1	Review Article
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4	An Over View of Dermatophytosis in Camels
5	
6	Abstract:
7	Dermatophytosis is a fungal infection of the skin caused by dermatophytes-filamentous fungi
8	which have ability to invade the epidermis and keratinized tissues such as hair, skin or nails.
9	Trichophyton verrucosum is the most common dermatophytes species isolated from camel. The
10	disease is characterized by circumscribed crusty hairless lesion, (1-2 cm) distributed over the
11	head, neck, shoulder, limbs and flanks. Dermatophytosis can be diagnosed by direct
12	examination, fungal culture, skin biopsy and molecular diagnosis methods. This review forecast

13 more light of the different aspects of this disease.

14 Key words:

- 15 Dermatophytosis, Camel, Clinical feature, Diagnosis. Treatment
- 16 17

#### 18 Introduction:

- 19 Camels in their natural habitat are exposed to severe stress conditions which make them
- susceptible to many diseases [1, 2]. In last decades camels were reported to be resistant to many
- disease causing agents [3, 4], now it has been realized that they are susceptible like other

22 livestock or even more, to the common disease causing pathogens [5-7].

23 Dermatophytes are among the most frequent causes of superficial skin infections in man and

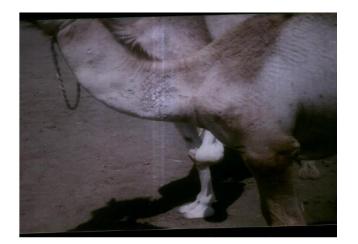
24 animals, known as Dermatophytosis (ringworm). It caused by fungi of three genera

25 *Microsporum, Trichophyton and Epidermophyton.* Ecologically, dermatophytes are classified to

three groups anthropophilic (mostly associated with humans), zoophilic (associated with

- animals) and geophilic (found in the soil). Dermatophytosis in camels is the most frequent
- 28 mycosis worldwide it has public health and economic importance. There are two forms of the
- disease sporadic as well as epidemic form [8-10]. .Ringworm occurs in camels less than 3 year
- 30 age and is characterized by circumscribed crusty hairless lesion, 1-2 cm in diameter distributed

31	over the head, neck, shoulder, limbs and flanks [1]. T. verrucosum is the most common cause of
32	dermatophytosis in camels [11].
33	Epidemiology:
34	Dermatophytosis was reported to be a common disease of camels worldwide [10,12]. Different
35	prevalence rate of the disease of 48 % [13] and 43.5% [14] were reported in camels, while [15]
36	reported lower prevalence of 8.58% in camels suffering from dermatophytosis. Camels less than
37	3 year age were more susceptible to the disease than older animals [1].
38	Predisposing factors:
39	1- Age animals less than three years old always get the infection.
40	2- Breed foreign breed is more susceptible to disease
41	3- Production system for example poor and crowded houses
42	4- Close confinement
43	5- Immunosuppression (including immunosuppressive treatment)[16,17]
44	Transmission
45	
46	The transmission of dermatophytosis is usually occurs by direct contact with infected host
47	(animals or humans) or asymptomatic carriers indirect contact with contaminated fomites besides
48	contact with soil [18,19].
49 50 51 52	Clinical features:
53	Ringworm in camels is characterized by circumscribed crusty hairless lesion, 1-2 cm in diameter
54	[1] on the head, the neck and shoulders with a possible extension to the flanks and legs, leading
55	sometimes to emaciation [20] (fig.1-8).
56	



58 Fig.1: Localized lesions of ringworm on camel neck [11]



60 Fig.2: Circular lesions on the neck of camel [56]



63 Fig.3: Alopecic ringworm lesions [57]



- 65
- 66 Fig.4: A young camel calf has Crusty and hairless lesions on the shoulder.[44]



- 67
- 68 Fig.5: Acamel from Elobied areas showing generalized lesions of ringworm
- 69 giving moth-eaten appearance of wool.[11]



- 70
- 71 Fig.6: Affected camel showed white hairless patches on different parts of the body. The
- 72 lesions typically consisted of an area of alopecia.[44]



- 73
- 74 Fig.7: Epidermophyton fast spreading lesions with circular patches[29]



### 76 Fig.8: Epidermophyton lesions giving just burning appearance [29]

- 77
- 78

#### 79 Mixed infection of dermatophytosis and other skin diseases:

- 80 Mixed infection of dermatophytosis and *Sarcoptic scabiei* has been reported by [21, 22]. Mixed
- 81 infection of dermatophytosis caused by *M.gpyseum* and *Dermatophilus congolensis* in dairy farm
- camel was recorded by [15] fig. 9.
- 83
- 84



- 85
- Fig.9: Hair matting and crusty, hairless lésions on the flanks of a camel calf
- 87

# 88 Etiology:

- 89 The disease in camel is mainly caused by *Trichophyton vertucosum* [12, 14, 23, 24, 25]. *T*.
- 90 mentagrophytes has been isolated by [11]. [26-28] were able to isolate T. schoenleinii. T.
- 91 *dankaliense* was isolated by [4]. [29] has been isolated *T. equinum T. concentricum*, *T.*
- 92 tonsurans, T. violaceum, T. soudanense, T. rubrum, M. canis, M. nanum and M. ferrugineum.
- 93 *M.gpseum* has been isolated by [15, 30, 31] *Epidermophyton floccosum* has been reported by
- 94 [32]
- 95
- 96

#### 97 Diagnosis:

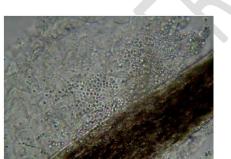
- 98 Dermatophytosis diagnosis is based on the clinical signs however in order to confirm the
- 99 diagnosis culturing and direct microscopic examination of skin scrapings from the periphery of
- the lesions should be indicated [33].

#### 101 **Collection of samples:**

- 102 Skin scraping samples from the cattle that were suspected to be infected with dermatophytes will
- 103 be collected on the basis of gross lesion on their body. After cleaning with ethyl alcohol
- 104 70%.hair and scrapings samples should be collected with forceps or scalpel just behind the
- 105 extending margin in the infected area. Samples can be kept in polyethylene bags [25].
- 106

#### 107 Direct examination:

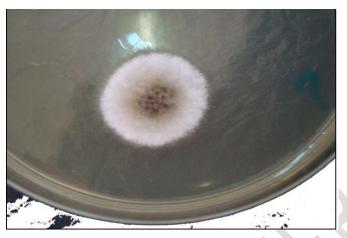
- 108 Each Sample from infected camel should be divided into two portions, one portion for direct
- 109 microscopic examination and the other for culture. Fungal hyphae and/or ectothrix spores are
- 110 determined to be seen in the direct examination when they appear to make hairs or hair
- 111 fragments thicker and rough with irregular surface.
- 112 Potassium Hydroxide (KOH) 10 or 20% is used as a clearing agent because it has keratinolytic
- activity [34-36]. Infected hairs appear pale, wide and filamentous compared with normal hairs
- 114 when examined at x4 or x10 magnification, appearing. Arthrospores can be visible on high
- 115 magnification (x40).(fig.10)
- 116
- 117



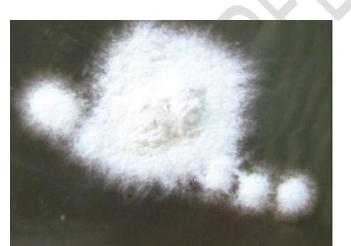
- 118
- Fig.10: KOH preparation showing hair surrounded with chain of large ectothrix spores
   X400[58]
- 121

- 122 Fungal culture:
- 124 Fungal culture is considered the 'gold standard' for diagnosis [37]. Sabouraud's dextrose agar
- 125 (SDA) containing cycloheximide, penicillin and streptomycin were used in most diagnostic
- 126 laboratories. Plates should be incubated at 25°C for 5 weeks. Dermatophytes test media (DTM) is

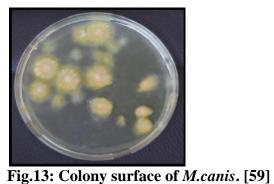
- 127 recommended as the best media for isolation of dermatophytes because the presence of the red
- 128 color indicated positive result, this can help in early identification of highly suspected cultures
- [38]. The isolates should be examined macroscopically and microscopically after staining with
- 130 lactophenol cotton blue using wet mount technique [39].(fig.11-20)
- 131 In addition to technique steps mentioned above, pigment production on corn meal agar, urease
- activity on urea agar base, growth at 37°C on SDA.(fig.21)
- 133



- 135 Fig.11: Colony of *T. verrucosum* on the modified SDA. [55]
- 136



- 137
- 138 Fig.12: Colony *T. mentagrophytes*: surface of colony show powder-like shape, white, loose
- 139 irregular mycelium on the edge. [58]
- 140
- 141
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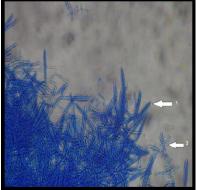


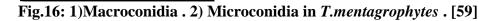
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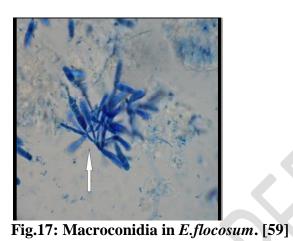
Fig.14: Colony of Microsporum gypseum. [60]

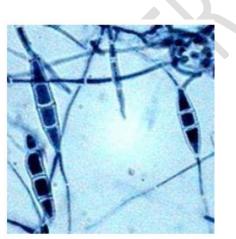


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151 Fig.15: Colony surface of *E.flocosum*[59]





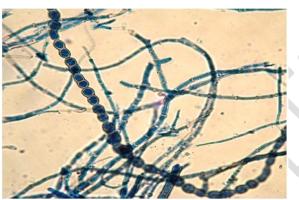




4 Fig.18: *Microsporum canis* microscopic observation in lactophenol cotton blue [61]



- Fig.19: Microsporum gypseum microscopic observation in lactophenol cotton blue [60]



- Fig.20: Microscopic appearance of *T. verrucosum*.[52]

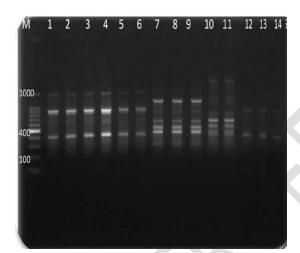


- Fig.21: Growth of *T.mentogrophytes* on urea agar after 4 days showing hydrolysis of the
- **urea.[62]**

#### 177 Molecular diagnosis:

- 178
- 179 Diagnosis with conventional methods is time-consuming because it might take up to 4 weeks or
- 180 longer to give the final results [40]. Furthermore, morphological identification may be confusing
- due to polymorphism of dermatophytes [41]. During the last decade, a wide variety of molecular
- techniques has become available as possible alternatives for routine identification of fungi in
- 183 clinical microbiology laboratories [42, 43].
- 184 Molecular identification for *Trichophyton* species isolated from camel skin lesions was done
- using (GACA) 4 all the strains were amplified simply resulting PCR bands ranged from 2-5.
- 186 Three profiles of *Trichophyton mentagrophytes* have been detected so *T.mentagrophytes* is
- 187 known to be a species complex [44].(fig.22)





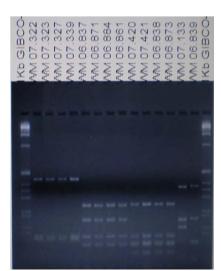
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- 190 Fig.22: Agarose gel electrophoresis for PCR using (GACA)4. M, molecular weight marker.
- 191 Lanes (1-6) first profile of *T.mentagrophytes* Lanes (7-9) second profile of *T.mentagrophytes*.
- 192 Lanes (10&11) third profile of *T. mentagrophytes* Lanes (12-14) *T.verrucosum*. [44]

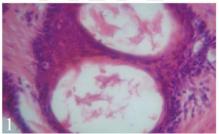
193 *Trichophyton* species isolated from camel and human were identified using restriction fragment

194 length polymorphism (RFLP), *Mva*1 was used as restriction enzyme. Five different patterns

- 195 of two to four bands were obtained. None of these different species gave the same profile pattern
- 196 [45]. (fig.23)

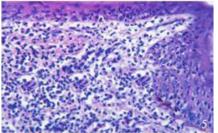


- 198 Fig.23: RFLP profiles of *Trichophyton* spp isolates. WM 07.322, WM 07.323, WM 07.327,
- 199 WM 07.339 were T.mentagrophytes var mentagrophytes, WM06.837, WM 06.871, WM
- 200 06.884, WM 06.861 were T.mentagrophytes var interdigitale, WM 07.420, WM07.421, WM
- 201 06.838, WM 06.873 were T.tonsurans, WM 07.133 T.mentagrophytes var erinacei and WM
- 202 **06.839** was *T.rubrum*.[45]
- 203
- 204 Skin biopsy:
- 205 Specimens from infected skin should be taken and fixed in 10% formaline solution then
- 206 dehydrated, cleared and embedded in paraffin wax, sectioned at 4 µm thickness should be stained
- by haematoxylin and eosin for microscopical examination [46]. haematoxylin and eosin staining
- 208 (H&E) may or may not identify dermatophytes and special stains such as periodic acid Schiff
- 209 (PAS) and Grocott methenamine silver (GMS) are needed.
- 210 Microscopically, the hair follicles and sweat glands exhibited cystic dilatation (fig. 24) and were
- 211 lined by atrophied epithelium.



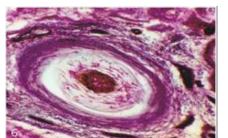
- Fig.24: Ring worm in camels, noticed destructed hair shaft, perifollculitis and cystic
- dilatation of the hair follicles (H&E X650). [46]

- Occasionally, perivascular dermatitis, and intra-epidermal pustules characterized by focal
- aggregation of neutrophils mixed with eosinophil and karyrrhectic debris were reported (fig. 25).



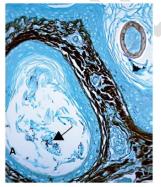
- Fig.25: Ring worm in camels, noticed intra epidermal pustules and acanthosis (H&E X400). [46]

- The branched fungal hyphae were seen when sections stained with PAS (fig.26)

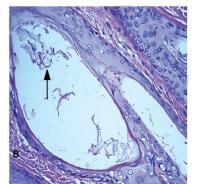


#### Fig.26: Ring worm in camels, noticed tricophyton hyphae positive for PAS-reaction (H&E X400). [46]

- The surface as well as intrafollicular hairs was colonised by large numbers of refractile or
- slightly basophilic arthrospores and hyphae. These were coloured bright magenta with periodic
- acid-Schiff stain and black with Gomori's methenamine silver stain. The Keratin-filled follicles
- ruptured leading to prominent furunculosis [22]. (fig.27,28)



- Fig.27: Refractile arthrospores and hyphae appear: bright magenta with Periodic Acid-
- Schiff stain [22].



# Fig.28: Refractile arthrospores and hyphae appear black with Gomori's Methenamine Silver stain [22].

239

240 **Treatment:** 

- 241 Optimal therapy of dermatophytosis requires a combination of topical antifungal therapy,
- 242 concurrent systemic antifungal therapy and environmental decontamination. The treatment
- should be continued until two consecutive negative cultures (at weekly or bi-weekly intervals)
- are obtained [47]. Topical treatments speed resolution of clinical lesions and may help prevent
- 245 zoonotic contagion. Systemic therapies that have prolonged residual activity in the skin and hair
- 246 provide the most effective treatments.

# 247 **Topical Therapy:**

- 248 1- 2% solution of tincture iodine [23].
- 249 2- 10% iodine ointment daily for three weeks [48].
- 250 3- Enilconazole Wash or spray with diluted emulsion (2000 ppm) four times at 3–4-day
  251 intervals [49].
- 252
- 253

255

256

# 254 Systemic Therapy:

1- Griseofulvin10 mg/kg body weight for 7 days in mild infections; in severe cases 2–3 weeks [49].

# 257 Environmental decontamination:

- Dermatophytes can remain viable in infected soil for many years [50-52], so 10% hypochlorite solution can be used as disinfectant [53].
- 260
- 261
- \_\_\_\_
- 262

263	Vaccination:
264	Live attenuated vaccine is used for prophylaxis and therapy for dermatophytosis caused by
265	T.verrucosum and T.mentagrophytes every five years [54].
266 267	Conclusion:
268	Dermatophytoses are the most common fungal infections in camels. Many studies were done
269	considering different aspects of the disease (eg. epidemiology, clinical presentation and
270	diagnosis, treatment, prevention, and control).Infected camel with dermatphytes can be a source
271	of infection to human this can lead to public health problem.
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