ABioaccumulation of Heavy Metals in Wwater and some Ffish Samples from Onuimo River, Imo State, Nigeria

Comment [b1]: Metals do not bioaccumulate in water and sediments but rather in fish so the title here is misleading see my suggestions

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6 ABSTRACT

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7 ABioaccumulation of some heavy metals (Cu, Zn, Cd and Cr) was determined in selected fish, Moon fish 8 (Citharinus citharus), Tilapia fish (Oreochronus niloticus), Mud fish (Clarias anguillaris), Cat fish (Clarias 9 gariepinus) and Carp fish (Labeo coulbie) and water samples from Onuimo River in Imo State in Nigeria. Concentrations of some heavy metals in water and fish samples were determined using atomic 10 absorption spectrophotometer (AA 500 PG) and the results obtained varied significantly among the fish 11 species. BioaAccumulation order of heavy metals in fish samples comprised of were found to follow the 12 13 sequence as Damsel fish > Cling fish > Dat fish > Tilapia fish > Cat fish.- Bioconcentration factor model 14 usedimplored in the present study showed the following a decreasing order as follows; Damsel fish = Cling fish > Cat fish > Dat fish > Tilapia fish and Damsel fish = Cling fish > Tilapia > Dat fish > Cat fish for 15 16 copper and zinc metals respectively. The concentration of Those of cCadmium and chromium were calculated due to the fact that there concentrations in water samples were below detection limits in all 17 18 water samples. Results of analyses of heavy metals in selected water samples revealed that Ceoncentrations of Cu, Cd, Zn and Cr cadmium and chromium wereas below permissibledetection limits 19 20 of some international regulatory bodies whereas those of zinc and copper were also lower than these 21 permissible limits.

Keywords: <u>Atomic absorption spectrophotometer</u>, <u>bioaccumulation</u>, <u>bioconcentration</u>, fish,
 heavy metals, Onuimo River, <u>Nigeriawater analysis</u>.

24 1. INTRODUCTION

Due to geometric increase in human population around the world, the rate at which fish is consumed globally had also increased owing to the fact that fish have both nutritional and therapeutic values. Fish not only serve as good sources of protein but are <u>rich in</u> unsaturated fatty acids <u>basically asrichly in</u> various essential minerals and vitamins which nourishes the body when consumed [1]. These vital nutrients makes fish a perfect health food which when consumed adequately encourages optimal heart functioning. The American Heart Association (AHA) recommended the consumption of fish for at least twice per week in order to reach the daily intake of omega - 3 fatty acids [1].

32 However, Eexcess consumption of heavy metal contaminated fish can result in bioaccumulation of these 33 toxic wastes in the body over time. Fish which are relatively situated at the top of the aquatic food chain 34 can accumulate heavy metals in their tissues either from food, water, and sediments or all. [2-4]. 35 Bioaccumulation of heavy metals in fish and their subsequent distribution toat various organs of the fish 36 are greatly interspecific. Some factors that can influence metal uptake by fish may include the following; 37 metal type, fish species and tissue, sex, size, age feeding behaviour, swimming pattern, reproductive cycle and geographical location etc [2, 5-7]. Entry of heavy metals into the organs of fish mainly takes 38 place by adsorption and absorption along kidney, liver, gut tract walls, muscular and gills surfaces. Thus, 39 40 the rate of accumulation becomes a function of uptake and depuration rates. This makes fish a good 41 indicator of heavy metal contamination in water [8-10].

Comment [b3]: No check the metals that you said you digested in the methods section:

Comment [b4]: Name them here please and put their scientific names also

Comment [b5]: I think you are using the term detection limits incorrectly
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Comment [b7]: Your results say otherwise

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Comment [b8]: This sentence says three confused and disjointed things

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Comment [b12]: reference Comment [b13]: reference 42 Essential heavy metals like zinc, iron, manganese and copper are vital for biological systems like 43 enzymatic activities but they can also be toxic at high concentration [11]. Non essential heavy metals like 44 nickel, chromium, cadmium, mercury, lead, arsenic, and silver-etc-have no known essential role in living 45 organisms. They exhibit extreme toxicity at very low concentration and have been regarded as major 46 threat to life of both organisms and humans. [12-14]. Toxicity effect occurs due to inability of excretory, 47 metabolic, storage and detoxification mechanisms to counter metal uptake [15]. This eventually leads to 48 histopathological and physiological changes [16-17]. Excessive hHeavy metals in fish_tissues can 49 invalidate their health benefits to humans who consume the fish. Several adverse effects of heavy metals 50 in fish samples have been extensively studied [18]. Some of the adverse effects are cardiovascular, renal 51 and peripheral vascular diseases, neurologic and non-behavioural disorders, cancer, proteinuria, liver 52 and kidney failure, gastrointestinal toxicity, nephropathy, hematologic disorder, encephalopathy, 53 pulmonary fibrosis, nasopharyngeal tumors, nephrotoxicity, tremor, nausea, and damage to fetus among 54 others [19-21]. As a result of these effects, many international monitoring programs have been 55 established in order to assess the quality of fish for human consumption and also to monitor the health of 56 aquatic ecosystem [22-24].

Heavy metals are elements with a relative density greater than 5.0 g/cm³ occuringare naturally as 57 58 components of the earth's crust with a relative density greater than 5.0 g/cm³. Metals The can enter the 59 environment through anthropogenic processes like mining, extraction and refining, combustion and 60 electro wining process, indiscriminate discharge of automobile and industrial wastes, fertilizer, pesticides 61 and herbicides application in farm lands, river run off- like hydrothermal vent [26] and volcanic eruptions 62 [27]. Due to great benefits of fish consumption, there is an urgent need for humans to ascertain the levels 63 of heavy metals in fish species that are consumed daily by both humans and animals to minimisein other 64 avoid health hazards associated with that come with consumption of heavy metals contaminated fishes. 65 Some researchers have reported presence of toxic metals in various fish samples in water bodies_such 66 as Onuimo in Nigeria [24, 28-35]. Onuimo has been one of the fresh water rivers in the South Eastern 67 part of the country. Onuiom River The river plays an important role in water supply (domestic and 68 industrial), flood control and fisheries and agricultural purposes to the rural communities that her 69 tributaries cut across. Presence of massive economic and agricultural activities around the river is 70 believed to be the major sources of pollution through river run offs. Hence presence of contaminants like 71 heavy metals and its bioaccumulation in water, soil, sediments, and aquatic organisms especially fishes 72 in river could cause adverse health risk to people who consume products from the river [36-37]. The main 73 objective of this work wasis to investigate the bioaccumulation of four heavy metals in selected fish 74 samples from Onuimo River namely; Moon fish (Citharinus citharus), Tilapia fish (Oreochronus niloticus), 75 Mud fish (Clarias anguillaris), Cat fish (Clarias gariepinus) and Carp fish (Labeo coulbie) as well as to 76 assess the levels of Cu, Cd, Cr and Zn in the water phase of the same river.

77 2. EXPERIMENTAL DETAILS

78 2.1. Site Description

Onuimo River is located in Umungwa Community in Obowo Local Government Area of Imo State, Nigeria
 and the river lies between Longitude 5°50 '56 N and latitude 7°14 ' 20 W. The River is also linked to

81 Umunachi River which is also in Obowo Local Government Area. The river is located close to Onuimo

82 industrial market and has been seen to be a repository for waste generated in the market. The river runs

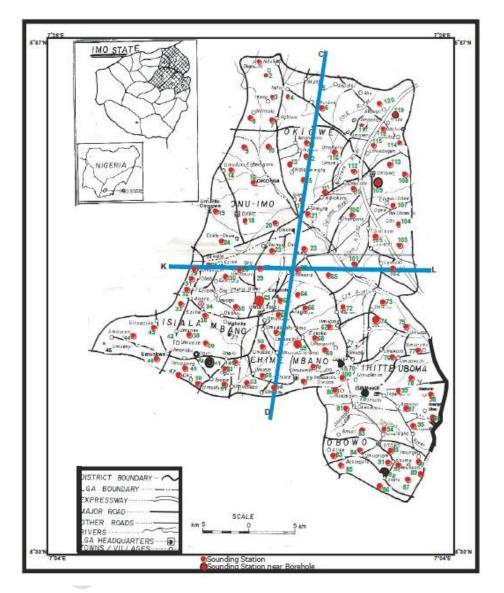
83 approximately ten kilometers across Umuahia down to Obowo where it is located.

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Comment [b16]: You describe the area with the aid of the map not the other way around

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86 Figure 1. Geological Map Showing Onuimo River

Onuimo River is located in Umungwa Community in Obowo Local Government Area of Imo State, Nigeria
 and the river lies between Longitude 5^a50 '56 N and latitude 7^a14 ' 20 W. The River is also linked to
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 industrial market and has been seen to be a repository for waste generated in the market. The river runs
 approximately ten kilometers across Umuahia down to Obowo where it is located.

92 2.2 Fish and Water Sampling

93 Fifteen commercial fish samples of the Moon fish (Citharinus citharus), Tilapia fish (Oreochronus 94 niloticus), Mud fish (Clarias anguillaris), Cat fish (Clarias gariepinus) and Carp fish (Labeo coulbie) were 95 harvested with a locally made gill net in the Onuimo River through the aid of local fishermen at the river. 96 The harvested fish samples were first washed with clean water at the point of harvest, separated 97 according to species and preserved in an ice chest. They were then taken to the laboratory and kept frozen at -20 °C in a refrigerator prior to analyses. On the other hand, water samples were collected 98 alongside where the fishes. Water samples were harvested with separate acid pre washed water plastic 99 100 250ml containers, labelled, enclosed in a chest and transported to the laboratory where they were further 101 refrigerated for a maximum of 48 hours prior to analysis.

102 2.3 Quality Control

All glass wares used for the analysis were all Pyrex, and the reagents used were of analytical grade. The glass wares were washed in 10% nitric acid, 0.5% potassium permanganate and thoroughly rinsed with deionized water and air dried prior to their various uses, were first washed with 10% nitric acid, 0.5% potassium permanganate and thoroughly with deionized water and air dried. Standard operating conditions were all observed to ensure accurate and precise analysis.

108 2.4 Determination of Heavy Metal Contents in Fish and Water Samples

109 In the laboratory, tThe fish samples were brought out of the refrigerator, rinsed thoroughly with deionized 110 water and were dried in an electric oven at a temperature range of 70 - 80 °C for three days. After drying, fish samplesthey were all separately brought out and were crushed to fine particle size using acid pre-111 washed mortar and pestle. Exactly 1 g of the grounded fish sample was accurately weighed and 112 113 transferred into a 250 mL conical flask. 10 mL of digestion mixture in the ration of (1:2:2) of perchloric, 114 nitric and sulphuric acids ware all added to the sample and heated on a hot plate in a fume cupboard at about 200 °C for 30 minutes until white fumes -formed disappeared disappeared indicating complete 115 digestion. The sample was allowed to cool after which 20 mL of distilled water was added to bring the 116 metals into solution. Sample was later filtered with Wwhatman filter paper under gravity into a 100 mL 117 volumetric flask. The filtrate was made up with deionized water up to the graduated mark of the 118 119 volumetric flask. The following metals (Cu, Zn, Fe, Pb, Cd and Hg) contents were analyzed using atomic 120 absorption spectrophotometer (AAS) of model AA500PG after selecting the various wavelengths at which 121 the heavy metals were determined. Also an analytical blank was prepared in a similar method which was 122 used in the calibration of the AAS machine. The analyses were validated by diluting the salt solutions of 123 the investigated heavy metals (Zn, Cu, Cr, Cd) in various concentrations of 0.2, 0.4, 0.6, 0.8 and 1.0 ppm 124 to enable the spectrophotometer measure the concentrations of the investigated heavy metals from their 125 sample solutions [30]. The same procedure was also repeated for all the fish samples and all analyses 126 were done in triplicate. Refrigerated water samples were brought out and allowed to defrost and were 127 transferred into separate beakers. It was followed by digestion of samples according to the procedure 128 stated by Standard Methods for Examination of Water and Waste water, 2nd edition, 2012 (APHA-AWWA-129 WEF) [38].

130 2.5 Statistical Analysis

131 Statistical data analysis was done using SPSS 16.0 software. Descriptive statistics was conducted on the 132 triplicate data to determine the mean, range and standard deviation. **Comment [b17]:** For how many hours before analysis

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133 2.6 Bioconcentration of Heavy Metals in Investigated Fish Samples

Bioconcentration is the process by which a chemical substance is absorbed by an organism from the ambient environment only through its respiratory and dermal surfaces. The degree to which

136 bioconcentration occur is expressed as the bioconcentration factor (BCF) and can only be measured

137 under controlled laboratory conditions in which dietary intakes of the chemicals are deliberately not

138 included. The competing uptake and elimination processes resulting in bioconcentration can be

139 represented mathematically by an organism-water-two-compartment model where the organism is

considered to be single compartment in which the chemical is homogeneously mixed [39].

$$\frac{dC_{B}}{dt} = (k_{1}C_{WD}) - (k_{2} + k_{E} + k_{M} + k_{G})C_{B}$$

where $C_B =$ chemical concentration in the organism (g.kg⁻¹),

 $t = unit time (d^{-1})$

 k_1 = chemical uptake rate constant from the water at the respiratory surface (L.kg⁻¹.d⁻¹)

Eq. 1

 C_{WD} = freely dissolved chemical concentration in the water (g.L¹)

 k_2, k_E, k_M, k_G = rate constants (d⁻¹) representing chemical elimination from the organism via the respiratory surface, fecal egestion, metabolic transformation and ngrowth dilution

when both C_B and C_{WD} no longer vary with exposure duration. That is, $\frac{dC_B}{dt} = 0$, the system

has reached a steady state and equation (1) can be rearranged to calculate the BCF as;

BCF =
$$\frac{C_B}{C_{WD}} = \frac{k_1}{(k_2 + k_E + k_M + k_G)}$$
 Eq. 2

Bioconcentration can also be calculated as the ratio of the chemical concentration in the organism to the chemical concentration in water at steady state [39].

$$BCF_{ss} = \frac{C_B}{C_{WD}} = \frac{Conc_{\cdot Biota}}{Conc_{\cdot Water}}$$
Eq. 3

141 The steady state calculation also refered to as the "Plateau Method" is only valid if the staedy state actually occur (USEPA 1996a; OECD 1996) [40-41].

BCF can also be determined kinetically as the ratio of the chemical uptake rate constant from water and the total elimination or depuration rate constant k_{τ} (d⁻¹),

$$BCF_{k} = \frac{k_{1}}{k_{T}}$$
, where $k_{T} = k_{2} + k_{E} + k_{M} + k_{G}$ Eq. 4

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146 147 **Comment [b20]:** This is unnecessary at most just tell us what you did and show us the formula

148 Table 1. An Overview of Regulatory Bioaccumulation Assessment Endpoint and Criteria [39]

| S/N o | Regulatory Agency | Bioaccumulation Criteria (log values endpoint | | Programme | | |
|----------|--|---|-------------------|-------------------------|--|--|
| 1 | Environmental Canada | K _{ow} | 2 100000 (5) | CEPA (1999)* | | |
| 2 | Environmental Canada | BCF | 2 5000 (8.7) | CEPA (1999) | | |
| 3 | Environmental Canada | BAF | 2 5000 (8.7) | CEPA (1999) | | |
| 4 | European Union 'bioaccumulative' | BCF | ≥ 2000 (3.8) | REACH⁺ | | |
| 5 | European Union 'very bioaccumulative' | BCF | ≥ 3000 (8.7) | REACH | | |
| 6 | United States 'bioaccumulative' | BCF | 1000 - 5000 (8.7) | TSCA † , TRI | | |
| 7 | United States 'very bioaccumulative' | BCF | ≥ 5000 (3.7) | TSCA, TRI | | |
| 8 | United Nations Environment Programme | K _{ow} | ≥ 100000 (\$) | Stockholm convection | | |
| 9 | United Nations Environment Programme | BCF | ≥ 8000 (8.7) | Stockholm convection | | |

*CEPA, Canadian Environmental Protection Act, 1999 (Government of Canada 2000). *Registration, Evaluation and Authorization of Chemicals (REACH) Annex XII (European Commission 2001)

Currently being used by the US Environmental Protection Agency in its Toxic Substance Control Act (TSCA) and Toxic Release Inventory (TRI) Program (USEPA 1976). Stockholm Convention on Persistent organic Pollutants (UNEP 2001).

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150 3.0 EXPERIMENTAL RESULTS AND DISCUSSION

151 Table 2. Levels of Heavy Metals in fish samples in River Onuimo compared to some International

152 Standards

| Stanuarus | | | | | | |
|-----------------------|----------------------|-------------------------|--------------------|----------------------|--|--|
| Sample | Cu | Cd | Cr | Zn | | |
| Damsel fish | | | | | | |
| А | 30.88 | 2.00 | 2.61 | 79.55 | | |
| В | 28.99 | 1.36 | 3.20 | 80.10 | | |
| С | 27.10 | 0.72 | 2.10 | 79.00 | | |
| Range | 27.10 – 30.88 | 0.72 – 2.00 | 2.02 – 3.20 | 79.00 – 80.10 | | |
| $\overline{X} + SD$ | 28.99 <u>+</u> 1.89 | 1.36 <u>+</u> 0.64 | 2.61 <u>+</u> 0.59 | 79.5 <u>+</u> 0.55 | | |
| Tilapia fish | | | | | | |
| Α | 33.56 | 2.10 | 1.74 | 100 | | |
| В | 23.29 | 1.14 | 1.40 | 61.37 | | |
| С | 13.02 | 0.18 | 2.08 | 22.64 | | |
| Range | 13.02 – 33.36 | 0.18 – 2.10 1.40 – 2.08 | | 22.64 – 100 | | |
| $\overline{X} \pm SD$ | 23.29 <u>+</u> 10.27 | 1.14 <u>+</u> 0.96 | 1.74 <u>+</u> 0.34 | 61.37 <u>+</u> 38.73 | | |
| Cat fish | | | | | | |
| A | 30.36 | 2.15 | 0.88 | 40.12 | | |
| В | 27.18 | 1.24 | 1.10 | 45.56 | | |
| С | 24.00 | 0.33 | 0.66 | 51.00 | | |
| Range | 24.00- 30.36 | 0.33 – 2.15 | 0.66 – 1.10 | 40.12 – 51.00 | | |
| $\overline{X} \pm SD$ | 27.18 ± 3.80 | 1.24±0.91 | 0.88 ± 0.22 | 45.56 🛨 5.44 | | |
| Dat fish | | | | | | |
| | | | | | | |

Comment [b21]: Format these tables for them to make sense

| А | 25.12 | 1.26 | 1.27 | 70.00 |
|----------------------------|---------------------|--------------------|--------------------|---------------------------|
| В | 24.66 | 1.88 | 0.53 | 60.44 |
| С | 24.20 | 2.40 | 2.01 | 50.88 |
| Range | 24.20 – 25.12 | 1.36 – 2.40 | 0.53 – 2.01 | 50.88 - 70.00 |
| $\overline{X} \pm SD$ | 24.66 ± 0.46 | 1.88 <u>+</u> 0.52 | 1.27 <u>±</u> 0.74 | 60.44 <mark>±</mark> 9.56 |
| Cling fish | | | | |
| A | 30.77 | 0.21 | 0.41 | 79.35 |
| В | 29.10 | 1.12 | 0.32 | 80.20 |
| С | 27.10 | 2.03 | 0.23 | 79.10 |
| Range | 27.10 - 30.77 | 0.12 - 2.03 | 0.23 - 0.41 | 79.10 - 80.20 |
| $\overline{X} \pm SD$ | 28.99±1.89 | 1.12±0.91 | 0.32 <u>+</u> 0.09 | 79.55±0.55 |
| ^[42] FEPA(2003) | 1.3 | - | 0.15 | |
| ^[43] WHO(2006) | 3.0 | - | 0.15 | -/ |
| ^[44] FU (2001) | 1.0 | - | 1.0 | |
| ^[45] EU (2008) | 0.5 – 1.0 | 0.5 – 1.0 | 2.0 | |
| ^[46] Indonèsia | 80 | - | - | 200 |
| ^[47] FAO (1983) | - | 2.0 | 1.0 | |
| | | | | |

^[42]Federal Environmental Protection Agency (2003)

^[43]World Health organization (2006)

^[44]European Union (2001) ^[45]European Union (2008)

^[46]Indonesia Decree of General Director of Food and Drug Supervision No.03725/B/SK/VII concerning, maximum limits of metals in food.

^[47]Food and Agricultural Organization of the United Nation (1983)

153

Previous research had shown that escalating human population, economic development and poor waste 154 155 management have affected adversely the ecosystem. This have significantly contributed to the current worldwide deterioration of water quality including seasonal accumulation of heavy metals like Cr, Cu, Zn, 156 Cd, As, Hg, Fe, etc [23,24]. An insight on the levels of these heavy metals in aquatic organism especially 157 fishes are of paramount important to basically for consumption by human and nature management. This 158 research work documents results of bioaccumulation of some heavy metals in some fish samples 159 together with their bioconcentration factor. 160

161 Results of the study conducted, as shown in Table 2 above, reveals that copper level in the investigated 162 fish samples ranges as follows; Damsel fish (27.10- 30.88) mg/kgir Cat fish (24.00- 30.36) mg/kgir Tilapia fish (13.02-33.56) mg/kg; Dat fish (24.20-25.12) mg/kg and Cling fish (27.10-30.77) mg/kg. -A trend of 163 164 mean concentrations of copper in mg/kg can be written as Tilapia (23.29) < Dat (24.66) < Cat (27.18) < 165 Damsel = Cling (28.99) mg/kg. -These mean values were found to have been higher than some standard 166 permissible limits like WHO (3.0mg/kg), FEPA (1.3 mg/kg), EU (2008) (1.0 mg/kg) and those reported in Cyprinus Carpio and Pelteobagrus Fluridraco [23], L._Coubie and M. Tapirus [31]. Indo-pacific king 167 Mackerel and Tiger tooth Crocker [48]. Although copper is recognized as an essential element, excessive 168 169 intake of it can lead to poisoning, nausea, nausea, diarrhea and fever, acute stomach pain and death 170 [23].

171 Cadmium is an environmental pollutant and a highly toxic metal which has no biological function in both 172 human system and aquatic organisms. Bioaccumulation of cadmiumCadmium can remain in human 173 system for decades and cannot be efficiently metabolized. It can cause kidney damage, lung cancer, 174 ostemalacia, testicular tissue destruction, high blood pressure, proteinuria, red blood destruction and nondescended testes in young males [19, 48-49]. Recent research showhas it that exposure to cadmium at 175

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even low concentration can increase the risk <u>ofte</u> hormonal cancer [50]. Another research on Long Island
also estimated <u>that</u> about 40% of breast cancer cases recorded in the United States might be associated
with elevated cadmium levels [51]. Results of <u>cadmiumvarious</u>_levels_<u>of cadmium as</u>-shown in Table 2
depicts that cadmium recorded highest mean value of 1.88 mg/kg in Dat fish and <u>the</u> least value of 1.12
mg/kg in Cling fish. The various mean values as shown in Table 2 were also higher than permissible limits
of EU (2008) but lower than that of FAO (1983).

182 Chromium another environmental pollutant showed an increasing trend in mg/kg as follows, Cling fish
(0.32) < Cat fish (0.88) < Dat fish (1.27) < Tilapia fish (1.74) < Damsel fish (2.61). Mean values of
chromium in Cling and Cat fishes (Table 2) were observed to be lower than permissible limit of FAO
(1983), EU (2008) and some literature values [52-53]. Chromium levels in the remaining fish samples
were higher than the permissible limits above and those reported in *Balistoides Vridiscens* [19].
Bioaccumulation of chromium in humans can lead to-the following health diseases; pulmonary fibrosis,
lung cancer (inhalation), cardiovascular, renal, gastrointestinal, hematological and neurological effects

189 [19].

190 Levels of zinc in the investigated fish recorded the least minimum value of 45.56 mg/kg in Cat fish and

191 highest value of 79.55 mg/kg in Damsel and Cling fishes. A trend of decrease in mean values of zinc in

the investigated fish samples can be seen as; Cat fish (45.56) \leq Dat fish (60.44) \leq Tilapia fish (61.37) \leq

Damsel fish = Cling fish (79.55). These mean values are also higher than some permissible limits of Indonesia maximum limits of metals in food (Table 2) and some literature studies [30, 31].

195 Table 3. Level of Heavy Metals in Water, Bioconcentration Factor (BCF) of Investigated Fish 196 Samples

| | | | | | | | | Bioconcentration Factor | | | | |
|-------|--------|--------|--------|--------|--------|--------|--------|-------------------------|------|-----|-----|-------|
| Metal | Conc. | [54] | [55] | [56] | [57] | [58] | [59] | Dam | Tila | Cat | Dat | Cling |
| | of | WHO | UNEP | USEPA | ECE | FTP- | ADWG | sel | pia | | | - |
| | water | (2011) | (2007) | (mg/L) | (1998) | CDW | (mg/L) | | | | | |
| | (mg/L) | limits | limits | | (mg/L) | (mg/L) | | | | | | |
| | , | (mg/L) | (mg/L) | | | | | | | | | |
| Cu | 0.065 | 2.000 | 2.000 | 1.300 | 2.000 | 1.000 | 2.000 | 446 | 358 | 418 | 379 | 446 |
| Cd | BDL | 0.003 | 0.003 | 0.005 | 0.005 | 0.005 | 0.002 | - | - | - | - | - |
| Cr | BDL | 0.050 | 0.050 | 0.010 | 0.050 | 0.005 | 0.050 | - | - | - | - | - |
| Zn | 0.112 | 3.000 | 3.000 | 0.500 | - | 50.00 | 3.000 | 710 | 548 | 407 | 540 | 710 |

^[54]World Health Organization (WHO, 2011)

^[55]United Nation Environmental Programme (2007)

^[56]United States Environmental Protection Agency (USEPA, 2011)

^[57]European Commission Environment (ECE, 1998)

^[58]Federal-Provincial-Territorial Committee on drinking Water (CDW), Health Canada (FTP-CDW, 2010)

^[59]Australian Drinking Water Guidelines (2011)

197

Results of heavy metals in investigated water samples ares shown in Table 3, and these depicts that 198 199 levels of copper (0.065 mg/L) and zinc (0.112 mg/L) were lower than permissible limits of World Health Organization limits(2011). Cadmium and chromium levels were observed to be below detection limit 200 201 (BDL). Results of Bioconcentration Factor (BCF) implored showed that copper levels have an increasinghas an increasing BCF order as followsvalues of : Tilapia fish (358) < Dat fish (379) < Cat fish 202 (418) < Damsel = Cling (446). Also zinc has an increasing BCF order which comprised; values of Cat fish 203 204 (407) < Dat fish (540) < Tilapia (548) < Damsel = Cling (710). These values were found to be lower than 205 the criteria (log value) of ≤2000 (3.3) for European Union bioaccumulative and criteria (log values) of 1000 (3) - 5000 (3.7) for United States bioaccumulative and other international regulatory assessment 206

endpoints and criteria as shown in Table 1. Bioconcentration factors of chromium and cadmium were not
 calculated because there levels in water samples were below detection limits.

209 Levels of heavy metals in water samples were also investigated and are reported in (Table 3). The results 210 showed that the mean concentration of copper in the investigated water samples had a value of 0.0065 211 mg/L which was_-observed to be lower than the permissible WHO limits of World Health Organization 212 (WHO, 2011) (2.0 mg/L), United States Environmental Protection Agency (USEPA) (1.3 mg/L), European 213 Commission Environment (2.0 mg/L) and Federal-Provincial-Territorial Committee on Drinking Water, 214 Health Canada (FTP-CDW, 2010) (1.0 mg/L). Mean concentration of zinc (0.112 mg/L) was also lower 215 than permissible limits of WHO (3.0 mg/L), USEPA (0.5 mg/L), FTP-CDW (50 mg/L) and UNEP (3.0 216 mg/L). Values of cadmium and chromium were observed to be below detection limit.

217 4. CONCLUSION

218 The results of the study provide valuable information on levels of heavy metals in water samples and their 219 bioaccumulation in some selected fish samples from Onuimo River, Imo State. The result also showed 220 that heavy metals sorption and bioaccumulation by fish varied with respect to fish species of fish and 221 other specific factors like the feeding pattern, weight and age. In this study the A trend of bioaccumulation 222 of heavy metals inby fish samples was can be deduced as follows; Damsel fish > Cling fish > Dat fish > 223 Tilapia fish > Cat fish. The bBioconcentration factor model usedimplored in the study showed a 224 decreasing order as follows; Damsel fish = Cling fish > Cat fish > Dat fish > Tilapia fish, and Damsel fish = 225 Cling fish > Tilapia for copper and zinc metals respectively. Also, the bioconcentration (BCF) values 226 obtained were lower than the standards of some International regulatory bodies. Levels of heavy metals 227 in investigated water samples were also lower than the permissible limits of WHO, USEPA, UNEP and 228 FTP-CDW limits. These results imply that River Onuimo is relatively less contaminated by heavy metals, 229 thus the water having little contamination by heavy metals is safe for human, agricultural and industrial 230 uses. Also rate of disposal of heavy metal containing waste into the river and other anthropogenic 231 activities which can contaminants the water should be discouraged. Better still, they should be adequately 232 recycled before disposal to avoid loading of heavy metals and other

233 COMPETING INTEREST

The authors have declared that there is no conflict of interest regarding the publication of this research work.

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Comment [b25]: Poor recommendations start again please

Comment [b26]: Follow the format of the journal

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