

2 **EVALUATION OF MEDIAN LETHAL DOSE AND**
3 **SUBCHRONIC ORAL TOXICITY ASSESSMENT OF**
4 **ETHANOLIC LEAF EXTRACT OF *PHYLLANTHUS***
5 ***AMARUS***

6 **ABSTRACT**

7
8 **Aims:** To determine the median lethal dose (LD₅₀) of crude ethanolic leaf extract of *Phyllanthus*
9 *amarus* and evaluate its sub-chronic oral toxicity in experimental mice (BALB/c strain).

10 **Study design:** One-factor, one-control, one-test group experimental design.

11 **Place and Duration of Study:** Department of Medical Biochemistry, Delta State University, Abraka,
12 Nigeria, between December, 2014 and November, 2015.

13 **Methodology:** Crude ethanolic leaf extract of *P. amarus* was prepared as previously described and
14 twenty (20) Swiss albino mice (BALB/c strain) were randomly and equally divided into two (2) groups
15 and administered 2000 mg/kg body weight (Group A) and 5000 mg/kg body weight (Group B of the
16 prepared extract as single oral dose in line with the limit dose method of determining LD₅₀. For the
17 sub-chronic oral toxicity study, ten (10) mice were assigned into control (n=5) and experimental
18 (n=5). The control animals were given placebo-normal saline, but the experimental mice were
19 administered with nocebo – 300 mg/kg body weight of *P. amarus* of crude ethanolic extract for
20 twenty one (21) days. Thereafter, the animals in each group were sacrificed and then, serum and
21 liver homogenate were obtained for the assay of total antioxidant capacity (TAC) and oxidative
22 damage (Malondialdehyde-MDA) Using documented methods. Liver tissue was also processed for
23 histopathological examination using H&E stain.

24 **Results:** Data showed LD₅₀ of the extract to be greater than 5000 mg/kg. Assessment of the herb's
25 sub-chronic oral toxicity indicates that the leaf extract significantly ($P=.03$) enhanced total
26 antioxidant capacity (TAC) in both serum (Control: TAC = 0.10±0.03 mM, Experimental: TAC =
27 0.33±0.05 mM) and liver (Control: TAC = 0.12±0.09 mM, Experimental: TAC = 0.34±0.06 mM) but
28 reduced ($P = .01$) the biomarker for liver tissue (Control: MDA = 41.89±3.36 µM, Experimental: MDA
29 = 4.67±4.04 µM). In addition, hepatic cells were invigorated by *P. amarus* treatment as suggested
30 by the histopathological features.

31 **Conclusion:** Collectively, *P. amarus* crude ethanolic leaf extract possesses high degree of
32 tolerance and hepatic tonic potential with no identifiable toxic or side effects.

33
34 **Keywords:** *Phyllanthus amarus*, Median Lethal Dose (LD₅₀), Sub-chronic Toxicity, Total Antioxidant
35 Capacity (TAC), Malondialdehyde (MDA).

36 **1.0 INTRODUCTION**

37 The use of plants, plant extracts or plant-derived chemicals to treat diseases is a therapeutic
38 modality that has been explored for centuries. Over 40,000 species of tropical flowering plants are
39 known to possess medicinal properties [1] and are currently in use for various medical conditions.
40 Majority of Africans patronize herbal or traditional medicine for their health needs. It is estimated that
41 70-80% of patients in Africa are treated by traditional healers and herbal practitioners [2]. Modern
42 medicine recognizes herbalism as a form of alternative medicine based on evidence derived from
scientific methods [3]. Herbal medicine is, thus, gaining popularity and one of such herbs receiving
wide patronage is *Phyllanthus amarus*.

Phyllanthus amarus is an herbal plant belonging to the Euphorbiaceae family. It has
approximately 800 species which are found in tropical and subtropical countries of the world [4,5]. The
plant has been found in Philippine, Cuba, Nigeria and India among others. Extract of the plant has
been reported to possess pharmacological effects such as antibacterial [4,6], antiviral [7], anticancer
[8], anti-amnesic [9], antioxidative [10], antimicrobial [11], antileptospiral [12], anticonvulsant [13] and

43 anti-inflammatory [14,15] activities. *Phyllanthus amarus* has been used as chemoprotective [16],
44 antimutagenic [17], nephroprotective, cardioprotective [18], hepatoprotective [19] and hypoglycemic
45 [20] agent. It is known to exhibit *in vivo* antiplasmodial property [21] in addition to its demonstrated
46 ability to invigorate the pancreas [22] and restore renal function altered by *Plasmodium berghei*
47 malarial parasite infection in experimental mice [21].

48 Lack of knowledge of the mechanisms and side effects of some herbal preparations as well
49 as safety regulations for their usage may have serious consequences [23]. Many consumers believe
50 that herbal medicines are “safe” because they are “natural”, but, several adverse effects of herbs have
51 been reported including allergic reactions, hepatotoxicity [24,25,26], nephrotoxicity [27,28,29], cardiac
52 toxicity [30,31], neurotoxicity [32,33], and even death [34].

53 Since *Phyllanthus amarus* is currently gaining recognition in alternative medical practice, it
54 has therefore become pivotal to evaluate the median lethal dose and subchronic toxicity of the
55 ethanolic leaf extract of the plant cultivar wildly grown in the tropical rain forest zone of Abraka, Delta
56 State, Nigeria. This freely growing variety of the plant is common and easily harvested in our
57 environment for medicinal use.

58 **2.0 MATERIALS AND METHODS**

59 **2.1 Harvesting and preparation of plant extract:** Fresh whole plants of *Phyllanthus*
60 *amarus* wildly growing in uncultivated land space in Abraka, Ethiope East Local Government Area of
61 Delta State, Nigeria were obtained in July, 2015 and authenticated (No: FHI: 109728) in the
62 Herbarium Unit, Forestry Research Institute of Nigeria, Ibadan. Crude ethanolic leaf extract of the
63 harvested fresh plant was prepared as earlier described [21]. The leaves were washed, air-dried and
64 pulverized using a sterile Electric blender (Kenwood Ltd, Hertfordshire, U.K) to produce a fine powder.
65 The ethanolic extract of the plant sample was prepared by soaking 100 g of dry powdered sample in
66 200 ml of ethanol for 24 hours. The extract was filtered using whatman filter paper and the filtered
67 extract were concentrated using the Soxhlet apparatus (Corning, U.S.A). The extract was evaporated
68 to dryness using rotary evaporator (Buchi R-210 Hana, China) under reduced pressure and dissolved
69 in distilled water which was then stored in a refrigerator until required for analysis.

70 **2.2 Experimental mice:** Forty (40) Swiss albino BALB/*c* mice of mixed sexes weighing between
71 21.1 to 28.2 g were used for the entire study. They were maintained at the Laboratory Animal Centre,
72 Faculty of Basic Medical Sciences, Delta State University, Abraka, Nigeria. The mice were fed on
73 growers' mash (Top Feeds, Sapele, Delta State, Nigeria), and were given clean drinking water *ad*
74 *libitum*. The animals were housed in plastic cages, under controlled condition of 12 hr light/12 hr dark
75 cycle at a temperature of 29±2°C. The animals were maintained in accordance with the guidelines
76 provided by the Research and Bioethics Committee of the Faculty of Basic Medical Sciences, Delta
77 State University, Abraka, Nigeria.

78 **2.3 Evaluation of lethal and effective doses (LD₅₀ and ED₅₀):** LD₅₀ and ED₅₀ were
79 determined by the limit dose method [35]. A total of thirty (30) mice (20 for LD₅₀ and 10 for ED₅₀) were
80 used. In the phase of LD₅₀ determination, the mice were divided into two groups of ten (10) mice
81 each. They were treated with ethanolic leaf extract of *Phyllanthus amarus* at doses of 2000 and 5000
82 mg/kg body weight as oral single dose. The animals were observed for 24 hours first and then, for
83 twenty one (21) days for any sign of toxicity and mortality.

84 **2.4 Subchronic Study:** For the subchronic study, the remaining ten (10) mice were divided into
85 Control (n = 5) and Experimental (n = 5) Groups. The Experimental Group was administered 300
86 mg/kg/d *P. amarus* ethanolic leaf extract as single daily dose for 21 days. The dosing regimen was
87 based on previous experience [22]. The animals were observed for any physical signs of toxicity,
88 morbidity and mortality. Body weights were measured weekly throughout the 21-day study period.

89 **2.5 Animal Sacrifice and Collection of Sample:** On the 21st day of the experiment, the
90 mice were fasted overnight and sacrificed the next day under chloroform anesthesia. The liver was
91 excised and whole blood was collected by heart puncture and centrifuged (Cent 80D, Serico, China)
92 to obtain serum which was used for the biochemical analyses of total antioxidant capacity (TAC) and
93 malondialdehyde (MDA) levels. The excised liver was fixed in 10% formol saline for histological
94 processing and examination. However, a portion (0.5 g) was homogenized and then, prepared for
95 biochemical assay.

96 **2.6 Biochemical Assay:** Total antioxidant capacity, TAC in serum and liver homogenate as
97 determined by the Trolox Equivalent Antioxidant Capacity (TEAC) method described by Miller *et*
98 *al.*[36] and MDA levels were estimated by the Thio-Barbituric Acid Reacting Substances (TBARS)
99 method earlier described by Ohkawa *et al.*[37]. TAC provides information on degree of antioxidant
100 defense, and MDA indicates a measure of membrane lipid peroxidation, and hence, oxidative
101 stress/damage.

102 **2.7 Histological Studies:** The portion of the liver tissue fixed in 10% formol saline was
 103 processed overnight using histokinette and embedded in paraffin wax. Three sections - four micron in
 104 thickness - were cut from each paraffin block.

105 **2.8 Light Microscopic Examination:** One section from each sample was stained with
 106 Heamatoxylin and Eosin (H&E) stain by the standard method for light microscopic (histological)
 107 examination.

108 **2.9 Ethical Approval:** The study was conducted in compliance to the guidelines provided by the
 109 Research and Bioethics Committee of the Faculty of Basic Medical Sciences, Delta State University,
 110 Abraka, Nigeria – the body that approved the study.

111 **2.10 Statistics:** Data were presented as Mean \pm S.D and analyzed by the Student's *t*-Test using
 112 SPSS software package version 20. Significant difference was set at $P=0.05$

113 3.0 RESULTS

114 Results obtained from evaluation of median lethal dose (LD_{50}) and subchronic oral toxicity
 115 study of the ethanolic leaf extract of *Phyllanthus amarus* grown freely in uncultivated land space in
 116 Abraka, Ethiope East Local Government Area of Delta State, Nigeria, are shown in Tables 1-2 and
 117 Figures 1-2.

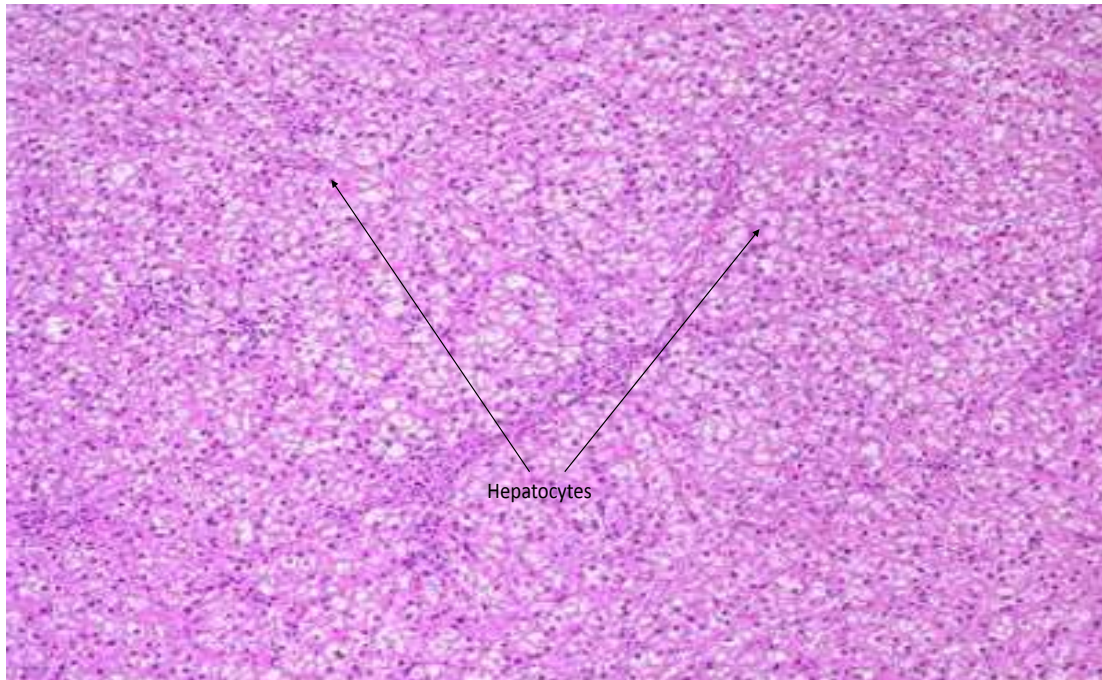
118 Table 1 shows the cage side physical observations of the control and experimental mice used in the
 119 determination of LD_{50} , while, Table 2 presents the biochemical data (TAC and MDA) obtained from
 120 both serum and liver tissues of the animals used to assess subchronic oral toxicity. Then, Figures 1-2
 121 are the histological features of the liver tissues excised from Control (Fig. 1) and *P. amarus* (300
 122 mg/kg/d for 21 days) treated mice (Fig. 2).

123

124 **Table 1: Cage side physical observations during the LD_{50} evaluation of *P. amarus***
 125 **ethanolic leaf extract**

	Considerations	Cage side physical observations after 24 hours and 21 days					
		2000 mg/kg		5000 mg/kg		Control (0 mg/kg)	
		24 hours	21 days	24 hours	21 days	24 hours	21days
1	Condition of fur	Normal	Normal	Normal	Normal	Normal	Normal
2	Skin appearance	Normal	Normal	Normal	Normal	Normal	Normal
3	Subcutaneous swelling	Nil	Nil	Nil	Nil	Nil	Nil
4	Abdominal distension	Nil	Nil	Nil	Nil	Nil	Nil
5	Eye dullness	Nil	Nil	Nil	Nil	Nil	Nil
6	Eye opacity	Nil	Nil	Nil	Nil	Nil	Nil
7	Pupil diameter	Normal	Normal	Normal	Normal	Normal	Normal
8	Colour/consistency of faeces	Normal	Normal	Normal	Normal	Normal	Normal
9	Teeth condition	Normal	Normal	Normal	Normal	Normal	Normal
10	Gait	Normal	Normal	Normal	Normal	Normal	Normal
11	Weight gain (%)	0.3	5.0	0.5	7.0	0.1	3.0
12	Mortality	0	0	0	0	0	0

126 Evidence from observations (Table 1) indicates that the LD_{50} of *P. amarus* crude ethanolic leaf extract
 127 is greater than 5000 mg/kg. Trial doses cannot be increased beyond 5000 mg/kg because that is the
 128 limit dose. Effective dose (ED_{50}) = 200 mg/kg. Hence, therapeutic index, $TI (LD_{50} / ED_{50}) = 25.0$



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Fig. 1: Photomicrograph of liver tissue from control mouse showing normal hepatocytes. Magnification $\times 100$ (H & E stain).

Table 2: Changes in total antioxidant capacity (TAC) and malondialdehyde levels (MDA) induced by subchronic oral toxicity study of *P. amarus* crude ethanolic leaf extract.

Sample	Assay	Control	<i>P. amarus</i> (300 mg/kg/d)	P- value
SERUM	TAC (mM)	0.10 \pm 0.03	0.32 \pm 0.05*	.03
	MDA (μ M)	40.33 \pm 3.36	21.02 \pm 1.59*	.02
LIVER	TAC (mM)	0.12 \pm 0.09	0.34 \pm 0.06*	.03
	MDA (μ M)	41.89 \pm 2.27	4.67 \pm 4.04*	.01

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Data are presented as Mean \pm SD for n=5

*Significantly different from comparable control values at $P < 0.05$

TAC-Total antioxidant capacity, MDA-Malondialdehyde.

The subchronic oral toxicity of *P. amarus* crude ethanolic leaf extract was studied by administering 300mg/kg/d of the plant extract to experimental BALB_c mice for 21 days.

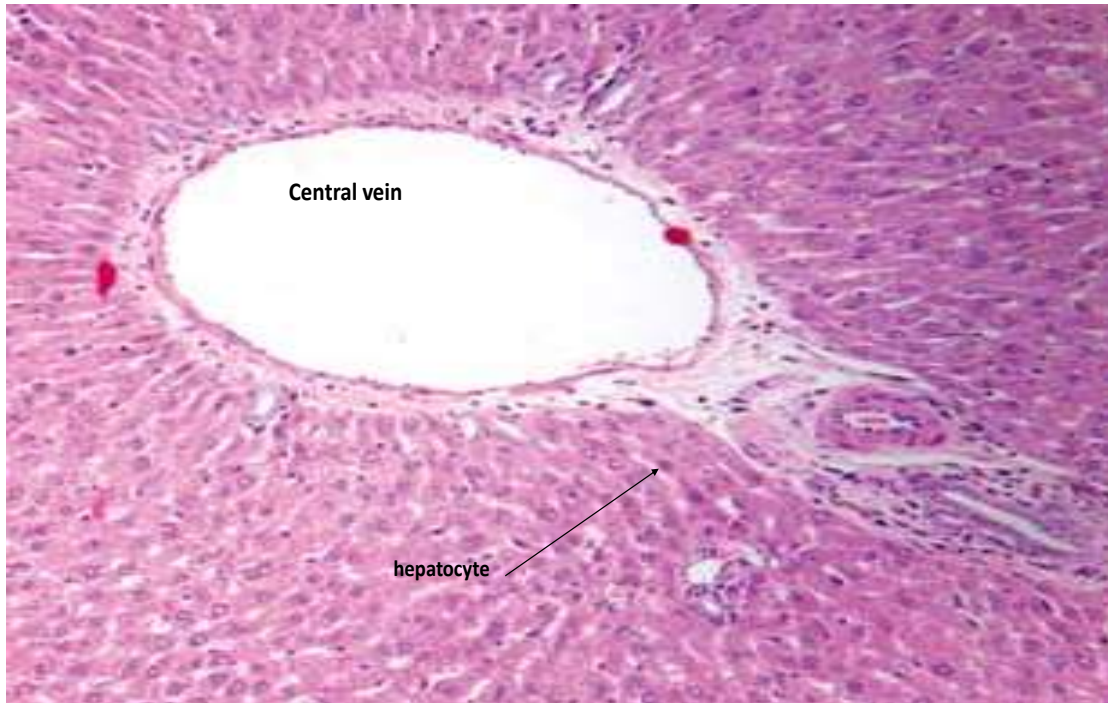


Fig. 2: Photomicrograph of liver tissue obtained from mouse administered 300mg/kg body weight of crude ethanolic leaf extract of *P. amarus* for 21 days, indicating normal histological features of invigorated hepatocytes and central vein. Magnification $\times 100$ (H & E stain).

4.0 DISCUSSION

This study attempted to evaluate the LD₅₀ and subchronic oral toxicity of the crude ethanolic leaf extract of *Phyllanthus amarus*. Result of the limit dose test indicates that the LD₅₀ of *P. amarus* crude ethanolic leaf extract is well above 5000 mg/kg with an ED₅₀ of 2000 mg/kg and hence, therapeutic index of 25. These observations show that the herb possesses very high phytotherapeutic efficacy with no demonstrated toxicity. These findings suggest that *Phyllanthus amarus* is safe and non-toxic with very high remedy potential in experimental mice. This agrees with previous documents [38].

Chronic toxicity study identifies and provides information on drugs that could possibly cause harm and pose health challenges [39]. The subchronic oral toxicity assessment of *P. amarus* crude ethanolic leaf extract during this study, reveals that the extract significantly ($P = .03$) boosted antioxidant defense activity in both blood and liver tissue with associated reduction ($P = .01$) in overall membrane damage. The liver is the organ involved in several metabolic functions and is therefore prone to xenobiotic-induced injury because of its central role in xenobiotic metabolism [40]. Histopathological examination of the liver shows that *P. amarus* administered at 300 mg/kg/d body weight for 21 days invigorated liver cells. Hepatotoxic drugs could cause peroxidation of liver cell membrane lipids and increase the amount of end products such as MDA [39].

Data suggest that *Phyllanthus amarus* extract has a measure of health benefits as shown by the significant decrease in malondialdehyde (MDA) levels and associated increase in total antioxidant capacity, TAC (Table 2). The decrease in malondialdehyde level may be as a result of the increased antioxidant activities of *Phyllanthus amarus* [41]. Increased antioxidant activity in cells causes a decrease in free radicals thereby reducing lipid peroxidation and malondialdehyde production. The reduction in both blood and liver malondialdehyde levels suggests that the extract may contain mixture of biomolecules with hydroxyl groups that perhaps prevented the abstraction of hydrogen atom from the double bond of lipid bilayers thereby preventing lipid peroxidation. This suggestion corroborates previous report on the *in vitro* analysis of the plant extract [42].

Phytochemical studies of *Phyllanthus amarus* extract have shown that the plant contains chemicals such as flavonoids, tannins, saponins, alkaloids, terpenoids, glycosides and phenols [42,21]. Flavonoids present in the plant have been shown to possess several pharmacological properties such as antioxidant activities and anti-inflammatory activities [20,43]. Flavonoid as an antioxidant has a rejuvenating effect on cells and tissues [44], Tannin has demonstrated high activities

181 against viral and bacterial infections as well as acting as strong antioxidant [45]. The antioxidant
182 activity of this plant phytochemicals may have contributed to the decrease in MDA levels observed in
183 this study. These findings are concurrent with previous studies conducted on the toxicological
184 assessment of *Phyllanthus amarus* [46].

185 **4.1 CONCLUSION**

186 Findings indicate that *Phyllanthus amarus* plant materials have no significant toxic effect in
187 Swiss albino mice.

188 **4.2 RECOMMENDATION**

189 Put together, the crude ethanolic leaf extract of *Phyllanthus amarus* is bestowed with very
190 high phytotherapeutic efficacy and vitalizing property with no recognizable toxic effect. Therefore, the
191 phytochemicals and nutrient quality of *P. amarus* need to be characterized for functional analysis.

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