

1 **An *in vivo* antiplasmodial activity of aqueous and ethanol crude plant extracts of**
2 ***Phyllanthus fraternus* using *plasmodium berghei* infected balb/c mice**

3 **Keywords**

4 *In vivo*, antiplasmodial activity, *Phyllanthus fraternus*, phytochemicals, chemosuppression,
5 *Plasmodium berghei*.

6
7 **Abstract**

8
9 **Background:** *Phyllanthus fraternus* is a tropical plant that has numerous pharmacological
10 activities such as blennorrhagia, colic, diabetes, dysentery, fever, flu, tumours, jaundice,
11 vaginitis, dyspepsia, anti-inflammatory, antioxidant, anticoagulant, anti-diabetic, antiviral and
12 analgesic. The study evaluated *in vivo* anti-plasmodial activity of aqueous and ethanol crude
13 plant extracts of *Phyllanthus fraternus* using *Plasmodium berghei* infected *Balb/c* mice.

14 **Methodology:** The preparation of the aqueous crude extract was done by boiling 195 g of the
15 dried plant material in 4 L of water for 30 minutes and cooled. The resultant extract was
16 filtered through a cotton wool and put in an oven at 50 °C to concentrate it before it was pre-
17 freeze and lyophilized into powder using a freeze dryer (Heto powder dry LL 300, Sapa).
18 Similarly the preparation of the ethanol crude extract was obtained by simple maceration of
19 195 g of dried sample of the plant in 2 L aqueous ethanol (1.4 L of ethanol plus 0.6 L of
20 distilled water) for 72 h. It was then filtered through cotton wool and subjected to rotary
21 evaporator (ILA CCA-1111 Japanese branch) to evaporate the ethanol and then pre-freeze
22 and freeze- dried. The crude extracts were screened for their phytochemical constituents
23 which showed the presence of secondary metabolites. The LD₅₀ of both extracts were
24 investigated using Sprague-Dawley rats and found to be greater than 5000 mg/kg. The *in*

25 *vivo* antiplasmodial activity (percentage parasitaemia (%P) and the percentage chemo-
26 suppression (%C)) of the extracts were evaluated using *Balb/c* mice.

27 **Results:** The aqueous and ethanol extracts established modest antiplasmodial activity in a
28 dose dependent manner. The standard drug (coartem 2 mg/kg) with percentage parasitaemia
29 (%P) of 28.57 ± 4.70 and 2.48 ± 0.48 caused percentage chemosuppression (%C) of 44.38 ± 7.63
30 and 81.27 ± 2.07 in day four and six respectively. The test groups (aqueous and ethanol
31 extracts) for two different doses (100 mg/kg and 200 mg/kg) each administered with
32 percentage parasitaemia (%P) 39.67 ± 1.35 , 39.58 ± 1.64 , 37.32 ± 2.37 , 36.23 ± 1.99 and
33 10.24 ± 1.32 , 9.33 ± 0.66 , 8.61 ± 0.96 , 7.27 ± 1.26 caused percentage chemosuppressions (%C) of
34 22.78 ± 2.20 , 22.96 ± 2.66 , 27.35 ± 3.84 , 29.48 ± 3.23 and 22.54 ± 9.93 , 29.43 ± 4.99 , 34.87 ± 6.66 ,
35 44.99 ± 5.98 in day four and six respectively. The aqueous extract demonstrated better
36 inhibition of *plasmodium* in doses 100 mg/kg and 200 mg/kg with chemosuppressions (27.35
37 ± 3.84 and 29.48 ± 3.23) respectively compared with the ethanol extract of the same doses
38 100 mg/kg and 200 mg/kg with chemosuppressions (22.78 ± 2.20 and 22.96 ± 2.66)
39 respectively. The activity of the standard drug, coartem at 2.0 mg/kg was significantly higher
40 ($p < 0.05$) with chemosuppression (44.38 ± 7.63) than those of the extracts. The extracts were
41 also screened for phytochemicals for which some were found in the extracts which have
42 previously been implicated as antiplasmodial agents. The LD_{50} of both extracts were
43 investigated and found to be greater than 5000 mg/kg.

44 **Conclusion:** The aqueous and ethanol crude plant extracts of *P. fraternus* possess
45 antiplasmodial activity and would be useful in the search for novel antimalarial agents.

46 **Introduction:**

47 The plant *Phyllanthus fraternus* belongs to the Family *Euphorbiaceae* and is commonly
48 called gulf leaf-flower, Chancapiedra, stone breaker, carry-me-seed, hurricane weed or

49 quinine weed. The plant also has local names such as Mache da goyo (Hausa), Gbogbonowun
50 lese (Yoruba), Ofobi okpabi (Krobo and Ga), Lume or Kpavideme (Ewe),
51 Awommaaguwakayi (Twi) [1]. It is an annual dicotyledonous herb which is small, erect and
52 grow in gutters, dumping places and along the road of 30 to 40 cm in height [2]. Traditional
53 herbalist in Ghana uses the whole plant for numerous pharmacological activities such as
54 blennorrhagia, colic, diabetes, dysentery, fever, flu, tumors, jaundice, vaginitis, and dyspepsia
55 [3]. From literature *Phyllanthus fraternus* possesses anti-inflammatory [1], antioxidant and
56 anticoagulant [2], antidiabetic [4-5], antiviral [6] and analgesic properties [7-8]. Through bites
57 of female *Anopheles* mosquitoes a parasite called *Plasmodium* species are transmitted into
58 human which result in malaria disease [9]. Antiplasmodial activity of different species of the
59 genus *Phyllanthus* have been determined elsewhere [10], but as far as literature can tell no
60 work have been done on an *in-vivo* of the aqueous and ethanol whole plant extracts of *P.*
61 *fraternus* against malaria. As a matter of fact, the existing orthodox drugs have lots of side
62 effects and the most efficacious among them are now becoming impotent to the parasite and
63 there is a need to research on new antimalarial plants (*P. fraternus*) [1].

64 *Plasmodium berghei* infected balb/c mice were employed in this study because they have
65 similar properties of genetics, anatomy and physiology with humans in terms of experimental
66 research. Especially mice are used due to their similarity of genomes that mimics humans and
67 also their cost effective. The other types of mammals normally used for animal model
68 experiments are rodents and these include; rats, gerbils, guinea pigs and hamsters. [11]. Even
69 though there is an advancement into modern medicines, underdeveloped countries still rely
70 massively on medicinal plants for their survival during disease attack. To get rid of malaria
71 infection in the underdeveloped countries, the World Health Organization aimed to include
72 traditional medicine for its preventive approach. Many medicinal plants have been employed
73 on the basis of their antimalarial properties by traditional herbalists but their effectiveness

74 have not been scientifically assessed [12]. The Herbalist in Ghana documented the plant *P.*
75 *fraternus* as antimalarial drug but as far as literature can ascertain, it had not been
76 scientifically assessed. There was no much adverse effects assigned to medicinal plants since
77 its existence and are also believed to be significant in terms of new source of chemical
78 substances with a therapeutic effects. Therefore this study aimed to evaluate an *in vivo*
79 antiplasmodial activity of aqueous and ethanol crude plant extracts of *P. fraternus* on *P.*
80 *berghei* infected *Balb/c* mice.

81

82 **Materials and Methods**

83 **Drugs and chemicals**

84 All drugs and chemicals used such as tetraoxosulphate (vi) acid (H_2SO_4), ammonium
85 hydroxide (NH_3OH (aq)), magnesium ribbon, 2 mL of hydrochloric acid (HCl), chloroform,
86 ammonia, ferric chloride, acetone, sodium picrate paper, Fehling solution A and B, 70%
87 ethanol, Giemsa stain, methanol and sodium chloride, were obtained from British Drug
88 House Ltd (Poole, England). Coartem was obtained from Troge Medical GMBH (Hamburg
89 Germany) were all of analytical grade unless otherwise stated.

90 **Plant Raw Materials and Herbal Standard**

91 *Phyllanthus fraternus* whole plant material (leaves, stems and roots) were obtained from the
92 Plant Production Department (PPD), of Centre for Scientific Research into Plant Medicine
93 (CSRPM) Mampong-Akuapem, Ghana and authenticated by Dr. Yaw Ameyaw, a botanist of
94 the production department.

95 **Animals**

96 Seven-week old female *Balb/c* mice (30 g) were obtained from the animal unit of the Centre
97 for Scientific Research into Plant Medicine (CSRPM), Mampong-Akuapem, in the Eastern

98 Region of Ghana. The animals were fed on powdered feed obtained from Ghana Agro Food
99 Company (GAFCO), Tema, Ghana. They were allowed free access to sterile distilled water.

100 **Preparation of Herbal Extracts**

101 The plant material was cut into small pieces and spread thinly on a flat clean tray to prevent
102 spoilage by moisture condensation and allowed to dry at room temperature for three (3) days.
103 The dried plant material (195 g) was boiled in 4 L of water for 30 minutes and cooled. The
104 resultant extract was filtered through a cotton wool and put in an oven at 50 °C to concentrate
105 it before it was pre- freeze and lyophilized into powder using a freeze dryer (Heto powder dry
106 LL 300, Sapa). The dry powder was weighed to determine the yield and stored in a desiccator
107 at room temperature. This was reconstituted in sterilized distilled water before use. 70%
108 ethanol extract was obtained by simple maceration of 195 g of dried sample of whole plant of
109 *P. fraternus* in 2 L aqueous ethanol (1.4 L of ethanol plus 0.6 L of distilled water) for 72 h. It
110 was filtered through cotton wool and subjected to rotary evaporator (ILA CCA-1111
111 Japanese branch) to evaporate the ethanol and then pre-freeze and freeze- dried.

112 **Malaria parasites and inoculum preparation**

113 *Plasmodium berghei* NK65 strain from the University of Copenhagen Denmark through the
114 Department of Immunology, Noguchi Memorial Institute of Medical Research (NMIMR),
115 University of Ghana, Accra, Ghana, was used for the experiment. The stock of parasitized
116 erythrocytes was obtained from infected Balb/c mice, with a minimum peripheral
117 parasitaemia of 20%, by cardiac puncture in heparin-coated tube. The cell concentration of
118 the stock was determined and diluted with physiological saline such that 0.2 mL of final
119 inoculum contained 10⁶ parasitized red blood cells (RBCs).

120

121 **Acute toxicity test**

122

123 The acute oral toxicity study was conducted to know the amount of dose to be given to the
124 animals. This was done by the Organization for Co-operation and Development (OECD)
125 guidelines 425 received from the Committee for the Purpose of Control and Supervision of
126 Experiments on Animals (CPCSEA) [11].

127 **Treatment of Animals**

128 Thirty six (36) mice were selected and put into six (6) groups of six per group. Each mouse
129 was inoculated intraperitoneal with the parasite *P. berghei*. Group 1 (Gp1) animals received
130 distilled water (negative control), group 2 (Gp 2) animals received 2 mg Coartem (positive
131 drug control), group 3 (Gp 3) and group (Gp 4) animals received 100 mg/kg and 200 mg/kg
132 of ethanol extract of whole plant of *P. fraternus* respectively, group 5 (Gp 5) and group 6
133 (Gp6) animals received 100 mg/kg and 200 mg/kg aqueous extract of whole plant of *P.*
134 *fraternus* respectively. All the drugs were orally administered to the animals (0.2 mL) 2-3 h
135 after the mice have been inoculated with the parasite over a period of 6 days.

136 **Monitoring of Parasitaemia and Antimalarial Activity**

137 On the fourth and sixth days after drug administration, thin blood smears were prepared using
138 blood from the tail vein of each mouse. Each smear was air-dried, fixed in methanol, air-dried
139 again, stained with 10% Giemsa for 10-15 minutes and examined under oil immersion with a
140 microscope. Each slide was observed at three different fields and the Red Blood Cells
141 (RBC_s) and total number of RBC_s for each field was recorded. The percentage parasitaemia
142 (% P) and the percentage chemo-suppression (% C) also known as the activity was estimated

143 according to the following formulae $\% P = 100 \times \frac{PRBC - TRBC}{TRBC}$ $\% C = 100 \times \frac{PCON - PTEST}{PCON}$

144 Where; *PRBC* is the number of parasitized Red Blood Cell (RBC).

145 *TRBC* is the total number of RBC counted per field.

146 *PCON* is the control parasitemia and *PTEST* is the test parasitemia.

147 **Statistical analysis**

148 Data were presented as means \pm SEM of n= 6 and analyzed using One-way ANOVA which
 149 was followed by students t-test. The $P \leq 0.05$ was considered statistically significant in all
 150 analysis.

151 **Results**

152 Phytochemical screening was carried out for aqueous and ethanol whole plant extracts of
 153 *Phyllanthus fraternus* which identified the presence and absence of groups of secondary
 154 metabolites using the standard method [13-14].The phytochemical screening of the extracts
 155 showed the presence of alkaloids, saponin, phenolics, reducing sugars, triterpenes and
 156 phytosterols in both extracts while cyanogenic glycoside and anthraquinones were absent in
 157 both extracts and flavonoids and polyuronides showed presence only in the aqueous extract
 158 (Table 1).

159

160 **Table 1: Phytochemical constituents of *Phyllanthus fraternus* whole plant extracts.**

| PHYTOCHEMICAL | EXTRACTS | |
|----------------|----------|---------|
| | Aqueous | Ethanol |
| Alkaloids | + | + |
| Saponins | + | + |
| Phenolics | + | + |
| Reducing Sugar | + | + |
| Polyuronide | + | - |
| Terpenoids | + | + |
| Flavonoids | + | - |
| Phytosterols | + | + |

| | | |
|----------------------|---|---|
| Anhthracenoside | - | - |
| Cyanogenic Glycoside | - | - |

161 (+) = Present and (-) = Absent

162

163 **Acute toxicity test**

164 The LD₅₀ of the extracts were identified and was greater than 5000 mg/kg and may be
 165 classified as practically non-toxic and within the acceptable margin of safety (Hodge and
 166 Sterner scale) at the recommended dose. Thus 1/50th and 1/25th (i.e.100 mg/kg and 200
 167 mg/kg) were selected for the study (Table 2).

168 **Table 2: Acute toxicity test for *Phyllanthus fraternus* whole plant of aqueous and**
 169 **ethanol extracts**

| | <i>Phyllanthus fraternus</i> whole plant | |
|--|--|---------------------|
| | Aqueous extract | Ethanol extract |
| Species and strain | Sprague-Dawley rats | Sprague-Dawley rats |
| Number of animals | Twelve (12) | Twelve (12) |
| Sex | Females | Females |
| Number. of groups | 3 (N=4) | 3 (N=4) |
| Route of administration | Oral | Oral |
| Formulation | Freeze dried | Freeze dried |
| Dose administered (mg/kg) | 1250, 2500, 5000 | 1250, 2500, 5000 |
| Period of observation | 48 hours | 48 hours |
| Number. of deaths | Zero (0) | Zero (0) |
| Approximate lethal dose(LD ₅₀) | >5000 mg/kg | >5000 mg/kg |
| Signs of toxicity | Nil | Nil |

170

171 **Percentages of Parasitaemia and Chemo-suppression of *Phyllanthus fraternus* whole**
172 **plant of aqueous and ethanol extracts in 4th and 6th days test**

173

174 The route of administration of the controls (Coartem and distilled water) were done at doses
175 of 2 mg/kg orally; aqueous and ethanol crude plant extracts of *P. fraternus* were given orally
176 at doses of 100 mg/kg and 200 mg/kg which significantly exerted *in vivo* antiplasmodial
177 activity on the *P. berghei* infected *Balb/c* mice in a dose-dependent fashion at day 4 and day
178 6 except ethanol crude extract at the dose of 100 mg/kg for day 4 and day 6 (Table 3).

179 **Table 3: Results of Percentage Parasitaemia and Chemosuppression of 4 and 6 days test**

| Extracts | Day four | | Day six | |
|-----------------------|------------------|----------------------|------------------|----------------------|
| | Parasitaemia (%) | Chemosuppression (%) | Parasitaemia (%) | Chemosuppression (%) |
| Concentration (mg/kg) | | | | |
| Control | 61.64±3.77 | 0.00 | 13.22±2.32 | 0.00 |
| Coartem 2 | 28.57±4.70 | 44.38±7.63 | 2.48±0.48 | 81.27±2.07 |
| PET 100 | 36.23±1.99 | 29.48±3.23 | 10.24±1.32 | 22.54±9.93 |
| PET 200 | 37.32±2.37 | 27.35±3.84 | 9.33±0.66 | 29.43±4.99 |
| PAQ 100 | 37.32±2.37 | 27.35±3.84 | 10.24±1.32 | 34.87±6.66 |
| PAQ 200 | 39.67±1.35 | 22.78±2.20 | 7.27±1.26 | 44.99±5.98 |

180

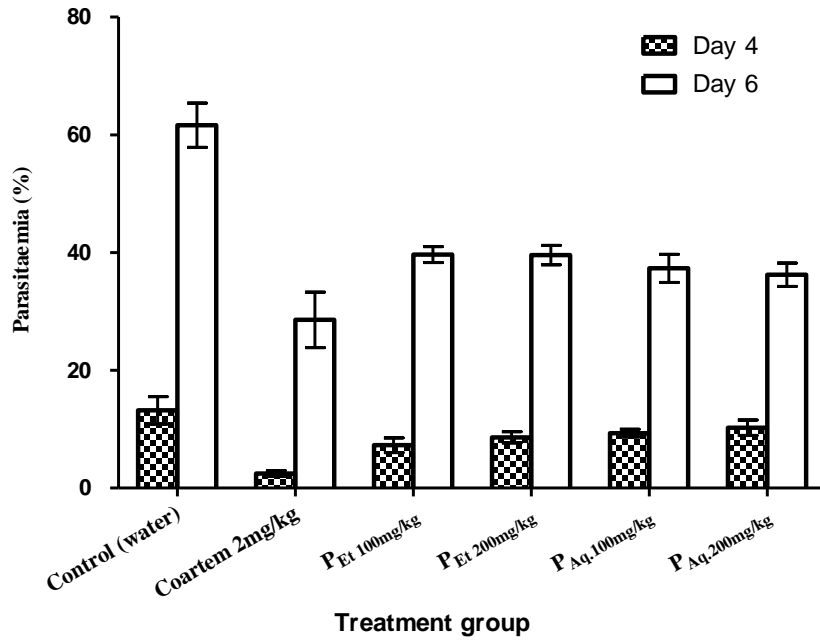
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182

183 **Graphs of Percentages of Parasitaemia and Chemo-suppression of *Phyllanthus fraternus***
184 **whole plant of aqueous and ethanol extracts in 4th and 6th days test**

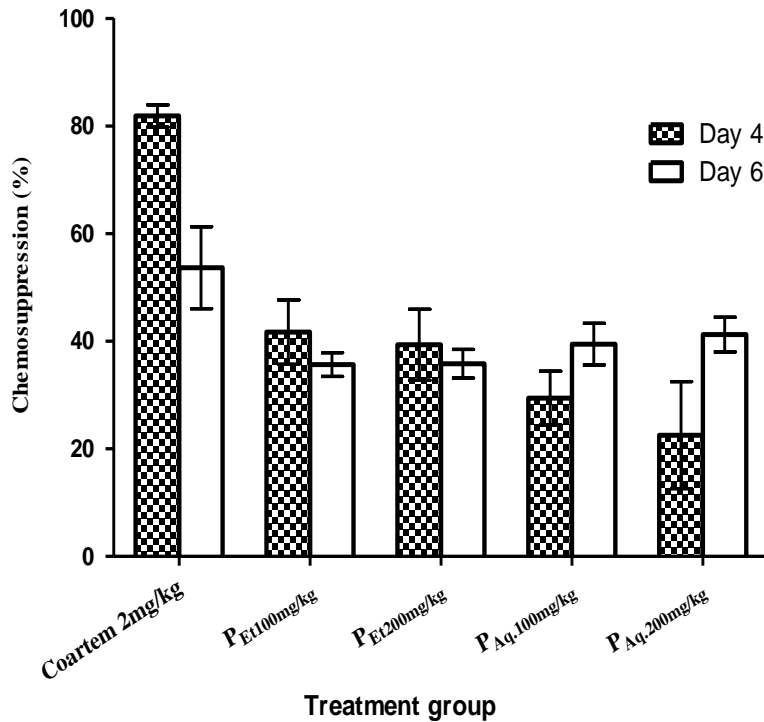
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186 The results obtained from Percentage Parasitaemia and Chemo-suppression of 4 and 6 days
187 test (Table 3) were represented graphically where PAq = Aqueous extract of *Phyllanthus*
188 *fraternus*, PEt = Ethanol extract of *Phyllanthus fraternus*, Results are means ± SEM of n= 6,
189 * = Values significantly different from Distilled water controls (p<0.050) and # = Value
190 significantly different from positive controls (p<0.050) (Figure 1 and 2).



191

192 Figure 1: Graph of the percentage parasitaemia of *Plasmodium berghei* infected balb/c mice at
 193 day four and six.



194

195 Figure 2: Graph showing the percentage Chemo-suppression of *Plasmodium berghei* infected
 196 *Balb/c* mice at day four and six.

197

198 **Discussions**

199 The phytochemical screening of the extracts showed the presence of alkaloids, saponin,
200 phenolics, reducing sugars, triterpenes and phytosterols in both extracts while cyanogenic
201 glycoside and anthraquinones were absent in both extracts and flavonoids and polyuronides
202 showed presence only in the aqueous extract. The result showed differences from reported
203 works by Sofowora; Olonisokan *et al.* [15-16]. The factors attributed to these differences were
204 as a result of environment of the plant, mode of extraction and the climatic conditions [17-
205 18]. Reports have shown that antiplasmodial activity of many agents were due to interference
206 with the reproductive system of the protozoa [19]. Several reports have implicated alkaloids
207 [20], terpenoids [21] and lignans [22-24] as antimalarial agents. The antiplasmodial activity
208 demonstrated by both extracts may be attributed to the presence of some of these
209 phytochemicals. The extracts showed modest antiplasmodial activity in a dose dependent
210 manner as manifested in the results (Table 3). The standard drug (coartem 2 mg/kg) with
211 percentage parasitaemia (%P) of 28.57 ± 4.70 and 2.48 ± 0.48 caused percentage
212 chemosuppression (%C) of 44.38 ± 7.63 and 81.27 ± 2.07 in day four and six respectively. From
213 (figure 1 and 2), the test groups (aqueous and ethanol extracts) for two different doses (100
214 mg/kg and 200 mg/kg) each administered with percentage parasitaemia (%P) of 39.67 ± 1.35 ,
215 39.58 ± 1.64 , 37.32 ± 2.37 , 36.23 ± 1.99 and 10.24 ± 1.32 , 9.33 ± 0.66 , 8.61 ± 0.96 , 7.27 ± 1.26
216 caused percentage chemosuppressions (%C) of 22.78 ± 2.20 , 22.96 ± 2.66 , 27.35 ± 3.84 ,
217 29.48 ± 3.23 and 22.54 ± 9.93 , 29.43 ± 4.99 , 34.87 ± 6.66 , 44.99 ± 5.98 in day four and six
218 respectively. The plant *P. fraternus* was observed to show intrinsic antiplasmodial activity by
219 its percentage chemosuppressions (%C) (figure 2) and even curative ability as compared to
220 that of the standard drug (coartem) but the relatively higher potency of the standard drug
221 (coartem) was not surprising since it is a first line drug used in treatment of malaria, its active
222 constituents are in refined state as compared to the crude extracts of the plants [25-27].

223 Generally, the low antiplasmodial activity could be attributed to the crude nature of the
224 extracts. The result (table 3) showed that the aqueous extract work better than the ethanol
225 extract the concentrations. The low percentage chemosuppression (%C) of the ethanol extract
226 could be as a result of the poor solubility nature of the active components in the organic
227 solvent and also the extract contain possible antagonistic compounds that hinders the activity
228 of the active ones and increasing the concentration of the extract also increases the
229 antagonistic components thereby reducing the activity of the extract. Further investigations
230 are warranted to ascertain the exact mechanisms by which *P. fraternus* aqueous extract exerts
231 these effects. Nevertheless, these findings lend some information to the use of *P. fraternus*
232 aqueous and ethanol extracts in the management of antiplasmodial activity.

233 **Conclusion**

234 *P. fraternus* aqueous and ethanol crude plant extracts from the results exhibited
235 antiplasmodial activity, thus supporting its traditional use in the management of malaria. A
236 product formulated from the plant could be beneficial as adjunct therapy for management of
237 *plasmodial* infections in Ghana.

238 **Ethical Approval:**

239

240 As per international standard or university standard written ethical approval has been collected and
241 preserved by the author(s).

242

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