

**Original Research Article****Phytochemical and Gastrointestinal study on the leaf extract of *Stachytarpheta angustifolia* Mill Vahl (Verbenaceae) in Rabbit****Jejunum****Abstract:**

*S. angustifolia* (Verbenaceae) is mostly prescribed by the folkloric healers for varieties of gastrointestinal disorder. This study was carried out to ascertain the gastrointestinal effect of the ethanol leaf and other various fractions (CHCl<sub>3</sub>, EtOAc, n- BuOH and residual aqueous) on rabbit Jejunum. The ethanol, n-butanol and residual aqueous of the extract exhibited dose concentration at (0.1, 0.2, 0.4 and 0.8mg/ml) dependent contraction of Jejunum which was blocked by atropine suggesting that the observed pharmacological actions was mediated through the muscarinic receptors. In contrast, chloroform and ethylacetate fraction of the leaf extract exhibit dose concentration dependent relaxation of the rabbit jejunum. Intreperitoneal LD<sub>50</sub> of the extract in mice was found to be 295.8mg/kg. Phytochemical screening of the leaf extract revealed the presence of carbohydrates, tannins, saponins, cardiac glycoside, sterols, flavonoids and terpenoids. The result indicated that, the plant extract possesses some pharmacological activity, hence justifying its use traditionally in alleviating gastrointestinal disorder.

**Keywords:** *Stachytarpheta angustifolia*, Phytochemistry, Gastrointestinal activity, Jejunum

**INTRODUCTION**

Despite the immense technological advancement in modern medicine, a lot of the Africans (approximately 80% of the population) still rely on traditional healing practices and medicinal plants for their daily health care needs (Akerle, O. 1991). The floral biodiversity of Africa provides the African traditional medical practitioner with an impressive ‘natural pharmacy’ from which plants are selected as remedies or as ingredients to prepare herbal medicine (phytomedicines) for various human ailments (WHO, 2005). The traditional preparations comprise of medicinal plants, mineral and organic matter, although the Ayurvedic medicine is essentially primitive but are also preventive in therapeutic approach (Sofowoa, 2008).

*Stachytarpheta angustifolia* is a medicinal plant that belongs to the family verbenaceae. It is a shrub of about 4ft high, with a soft and cylindrical bark. They are mostly simple, slightly branch and often succulent. The flowers are mostly pale blue with or without Centre (Dalziel, 2002: Hutchinson, 1963). The plant is commonly known as the Devils coach whip while the Hausa’s called it Wutsiyar kadangare and the yoruba’s called it Iru alangba (Adjanohoun *et al*, 1991, Jinju, 1990). In Brazil the triturated fresh leaf of the plant is applied locally for the

38 treatment of ulcer and also a good remedy against rheumatism. This plant is reported to  
39 contain a glucosidal substance 'stachytarphine' which is reputed to be Abortifacient (Watt and  
40 Breyer Brand Wijk, 1963). The cold infusion of the plant is taken as a remedy against  
41 gonorrhoea and other forms of venerable infectious diseases. It is also taken as a vermifuge or  
42 purging vehicle for other vermifuge. The leaf from the plant is boil and taken as a remedy  
43 against diabetes in the northern part of Nigeria (Dalziel 2002, Jinju 1990). The alcohol  
44 extract of the leaf portion of the plant has been reported to show some antimicrobial activities  
45 against *Mycobacterium tuberculosis*, *Staphylococcus aureus* and *Escherichia coli*, but give a  
46 negative result in ant malaria test (Watt and Breyer Brandwijk, 1963).

47 The effect of this widely used plant on the gastrointestinal smooth muscle is unknown. The  
48 present study was undertaken to evaluate the pharmacological effect of the various extract of  
49 *S. angustifolia* on smooth muscles.

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## 51 MATERIALS AND METHODS

### 52 **Plant Material**

53 The whole plant material *Stachytarpheta angustifolia* (mill) vahl verbenaceae was collected  
54 from a farm land in Basawa village outskirts of Zaria, Kaduna state. The plant was identified  
55 and authenticated at the herbarium Biological sciences Department, Ahmadu Bello  
56 University Zaria, Nigeria. Herbarium sample was made and voucher deposited with (No. DC  
57 90188).

58

### 59 **Animals**

60 Four adult's rabbits weighing 3.0-3.8kg were obtained from the animal house Department of  
61 Pharmacology, Ahmadu Bello University, Zaria. They were given access to standard animal  
62 feed and water *ad libitum*.

### 63 **Drugs.**

64 Acetylcholine was freshly prepared to desired concentrations with distilled water just before used.  
65 The extracts were also freshly prepared using distilled water.

### 66 **Phytochemical Screening**

67 The air-dried powdered material of the leaf (360g) was subjected to exhaustive extraction  
68 with petroleum ether 60°C – 80°C and subsequently with 95% ethanol using cold maceration  
69 techniques. The petroleum ether and ethanol extract were concentrated using rotary evaporator to  
70 afford 25.45g and 47.34g respectively (Richard, 1998).

71 The ethanol extract portion (30g) was suspended in water (500ml) and partitioned  
72 exhaustively with solvent of increasing polarity chloroform, ethyl acetate and n-butanol  
73 respectively. The various partition portions of the extracts were concentrated *in vacuo*

74 (Yaling *et al*, 2003, Shengmin *et al*, 2001). The various partition portion of the extracts were  
75 subjected to phytochemical screening using standard protocols ( Sofowora, 2008, Trease and  
76 Evans 2002).

#### 77 ***Toxicity Studies on S. angustifolia (LD<sub>50</sub>)***

78 A total of 13 mice were used for the experiment. In the first phase, three doses of the extract  
79 were administered to three groups each containing three mice. In the second phase, more  
80 specific doses were administered to group each containing one mouse. The median lethal  
81 dose (LD<sub>50</sub>) value was determined as the geometric mean of the highest non-lethal dose and  
82 the lowest lethal dose of which there is 1/1 and 0/1 survival (Lorke, 1983)

83

#### 84 ***Pharmacological Studies on Isolated Rabbit Jejunum***

85 The method described by Schlemper *et al.*, (1996) and modified by Amos *et al.*, (2000) was  
86 adopted. The four adult rabbits obtained were starved overnight prior to the experiment. The  
87 animals were sacrificed by a blow on their head, exsanguinated and their abdomen cut open.  
88 Segments of their jejunum 3.0cm long were placed separately in 25ml organ baths containing  
89 Tyrode's solution containing 136.8mMNacl, 2.7mMKcl, 1.3mMCacl, 12mMNaHCO<sub>3</sub>,  
90 0.5mMMgcl<sub>2</sub>, 0.14mMNa<sub>2</sub>HPO<sub>4</sub> and 5.5mMglucose well aerated and maintained at 37°C. An  
91 initial tension of 1.0g was applied to the tissue and a 60min period of stabilization was  
92 allowed. During this time, the physiological solution was changed every 15min after which  
93 the effect of acetylcholine at final bath concentration of (6.4x10<sup>-3</sup>M) was evaluated and the  
94 tissue was equilibrated for 60mins before use. Dose response curve for acetylcholine (4.0x10<sup>-3</sup>  
95 -6.4x10<sup>-3</sup>) bath concentrations was obtained. The contractile responses of the spasmogen  
96 were recorded on the kymograph paper by means of a frontal writing lever in Ugo basile  
97 unirecorder 7050(GMBH, German). The tissue was washed three times with physiological  
98 solution and allowed to rest before the addition of the subsequent spasmogen. The direct  
99 effect of different portion of the extracts (4.0x10<sup>-3</sup>-6.4x10<sup>-3</sup>) bath concentrations were  
100 investigated after allowing the tissue to rest for 30 sec. Similarly, the effect of the extract was  
101 investigated on submaximal dose of acetylcholine (Fig.1) so as to study the effect of the  
102 extract on these spasmogen.

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108 **Results**

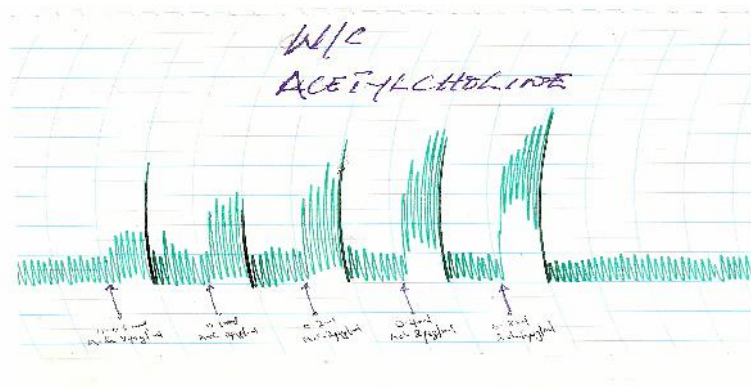
109 Table 1 Phytochemical screening of the Leaves extract of *S. angustifolia*

CONSTITUENTS	TEST	OBSERVATION	PORTIONS OF EXTRACTS					
			Ps	Es	Cl	Eta	n-But	Aq
Carbohydrate			Ps	Es	Cl	Eta	n-But	Aq
General Test	Molisch	Red colouring	-	+	-	-	-	++
Sugar Test	Aniline	Red colour	-	-	-	-	-	+++
Sugar (Monosaccharide)	Barfoed's	Red ppt	-	+	-	-	-	++
Red. Sugar	Fehling's	Red ppt	-	+	-	-	-	++
Tannins	Lead Ethanoate	White ppt	-	++	-	+	++	++
	Iron (III) Chloride	Blue – Black	-	+	-	+	++	+
	Ethanoic acid	White ppt	-	+	-	+	-	-
	Methanol's	Red ppt	-	++	-	-	++	+
Saponins	Frothing	Persist frothing	-	++	-	+	++	-
Sterols	Liberman B.	Blue or green	++	++	-	+	++	+
Saponin Glycoside	Fehlings Solution	Red ppt		++	-	+	++	-
	Tetraoxosulphate(iv) acid	Brick red	-	++	-	+	++	-
Phlobatannins	Hydrochloric Acid	Red ppt	-	++	-	-	+	-
Carotenoids	Carr price's	Blue to red colour	-	++	-	-	-	++
Emodol	Borntrager's	Red colour	-	-	-	-	-	++
Flavones aglycones	Shibata's	Red to Orange	-	-	-	-	-	-
Terpenoids	Liebermann B.	Pink to Red colour	++	++	+	+	+	-
	Dragendoff's	Orange red ppt	-	-	-	-	-	-
Alkaloids	Mayer's	Buff ppt	-	-	-	-	-	-
	Wagner's	Dark brown ppt	-	-	-	-	-	-
Flavonoids	Shinoda	Dee red	-	-	-	-	-	-
	Tetraoxosulphate (vi) acid	Deep Yellow	-	-	-	-	-	-
Cardiac glycoside	Legal's	Deep red colour	-	++	+	++	++	+
	Kedd's	Violet colour	-	+	+	+	+	-
	Keller – kilanis	Reddish brown	-	++	+	++	++	+
	Baljet	Orange to Deep red	-	+	+	+	++	-
	Lieberman	Bluish green	-	++	+	+	++	-

110 **Key: - = Absent, + = Fairly present, ++ = Moderately present and +++ = Highly present**

111 Ps=pet-ether, Es=Ethanolic, Cl=Chloroform, Eta=Ethylacetate,n-But=n-Butanol, Aq=Aqueous

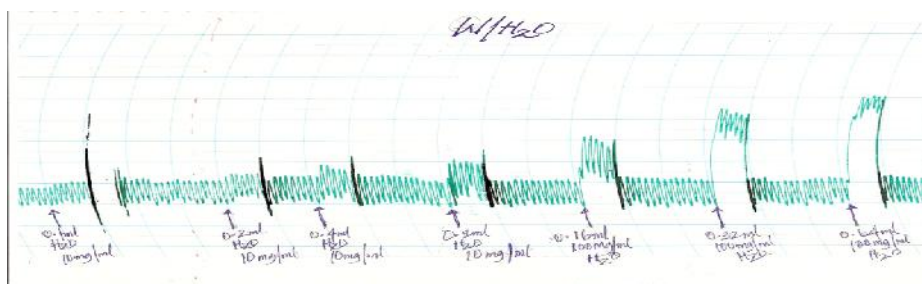
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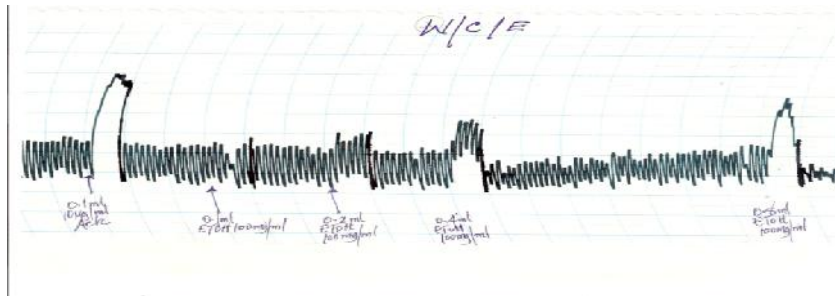
118 Fig: 1 Effect of contraction produce by Acetylcholine on Isolated rabbit jejunum.

119 Fig: 2 Effect of contraction produced by the Aqueous whole plant extract on isolated rabbit jejunum

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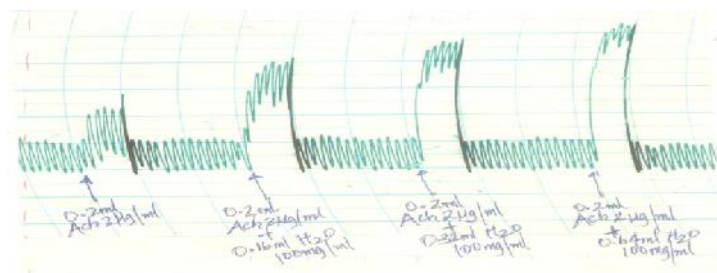


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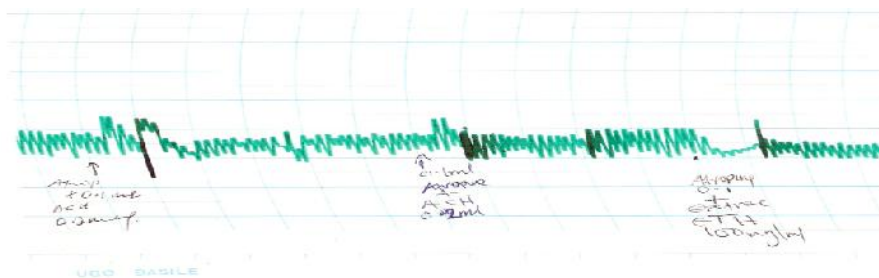


122 Fig: 3 Effect of contraction produced by ethanol whole plant extract on the Isolated rabbit jejunum.

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124 Fig.4 Effect of contraction produced by Aqueous whole plant extract pre contracted with  
125 Acetylcholine on rabbit jejunum



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127 Fig:5 Effect of contraction produced by Atropine on tissues pre –contracted with Aqueous portion of  
128 the extract.

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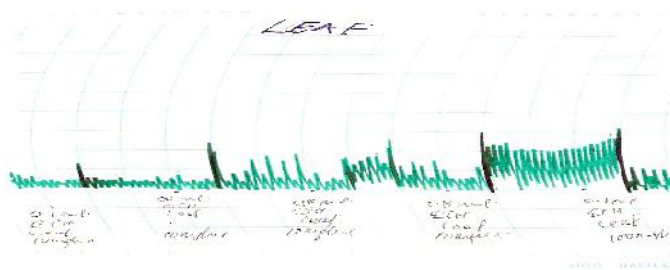
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135 Fig: 6 Effect of contraction produced by the Leaf extract on isolated rabbit jejunum.

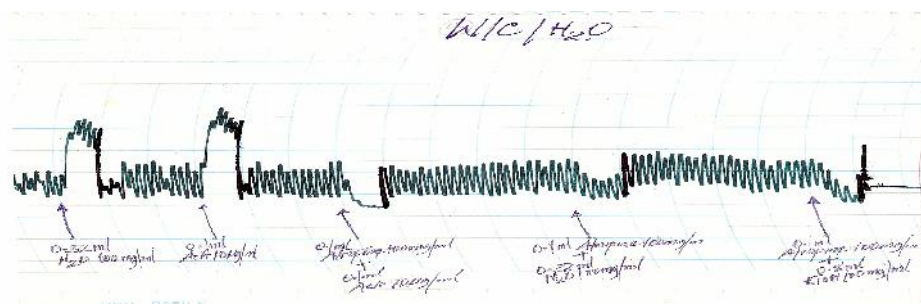
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141 Fig: 7: Effect of Atropine on tissue Pre-contracted with Aqueous Leaf extract on isolated rabbit  
142 jejunum

143 **DISCUSSION**

144 The result of phytochemical screening reveals the presence of terpenoids, steroids, saponins,  
145 tannins, cardiac glycoside, flavonoid and carbohydrate (Table 1.). The standard solution of  
146 acetylcholine at various concentrations produces contraction dependent on rabbit jejunum  
147 (Fig:1). The result on (Fig: 2 and 3) shows the aqueous and ethanol portion of the whole  
148 plant extracts inducing concentration contraction dependent of the rabbit jejunum. The  
149 aqueous portion of the extract pre contracted with acetylcholine on rabbit jejunum in (Fig: 4)  
150 was observed to potentiate the contraction of rabbit jejunum. In (Fig: 5 ) of the result above  
151 shows the blocking effect of the contraction, this is as a result of Atropine pre contracted with  
152 ethanol portion of the extract on the rabbit jejunum. Fig: 6 show the induced dose dependent  
153 contraction of the rabbit jejunum exhibited by the ethanolic portions of the leaf. The  
154 contractions observed by the extracts on the tissues were similar to those produced by

155 Acetylcholine ( Amos *et al.*, 2003; Mitchelson F.J 1984). The leaf extract portion pre-  
 156 contracted with Atropine was also found to block the response of the spasmogen contraction  
 157 as in (fig. 7). Acetylcholine induced contraction of the smooth muscle results from the  
 158 activation of muscarinic receptors and the differences in the muscarinic receptors are known  
 159 to exist (Vongtau *et al.*, 200; Bonner, 1989).

160 The inhibitory effects of the extract induced contraction by the non-selective  
 161 muscarinic antagonist i.e. atropine observed in our study hence agrees with those of (Akah *et*  
 162 *al.*, 1997, Schlemper *et al.*, 1996). The attenuated rhythmic contractions of the isolated tissue  
 163 produced in our previous study by various extracts, signifies that the action might be  
 164 mediated through the cholinergic receptors (Amos *et al.*, 2003). The medium inhibitory  
 165 contraction of the extract on each of the spasmogen was observed to be as result of the  
 166 extract antagonizing the muscarinic receptors (Augustine *et al.*, 2003; Pohocha *et al.*,  
 167 2001). The extract was found to act through the musculotropic route on the rabbit jejunum.  
 168 This further confirms its activities via the musculotropic route (Augustine, *et al.*, 2003,  
 169 Amos *et al.*, 2000). The active principles presents in the extracts are apparently acting on  
 170 the tissue through the cholinergic receptors and hence are responsible for the actions on the  
 171 tissue (Bolton TB 1979a, Bolton TB 1979b).

172

## 173 CONCLUSION

174 The study indicates that, the Aqueous and Ethanol portion of the whole plant extract contains  
 175 active components which can induce concentration dependent contraction of the rabbit  
 176 jejunum. The contraction observed suggest that, they are inactivated in the presence of other  
 177 portion of the principles (fig: 6). The active principles contain in the plant *S. angustifolia* are  
 178 apparently mediated through muscarinic receptors other than MI receptors. Therefore, the  
 179 study has now justifies the use the plant by the folkloric healers in the treatment of various  
 180 gastrointestinal disorder.

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