# Original Research Article Original Research Article COMPARATIVE PROXIMATE, VITAMINS COMPOSITION OF LEAVES OF FOUR SELECTED TROPICAL NUTRITIONAL PLANTS NAMELY: Ocimum gratissimum, Piper guineense, Gongronema latifolium and Vernonia amygdalina.

#### 10 ABSTRACT

7 8 9

11

Aim: The aim of the study was to carry out a comparative analysis of the proximate, vitamin 12 and mineral composition of the leaves of four selected tropical nutritional plants namely: 13 Ocimum gratissimum, Piper guineense, Gongronema latifolium and Vernonia amygdalina. 14 Methodology: The macro and micronutrients in the plant leaves were extracted by cold 15 maceration in ethanol and subjected to quantitative proximate, vitamin and mineral analysis. 16 **Results:** For all four plants, carbohydrates was the major macronutrient constituents (range 17 49.61-64.09% dry wt.) followed by fats (15.06-29.43%), ash (1.81-14.82%) and fiber (2.92-18 7.53%) in that order. G. latifolium had the highest carbohydrate (64.09% dry weight) and 19 protein (12.53%) composition while V. amygdalina had the highest fat (29.43%) composition. 20 21 Results of Ash analysis of the four leaves showed P.guineense to have the highest total mineral content (14.82%) followed by V. amygdalina (10.75%), O. gratissimum (4.60%) and 22 G. latifolium (1.87%) in that order. O. gratissimum and P.guineense had the highest 23 composition of fiber (7.53% and 7.22% respectively) closely followed by G. latifolium (6.03%) 24 and V. amygdalina (2.92%). Vitamin analysis revealed that leaves of the four vegetable 25 plants contained high levels of vitamin C (range 18.1-45.4 mg/100g) and appreciable 26 quantities of vitamins A (0.3-1.2mg/100g) and E (0.67-0.9 mg/100g). V. amygdalina leaf 27 contained the highest concentration of vitamin C (45.4 mg/100g) and A (1.2 mg/100g) while 28 O. gratissimum had the highest vitamin E content (0.9 mg/100g). The mineral assay 29 indicated that the leaves of the plants contain high levels of Magnesium (Mg)(3.6-24.8 30 mg/100g), Phosphorus (P) (2.8-34.3), Calcium (Ca) (12.1-19.0) and copper (Cu) (5.8-18.5) 31 relative to their Zinc (Zn) (1.1-2.1), Potassium (K) (2.1-6.9) and Sodium (Na) (4.3-8.1) 32 contents. **Conclusion**: In conclusion, these plants were shown to be rich in carbohydrates, 33 proteins and fats, vitamins and minerals justifying their use in diets. The plants were 34 particularly rich in vitamins and mineral with antioxidant properties and could explain the 35 therapeutic uses of the various preparations of these leafy vegetables, in traditional 36 37 medicine, for the treatment and management of diseases that have their etiology and pathophysiology in free radical generation and oxidative stress. 38

39 40

41 Key words: Proximate, Vitamins, Minerals, nutritional plants, Ocimum gratissimum, Piper 42 guineense, Gongronema latifolium and Vernonia amygdalina.

- 43 44
- 45 46
- 47
- 48
- 49
- 50
- 51
- 52
- 53

#### 54 **1. INTRODUCTION**

55

Plants since prehistoric times, have been used as spices, food and medicine in all cultures 56 57 [1-3]. More than 70% of people in developing countries depend on plants (vegetables and fruits) for regular dietary needs [4]. It is well known that apart from energy needs, many 58 59 plants and foods are ingested because of their perceived medicinal and health benefit. Indeed a significant amount of research has shown a correlation between a healthy diet and 60 lifestyle and significant reductions in diseases and associated chronic conditions [5]. Plants 61 62 are also a rich source of vitamins and minerals. Studies have identified a vast majority of vitamins with antioxidant properties from vegetable plants like vitamins A, C and E [6]. 63 64 Selective intake of food containing these vitamins, minerals and phytochemicals can prevent 65 the onset of degenerative diseases like cardiovascular diseases, cancer and diabetes.

66

Given the plethora of vegetable plants available it becomes difficult to identify which plant 67 should be added to our diet to address particular nutrient deficiency or ameliorate particular 68 ailments. Moreover, some plants may contain appreciable levels of anti-nutrients rendering 69 70 them unsafe for human consumption. It is with a view to establishing the relative proximate, 71 vitamins and mineral composition in some commonly used vegetable leafs in the southern 72 region of Nigeria namely Ocimum gratissimum, Piper guineense, Gongronema latifolium and Vernonia amygdalina, that the current study is being carried out. The focus on the four 73 plants stems from their common use as vegetables and spices in soups in the southern part 74 75 of Nigeria. The plants have also been employed in ethnobotany for the treatment of various 76 diseases. A comparative analysis of the proximate, vitamin and mineral composition of the four plants will provide a bio-rational basis for the choice of the plants for addressing some 77 78 nutrient deficiency. Earlier work in our laboratory had carried out a comparative analysis of 79 the phytochemical composition of the four plants [7].

80

O. gratissimum commonly called African basil and belonging to the family Lamiaceae, is a
 herbaceous perennial flowering plant which is woody at its base. The leaf is called scent leaf
 because it possesses a pleasant aroma which is responsible for its use as spice and
 condiments in cooking. It is widely distributed in tropical Africa and Asia, especially India.
 The plant is economically important for its food flavoring (as spice and condiments) [8] and
 essential oil which has been widely used in food industries [9,10].

87

*P. guineense* (family *Piperaceace*) is a climbing perennial plant native to the tropical regions
of Central and Western Africa. It is commonly referred to as Ashanti pepper, West African
pepper or African black pepper. *P. guineense* is economically important for its culinary uses
as well as medicinal, cosmetic and insecticidal uses [11]. It is a highly spicy plant and the
leaves have a pungent taste and pleasant aroma when crushed. It thus imparts "heat",
"pungency" and a spicy aroma to classic West African soups (stews). The plant oils is used
as aromatics in the drink industry [12].

95

G. latifolium, commonly called "utazi," "aroeke" in the South Eastern and South Western parts of Nigeria respectively, belongs to the family *Asclepiadaceae*. It is primarily used as spice and vegetable for cooking and in traditional medicine [13]. A non-wood forest plant, it is native to West Africa and widely distributed elsewhere in tropical Africa and subtropical Asia.

101

102 V. amygdalina, popularly called bitter leaf, belongs to the family Asteraceae. It is widely used
 103 in the West African sub-region for a number of medicinal and nutritional purposes [14,15]. It
 104 has also been employed as a digestive tonic and appetizer [16].

- 105
- 106
- 107

## 108 2. MATERIALS AND METHOD

109

## 110 **2.1 Plant Materials**

Mature leaf samples of O. gratissimum, P. guineense, G. latifolium and V. amygdalina were
harvested from local farms in Cross River State, South-south Nigeria.

# 114115 **2.2 Methods**

#### 116 **2.2.1 Extraction Procedure**

117

Fresh leaves of each plant were washed and air dried at room temperature (25°C) for two 118 weeks. The dried leaves were pulverized using a mechanical grinder. A weighed quantity, 119 120 200g, of each plant material was extracted by cold maceration in absolute ethanol for 48 hours. The extracts were double filtered, first with a white muslin cloth then with Whatman 121 no.1 filter paper. The resulting ethanol leaf extracts were concentrated in vacuum using a 122 rotary evaporator (at temperatures between 40°C and 45°C) to obtain a semi-solid mass. 123 Weighed quantities of each extract were used in the macro and micro nutrient analysis 124 125 according to experimental protocol.

#### 127 2.2.2 Proximate Analysis

128

126

Proximate composition of the leaf extracts was determined using methods prescribed by the Association of Official Analytical Chemists (AOAC) [17] and the Food and Agriculture organization (FAO) [18].

#### 133 **2.2.3 Determination of Mineral Composition**

134

Potassium and sodium were determined by the Flame photometric method while iron, copper, zinc, calcium and magnesium were determined by atomic absorption spectrophotometric method as described by James [19] and the Association of Official Analytical Chemists, AOAC [20]. Phosphorus was determined spectrophotometrically by the vanadomolybdate yellow method.

#### 141 **2.2.4 Determination of Some Vitamins**

142
143 Vitamin A and E concentration was determined by the spectrophotometric method as
144 described by Pearson [21]. Vitamin C was determined by the method of AOAC [22].

145

# 146 2.2.5 Statistical Analysis147

148 Data was presented as mean ±standard error of mean. Quantitative data generated were 149 analyzed by Anova to test the significance of the data at 5% confidence limit (p<0.05)

# 150151 3. RESULTS & DISCUSSIONS

- 152153 **3.1 Proximate Analysis**
- 154

The result of the proximate composition of the fresh leaves of the four plants is shown in Figure 1. For all four plants, carbohydrates was the major macronutrient constituents (range 49.61-64.09% dry wt.) followed by fats (15.06-29.43%), proteins (7.28-12.53%), ash (1.81-14.82%) and fiber (2.92-7.53%) in that order.

159

160 *G. latifolium* had the highest carbohydrate composition (64.09% dry weight) followed by *O.* 161 *gratissimum* (60.19%), *P.guineense* (59.04%) and finally *V. amygdalina* (49.61%). A report 162 by Asaolu et al. [23], on the proximate and mineral composition of Nigerian leafy vegetables, puts a range of 1.22-8.65% dry weight for the three plants O. gratissimum, V. amygdalina 163 and G. latifolium. It is worth noting that while carbohydrates was the major constituent in our 164 165 study, protein was the major constituent in the report by Asaolu et al. [23]. The variation in composition may be as a result of variation in soil nutrient, environmental factors, age of 166 plant at harvest, geographic location, diurnal and seasonal variations, method of cultivation, 167 168 time of harvesting and procedures in extraction and preparation. Dietary carbohydrate is a major macronutrient for both humans and omnivorous animals; human adults in the Western 169 170 countries obtain approximately half their daily caloric requirements from dietary carbohydrate while it is the major source of energy in other countries [24]. Carbohydrate is stored as 171 172 glycogen, and although it is important for short-term energy needs, it is of very limited 173 capacity for providing for energy needs beyond a few hours.

174

Fats, the second highest macro nutrient in the four plants (15.06-29.43% dry wt.), constitute
the highest energy in humans. *V. amygdalina* had the highest fat composition followed by *O. gratissimum*, *G. latifolium* and *P.guineense*. Asaolu *et al.* [23] reported a range of (3.519.05%) while Okafor [25] reported a range of 4.5-18.77% for the three plants, *P.guineense*, *G. latifolium* and *V. amygdalina*.

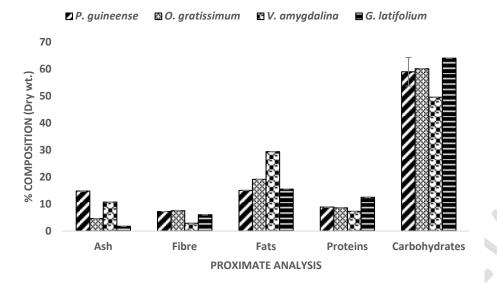
180

Protein is the second largest store of energy in the body after adipose tissue fat stores [26]. The result of macronutrient analysis revealed that all the four plants were a fairly rich source of protein (7.28-12.53% dry wt.) and may be used as a protein supplement for patients with protein deficiency diseases. *G. latifolium* had the highest protein composition followed by *P.guineense*, *O. gratissimum* and *V. amygdalina* in that order. A similar report by Asaolu *et al.* [23] and Okafor [25] put the range at 50.94-66.71% and 18.54-62.66% dry wt. respectively.

188

Ash, which refers to the inorganic residue remaining after ignition or complete oxidation of organic matter in a food sample, is a measure of the total amount of minerals present within the food [27]. Results of Ash analysis (1.81-14.82%) of the four leaves shows *P.guineense* to have the highest total mineral content followed by *V. amygdalina, O. gratissimum* and *G. latifolium* in that order. Asaolu *et al.* [23] and Okafor [25] reported a range of 9.01-13.01% and 10.13-15.56% respectively.

Fibre is a measure of the quantity of indigestible cellulose, pentosans, lignin and other like components in foods. Insoluble fibers can help promote bowel health and regularity. It also support insulin sensitivity and may help reduce the risk of diabetes. The fibre content in this study ranged from 2.92-7.53%. *O. gratissimum* and *P.guineense* had the highest composition of crude fiber closely followed by *G. latifolium* and *V. amygdalina* (Figure 1). The range for three of these plants as reported by Asaolu *et al.* [23] was 4.02-12.08% dry wt.



207

209

Fig 1: Proximate Analysis of crude leaf extracts of *P.guineense*, *O.gratissimum*, and *V. amygdalina* and *G.latifolium*. Values (% dry wt.) are expressed as mean <u>+</u> SEM.

### 208 3.2 Vitamins

The protective action of fruit and vegetables has been attributed to the presence of 210 antioxidants, especially vitamins known to have antioxidant properties like ascorbic acid, a-211 tocopherol and beta-carotene [28-30]. The results of this study (Figure 2) revealed that 212 leaves of the four vegetable plants contain appreciable concentration of vitamin C (range 213 18.1-45.4 mg/100g), vitamin E (0.67-0.9mg/100g) and beta-carotene (vitamin A)(0.3-214 215 1.2mg/100g). V. amygdalina leaf contained the highest concentration of vitamin C (45.4mg/100g) and vitamin A (1.2mg/100g). Other reports have also shown the plant to be 216 217 rich in Vitamin C and A [30,31](13.41 and 197.5 mg/100g respectively for vitamin C respectively and a carotenoid value of 30mg/100g [31]). Odukova et al. [30] also reported a 218 Vitamin C value of 187.11mg/100g for G.latifolium [30]. These results seem to suggest that 219 fresh leaves of the plants are good sources of vitamins with antioxidant activities. Vegetable 220 leaves/ Spices provide a variety of vitamins and minerals as well as macronutrients to the 221 diet [32]. These vitamins with antioxidant properties may be partly responsible for the 222 223 antioxidant properties of the leaves. Vitamin C is an antioxidant which helps to protect the body against cancer and other degenerative diseases such as arthritis and type 2 diabetes 224 mellitus and also strengthens the immune system [33]. Vitamin C has also been shown to 225 facilitate iron absorption by its ability to reduce inorganic ferric ion to the ferrous form [34]. 226 This suggests that the vegetable leaves may be beneficial to people suffering from iron-227 228 deficiency anemia. Vitamin E ( $\alpha$ -tocopherol) appears to be the most important lipid soluble antioxidant protecting membranes from lipid peroxidation by acting as a chain-breaking 229 antioxidant [35]. It also limits the oxidation of LDL cholesterol and may help prevent or delay 230 the development of atherosclerosis and/or coronary heart disease (CHD) [36]. This probably 231 explains why high vitamin E intake is associated with lower rates of heart diseases. Beta-232 carotene is a lipid-soluble antioxidant. It is the precursor of vitamin A, so it is necessary for 233 the production and re-synthesis of rhodopsin. High levels of beta-carotene intake have been 234 235 correlated with lower risk of lung cancer, coronary heart disease, stroke and age-related eye 236 disease [37].

237

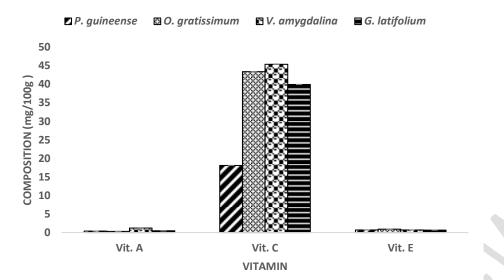


Fig 2: Quantitative Analysis of some Vitamins in *P.guineense, O.gratissimum,* and *V. amygdalina* and *G.latifolium.* Values are expressed as mean <u>+</u> SEM.

# 241242 **3.3 Minerals**

#### 243

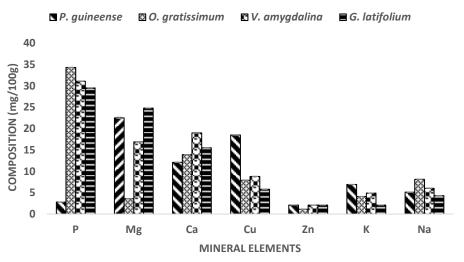
263

The results of the quantitative analysis of mineral elements (Figure 3) indicate that the 244 leaves of the plants contain high levels of Magnesium (Mg)(3.6-24.8 mg/100g), Phosphorus 245 (P)(2.8-34.3) and Calcium (Ca)(12.1-19.0) and copper (Cu)(5.8-18.5), relative to their Zinc 246 (Zn)(1.1-2.1), Potassium (K)(2.1-6.9) and Sodium (Na)(4.3-8.1) contents. A similar report by 247 Asaolu et al. [23] for the three plants O. gratissimum, V. amygdalina and G. latifolium gave 248 the range as Mg (61.08-92.51 mg/100g), P (12.52-29.42), Ca (64.8-72.65), Cu (ND-5.69), Zn 249 (6.85-18.15), K (72.25-99.01) and Na (32.97-84.10) mg/100g. The report by Okafor [25] puts 250 251 the range (converted from parts per million) for three of the plants, P. quineense, G. latifolium 252 and *V. amygdalina* as Mg (5.6-14.7), Ca (0.71-46.0), Cu (0.01-0.015), Zn (0.081-0.205), K (30.4-33.6) and Na (3.5-5.8) mg/100g. In addition to the numerous biological roles these 253 minerals play, they also serve as co-factors in certain biochemical reactions including those 254 involving antioxidant enzymes. Magnesium serves as a co-factor for the enzyme catalase, a 255 primary antioxidant that detoxifies hydrogen peroxide by dismutation to water and oxygen. 256 Similarly Copper and Zinc, are vital co-factor of the different forms of SOD found in plants 257 and animals [38]. Superoxide dismutase (SOD) is a primary antioxidant enzyme that 258 259 catalyses the dismutation or disproportion of superoxide anion radicals ( $O_2$ -) to hydrogen peroxide and molecular oxygen [39]. It is therefore suggested that these minerals contribute 260 261 to the antioxidant properties of the plants probably by boosting the levels of antioxidant enzymes such as SOD and catalase. 262

Except for *P. guineense*, Phosphorus (P) was the major constituents of the mineral elements 264 assayed. O.gratissimum had the highest phosphorus content closely followed by V. 265 amygdalina and G.latifolium in that order. Phosphorus is an ubiquitous mineral in the human 266 body and has diverse functions ranging from the transfer of genetic information to energy 267 utilization [40]. It forms the backbone of DNA and RNA, it is an essential component of 268 phospholipids that form all membrane bilayers and is an integral component of the body's 269 key energy source, adenosine triphosphate (ATP). Phosphorus also plays a vital role in the 270 dissociation of oxygen from hemoglobin, it is the main intracellular buffer and therefore is 271 essential for pH regulation of the human body and is a key component of the second 272 273 messenger molecules such as cyclic adenosine monophosphate (cAMP), cyclic guanine monophosphate (cGMP) and inositol polyphosphates. Taken together with the equally high 274 level of carbohydrates, the four plants are a very good source of energy. 275

- Comparatively, *G.latifolium* had the highest Mg content with *O.gratissimum* having the
   lowest. *G.latifolium* is thus the plant of choice to address Mg deficiency. Mg plays an
   essential role in a wide range of fundamental biological reactions. Apart from its cofactor
   role, it is involved in bone mineralization, the building of proteins, muscle contraction, nerve
   transmission and immune system health [4,41]
- 282 Calcium (Ca) is the most tightly regulated ion in the extracellular fluid (ECF). In higher 283 mammals, the most obvious role of calcium is structural or mechanical being responsible for the mass, hardness, and strength of the bones and teeth [42]. Calcium is also involved in 284 285 cell movement, muscle contraction, nerve transmission, glandular secretion, and even cell division where it acts as both a signal transmitter from the outside of the cell to the inside 286 287 and as an activator or stabilizer of the functional proteins involved. Calcium also plays a role 288 in the regulatory activities of parathyroid hormone [PTH], calcitonin [CT], and a key activity of vitamin D. Ca was more predominant in V. amygdalina followed by G.latifolium, 289 290 O.gratissimum and P.guineense in that order. 291
- Copper is a constituent of many enzymes including superoxide dismutase. It is also required
   for iron metabolism [4,43]. It was more prevalent in *P.guineense*.
- 295 Zinc plays a catalytic, structural, and regulatory role in the body [44]. Zinc is essential for general growth and proper development of the reproductive organs and for normal 296 functioning of the prostate gland. Apart from SOD, Zinc is a co-factor of over 300 enzymes 297 including carbonic anhydrase, which is crucial to maintenance of acid-base balance in the 298 299 blood, and alcohol dehydrogenase that break down alcohol. It is also a component of insulin 300 and plays a role in its processing, storage, secretion and action [45]. The Zinc content of P. guineense may be responsible for the observed stimulated sexual behaviors of mature male 301 rats fed with extract of *P. guineense* [46]. The level of the mineral was pretty much the same 302 in V. amygdalina, G.latifolium and P. guineense (2.1 mg/100g). O.gratissimum had the 303 lowest level of the mineral (1.1 mg/100g). 304
- 305

Sodium (Na) and potassium (K) (and chloride ions Cl<sup>-</sup>) are the major electrolytes located in all body fluids. While sodium is extracellular, pottassium is intracellular. They are responsible for the maintenance of acid/base balance, nerve transmission, muscle contraction and regulation of fluid movement in and out of cells [47]. *P. guineense* had the highest amount of potassium while *O.gratissimum* had the highest level of sodium.



- 311 312
- 313
- Fig 3: Quantitative Analysis of some Minerals in *P.guineense, O.gratissimum, V. amygdalina*
- and *G.latifolium*. Values are expressed as mean  $\pm$  SEM.

## 317 4. Conclusion

318

In summary, the four plants, P. quineense, O. gratissimum, V. amygdalina and G. latifolium, 319 have been shown to be rich in carbohydrates, proteins and fats, vitamins and minerals 320 justifying their use in diets. The plants are particularly rich in vitamins and mineral. Taken 321 together with earlier work on the comparative phytochemical analysis of these plants [7], the 322 findings have good correlation with the therapeutic uses of the various preparations of these 323 324 leafy vegetables in traditional medicine for the treatment and management of diseases that have their etiology and pathophysiology in free radical generation and oxidative stress like 325 diabetes, arthritis, rheumatism, eye problems and infectious diseases such as AIDS. 326 Increased consumption of the leaves of these plants is therefore recommended, especially 327 as they have been shown to contain low levels of anti-nutrients [7], for optimized health and 328 329 wellness.

- 331 **COMPETING INTERESTS**
- 332

334

330

333 The Authors declare that no competing interests exist.

## 335 **References**

- Farnsworth NR, Akerele O, Bingel AS, Soejarto DD, Guo Z. World Health Organ. 1985;
  63: 965.
- Bandaranayake, WM. Quality Control, Screening, Toxicity, and Regulation of Herbal Drugs. In: Ahmad I, Aqil F, and Owais M, editors. Modern Phytomedicine. Turning Medicinal Plants into Drugs. WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim; 2006; pp25-57.
- 343 3. Odugbemi TA. A textbook of Medicinal Plants from Nigeria. Lagos: University of Lagos
   344 Press; 2008.
- Nahak G, Suar M, Sahu RK. Antioxidant Potential and Nutritional values of vegetables:
   A Review. Research Journal of Medicinal Plant. 2014; 8(2): 50-81.
- Foe K. Plant-Based diets and phytonutrients: Potential health benefits and disease
   prevention. Arch Med. 2017; 9(6): 7.
- Godber JS. Nutrient bioavailability in humans and experimental animals. J Food quality.
  1990; 13: 21-36.
- Mgbeje BIA, Umoh EU and Emmanuel-Ikpeme C. Comparative Analysis of Phytochemical Composition of Four Selected Tropical Medicinal Plants Namely: *Ocimum gratissimum, Piper guineense, Gongronema latifolium* and Vernonia *amygdalina*. Journal of Complementary and Alternative Medical Research. 2019: 7(3): 1-11.
- Akinmoladun AC, Ibukun EO, Afor ED, Obuotor EM & Farombi, EO. Phytochemical constituent and antioxidant activity of extract from the leaves of *Ocimum gratissimum*. Science Research Essay. 2007; 2(5): 163-166.
- Lachowicz KJ, Jones GP, Briggs DR, Bienuenu FE, Palmer MV, Tings SST & Hunter
   MO. Characteristics of essential oils from basil (*Ocimum basillicum*) grown in Australia.
   J. Agric. Food Chem. 1996; 144: 877-881.
- Machale KW, Niranjan KU & Pangarkar VG. Recovery of dissolved essential oils form
   condensate waters of basil and *Menthe arvensis* distillation. J. Chem. Tech. Biotech.
   1997; 69: 362-366.
- 365 11. Okwute SK. Plants derived pesticidal and antimicrobial agents for use in Agriculture. A
   366 review of phytochemical and biological studies on some Nigeria plants. J of Agric.
   367 Science and Technology. 1992; 2(1): 62-70.

- Rehm SS & Espig GT. The cultivated plants of the tropics and subtropics cultivation,
   economic value, utilization. Germany: Verlay Josef Margraf; 1991, 552 pps. ISBN 3 8236-1169-0.
- 13. Ugochuwku NH, Babady NF, Cobourne MN & Gasset SR. The effect of *Gongronema latifolim* extracts on serum lipid profile and oxidative stress in hepatocytes of diabetic
   rats. J Biosci. 2003; 28(1): 1-5.
- Igile GO, Fafunsho M, Fasanmade A, Burda S, Jurzysta M, & Oleszek W. Toxicity of
   *Vernonia amygdalina* leaves, extracts and purified saponins in mice. Proc. Eurp. Food
   Tox. 1994; 2: 394-399.
- 377 15. Okafor JC. Conservation and use of traditional vegetable from woody forest species in
  378 south eastern Nigeria. Fame Agriculture Centre, Enugu, Nigeria; 2005. Pp 55-59.
  379 Available online at <a href="http://www.biodiversityinternational.org">http://www.biodiversityinternational.org</a>.
- 16. Iwu MM. Handbook of African medicinal plants. 1<sup>st</sup> ed. Florida: CRC Press; 1993; pp.
   221-222.
- AOAC. Official Methods of Analysis 14th ed. Association of Official Analytical Chemists,
   Washington DC; 1984.
- 18. FAO. Manuals of food quality control. Food analysis: general techniques, additives,
   contaminants and composition. FAO Food and Nutrition Paper. 1986; 14(7): 203-232.
- James CS. Analytical Chemistry of Food. New York: Chapman Hall; 1995. Available at <a href="http://dx.doi.org/10.1007/978-1-4615-2165-5">http://dx.doi.org/10.1007/978-1-4615-2165-5</a>.
- AOAC. Official methods of analysis. 15th ed. Association of Official Analytical Chemists
   Washington D.C; 1990 <u>http://dx.doi.org/10.3923/pjn.2009.1204.1208.</u>
- 21. Pearson, D. Chemical analysis of food. 7th ed. Edinburgh, New York: Churchill
   Livingstone; 1976, Pp. 7-9.
- AOAC. Official Methods of Analysis. 13th ed. Association of Official Analytical Chemists,
   Washington D.C; 1980.
- Asaolu SS, Adefemi OS, Oyakilome IG, Ajibulu KE & Asaolu MF. Proximate and Mineral
   Composition of Nigerian Leafy Vegetables. Journal of Food Research; 2012; 1(3): 214 218.
- Keim NL, Levin RJ, and Havel PJ. Carbohydrates. In: Ross AC, Caballero B, Cousins
   RJ, Tucker KL, Ziegler TR, editors. Modern Nutrition in health and disease. 11th edition.
   Philadelphia: Lippincott Williams & Wilkins; 2014; pp 36-57.
- 400 25. Okafor JC. Conservation and use of traditional vegetable from woody forest species in south eastern Nigeria. In Guarino, L., editor. Traditional African Vegetables. Promoting 401 the conservation and use of underutilized and neglected crops. 16. Proceedings of the 402 IPGRI International Workshop on Genetic Resources of Traditional Vegetables in Africa: 403 Conservation and Use, 29-31 August 1995, ICRAF-HQ, Nairobi, Kenya. Institute of 404 Plant Genetics and Crop Plant Research, Gatersleben/International Plant Genetic 405 Institute, Rome, Italy; 1997. Available online at 406 Resources http://www.biodiversityinternational.org 407
- 408 26. Cahill GF. Starvation in man. N Engl J Med. 1970; 282:668–75.
- 409 27. Ismail BP. Ash content determination. In: Nielsen S, ed. Food Analysis Laboratory
   410 Manual. Food science Text series. Cham: Springer; 2017; Pp 117-119.
- 28. Cao GH, Sofic EE & Prior RL. Antioxidant capacity of tea and common vegetable.
  Journal of Agriculture and Food Chemistry. 1996; 44: 3426-3430.
- 413 29. Grivetti LE & Ogle BM. Value of traditional foods in meeting macro-and micro nutrients
   414 needs: the wild plant connection. Nutrition Research and Reviews. 2000; 13: 31-46.
- 30. Odukoya OA, Inya-Agha SI, Segun FI, Sofidiya MO & Ilori OO. (2007). Antioxidant
  activity of selected Nigeria green leafy vegetables. American Journal of food
  Technology. 2007; 2(3): 169-175.
- 418 31. Aregheore EM. Nutritive Value and Inherent Anti-nutritive Factors in Four Indigenous
  419 Edible Leafy Vegetables in Human Nutrition in Nigeria: A Review. Journal of Food
  420 Resource Science; 2012; 1(1): 1-14.
- 421 32. Hiza H. Availability of Spices on the rise in the U.S. food supply. US Department of
   422 Agriculture, Centre for Nutrition Policy and Promotion. Nutrition insight. 2008; 39.

- 33. Mensah JK, Okoli RI, Ohaju-Obodo JO & Eifediyi KB. Phytochemical, nutritional and medicinal properties of some leafy vegetable consumed by Ede people of Nigeria.
  African Journal of Biotechnology. 2008; 7(14): 2304-2309. Available online at http://www.academicjournals.org/AJB.
- 427 34. Charttejea MN & Shinde RQ. Textbook of medical Biochemistry. 6th Ed. New Delhi:
  428 Jaypee Brothers; 2005; Pp. 124 -132.
- 35. Traber MG & Atkinson JC. Vitamin E, antioxidants and nothing more. Free Ecological
  Biology and Medicine. 2007; 43(1): 4-15.
- 431 36. Jialal IW & Fuller CJ. Effect of vitamin E, Selenium and beta-Carotene on LDL oxidation
  432 and atherosclerosis. Canadian Journal of Cardiolog. 1995; 11: 97G-103G.
- 433 37. Johnson EJ. Aging and carotene nutritive. Age. 1993; 16: 59-66.
- 434 38. Ahmad, S. Oxidative Stress and antioxidant defenses in Biology. New York: Chapman
  435 and hall; 1995; Pp. 1-18, 238-258.
- 39. Zelko IB, Mariani TF & Folz RM. Superoxide dismutase multigene family: a comparison
  of the CU/Zn-SOD (SODI), Mn-SOD, (SOD2) and EC-SOD (SOD3) gene structures,
  evolution and expression. Free Radical Biology and Medicine. 2002; 33(3): 337-349.
- 439 40. O'Brien KO, Kerstetter JE, Insogna KL. Phosphorus. In: Ross AC, Caballero B, Cousins
  440 RJ, Tucker KL, Ziegler TR, editors. Modern Nutrition in health and disease. 11th edition.
  441 Philadelphia: Lippincott Williams & Wilkins; 2014; pp 150-158.
- 41. Rude RK. Magnesium. In: Ross AC, Caballero B, Cousins RJ, Tucker KL, Ziegler TR,
  editors. Modern Nutrition in health and disease. 11th edition. Philadelphia: Lippincott
  Williams & Wilkins; 2014; pp 159-175.
- 42. Weaver CM & Heany RP. Calcium. In: Ross AC, Caballero B, Cousins RJ, Tucker KL,
  Ziegler TR, editors. Modern Nutrition in health and disease. 11th edition. Philadelphia:
  Lippincott Williams & Wilkins; 2014; pp 133-149.
- 43. Collins JF. Copper. In: Ross AC, Caballero B, Cousins RJ, Tucker KL, Ziegler TR, editors. Modern Nutrition in health and disease. 11th edition. Philadelphia: Lippincott Williams & Wilkins; 2014; pp 206-216.
- 44. King JC and Cousins RJ. Zinc. In: Ross AC, Caballero B, Cousins RJ, Tucker KL,
  Ziegler TR, editors. Modern Nutrition in health and disease. 11th edition. Philadelphia:
  Lippincott Williams & Wilkins; 2014; pp 189-205.
- 454 45. Li YV. Zinc and Insulin in pancreatic beta-cells. Endocrine. 2014; 45(2): 178-189.
- 46. Kamtchouing P, Mbongue GYF, Dimo T, Watcho P, Jatsa HB & Sokeng SD. Effect of *Aframomum melegueta* and *Piper guineense* on sexual behavior of mate rats.
  Behavioural Pharmacology. 2002; 13: 243-247.
- 47. Vasudevan DM and Sreekumari S. Textbook of Biochemistry for Medical Students. 5<sup>th</sup>
   Ed. New Delhi: JP Medical Ltd; 2007; pp. 373-378.
- 460 461