.  
365 Journal of Agriculture and Food Chemistry. 1996; 44: 3426-3430.
- 366 27. Grivetti LE & Ogle BM. Value of traditional foods in meeting macro-and micro nutrients  
367 needs: the wild plant connection. Nutrition Research and Reviews. 2000; 13: 31-46.
- 368 28. Odukoya OA, Inya-Agha SI, Segun FI, Sofidiya MO & Ilori OO. (2007). Antioxidant  
369 activity of selected Nigeria green leafy vegetables. American Journal of food  
370 Technology. 2007; 2(3): 169-175.
- 371 29. Palaniswamy UR, Caporuscio CX & Stuarts JD. A Chemical analysis of antioxidant  
372 Vitamins in fresh curry leaf (*Murraya Koenigii*) by reversed phase HPLC with UV  
373 detection. Acta Horticulture (ISHS). 2003; 620: 475-478. Available online at  
374 <http://doi.org/10.17660/ActaHortic.2003.620.57>
- 375 30. Hiza H. Availability of Spices on the rise in the U.S. food supply. US Department of  
376 Agriculture, Centre for Nutrition Policy and Promotion. Nutrition insight. 2008; 39.
- 377 31. Mensah JK, Okoli RI, Ohaju-Obodo JO & Eifediyi KB. Phytochemical, nutritional and  
378 medicinal properties of some leafy vegetable consumed by Ede people of Nigeria.  
379 African Journal of Biotechnology. 2008; 7(14): 2304-2309. Available online at  
380 <http://www.academicjournals.org/AJB>.
- 381 32. Charttejea MN & Shinde RQ. Textbook of medical Biochemistry. 6th Ed. New Delhi:  
382 Jaypee Brothers; 2005; Pp. 124 -132.
- 383 33. Traber MG & Atkinson JC. Vitamin E, antioxidants and nothing more. Free Ecological  
384 Biology and Medicine. 2007; 43(1): 4-15.
- 385 34. Jialal IW & Fuller CJ. Effect of vitamin E, Selenium and beta-Carotene on LDL oxidation  
386 and atherosclerosis. Canadian Journal of Cardiol. 1995; 11: 97G-103G.



- 387 35. Johnson EJ. Aging and carotene nutritive. *Age*. 1993; 16: 59-66.  
388 36. Ahmad, S. Oxidative Stress and antioxidant defenses in Biology. New York: Chapman  
389 and hall; 1995; Pp. 1-18, 238-258.  
390 37. Zelko IB, Mariani TF & Folz RM. Superoxide dismutase multigene family: a comparison  
391 of the CU/Zn-SOD (SOD1), Mn-SOD, (SOD2) and EC-SOD (SOD3) gene structures,  
392 evolution and expression. *Free Radical Biology and Medicine*. 2002; 33(3): 337-349.  
393 38. O'Brien KO, Kerstetter JE, Insogna KL. Phosphorus. In: Ross AC, Caballero B, Cousins  
394 RJ, Tucker KL, Ziegler TR, editors. *Modern Nutrition in health and disease*. 11th edition.  
395 Philadelphia: Lippincott Williams & Wilkins; 2014; pp 150-158.  
396 39. Rude RK. Magnesium. In: Ross AC, Caballero B, Cousins RJ, Tucker KL, Ziegler TR,  
397 editors. *Modern Nutrition in health and disease*. 11th edition. Philadelphia: Lippincott  
398 Williams & Wilkins; 2014; pp 159-175.  
399 40. Weaver CM & Heany RP. Calcium. In: Ross AC, Caballero B, Cousins RJ, Tucker KL,  
400 Ziegler TR, editors. *Modern Nutrition in health and disease*. 11th edition. Philadelphia:  
401 Lippincott Williams & Wilkins; 2014; pp 133-149.  
402 41. Collins JF. Copper. In: Ross AC, Caballero B, Cousins RJ, Tucker KL, Ziegler TR,  
403 editors. *Modern Nutrition in health and disease*. 11th edition. Philadelphia: Lippincott  
404 Williams & Wilkins; 2014; pp 206-216.  
405 42. King JC and Cousins RJ. Zinc. In: Ross AC, Caballero B, Cousins RJ, Tucker KL,  
406 Ziegler TR, editors. *Modern Nutrition in health and disease*. 11th edition. Philadelphia:  
407 Lippincott Williams & Wilkins; 2014; pp 189-205.  
408 43. Li YV. Zinc and Insulin in pancreatic beta-cells. *Endocrine*. 2014; 45(2): 178-189.  
409 44. Kamtchouing P, Mbongue GYF, Dimo T, Watcho P, Jatsa HB & Sokeng SD. Effect of  
410 *Aframomum melegueta* and *Piper guineense* on sexual behavior of mate rats.  
411 *Behavioural Pharmacology*. 2002; 13: 243-247.  
412 45. Vasudevan DM and Sreekumari S. *Textbook of Biochemistry for Medical Students*. 5<sup>th</sup>  
413 Ed. New Delhi: JP Medical Ltd; 2007; pp. 373-378.  
414