

1 **BIOREMEDIATION OF INDUSTRIAL EFFLUENT USING CYANOBACTERIAL**
2 **SPECIES: *PHORMIDIUM MUCICOLA* AND *ANABAENA AEQUALIS***

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6
7 **ABSTRACT**
8

Industries discharge effluent into different water body subjected to severe levels of pollution that can cope with the high pollution load in the water. Textile and Pharmaceutical industry, Mandideep, Bhopal has discharge industrial effluent into river. The main objective of the present study was to investigate the biodegradation and biosorption capacity of some potential cyanobacterial species; *Phormidium mucicola* and *Anabaena aequalis* dominating the river ecosystem. Heavy metals contaminants polluting the Industrial effluents. The effluents were subjected to biological treatment using axenic cyanobacterial strains as batch system for 7 days. Removal efficiencies of the different contaminants were evaluated and compared. Results confirmed the high efficiencies of the investigated species for the removal of the target contaminants which were species and contaminant-dependent. BOD and COD recorded 91.18 and 82.54% as maximum Removal efficiencies achieved by *Anabaena aequalis*. The highest Removal efficiencies of the Total suspended solids recorded 53.23% achieved by *Phormidium mucicola*, while 41.61% was recorded as the highest TDS. Removal efficiencies achieved by *Phormidium mucicola*. Concerning the contaminant metals, *Phormidium mucicola* showed the highest biosorption capacity where 86.12 and 94.63% Removal efficiencies were achieved for Zn and Cu, respectively. In conclusion, results of the study confirmed the advantageous potential of using the tested cyanobacterial species for the bioremediation of industrial effluent and clearly showed the quality improvement of the discharged effluent which in turn will eliminate or at least minimize the expected deterioration of the receiving environment.

9 **Keywords:** algae, bioremediation, cyanobacterial species, heavy metals, industrial effluent

10 **1. INTRODUCTION**

11 Textile and Pharmaceutical industrial effluents discharge directly into river or other water
12 source like, close Water Lake in Bhopal city. Beside nutrients, the river water and sediments
13 showed terrible levels of organic matter, and heavy metals in worldwide. This is mainly due
14 to continuous discharge of huge quantities of the effluents lead to deterioration in the water

15 quality of this river (El-Bestawy E., 1993; El-Bestawy E. et al., 2007; Mansy And El-Bestawy,
16 2002). Such pollutant with time and the shift of bacterial and algal populations toward more
17 resistant species such as the planktonic cyanobacteria that dominate the river water
18 especially in the warm seasons. These species characterized by great ability to tolerate such
19 high levels of pollution and proved high efficiency for degrading highly organic contaminants
20 and accumulating heavy metals (El-Bestawy E et al.,2007; Podda et al.,2000 and Palmer,
21 C.M. 1980). Therefore, they could be efficiently used in advanced technologies for
22 bioremediation of the industrial effluents.

23 Cyanobacteria are gram-negative photosynthetic Some lakes are naturally eutrophic, but in
24 many other prokaryotes. They can be found in a wide range of water bodies the excess
25 nutrient input is of anthropogenic habitats from ice fields to hot springs and deserts. Origin,
26 resulting from municipal wastewater discharge or Morphologically, physiologically and
27 metabolically, this runoff from agricultural land. Cyanobacteria have a group is one of the
28 most diverse groups of prokaryotes number of special properties that determine their (Abd
29 Allah LS 2006). The rapid evolution of cyanobacteria in different water importance, relative
30 success and predominance during the land environments is related to their capacity for both
31 growth season in phytoplankton communities. However, aerobic and anaerobic
32 photosynthesis. cyanobacteria are located in thylakoids lying free in the summer period:
33 water temperature above 25°C, low light cytoplasm near the cell periphery intensity in water,
34 low N: P ratio and stability of the water.

35 Any change in pH of water bodies as a result of influx of effluent; can cause serious change
36 in water chemistry, which can affect resources especially around the coastal areas. These
37 effects on water bodies can be very significant. Traditional method for the clean up of
38 pollutants usually involve, the removal of unwanted materials through sedimentation and
39 filtration, and subsequent chemical treatment such as flocculation, neutralization and electro-
40 dialysis before disposal.

41 Many species of cyanobacteria possess gas nitrogenase, they convert N directly into
42 ammonium in vesicles, which enable regulation of the buoyancy aerobic conditions.
43 Recently, there has been increasing interest about using cyanobacteria as pollution control
44 agents since they possess many advantages over other microorganisms isolated from soil.
45 Their photoautotrophic nature and the ability of some species to fix atmospheric nitrogen
46 enable them to be producers, as opposed to consumers, and make their growth and
47 maintenance inexpensive [Castenholz et al., 1989; Somashekar, R.K. and Ramaswamy,
48 S.N.1983). Metabolic activities are not affected by the decrease in the levels of the
49 biodegradable pollutants that they may break down. Cyanobacteria have been used
50 efficiently as a low-cost method for remediating all industrial effluents as well as

51 transformation and removal of heavy metals (Lefebvre et al., Budd K 2007; Podda et
52 al.,2000). Remediation capabilities of cyanobacteria toward environmental pollutants can be
53 improved and enhanced through genetic engineering technologies (Kuritz and Wolk 1995;
54 Mansy and El-Bestawy E. 2002 and Palmer, C.M. 1980). However, the beneficial application
55 of cyanobacteria in remediation of contaminated waters and industrial effluents is still not
56 optimally manipulated (Jeganathan, 2006 and Kannan, 2006). The main objective of the
57 present study was to investigate the remediation capacity of some potential cyanobacterial
58 species isolated from Textile and Pharmaceutical industrial effluent (Gohl and Vilensky 1987;
59 James et al., 1979; Stewart et al., 1970; Tien and Kirk 1984).

60 2. MATERIAL AND METHODS

61 Survey of different sites of industrial effluent for identification of different algal forms from
62 taxonomic point of view will be undertaken. Collecting sample (effluent and cyanobacteria)
63 from two industrial effluents such as Textile and pharmaceuticals industries, Mandideep,
64 Bhopal, India. Effluent samples and cyanobacteria were collected in large sterilized
65 containers and polythene bags respectively. Thus, it is expected that the effluents contain
66 industrial pollutants such as heavy metals which are not likely to be removed by that primary
67 treatment of the industries. Grab samples representing all effluent entering the plant during
68 24 h were collected from both plants to avoid the fluctuation in the flow and the strength of
69 the effluent.

70 Physico-chemical characteristics of waste waters were carried out by standard methods
71 (APHA, 1995).Such as biochemical oxygen demand (BOD); chemical oxygen demand
72 (COD); total suspended solids (TSS); total dissolved solids (TDS); and two heavy metals
73 (Zn, Cu) were characterized before and after treatment to determine the effectiveness of the
74 remediation process(5,6). All the investigated parameters were determined using the
75 standard techniques described by (Celesseri et al., 1999) in the standard methods for the
76 examination of water and effluent water.

77 Standard microbiological methods were followed for the isolation of cyanobacteria. Algal
78 samples were microscopically examined and the selected cyanobacterial species were
79 grown in Chu No.10 (1942) medium were used as culture medium. They have the following
80 composition (macronutrients).Ca(NO₃)₂ ,0.04 ; K₂HPO₄ ,0.01- 0.005 ;MgSO₄. 7H₂O, 0.025;
81 Na₂CO₃ , 0.02; NaSiO₃ , 0.025; EDTA, 0.008 and Solution B(micronutrients) contained (in
82 g/l): Na₂.EDTA, 4.36; FeCl₃. 6H₂O, 3.15; CuSO₄.5H₂O, 0.01; ZnSO₄.7H₂O, 0.022;
83 CoCl₂.6H₂O, 0.01; MnCl₂.4H₂O, 0.18; NaMoO₃.2H₂O, 0.006 and the pH was adjusted to
84 7.2 with HCl. Each component of solution A was separately prepared as stock solution while
85 all the components of (solution B) were prepared as a mixture. (Solution A) component were
86 sterilized by autoclaving separately at 121⁰C for 20 min. Micronutrient solution was sterilized

87 by filtration through 0.22 μm polycarbonate membrane to avoid interaction and precipitation
88 of heavy metals. Chu No.10 media was freshly prepared from A and B where 1.0 ml of each
89 component of solution A and 1.0 ml of Solution B were combined and diluted to 1.0 l,
90 sterilized as mentioned and used for selective culturing of the selected species. After
91 inoculation, all the selected species were incubated at room temperature (28°C) and day
92 light with manual shaking every 24 h to avoid adhesion of the algae on the walls of the glass
93 vessels until heavy growth appeared within 3 weeks.

94 Identification was confirmed based upon the keys given by (Geitler, 1932 and Desikachary,
95 1959) for microscopic parameters. The isolated cyanobacteria were identified with the help
96 of classical manuals. Two different cyanobacterial species; *Anabaena aequalis* and
97 *Phormidium mucicola*; were investigated as free-living cells for their ability for organic matter
98 biodegradation and heavy metal removal from the effluent. They were selected based on
99 their dominance and survival in the highly polluted water of Pharmaceutical industries and
100 textile industries where they acquired high resistance and acclimatized to deal with high
101 loads of different contaminants. They were also proven high ability for degradation of the
102 heavy metals. Therefore, the selected species were considered promising candidates for
103 biological treatment of the industrial effluents. They were kindly provided as axenic strains by
104 the algal culture collection at the lab of phycology, where they were identified using the
105 classical methods.

106 **2.1 Axenity and bioassay**

107 Unialgal cultures usually remained contaminated with bacteria and therefore to free them
108 from bacteria is a pre requisite for further studies (Ash and Jenkins, 2006; Anagnostidis, and
109 Komárek, 1985). The cultures were made bacteria free by ultraviolet irradiation (2537Å) for
110 varying periods and inoculated in the medium. Selected species were provided as axenic
111 cultures. However, before using these strains in the bioremediation of the contaminated
112 industries effluents, their axenity was checked using agar phototactic response method.
113 Semi solid standard agar medium was prepared and aliquoted into test tubes and sterilized.
114 Each tube was inoculated with 100 μl of cyanobacterial culture (two replicates per culture).
115 Light was prevented to reach the top 10 cm of the tube using aluminum foil. All the tubes
116 were incubated in optimal conditions (28°C) in an illuminated incubator. Based on the
117 phototactic response phenomena the cyanobacterial filaments were grown toward light
118 direction through the semi solid agar, but bacteria did not grown. After 7 days incubation of
119 the agar column was dragged out the tube on sterilized Petri dish. The agar column was
120 sliced into ten slices 1 cm per each. Each slice was stranded longitudinally and transversally
121 cut under common sterilized conditions to separate the algal filaments surrounded with a

122 small piece of agar. Each agar piece involving cyanobacterial growth was inoculated into
 123 standard selective liquid medium. After incubation, each inoculated culture was tested for
 124 contamination using general bacterial medium (nutrient agar). In bioremediation bioassay,
 125 the tested species were checked for their axenicity, and the liquid cultures were tested by
 126 plating on bacterial nutrient medium and incubating at 28⁰C for 7 days. Only axenic cultures
 127 were involved in the assays. The selected species were inoculated individually in 100 ml
 128 culturing medium (three replicates) and incubated for 2 weeks till heavy growth was
 129 obtained. Effluents water from both industries textile and pharmaceutical was dispensed
 130 (900 ml each) in 18 sterilized conical flasks, nine flasks for each effluent. Each culture (100
 131 ml) was separately seeded at a final volume of 1 l each (three replicates/strain/effluent) and
 132 incubated under the previously mentioned conditions for 7 days. Another six flasks (three
 133 flasks for each industry) were supplied by 1.0 l each of the effluent of both industries without
 134 seeding with cyanobacteria to serve as control for the bioassay. They were incubated under
 135 the same conditions. For the determination of heavy metals and other parameters residues,
 136 samples were collected at 24 h interval. At each sampling time, 130 ml from each flask were
 137 aseptically drawn, where all the investigated parameters were determined and their removal
 138 efficiencies using the selected species were calculated.

139 3. RESULTS AND DISCUSSION

140 Table 1 Residue concentrations (RC) of the quality parameters from the contaminated
 141 industrial effluents using the selected cyanobacteria at different exposure time

142

143 Textile industry effluent (control)

144 Time(day)	BOD	COD	TSS	TDS	ZN	CU
145 Raw water	140	360	167	1,150	0.11	0.04
146 2	89	289	132	887	0.09	0.01
147 3	80	243	100	766	0.05	0.02
148 4	81	200	101	756	0.12	0.04
149 5	70	187	98	611	0.09	0.05
150 6	120	387	154	900	0.14	0.06
151 7	143	331	143	998	0.12	0.07

152 Pharmaceutical industry effluent (control)

153 Time (days)	BOD	COD	TSS	TDS	ZN	CU
154 Raw water	198	445	387	1430	0.01	0.03
155 2	187	465	411	1457	0.04	0.01
156 3	199	412	376	1432	0.03	0.04

UNDER PEER REVIEW

157	4	176	398	311	1200	0.01	0.04
158	5	166	345	298	1100	0.03	0.06
159	6	178	411	321	1289	0.05	0.02
160	7	200	378	365	1008	0.02	0.01

161

162

163 Textile industries effluent

164 Anabaena aequalis

Phormidium mucicola

165	Days	BOD	COD	TSS	TDS	ZN	CU	BOD	COD	TSS	TDS	ZN	CU
166	Raw water	149	312	200	1012	0.11	0.05	148	324	234	1134	0.12	0.07
167	2	141	301	199	1011	0.09	0.05	143	321	223	1124	0.10	0.07
168	3	132	298	189	1001	0.06	0.03	132	309	212	1120	0.5	0.05
169	4	121	288	170	998	0.05	0.02	122	289	206	1103	0.5	0.03
170	5	100	198	150	988	0.03	0.02	101	281	189	1087	0.3	0.02
171	6	76	122	132	986	0.03	0.02	98	256	167	1079	0.1	0.01
172	7	62	98	112	983	0.02	0.01	78	249	156	1012	0.1	0.01

173

174

175 Pharmaceutical industries effluent

176 Anabaena aequalis

Phormidium mucicola

177	Days	BOD	COD	TSS	TDS	ZN	CU	BOD	COD	TSS	TDS	ZN	CU
178	Raw water	254	432	321	1349	0.08	0.07	231	421	401	1401	0.06	0.07
179	2	233	428	311	1323	0.06	0.05	211	421	387	1400	0.04	0.06
180	3	212	412	306	1321	0.06	0.04	201	401	381	1387	0.04	0.04
181	4	198	398	285	1309	0.04	0.04	192	356	372	1345	0.02	0.03
182	5	167	378	265	1287	0.03	0.02	187	241	324	1302	0.01	0.03
183	6	121	321	209	1270	0.02	0.01	123	209	283	1265	0.01	0.01
184	7	98	243	187	1201	0.02	0.00	98	198	223	1230	0.00	0.01

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186

187

188 3.1 Industrial effluent characteristics

189 Effluent produced by the two industries was characterized (Table1, control).BOD, COD,
 190 TSS, TDS, Zn and Cu recorded averages of 140, 360, 167, 1150, 0.11and 0.04 mg/l,
 191 respectively, in the effluent of the textile industry. Significantly higher levels for almost all the
 192 tested parameters were detected in the pharmaceutical effluent where 198, 445, 387, 1430,

193 0.01, and 0.03 mg/l were recorded as average. However, Zn recorded much lower average
194 in the pharmaceutical effluent (0.01mg/l) compared to that of the textile effluent (0.11 mg/l)
195 while no significant differences were recorded in the Cu levels among the two industry (0.04
196 and 0.03 mg/l in the textile and pharmaceutical effluents). Nitrogen and phosphorus content
197 in both effluents (El-Bestawy E et al., 2005 and Ellis 1977) along with the toxic industrial
198 contaminants suppressed the growth of cyanobacteria or any other algae.

199 3.2 Treatability and removal efficiency of effluent

200 3.2.1 Contaminants

201 Residue levels of the selected quality parameters were determined (Table 1) and the
202 removal efficiencies (RE %) as results of the biological treatment using the selected species
203 were calculated. As a general trend, the two tested species exhibited positive correlation
204 between their RE% of the all the tested parameters and the exposure time up to the last
205 exposure day for both types of effluents.

206 3.2.2 Organic matter removal

207 Biochemical oxygen demand Removal of BOD from industrial effluents of both industries
208 using the selected algae revealed the following points:

209 1. High REs% were obtained for BOD removal from industrial effluent by the selected
210 species with *Anabaena aequalis* (90.65%) and finally *Phormidium mucicola* (81.9%).

211 2. Despite the RE variations of BOD achieved by the tested species, RC(s) of the BOD in
212 the industrial effluent reached acceptable limits (19, 20 and 32 mg/l by *Anabaena aequalis*
213 and *Phormidium mucicola*, respectively) after 7 exposure of days which is much lower than
214 the maximum permissible limit (MPL) of 60 mg/l stated by the Environmental Laws for safe
215 discharge into surface water courses. When these figures compared with those obtained by
216 the control it was showed that the natural microbial population of the effluent achieved a
217 maximum removal of 50% after 5 days equivalent to 60.7 mg/l (MPL of the BOD) after which
218 there was a sharp decline in the efficiency associated by increasing the RC reaching 140
219 mg/l and 3.20% RE after 7 exposure of days.

220 3. Comparing BOD removal by the selected cyanobacteria from the two plants revealed very
221 high efficiency for all of them in the degradation of biodegradable organic matter which is
222 stimulated by increasing the levels of the pollutant in the wastewater.

223

224 Chemical oxygen demand Removal of COD from the industrial effluents using the selected
225 species revealed the following points:

226 1. *Anabaena aequalis* considered the most effective for removing COD from the industrial
227 effluent achieving the maximum RE of 83.68% compared to the RE achieved by *Phormidium*

228 *mucicola* (45.00%) after 7 exposure days. However, *Phormidium mucicola* exhibited higher
229 COD RE% within the first 24 h compared to *Anabaena aequalis*.

230 2. The lowest residue concentration of 100 mg/l was achieved by *Anabaena aequalis* which
231 is the maximum acceptable limit stated by the law (MPL for COD = 100 mg/l) while
232 *Phormidium mucicola* could not bring the COD levels of the effluent to better quality. They
233 recorded 180 and 246 mg/l, respectively, and required longer exposures. The highest RE%
234 achieved by the control culture recorded 50.11% (170.59 mg/l) after 5 exposure days.

235 3. Similar to BOD removal, the natural microorganisms in the control culture were inhibited
236 by the high Strength of the industrial effluent leading to reduction in the COD removal.

237 4. Although high REs% of the COD was achieved by the selected species, none of them
238 could bring the COD levels in the effluent below the MPL during the investigated exposure
239 time (1 week). This may be attributed to the need for longer time for achieving the proper
240 quality. It could also result from the inhibition in cyanobacterial growth due to the higher COD
241 levels in the pharmaceutical effluent compared to that of the textile effluent.

242 **3.2.3 Solids removal**

243 Total suspended solids (TSS) Removal of TSS from the industrial effluents using the
244 selected species revealed the following points:

245 1. The highest recorded TSS REs% in the effluent recorded 42.0 and 29.12% achieved by
246 *Anabaena aequalis*, and *Phormidium mucicola* (125 and 133 mg/l RC), respectively, after 7
247 days.

248 2. According to the law, 60 mg/l is stated as the MPL of the TSS; therefore none of the
249 tested species reached the required efficiency to bring the TSS in the effluents below the
250 MPL during the tested exposure time. This indicates that they required longer time, heavier
251 biomass or different application using the same species to achieve that quality.

252 3. In contrast to cyanobacteria, the indigenous bacteria of the control culture achieved higher
253 TSS removal from the textile effluent compared to that of the pharmaceutical effluent.

254

255 Total dissolved solids (TDS) Removal of TDS from the industrial effluent using the selected
256 species revealed the following points:

257 1. The maximum TDS REs% obtained for the effluent by the tested species ranged between
258 a maximum of 16.66% (1,078 mg/l) achieved by *Phormidium mucicola* and a minimum of
259 12.00% (1,119 mg/l) obtained by *Anabaena aequalis* after 7 exposure days. *Phormidium*
260 *mucicola* exhibited higher TDS RE% at the shorter exposures (up to 2nd day) compared to
261 *Anabaena aequalis*.

262 2. Similar behavior for TSS removal was shown by bacteria of the control culture where
263 higher TDS removal was achieved from the textile effluent compared to that of the
264 pharmaceutical effluent.

265 3. Since the TDS content in the effluents were lower than the MPL of the TDS (2,000 mg/l),
266 the residual concentrations of the TDS produced in the final effluents by all the tested
267 species as well as the two controls improved and still within the safe range for discharging.

268 **3.2.4 Heavy metals removal**

269 Results revealed the following points:

270 1. *Phormidium mucicola* recorded the highest REs% for Zn from EWTP (86.12) and
271 *Anabaena aequalis* (70.88%) recording RCs of 0.0247, and 0.0370 mg/l by the three
272 species, respectively, after 7 days .

273 2. Although low Zn levels were detected in the pharmaceutical effluent, lower Zn REs were
274 achieved compared to those obtained for the textile effluent. Zn removal recorded 78.2, and
275 65.00% achieved as the highest Zn REs% by *Phormidium mucicola*, and *Anabaena*
276 *aequalis*, respectively (0.0123, and 0.0182 mg/l, respectively) after 7 days.

277 3. Although all the average levels of Zn for both effluents were below the MPL of 5 mg/l
278 before the treatment, Zn levels were reduced producing much better effluent quality. Zinc
279 removal was stimulated by increasing its level in the wastewater.

280 4. Concerning Cu , much higher REs% were recorded for effluent compared to those
281 obtained for Zn removal regardless it's high toxicity. This may be attributed to the high
282 resistance of the selected members which was stimulated by increasing Cu levels in the
283 wastewater. 94.63, 90.99 and 90.64% RE of Cu were achieved by *Phormidium mucicola*,
284 and *Anabaena aequalis* (0.0031, and 0.0054 mg/l RC), respectively, after 7 days.

285 **4. CONCLUSION**

286 In conclusion, results confirmed that the most effective species for BOD, COD, TSS, TDS
287 Zn and Cu removal from the effluents of the two industries are in the following order
288 *Phormidium mucicola* and *Anabaena aequalis* which may be attributed to the selective
289 uptake of the investigated pollutants by the tested cyanobacterial species.

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