

1 **BIOREMEDIATION OF INDUSTRIAL EFFLUENT USING CYANOBACTERIAL**

2 **SPECIES: *PHORMIDIUM MUCICOLA* AND *ANABAENA AEQUALIS***

3 Sanjay Kumar Dubey 1^{*}, Preeti Vyas 2², Vaishali Gupta³ and Jaishree Dubey⁴

4 (Lab of Phycology, Department of Botany,

5 Dr. Hari Singh Gour University, Sagar, India - 470003

6 **E-mail: dubeysanjay83@gmail.com)**

7
8
9
10 **ABSTRACT**

11

Industries discharge effluent into different water body subjected to severe levels of pollution that can cope with the high pollution load in the water. Textile and Pharmaceutical industry, Mandideep, Bhopal has discharge industrial effluent into river. The main objective of the present study was to investigate the biodegradation and biosorption capacity of some potential cyanobacterial species; *Phormidium mucicola* and *Anabaena aequalis* dominating the river ecosystem. Heavy metals contaminants polluting the Industrial effluents. The effluents were subjected to biological treatment using axenic cyanobacterial strains as batch system for 7 days. Removal efficiencies of the different contaminants were evaluated and compared. Results confirmed the high efficiencies of the investigated species for the removal of the target contaminants which were species and contaminant-dependent. BOD and COD recorded 91.18 and 82.54% as maximum Removal efficiencies achieved by *Anabaena aequalis* . The highest Removal efficiencies of the Total suspended solids recorded 53.23% achieved by *Phormidium mucicola*, while 41.61% was recorded as the highest TDS. Removal efficiencies achieved by *Phormidium mucicola*. Concerning the contaminant metals, *Phormidium mucicola* showed the highest biosorption capacity where 86.12 and 94.63% Removal efficiencies were achieved for Zn and Cu, respectively. In conclusion, results of the study confirmed the advantageous potential of using the tested cyanobacterial species for the bioremediation of industrial effluent and clearly showed the quality improvement of the discharged effluent which in turn will eliminate or at least minimize the expected deterioration of the receiving environment.

12 **Keywords:** algae, bioremediation, cyanobacterial species, heavy metals, industrial effluent

* Tel. +919981798209.

E-mail address: dubeysanjay83@gmail.com.

13 **1. INTRODUCTION**

14 Textile and Pharmaceutical industrial effluents discharge directly into river or other water
15 source like, close Water Lake in Bhopal city. Beside nutrients, the river water and sediments
16 showed terrible levels of organic matter, and heavy metals in worldwide. This is mainly due
17 to continuous discharge of huge quantities of the effluents lead to deterioration in the water
18 quality of this river (El-Bestawy E., 1993; El-Bestawy E.et al., 2007; Mansy And El-Bestawy,
19 2002). Such pollutant with time and the shift of bacterial and algal populations toward more
20 resistant species such as the planktonic cyanobacteria that dominate the river water
21 especially in the warm seasons. These species characterized by great ability to tolerate such
22 high levels of pollution and proved high efficiency for degrading highly organic contaminants
23 and accumulating heavy metals (El-Bestawy E et al.,2007; Podda et al.,2000 and Palmer,
24 C.M. 1980). Therefore, they could be efficiently used in advanced technologies for
25 bioremediation of the industrial effluents.

26 Cyanobacteria are gram-negative photosynthetic Some lakes are naturally eutrophic, but in
27 many other prokaryotes. They can be found in a wide range of water bodies the excess
28 nutrient input is of anthropogenic habitats from ice fields to hot springs and deserts. Origin,
29 resulting from municipal wastewater discharge or Morphologically, physiologically and
30 metabolically, this runoff from agricultural land. Cyanobacteria have a group is one of the
31 most diverse groups of prokaryotes number of special properties that determine their (Abd
32 Allah LS 2006). The rapid evolution of cyanobacteria in different water importance, relative
33 success and predominance during the land environments is related to their capacity for both
34 growth season in phytoplankton communities. However, aerobic and anaerobic
35 photosynthesis. cyanobacteria are located in thylakoids lying free in the summer period:
36 water temperature above 25°C, low light cytoplasm n ear the cell periphery intensity in water,
37 low N: P ratio and stability of the water.

38 Any change in pH of water bodies as a result of influx of effluent; can cause serious change
39 in water chemistry, which can affect resources especially around the coastal areas. These
40 effects on water bodies can be very significant. Traditional method for the clean up of
41 pollutants usually involve, the removal of unwanted materials through sedimentation and
42 filtration, and subsequent chemical treatment such as flocculation, neutralization and electro-
43 dialysis before disposal.

44 Many species of cyanobacteria possess gas nitrogenase, they convert N directly into
45 ammonium in vesicles, which enable regulation of the buoyancy aerobic conditions.
46 Recently, there has been increasing interest about using cyanobacteria as pollution control
47 agents since they possess many advantages over other microorganisms isolated from soil.
48 Their photoautotrophic nature and the ability of some species to fix atmospheric nitrogen

49 enable them to be producers, as opposed to consumers, and make their growth and
50 maintenance inexpensive [Castenholz et al., 1989; Somashekar, R.K. and Ramaswamy,
51 S.N.1983). Metabolic activities are not affected by the decrease in the levels of the
52 biodegradable pollutants that they may break down. Cyanobacteria have been used
53 efficiently as a low-cost method for remediating all industrial effluents as well as
54 transformation and removal of heavy metals (Lefebvre et al., Budd K 2007; Podda et
55 al.,2000). Remediation capabilities of cyanobacteria toward environmental pollutants can be
56 improved and enhanced through genetic engineering technologies (Kuritz and Wolk 1995;
57 Mansy and El-Bestawy E. 2002 and Palmer, C.M. 1980). However, the beneficial application
58 of cyanobacteria in remediation of contaminated waters and industrial effluents is still not
59 optimally manipulated (Jeganathan, 2006 and Kannan, 2006). The main objective of the
60 present study was to investigate the remediation capacity of some potential cyanobacterial
61 species isolated from Textile and Pharmaceutical industrial effluent (Gohl and Vilensky 1987;
62 James et al., 1979; Stewart et al., 1970; Tien and Kirk 1984).

63 **2. MATERIAL AND METHODS**

64 Survey of different sites of industrial effluent for identification of different algal forms from
65 taxonomic point of view will be undertaken. Collecting sample (effluent and cyanobacteria)
66 from two industrial effluents such as Textile and pharmaceuticals industries, Mandideep,
67 Bhopal, India. Effluent samples and cyanobacteria were collected in large sterilized
68 containers and polythene bags respectively. Thus, it is expected that the effluents contain
69 industrial pollutants such as heavy metals which are not likely to be removed by that primary
70 treatment of the industries. Grab samples representing all effluent entering the plant during
71 24 h were collected from both plants to avoid the fluctuation in the flow and the strength of
72 the effluent.

73 Physico-chemical characteristics of waste waters were carried out by standard methods
74 (APHA, 1995).Such as biochemical oxygen demand (BOD); chemical oxygen demand
75 (COD); total suspended solids (TSS); total dissolved solids (TDS); and two heavy metals
76 (Zn, Cu) were characterized before and after treatment to determine the effectiveness of the
77 remediation process(5,6). All the investigated parameters were determined using the
78 standard techniques described by (Celesseri et al., 1999) in the standard methods for the
79 examination of water and effluent water.

80 Standard microbiological methods were followed for the isolation of cyanobacteria. Algal
81 samples were microscopically examined and the selected cyanobacterial species were
82 grown in Chu No.10 (1942) medium were used as culture medium. They have the following
83 composition (macronutrients).Ca(NO₃)₂ ,0.04 ; K₂HPO₄ ,0.01- 0.005 ;MgSO₄. 7H₂O, 0.025;
84 Na₂CO₃ , 0.02; NaSiO₃ , 0.025; EDTA, 0.008 and Solution B(micronutrients) contained (in

85 g/l): Na₂EDTA, 4.36; FeCl₃·6H₂O, 3.15; CuSO₄·5H₂O, 0.01; ZnSO₄·7H₂O, 0.022;
86 CoCl₂·6H₂O, 0.01; MnCl₂·4H₂O, 0.18; NaMoO₃·2H₂O, 0.006 and the pH was adjusted to
87 7.2 with HCl. Each component of solution A was separately prepared as stock solution while
88 all the components of (solution B) were prepared as a mixture. (Solution A) component were
89 sterilized by autoclaving separately at 121⁰C for 20 min. Micronutrient solution was sterilized
90 by filtration through 0.22 μm polycarbonate membrane to avoid interaction and precipitation
91 of heavy metals. Chu No.10 media was freshly prepared from A and B where 1.0 ml of each
92 component of solution A and 1.0 ml of Solution B were combined and diluted to 1.0 l,
93 sterilized as mentioned and used for selective culturing of the selected species. After
94 inoculation, all the selected species were incubated at room temperature (28⁰C) and day
95 light with manual shaking every 24 h to avoid adhesion of the algae on the walls of the glass
96 vessels until heavy growth appeared within 3 weeks.

97 Identification was confirmed based upon the keys given by (Geitler, 1932 and Desikachary,
98 1959) for microscopic parameters. The isolated cyanobacteria were identified with the help
99 of classical manuals. Two different cyanobacterial species; *Anabaena aequalis* and
100 *Phormidium mucicola*; were investigated as free-living cells for their ability for organic matter
101 biodegradation and heavy metal removal from the effluent. They were selected based on
102 their dominance and survival in the highly polluted water of Pharmaceutical industries and
103 textile industries where they acquired high resistance and acclimatized to deal with high
104 loads of different contaminants. They were also proven high ability for degradation of the
105 heavy metals. Therefore, the selected species were considered promising candidates for
106 biological treatment of the industrial effluents. They were kindly provided as axenic strains by
107 the algal culture collection at the lab of phycology, where they were identified using the
108 classical methods.

109 **2.1 Axenity and bioassay**

110 Unialgal cultures usually remained contaminated with bacteria and therefore to free them
111 from bacteria is a pre requisite for further studies (Ash and Jenkins, 2006; Anagnostidis, and
112 Komárek, 1985). The cultures were made bacteria free by ultraviolet irradiation (2537Å) for
113 varying periods and inoculated in the medium. selected species were provided as axenic
114 cultures. However, before using these strains in the bioremediation of the contaminated
115 industries effluents, their axenity was checked using agar phototactic response method.
116 Semi solid standard agar medium was prepared and aliquoted into test tubes and sterilized.
117 Each tube was inoculated with 100 μl of cyanobacterial culture (two replicates per culture).
118 Light was prevented to reach the top 10 cm of the tube using aluminum foil. All the tubes
119 were incubated in optimal conditions (28⁰C) in an illuminated incubator. Based on the

120 phototactic response phenomena the cyanobacterial filaments were grown toward light
 121 direction through the semi solid agar, but bacteria did not grown. After 7days incubation of
 122 the agar column was dragged out the tube on sterilized Petri dish. The agar column was
 123 sliced into ten slices 1 cm per each. Each slice was stranded longitudinally and transversally
 124 cut under common sterilized conditions to separate the algal filaments surrounded with a
 125 small piece of agar. Each agar piece involving cyanobacterial growth was inoculated into
 126 standard selective liquid medium. After incubation, each inoculated culture was tested for
 127 contamination using general bacterial medium (nutrient agar).In bioremediation bioassay,
 128 the tested species were checked for their axenity, and the liquid cultures were tested by
 129 plating on bacterial nutrient medium and in cubating at 28⁰C for 7days. Only axenic cultures
 130 were involved in the assays. The selected species were inoculated individually in 100 ml
 131 culturing medium (three replicates) and incubated for 2 weeks till heavy growth was
 132 obtained. Effluents water from both industries textile and pharmaceutical was dispensed
 133 (900 ml each) in 18 sterilized conical flasks, nine flasks for each effluent. Each culture (100
 134 ml) was separately seeded at a final volume of 1 l each (three replicates/strain/effluent) and
 135 incubated under the previously mentioned conditions for 7 days. Another six flasks (three
 136 flasks for each industry) were supplied by 1.0 l each of the effluent of both industries without
 137 seeding with cyanobacteria to serve as control for the bioassay. They were incubated under
 138 the same conditions. For the determination of heavy metals and other parameters residues,
 139 samples were collected at 24 h interval. At each sampling time, 130 ml from each flask were
 140 aseptically drawn, where all the investigated parameters were determined and their removal
 141 efficiencies using the selected species were calculated.

142 3. RESULTS AND DISCUSSION

143 Table 1 Residue concentrations (RC) of the quality parameters from the contaminated
 144 indusrial effluents using the selected cyanobacteria at different exposure time

145

146 Textile industry effluent (control)

147 Time(day)	BOD	COD	TSS	TDS	ZN	CU
148 Raw water	140	360	167	1,150	0.11	0.04
149 2	89	289	132	887	0.09	0.01
150 3	80	243	100	766	0.05	0.02
151 4	81	200	101	756	0.12	0.04
152 5	70	187	98	611	0.09	0.05
153 6	120	387	154	900	0.14	0.06
154 7	143	331	143	998	0.12	0.07

155 Pharmaceutical industry effluent (control)

156	Time (days)	BOD	COD	TSS	TDS	ZN	CU
157	Raw water	198	445	387	1430	0.01	0.03
158	2	187	465	411	1457	0.04	0.01
159	3	199	412	376	1432	0.03	0.04
160	4	176	398	311	1200	0.01	0.04
161	5	166	345	298	1100	0.03	0.06
162	6	178	411	321	1289	0.05	0.02
163	7	200	378	365	1008	0.02	0.01

164

165

166 Textile industries effluent

167	Anabaena aequalis							Phormidium mucicola						
168	Days	BOD	COD	TSS	TDS	ZN	CU	BOD	COD	TSS	TDS	ZN	CU	
169	Raw water	149	312	200	1012	0.11	0.05	148	324	234	1134	0.12	0.07	
170	2	141	301	199	1011	0.09	0.05	143	321	223	1124	0.10	0.07	
171	3	132	298	189	1001	0.06	0.03	132	309	212	1120	0.5	0.05	
172	4	121	288	170	998	0.05	0.02	122	289	206	1103	0.5	0.03	
173	5	100	198	150	988	0.03	0.02	101	281	189	1087	0.3	0.02	
174	6	76	122	132	986	0.03	0.02	98	256	167	1079	0.1	0.01	
175	7	62	98	112	983	0.02	0.01	78	249	156	1012	0.1	0.01	

176

177

178 Pharmaceutical industries effluent

179	Anabaena aequalis							Phormidium mucicola						
180	Days	BOD	COD	TSS	TDS	ZN	CU	BOD	COD	TSS	TDS	ZN	CU	
181	Raw water	254	432	321	1349	0.08	0.07	231	421	401	1401	0.06	0.07	
182	2	233	428	311	1323	0.06	0.05	211	421	387	1400	0.04	0.06	
183	3	212	412	306	1321	0.06	0.04	201	401	381	1387	0.04	0.04	
184	4	198	398	285	1309	0.04	0.04	192	356	372	1345	0.02	0.03	
185	5	167	378	265	1287	0.03	0.02	187	241	324	1302	0.01	0.03	
186	6	121	321	209	1270	0.02	0.01	123	209	283	1265	0.01	0.01	
187	7	98	243	187	1201	0.02	0.00	98	198	223	1230	0.00	0.01	

188

189

190

191 **3.1 Industrial effluent characteristics**

192 Effluent produced by the two industries was characterized (Table1, control).BOD, COD,
193 TSS, TDS, Zn and Cu recorded averages of 140, 360, 167, 1150, 0.11and 0.04 mg/l,
194 respectively, in the effluent of the textile industry. Significantly higher levels for almost all the
195 tested parameters were detected in the pharmaceutical effluent where 198, 445, 387, 1430,
196 0.01, and 0.03 mg/l were recorded as average. However, Zn recorded much lower average
197 in the pharmaceutical effluent (0.01mg/l) compared to that of the textile effluent (0.11 mg/l)
198 while no significant differences were recorded in the Cu levels among the two industry (0.04
199 and 0.03 mg/l in the textile and pharmaceutical effluents).Nitrogen and phosphorus content
200 in both effluents (El-Bestawy E et al., 2005 and Ellis 1977) along with the toxic industrial
201 contaminants suppressed the growth of cyanobacteria or any other algae.

202 **3.2 Treatability and removal efficiency of effluent**

203 **3.2.1 Contaminants**

204 Residue levels of the selected quality parameters were determined (Table 1) and the
205 removal efficiencies (RE %) as results of the biological treatment using the selected species
206 were calculated. As a general trend, the two tested species exhibited positive correlation
207 between their RE% of the all the tested parameters and the exposure time up to the last
208 exposure day for both types of effluents.

209 **3.2.2 Organic matter removal**

210 Biochemical oxygen demand Removal of BOD from industrial effluents of both industries
211 using the selected algae revealed the following points:

212 1. High REs% were obtained for BOD removal from industrial effluent by the selected
213 species with *Anabaena aequalis* (90.65%) and finally *Phormidium mucicola* (81.9%).

214 2. Despite the RE variations of BOD achieved by the tested species, RC(s) of the BOD in
215 the industrial effluent reached acceptable limits (19, 20 and 32 mg/l by *Anabaena aequalis*
216 and *Phormidium mucicola*, respectively) after 7 exposure of days which is much lower than
217 the maximum permissible limit (MPL) of 60 mg/l stated by the Environmental Laws for safe
218 discharge into surface water courses. When these figures compared with those obtained by
219 the control it was showed that the natural microbial population of the effluent achieved a
220 maximum removal of 50% after 5 days equivalent to 60.7 mg/l ([MPL of the BOD) after which
221 there was a sharp decline in the efficiency associated by increasing the RC reaching 140
222 mg/l and 3.20% RE after 7 exposure of days.

223 3. Comparing BOD removal by the selected cyanobacteria from the two plants revealed very
224 high efficiency for all of them in the degradation of biodegradable organic matter which is
225 stimulated by increasing the levels of the pollutant in the wastewater.

226

227 Chemical oxygen demand Removal of COD from the industrial effluents using the selected
228 species revealed the following points:

229 1. *Anabaena aequalis* considered the most effective for removing COD from the industrial
230 effluent achieving the maximum RE of 83.68% compared to the RE achieved by *Phormidium*
231 *mucicola* (45.00%) after 7 exposure days. However, *Phormidium mucicola* exhibited higher
232 COD RE% within the first 24 h compared to *Anabaena aequalis*.

233 2. The lowest residue concentration of 100 mg/l was achieved by *Anabaena aequalis* which
234 is the maximum acceptable limit stated by the law (MPL for COD = 100 mg/l) while
235 *Phormidium mucicola* could not bring the COD levels of the effluent to better quality. They
236 recorded 180 and 246 mg/l, respectively, and required longer exposures. The highest RE%
237 achieved by the control culture recorded 50.11% (170.59 mg/l) after 5 exposure days.

238 3. Similar to BOD removal, the natural microorganisms in the control culture were inhibited
239 by the high Strength of the industrial effluent leading to reduction in the COD removal.

240 4. Although high REs% of the COD was achieved by the selected species, none of them
241 could bring the COD levels in the effluent below the MPL during the investigated exposure
242 time (1 week). This may be attributed to the need for longer time for achieving the proper
243 quality. It could also result from the inhibition in cyanobacterial growth due to the higher COD
244 levels in the pharmaceutical effluent compared to that of the textile effluent.

245 **3.2.3 Solids removal**

246 Total suspended solids (TSS) Removal of TSS from the industrial effluents using the
247 selected species revealed the following points:

248 1. The highest recorded TSS REs% in the effluent recorded 42.0 and 29.12% achieved by
249 *Anabaena aequalis*, and *Phormidium mucicola* (125 and 133 mg/l RC), respectively, after 7
250 days.

251 2. According to the law, 60 mg/l is stated as the MPL of the TSS; therefore none of the
252 tested species reached the required efficiency to bring the TSS in the effluents below the
253 MPL during the tested exposure time. This indicates that they required longer time, heavier
254 biomass or different application using the same species to achieve that quality.

255 3. In contrast to cyanobacteria, the indigenous bacteria of the control culture achieved higher
256 TSS removal form the textile effluent compared to that of the pharmaceutical effluent.

257

258 Total dissolved solids (TDS) Removal of TDS from the industrial effluent using the selected
259 species revealed the following points:

260 1. The maximum TDS REs% obtained for the effluent by the tested species ranged between
261 a maximum of 16.66% (1,078 mg/l) achieved by *Phormidium mucicola* and a minimum of
262 12.00% (1,119 mg/l) obtained by *Anabaena aequalis* after 7 exposure days. *Phormidium*

263 *mucicola* exhibited higher TDS RE% at the shorter exposures (up to 2nd day) compared to
264 *Anabaena aequalis*.

265 2. Similar behavior for TSS removal was shown by bacteria of the control culture where
266 higher TDS removal was achieved from the textile effluent compared to that of the
267 pharmaceutical effluent.

268 3. Since the TDS content in the effluents were lower than the MPL of the TDS (2,000 mg/l),
269 the residual concentrations of the TDS produced in the final effluents by all the tested
270 species as well as the two controls improved and still within the safe range for discharging.

271 **3.2.4 Heavy metals removal**

272 Results revealed the following points:

273 1. *Phormidium mucicola* recorded the highest REs% for Zn from EWTP (86.12) and
274 *Anabaena aequalis* (70.88%) recording RCs of 0.0247, and 0.0370 mg/l by the three
275 species, respectively, after 7 days .

276 2. Although low Zn levels were detected in the pharmaceutical effluent, lower Zn REs were
277 achieved compared to those obtained for the textile effluent. Zn removal recorded 78.2, and
278 65.00% achieved as the highest Zn REs% by *Phormidium mucicola*, and *Anabaena*
279 *aequalis*, respectively (0.0123, and 0.0182 mg/l, respectively) after 7 days.

280 3. Although all the average levels of Zn for both effluents were below the MPL of 5 mg/l
281 before the treatment, Zn levels were reduced producing much better effluent quality. Zinc
282 removal was stimulated by increasing its level in the wastewater.

283 4. Concerning Cu , much higher REs% were recorded for effluent compared to those
284 obtained for Zn removal regardless it's high toxicity. This may be attributed to the high
285 resistance of the selected members which was stimulated by increasing Cu levels in the
286 wastewater. 94.63, 90.99 and 90.64% RE of Cu were achieved by *Phormidium mucicola*,
287 and *Anabaena aequalis* (0.0031, and 0.0054 mg/l RC), respectively, after 7 days.

288 **4. CONCLUSION**

289 In conclusion, results confirmed that the most effective species for BOD, COD, TSS, TDS
290 Zn and Cu removal from the effluents of the two industries are in the following order
291 *Phormidium mucicola* and *Anabaena aequalis* which may be attributed to the selective
292 uptake of the investigated pollutants by the tested cyanobacterial species.

293

294 **ACKNOWLEDGEMENTS**

295 The authors are thankful to the Lab of Phycology. Dr. Hari Singh Gour central University,
296 Sagar, India for providing them necessary facilities and support to carry out this work.

297

298

299 **REFERENCES**

- 300 1. Abd Allah L S. Metal-binding ability of cyanobacteria: the responsible genes and optimal
301 applications in bioremediation of polluted water for agricultural use. Ph.D. Thesis,
302 Department of Environmental Studies, Institute of Graduate Studies and research,
303 Alexandria University, Alexandria, Egypt; 2006
- 304 2. Ash N, Jenkins M. Biodiversity and poverty reduction: the importance of biodiversity for
305 ecosystem services. Final report prepared by the United Nations Environment Programme
306 WorldConservation Monitoring Centre (UNEP-WCMC) for the Department for International
307 Development (DFID);2006
- 308 3. Anagnostidis, K. and Komárek, J. Modern approaches to the classification system of
309 cyanophytes. É. Introduction. Arch.Hydrobiol. Suppl.,1985; 71, 91–302.
- 310 4. APHA.Standard Methods for the Examination of Water and Waste Water, 19th edn.,
311 American Public Health Association.Washington D.C.;1995
- 312 5. Boominathan, M. Bioremediationstudies on dairy effluent using cyanobacteria. Ph.D.
313 Thesis. Bharathidasan University.Tiruchirapalli. Tamil- Nadu. India.;2005
- 314 6. Cairns, J. Jr. & Dickson,K.L. Asimple method for the biological assessment of the effects
315 of waste discharge on aquaticbottom dwelling organisms. J. Wat. Poll.Control Fed. 43: 722-
316 725.; 1971
- 317 7. Castenholz RW, Waterbury JM. Oxygenic photosynthetic bacteria. Group I.
318 Cyanobacteria. In: Hensyl WR (ed) Bergy's manual of systematic bacteriology. Williams and
319 Wilkins, Baltimore, 1989; 1710–1727
- 320 8. Celesseri LS, Greenberg CG, Eaton AD. Standard method for the examination of water
321 and wastewater, 20th edn. American Public Health Association (APHA), USA. ISBN
322 0875532357;1999
- 323 9. Desikachary, T. V. Cyanophyta, Indian Council of AgriculturalResearch, New Delhi, p.
324 686.; 1959.
- 325 10. Celesseri LS, Greenberg CG, Eaton AD. Standard methodfor the examination of water
326 and wastewater, 20th edn. AmericanPublic Health Association (APHA), USA. ISBN
327 0875532357; 1999

- 328 11. El-Bestawy E, Hussein H, Baghdadi H, El-Saka M. Comparison between biological and
329 chemical treatment of wastewater containing nitrogen and phosphorus. *J Ind Microbiol*
330 *Biotechnol*, 2005; 32:195–203
- 331 12. Ezeronye OU, Ubalua AO. Studies on the effect of abattoir and industrial effluent on the
332 heavy metal and microbial quality of Abariver in Abia state of Nigeria. *Afr. J. Biotechnol.*
333 2005; 4(3): 266-272
- 334 13. El-Bestawy E. Studies on the occurrence and distribution of pollutant metals in
335 freshwater phytoplankton and bacteria in Lake Mariut, Alexandria, Egypt. Ph.D. Thesis,
336 Department of Environmental Biology, Faculty of Sciences, Manchester University,
337 Manchester; 1993
- 338 14. El-Bestawy E, Abd El-Salam AZ, Abd El-Rahman HM. Potential use of environmental
339 cyanobacterial species in bioremediation of Lindane-contaminated effluents. *Int Biodeterior* ,
340 *Biodegradation*, 2007; 59(3):180–192
- 341 15. Ellis BE. Degradation of phenolic compounds by freshwater algae. *Plant Sci Lett.* 1977;
342 8:213–216
- 343 16. Gohl EPG. & Vilensky, L.D. *Textile Science*. CBS Publishers and Distribution. Delhi –
344 India. 1987; 107-109.
- 345 17. James, Evison, A. & L. (eds.). *Biological indicators of water quality*. John Wiley and Sons.
346 New York.; 1979
- 347 18. Jeganathan, K. *Bioremediation studies on oil refinery industry effluent using Oscillatoria*
348 *earli Gartner*. M.Phil.dissertation. Bharathidasan University.Tiruchirapalli.; 2006
- 349 19. Kannan, S. *Biodiversity of cyanobacteria in freshwater ponds of Poondi.Thanjavur.*
350 M.Phil. dissertation.Bharathidasan University. Tiruchirapalli; 2006
- 351 20. Kuritz T, Wolk C.P. Use of filamentous cyanobacteria for biodegradation of organic
352 pollutants. *Appl Environ Microbiol*, 1995; 61:234–238
- 353 21. Lefebvre DD, Kelly D, Budd K .Biotransformation of Hg(II) by cyanobacteria. *Appl*
354 *Environ Microbiol*, 2007; 73(1): 243–249. doi:10.1128/AEM.01794-06

- 355 22. Mansy AH, El-Bestawy E. Toxicity and biodegradation of fluometuron by selected
356 members of cyanobacteria. World J Microbiol Biotechnol, 2002; 18:125–131
- 357 23. Mansy AH, El-Bestawy E. Toxicity and biodegradation of fluometuron by selected
358 members of cyanobacteria. World J Microbiol Biotechnol, 2002; 18:125–131
- 359 24. Podda F, Zuddas P, Minacci A, Pepi M, Baldi F. Heavy metal coprecipitation with
360 hydrozincite $[Zn_5(CO_3)_2(OH)_6]$ from mine waters caused by photosynthetic microorganisms.
361 Appl Environ Microbiol, 2000 ; 66(11):5092–5098
- 362 25. Palmer, C.M. Algae and Water Pollution. Castelhouse Publications Ltd. USA.; 1980
- 363 26. Podda F, Zuddas P, Minacci A, Pepi M, Baldi F. Heavy metal coprecipitation with
364 hydrozincite $[Zn_5(CO_3)_2(OH)_6]$ from mine waters caused by photosynthetic microorganisms.
365 Appl Environ Microbiol, 2000; 66(11):5092–5098
- 366 27. Somashekar, R.K. & Ramaswamy, S.N. Algal indicators of paper mill wastewater.
367 Phykos. 1983; 22: 161-166
- 368 28. Stewart, W.D.P. & Parsons, M.W. Effect of aerobic and anaerobic conditions on growth
369 and metabolism of bluegreen algae. Proc. Roy. Soc. Lond. 1970; 175: 293-311
- 370 29. Tien M, Kirk TK. Lignin-degrading enzyme from *Phanerochaete chrysosporium*:
371 purification, characterization and catalytic properties of a unique H_2O_2 - requiring oxygenase.
372 Proc. Natl. Acad. Sci. 1984; 81: 2280 – 2284
- 373 30. Geitler, L. Cyanophyceae. In: Rabenhorst's Kryptogamen flora. Akademische
374 verlagsgesellschaft Leipzig. 1932 ; 1196