1 An *in vivo* antiplasmodial activity of aqueous and ethanol crude plant extracts of

2 Phyllanthus fraternus using plasmodium berghei infected balb/c mice

3 Keywords

4 In vivo, antiplasmodial activity, Phyllanthus fraternus, phytochemicals, chemosupression,

- 5 *Plasmodium berghei*.
- 6

7 Abstract

8

Background: *Phyllanthus fraternus* is a tropical plant that has numerous pharmacological
activities such as blennorrhagia, colic, diabetes, dysentery, fever, flu, tumors, jaundice,
vaginitis, dyspepsia, anti-inflammatory, antioxidant, anticoagulant, antidiabetic, antiviral and
analgesic. The study evaluated *in vivo* anti-plasmodial activity of aqueous and ethanol crude
plant extracts of *Phyllanthus fraternus* using *Plasmodium berghei* infected *Balb/c* mice.

Methodology: The preparation of the aqueous crude extract was done by boiling 195 g of the 14 15 dried plant material in 4 L of water for 30 minutes and cooled. The resultant extract was filtered through a cotton wool and put in an oven at 50 °C to concentrate it before it was pre-16 freeze and lyophilized into powder using a freeze dryer (Heto powder dry LL 300, Sapa). 17 Similarly the preparation of the ethanol crude extract was obtained by simple maceration of 18 195 g of dried sample of the plant in 2 L aqueous ethanol (1.4 L of ethanol plus 0.6 L of 19 distilled water) for 72 h. It was then filtered through cotton wool and subjected to rotary 20 evaporator (ILA CCA-1111 Japanese branch) to evaporate the ethanol and then pre-freeze 21 22 and freeze- dried. The crude extracts were screened for their phytochemical constituents which showed the presence of secondary metabolites. The LD_{50} of both extracts were 23 24 investigated using Sprague-Dawley rats and found to be greater than 5000 mg/kg. The in

vivo antiplasmodial activity (percentage parasitemia (%P) and the percentage chemosuppression (%C)) of the extracts were evaluated using *Balb/c* mice.

Results: The aqueous and ethanol extracts established modest antiplasmodial activity in a 27 dose dependent manner. The standard drug (coartem 2 mg/kg) with percentage parasitaemia 28 (%P) of 28.57 ± 4.70 and 2.48 ± 0.48 caused percentage chemosuppression (%C) of 44.38 ± 7.63 29 and 81.27±2.07 in day four and six respectively. The test groups (aqueous and ethanol 30 extracts) for two different doses (100 mg/kg and 200 mg/kg) each administered with 31 percentage parasitemia (%P) 39.67±1.35, 39.58±1.64, 37.32±2.37, 36.23±1.99 and 32 10.24±1.32, 9.33±0.66, 8.61±0.96, 7.27±1.26 caused percentage chemosuppressions (%C) of 33 22.78±2.20, 22.96±2.66, 27.35±3.84, 29.48±3.23 and 22.54± 9.93, 29.43±4.99, 34.87±6.66, 34 44.99 ±5.98 in day four and six respectively. The aqueous extract demonstrated better 35 inhibition of *plasmodium* in doses 100 mg/kg and 200 mg/kg with chemosuppressions (27.35 36 37 \pm 3.84 and 29.48 \pm 3.23) respectively compared with the ethanol extract of the same doses 100 mg/kg and 200 mg/kg with chemosuppressions (22.78 \pm 2.20 and 22.96 \pm 2.66) 38 39 respectively. The activity of the standard drug, coartem at 2.0 mg/kg was significantly higher 40 (p < 0.05) with chemosuppression (44.38±7.63) than those of the extracts. The extracts were also screened for phytochemicals for which some were found in the extracts which have 41 previously been implicated as antiplasmodial agents. The LD₅₀ of both extracts were 42 investigated and found to be greater than 5000 mg/kg. 43

44 Conclusion: The aqueous and ethanol crude plant extracts of *P. fraternus* possess
45 antiplasmodial activity and would be useful in the search for novel antimalarial agents.

46 Background

The plant *Phyllanthus fraternus* belongs to the Family *Euphorbiaceae* and is commonly
called gulf leaf-flower, Chancapiedra, stone breaker, carry-me-seed, hurricane weed or

49 quinine weed. The plant also has local names such as Mache da goyo (Hausa), Gbogbonowun Ofobi okpabi (Krobo and Ga), Lume or Kpavideme (Ewe), lese (Yoruba), 50 Awommaaguwakyi (Twi) [1]. It is an annual dicotyledonous herb which is small, erect and 51 52 grow in gutters, dumping places and along the road of 30 to 40 cm in height [2]. Traditional herbalist in Ghana uses the whole plant for numerous pharmacological activities such as 53 blennorrhagia, colic, diabetes, dysentery, fever, flu, tumors, jaundice, vaginitis, and dyspepsia 54 [3]. From literature Phyllanthus fraternus possesses anti-inflammatory [1], antioxidant and 55 anticoagulant [2], antidiabetic [4-5], antiviral [6] and analgesic properties [7-8]. Through bites 56 57 of female Anopheles mosquitoes a parasite called Plasmodium species are transmitted into human which result in malaria disease [9]. Antiplasmodial activity of different species of the 58 genus Phyllanthus have been determined elsewhere [10], but as far as literature can tell no 59 60 work have been done on an *in-vivo* of the aqueous and ethanol whole plant extracts of P. 61 fraternus against malaria. As a matter of fact, the existing orthodox drugs have lots of side effects and the most efficacious among them are now becoming impotent to the parasite and 62 63 there is a need to research on new antimalarial plants (*P. fraternus*) [1].

Plasmodium berghei infected balb/c mice were employed in this study because they have 64 similar properties of genetics, anatomy and physiology with humans in terms of experimental 65 research. Especially mice are used due to their similarity of genomes that mimics humans and 66 also their cost effective. The other types of mammals normally used for animal model 67 68 experiments are rodents and these include; rats, gerbils, guinea pigs and hamsters. [11]. Even though there is an advancement into modern medicines, underdeveloped countries still rely 69 massively on medicinal plants for their survival during disease attack. To get rid of malaria 70 71 infection in the underdeveloped countries, the World Health Organization aimed to include traditional medicine for its preventive approach. Many medicinal plants have been employed 72 on the basis of their antimalarial properties by traditional herbalists but their effectiveness 73

have not been scientifically assessed [12]. The Herbalist in Ghana documented the plant *P*. *fraternus* as antimalarial drug but as far as literature can ascertain, it had not been
scientifically assessed. There was no much adverse effects assigned to medicinal plants since
its existence and are also believed to be significant in terms of new source of chemical
substances with a therapeutic effects. Therefore this study aimed to evaluate an *in vivo*antiplasmodial activity of aqueous and ethanol crude plant extracts of *P. fraternus* on *P. berghei* infected *Balb/c* mice.

81

82 Materials and Methods

83 **Drugs and chemicals**

All drugs and chemicals used such as tetraoxosulphate (vi) acid (H₂SO₄), ammonium hydroxide (NH₃OH (aq)), magnesium ribbon, 2 mL of hydrochloric acid (HCl), chloroform, ammonia, ferric chloride, acetone, sodium picrate paper, fehling solution A and B, 70% ethanol, giemsa stain, methanol and sodium chloride, were obtained from British Drug House Ltd (Poole, England). Coartem was obtained from Troge Medical GMBH (Hamburg Germany) were all of analytical grade unless otherwise stated.

90 Plant Raw Materials and Herbal Standard

Phyllanthus fraternus whole plant material (leaves, stems and roots) were obtained from the
Plant Production Department (PPD), of Centre for Scientific Research into Plant Medicine
(CSRPM) Mampong-Akuapem, Ghana and authenticated by Dr. Yaw Ameyaw, a botanist of
the production department.

95 Animals

96 Seven-week old female Balb/c mice (30 g) were obtained from the animal unit of the Centre
97 for Scientific Research into Plant Medicine (CSRPM), Mampong-Akuapem, in the Eastern

Region of Ghana. The animals were fed on powdered feed obtained from Ghana Agro Food
Company (GAFCO), Tema, Ghana. They were allowed free access to sterile distilled water.

100 Preparation of Herbal Extracts

The plant material was cut into small pieces and spread thinly on a flat clean tray to prevent 101 spoilage by moisture condensation and allowed to dry at room temperature for three (3) days. 102 The dried plant material (195 g) was boiled in 4 L of water for 30 minutes and cooled. The 103 resultant extract was filtered through a cotton wool and put in an oven at 50 °C to concentrate 104 105 it before it was pre- freeze and lyophilized into powder using a freeze dryer (Heto powder dry LL 300, Sapa). The dry powder was weighed to determine the yield and stored in a desiccator 106 at room temperature. This was reconstituted in sterilized distilled water before use. 70% 107 ethanol extract was obtained by simple maceration of 195 g of dried sample of whole plant of 108 P. fraternus in 2 L aqueous ethanol (1.4 L of ethanol plus 0.6 L of distilled water) for 72 h. It 109 110 was filtered through cotton wool and subjected to rotary evaporator (ILA CCA-1111 Japanese branch) to evaporate the ethanol and then pre-freeze and freeze- dried. 111

112 Malaria parasites and inoculum preparation

Plasmodium berghei NK65 strain from the University of Copenhagen Denmark through the Department of Immunology, Noguchi Memorial Institute of Medical Research (NMIMR), University of Ghana, Accra, Ghana, was used for the experiment. The stock of parasitized erythrocytes was obtained from infected Balb/c mice, with a minimum peripheral parasitemia of 20%, by cardiac puncture in heparin-coated tube. The cell concentration of the stock was determined and diluted with physiological saline such that 0.2 mL of final inoculum contained 10⁶ parasitized red blood cells (RBCs).

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121 Acute toxicity test

The acute oral toxicity study was conducted to know the amount of dose to be given to the animals. This was done by the Organization for Co-operation and Development (OECD) guidelines 425 received from the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA) [11].

127 Treatment of Animals

Thirty six (36) mice were selected and put into six (6) groups of six per group. Each mouse 128 was inoculated intraperitoneal with the parasite P. berghei. Group 1 (Gp1) animals received 129 distilled water (negative control), group 2 (Gp 2) animals received 2 mg Coartem (positive 130 131 drug control), group 3 (Gp 3) and group (Gp 4) animals received 100 mg/kg and 200 mg/kg of ethanol extract of whole plant of *P. fraternus* respectively, group 5 (Gp 5) and group 6 132 (Gp6) animals received 100 mg/kg and 200 mg/kg aqueous extract of whole plant of P. 133 134 fraternus respectively. All the drugs were orally administered to the animals (0.2 mL) 2-3 h after the mice have been inoculated with the parasite over a period of 6 days. 135

136 Monitoring of Parasitemia and Antimalarial Activity

On the fourth and sixth days after drug administration, thin blood smears were prepared using blood from the tail vein of each mouse. Each smear was air-dried, fixed in methanol, air-dried again, stained with 10% giemsa for 10-15 minutes and examined under oil immersion with a microscope. Each slide was observed at three different fields and the Red Blood Cells (RBC_S) and total number of RBC_S for each field was recorded. The percentage parasitemia (% P) and the percentage chemo-suppression (% C) also known as the activity was estimated

143 according to the following formulae %
$$P = 100 \times \frac{PRBC - TRBC}{TRBC}$$
 % $C = 100 \times \frac{PCON - PTEST}{PCON}$

- 144 Where; *PRBC* is the number of parasitized Red Blood Cell (RBC).
- 145 *TRBC* is the total number of RBC counted per field.
- 146 PCON is the control parasitemia and PTEST is the test parasitemia.
- 147 Statistical analysis

148 Data were presented as means \pm SEM of n= 6 and analyzed using One-way ANOVA which 149 was followed by students t-test. The P \leq 0.05 was considered statistically significant in all 150 analysis.

151 **Results**

Phytochemical screening was carried out for aqueous and ethanol whole plant extracts of *Phyllanthus fraternus* which identified the presence and absence of groups of secondary metabolites using the standard method [13-14].The phytochemical screening of the extracts showed the presence of alkaloids, saponin, phenolics, reducing sugars, triterpenes and phytosterols in both extracts while cynogenic glycoside and anthraquinones were absent in both extracts and flavonoids and polyuronides showed presence only in the aqueous extract (Table 1).

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PHYTOCHEMICAL	EXTRACTS		
	Aqueous	Ethanol	
Alkaloids	+	+	
Saponins	+	+	
Phenolics	+	+	
Reducing Sugar	+	+	
Polyuronide	+	-	
Terpenoids	+	+	
Flavonoids	+	-	
Phytosterols	+	+	

160 **Table 1: Phytochemical constituents of** *Phyllanthus fraternus* whole plant extracts.

Anhthracenoside	-	-
Cyanogenic Glycoside	-	-

(+) = Present and (-) = Absent

163 Acute toxicity test

164	The LD_{50} of the extracts were identified and was greater than 5000 mg/kg and may be
165	classified as practically non-toxic and within the acceptable margin of safety (Hodge and
166	Sterner scale) at the recommended dose. Thus $1/50^{\text{th}}$ and $1/25^{\text{th}}$ (i.e.100 mg/kg and 200
167	mg/kg) were selected for the study (Table 2).

Table 2: Acute toxicity test for *Phyllanthus fraternus* whole plant of aqueous and **ethanol extracts**

	Phyllanthus fraternus v	whole plant
	Aqueous extract	Ethanol extract
Species and strain	Sprague-Dawley rats	Sprague-Dawley rats
Number of animals	Twelve (12)	Twelve (12)
Sex	Females	Females
Number. of groups	3 (N=4)	3 (N=4)
Route of administration	Oral	Oral
Formulation	Freeze dried	Freeze dried
Dose administered (mg/kg)	1250, 2500, 5000	1250, 2500, 5000
Period of observation	48 hours	48 hours
Number. of deaths	Zero (0)	Zero (0)
Approximate lethal dose(LD ₅₀)	>5000 mg/kg	>5000 mg/kg
Signs of toxicity	Nil	Nil

Percentages of Parasitemia and Chemo-suppression of *Phyllantus fraternus* whole plant of aqueous and ethanol extracts in 4th and 6th days test

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The route of administration of the controls (Coartem and distilled water) were done at doses of 2 mg/kg orally; aqueous and ethanol crude plant extracts of *P. fraternus* were given orally at doses of 100 mg/kg and 200 mg/kg which significantly exerted *in vivo* antiplasmodial activity on the *P. berghei* infected *Balb/c* mice in a dose-dependent fashion at day 4 and day 6 except ethanol crude extract at the dose of 100 mg/kg for day 4 and day 6 (Table 3).

9					
	Extracts	Day four		Day six	
	Concentration	Parasitemia	Chemosuppression	Parasitemia	Chemosuppression
	(mg/kg)	(%)	(%)	(%)	(%)
	Control	61.64±3.77	0.00	13.22 ± 2.32	0.00
	Coartem 2	28.57±4.70	44.38±7.63	2.48±0.48	81.27±2.07
	PET 100	36.23±1.99	29.48±3.23	10.24±1.32	22.54±9.93
	PET 200	37.32±2.37	27.35±3.84	9.33±0.66	29.43±4.99
	PAQ 100	37.32±2.37	27.35±3.84	10.24±1.32	34.87±6.66
	PAQ 200	39.67±1.35	22.78±2.20	7.27±1.26	44.99±5.98

179	Table 3: Results of Percentage	Parasitemia and	Chemosuppression	of 4 and 6 days test

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Graphs of Percentages of Parasitemia and Chemo-suppression of *Phyllantus fraternus* whole plant of aqueous and ethanol extracts in 4th and 6thdays test

186 The results obtained from Percentage Parasitemia and Chemo-suppression of 4 and 6 days

test (Table 3) were represented graphically where PAq = Aqueous extract of *Phyllanthus*

188 *fraterus*, PEt = Ethanol extract of *Phyllanthus fraternus*, Results are means \pm SEM of n= 6, *

189 = Values significantly different from Distilled water controls (p<0.050) and $^{\#}$ = Value

significantly different from positive controls (p<0.050) (Figure 1 and 2).

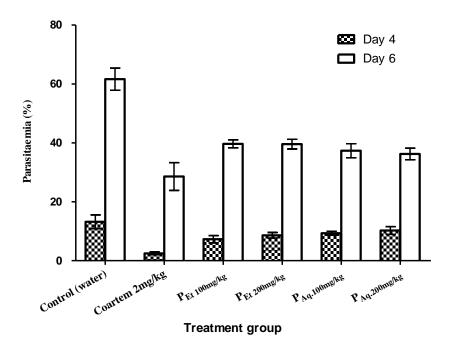


Figure1: Graph of the percentage parasitemia of *Plasmodium berghei* infected balb/c mice atday four and six.

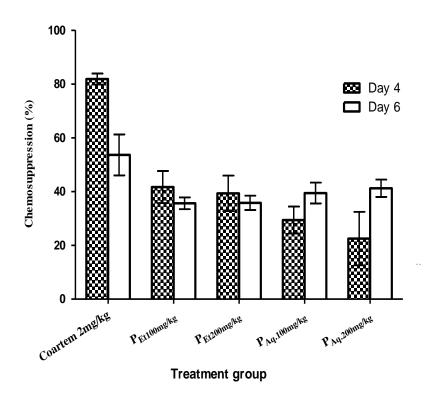


Figure 2: Graph showing the percentage Chemo-suppression of *Plasmodium berghei* infected *Balb/c* mice at day four and six.

198 Discussions

199 The phytochemical screening of the extracts showed the presence of alkaloids, saponin, phenolics, reducing sugars, triterpenes and phytosterols in both extracts while cynogenic 200 201 glycoside and anthraquinones were absent in both extracts and flavonoids and polyuronides showed presence only in the aqueous extract. The result showed differences from reported 202 works by Sofowora; Olonisokan et al. [15-16]. The factors attributed to these differences were 203 as a result of environment of the plant, mode of extraction and the climatic conditions [17-204 18]. Reports have shown that antiplasmodial activity of many agents were due to interference 205 with the reproductive system of the protozoa [19]. Several reports have implicated alkaloids 206 207 [20], terpenoids [21] and lignans [22-24] as antimalarial agents. The antiplasmodial activity demonstrated by both extracts may be attributed to the presence of some of these 208 phytochemicals. The extracts showed modest antiplasmodial activity in a dose dependent 209 210 manner as manifested in the results (Table 3). The standard drug (coartem 2 mg/kg) with (%P) of 28.57 ± 4.70 and 2.48 ± 0.48 caused percentage 211 percentage parasitemia 212 chemosuppression (%C) of 44.38±7.63 and 81.27±2.07 in day four and six respectively. 213 From (figure 1 and 2), the test groups (aqueous and ethanol extracts) for two different doses (100 mg/kg and 200 mg/kg) each administered with percentage parasitemia (%P) of 214 39.67±1.35, 39.58±1.64, 37.32±2.37, 36.23± 1.99 and 10.24±1.32, 9.33±0.66, 8.61±0.96, 215 7.27±1.26 caused percentage chemosuppressions (%C) of 22.78±2.20, 22.96±2.66, 27.35± 216 3.84, 29.48±3.23 and 22.54±9.93, 29.43±4.99, 34.87±6.66, 44.99± 5.98 in day four and six 217 respectively. The plant P. fraternus was observed to show intrinsic antiplasmodial activity by 218 its percentage chemosuppressions (%C) (figure 2) and even curative ability as compared to 219 that of the standard drug (coartem) but the relatively higher potency of the standard drug 220 (coartem) was not surprising since it is a first line drug used in treatment of malaria, its active 221 constituents are in refined state as compared to the crude extracts of the plants [25-27]. 222

223 Generally, the low antiplasmodial activity could be attributed to the crude nature of the extracts. The result (table 3) showed that the aqueous extract work better than the ethanol 224 extract the concentrations. The low percentage chemosuppression (%C) of the ethanol extract 225 226 could be as a result of the poor solubility nature of the active components in the organic solvent and also the extract contain possible antagonistic compounds that hinders the activity 227 of the active ones and increasing the concentration of the extract also increases the 228 antagonistic components thereby reducing the activity of the extract. Further investigations 229 are warranted to ascertain the exact mechanisms by which *P. fraternus* aqueous extract exerts 230 231 these effects. Nevertheless, these findings lend some information to the use of P. fraternus aqueous and ethanol extracts in the management of antiplasmodial activity. 232

233 Conclusion

P. fraternus aqueous and ethanol crude plant extracts from the results exhibited
antiplasmodial activity, thus supporting its traditional use in the management of malaria. A
product formulated from the plant could be beneficial as adjunct therapy for management of *plasmodial* infections in Ghana.

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