

Hematological and Biochemical Changes on Antidepressants Amitriptyline Induced Cardiac Toxicity in Male Rats

ABSTRACT

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Objects: Amitriptyline is a widely used tricyclic antidepressant. Amitriptyline is well-known for its cardiovascular side effects and toxicity in psychiatric patients. However, the mechanisms underlying the cardiovascular side effects of amitriptyline remain largely undefined.

Aims: This study aim to show the hematological and biochemical changes on Amitriptyline induced cardiac toxicity in male rats.

Methodology: A total of 20 male albino rats were equally divided into two groups: the first was control and the second was amitriptyline intoxicated group.

Results: A total of 20 male albino rats were equally divided into two groups: the first was control and the second was amitriptyline intoxicated group. Our results revealed that; a significant increase in sodium ions, alkaline phosphatase, AST, lipid profiles (cholesterol, triglycerides, LDL, HDL), and cardiac enzymes (CK-Mb, CPK, LDH, myoglobin) and in significant decrease in platelets, white and red blood cells, potassium ions, and total proteins in treated rats with amitriptyline as compared to control.

Conclusion: Amitriptyline toxicity is life-threatening and can cause acute myocarditis in addition to the known cardio toxic profile of tricyclic anti-depressant medications. Physicians should be aware of this rare entity as a differential diagnosis for myocarditis with an unknown etiology.

Keywords: Amitriptyline; Antidepressant; cardiac enzymes; Lipid profiles; Electrolytes; Rats.

1. INTRODUCTION

Cardiac dysfunction is the manifestation of multifactorial diseases, such as chronic heart failure, renal failure, metabolic disorders and cancer [1, 2, 3, 4, 5]. No other disease affecting humans is more severe and is associated with a greater mortality rate than psychiatric patients [6]. Many antidepressants have unwanted cardiovascular effects, so choosing between them may not be straightforward in patients with heart disease. Amitriptyline is one of the classical and typical tricyclic antidepressants which are work on central nervous system to increase the level of certain chemicals in your brain [7]; Amitriptyline is also currently used for the treatment of patients with migraine [8].

Amitriptyline can cause dizziness and drowsiness during the first few hours after you take it [9]. Amitriptyline acts as a serotonin-norepinephrine reuptake inhibitor, thereby increasing the concentration of these transmitters in the synapse [10]. Given the wide use of amitriptyline as an antidepressant medication, it is important to discuss this case of its overdose presenting as myocarditis with pericardial involvement and provide a brief review of amitriptyline-induced cardio toxicity [11]. Therefore, the aim of this study is to show the changes in some hematological and biochemical parameters after the treatments of male rats with antidepressant amitriptyline.

2. MATERIAL AND METHODS

2.1 Experimental animals

The experiment was performed on 20 male albino rats (*Rattus norvegicus*) weighing 150 g (± 10) and of 9-10 weeks' age. They were obtained from animal house of National Research Center (Dokki, Giza, Egypt). The rats were housed in

suitable plastic cages for one week before the experimental work for acclimation with a new room conditions and maintained on a standard rodent diet, with water available ad libitum. During the experiment animal behavior was noticed and body weight at the beginning and the end of experiment were measured. Animal maintenance and treatments were conducted in accordance with the Faculty of Science, Tanta University guide for animal, as approved by Institutional Animal Care and Use Committee (IACUC-SCI-TU-0050).

2.2 Experimental groups

The rats were randomly and equally divided into 2 groups (10 rats each). G1: control group that included animals that did not receive any treatment during the experimental period. G2: Amitriptyline (Tryptizol; El Kahira Pharm And Chem Ind Co) group in which rats were injected intraperitoneally with Amitriptyline (70 mg/kg body weight/daily) for four weeks [12].

2.3 Blood and serum samples

Heparinized blood was collected from the inferior vena cava for complete blood picture (CBC) according to Basuony [13]. Serum was collected from the inferior vena cava and separated by centrifugation at 3000 rpm for 15 minutes. The collected serum was stored at -18° C until analysis for estimation of some blood parameters (CK-MB, CPK, LDH, Myoglobin, Cholesterol, LDL, HDL, Tg, total protein, Na and K).

In keeping with the approach proposed by Whitaker [14], kits from Vitro Scient (Cairo, Egypt) were used to undertake the kinetic technique for measurement of the activity of serum lactate dehydrogenase (LDH). Meanwhile, the approach recommended by Zilva and Pannall [15] was adopted to perform an akinetic technique with kits from the same company (Vitro Scient) to measure serum levels of creatine kinase (CK). The approach suggested by Bishop et al. [16] was applied with the assay kit provided by Bioassay Systems (Hayward, CA, USA) to measure the serum levels of creatine-myoglobin (CK-MB). Last but not least, the approach proposed by Cummins et al. [17] was used with the assay kit provided by ReactivosSpinreact (Girona, Spain) to measure the serum levels of myoglobin.

The activities of serum AST were assayed by the colorimetric method according to Reitman and Frankel [18]. Serum ALP activity was measured according to Belfield and Goldberg [19] by using commercial kit. Serum levels of total protein were determined by using commercial kit according to Bowers and Wong [20]. Serum potassium, sodium ions levels were determined by using commercial kits (Sensa core electrolyte, India) according to Abd Eldaim et al. [21]. The concentration of cholesterol, triglyceride, high-density lipoprotein-cholesterol (HDL-C) and low density lipoprotein-cholesterol (LDL-C) were determined with Kits from ELLTECH according to Aldubayan et al. [22].

2.4 Statistical Analysis

Data were expressed as mean values \pm SE and statistical analysis was performed using unpaired t-test to assess significant differences among treatment groups. The criterion for statistical significance was set at $p < 0.05$ for the biochemical data. All statistical analyses were performed using SPSS statistical version 21 software package (SPSS® Inc., USA).

3. RESULTS

3.1 Toxicity

Many of side effects were appeared on rats after Amitriptyline administration such as weakness, loss of activity, increased perspiration, and increased or decreased appetite.

3.2 Biochemical investigations

Table (1) revealed that; CK-MB, CPK, LDH, and myoglobin levels in serum increased ($P < 0.05$) in the amitriptyline group compared with the control group. As well, AST and ALP levels in serum increased ($P < 0.05$) in the amitriptyline group compared with the control group (Table 1). Also, groups intoxicated by amitriptyline showed a significant increase in levels of serum cholesterol at ($p < 0.0001$), serum LDL at ($P < 0.0001$), serum HDL at ($p < 0.0001$) and serum triglycerides at ($p < 0.0001$) compared with control group (Table 2). At the same time, total protein levels decreased ($P < 0.05$) in the amitriptyline group compared with the control group (Table 3). Table 3 showed that; R.B.C.s, W.R.C.s and PLT indices levels in Amitriptyline group were poorly affected and showed low significant decrease when compared with control group.

Table 1. Changes in serum cardiac enzymes (CK-MB, CPK, LDH, Myoglobin, Aspartate transaminase (AST) and alkaline phosphatase (ALP) levels in experimental groups.

Particulars	Control group	Amitriptyline group
CK-MB(ng/ml)	0.1382 \pm 0.001****	0.2958 \pm 0.004
CPK(U/l)	3517 \pm 75.08****	5174 \pm 186.7
LDH10-5(U/l)	172.8 \pm 13.75****	565.8 \pm 24.87
Myoglobin (ng/ml)	13.88 \pm 0.259****	17.78 \pm 0.128
AST (U/L)	154.6 \pm 4.366****	281.4 \pm 5.105

* The significant difference was analyzed by T-test unpaired. Values are expressed as mean \pm SEM. T-test was significant at $p < 0.05$. T-test unpaired was significant from corresponding amitriptyline at NSP=0.1234; *P=0.0332; **P=0.0021; ***P=0.0002; ****P<0.0001. G1, control group; G2, Amitriptyline group.

Table 2. Changes in serum lipid profiles (cholesterol, Tg, HDL, LDL and VLDL) levels in experimental groups.

Particulars	Control group	Amitriptyline group
Cholesterol (mg/dl)	93.2±1.772****	178±5.376
Tg (mg/dl)	110.4 ± 1.631****	191.2 ± 4.913
HDL (mg/dl)	14.2 ± 0.3742****	19.6 ± 0.6
LDL (mg/dl)	54.92 ± 1.883****	127 ± 4.122
VLDL (mg/dl)	22.08 ± 0.3262****	38.24 ± 0.9826

* The significant difference was analyzed by T-test unpaired. Values are expressed as mean± SEM. T-test was significant at $p<0.05$. T-test unpaired was significant from corresponding amitriptyline at NSP=0.1234; *P=0.0332; **P=0.0021; ***P=0.0002; ****P<0.0001. G1, control group; G2, Amitriptyline group.

Table 3. Changes in serum total protein, sodium ions (Na⁺), potassium ions (K⁺), RBCs, WBCs and PLTs levels in experimental groups.

Particulars	Control group	Amitriptyline group
Total protein(g/dl)	7.01 ± 0.074****	5.326 ± 0.058
Na (mEq/l)	136 ± 0.257****	152.3 ± 2.29
K (mEq/l)	5.054 ± 0.0206****	4.132 ± 0.0356
R.B.Cs (mill./cmm)	7.14 ± 0.27*	6.09 ± 0.31
W.B.Cs (103UI)	6.35 ± 0.50*	3.68 ± 0.36

* The significant difference was analyzed by T-test unpaired. Values are expressed as mean± SEM. T-test was significant at $p<0.05$. T-test unpaired was significant from corresponding amitriptyline at NSP=0.1234; *P=0.0332; **P=0.0021; ***P=0.0002; ****P<0.0001. G1, control group; G2, Amitriptyline group.

4. DISCUSSION

Amitriptyline overdose may cause significant central nervous system effects, the presumed mechanism of action in overdose deaths is cardiac in nature, involving tachycardia, widening of the qrs complex and various arrhythmias [23]. Consequently, care must be taken when interpreting postmortem amitriptyline levels. Deaths related to the toxic effects of amitriptyline are typically suicidal in nature [24].

In this study these results were in accordance with boles et al. [25] who showed that the amitriptyline doses have different effects on enzymatic levels. The higher dose (25 mg/kg) increased the creatine phosphokinase, creatine kinase-mb, total protein, lactate dehydrogenase and myoglobin levels. Experimentally, acosta and ramos [26] have shown that amitriptyline induced severe abnormalities of the sarcolemma integrity using ldh release of cultured myocardial cells as a criterion for cell damage. Also, in current study there was significantly increase in cholesterol, triglycerides and ldl and an increase in hdl levels in amitriptyline group (g2).

Gurer et al. [27], who explained that amitriptyline, affect the heart muscle due to changes in the level of cholesterol and triglycerides. Also, kaur et al. [28] showed that amitriptyline may be responsible for cardiac dysfunction. Concentrations of cholesterol and triglycerides in the serum they were significantly elevated in rabbits managed antidepressants involved. This study shows that enzymes ast and alp were significantly elevated by amitriptyline intoxication in amitriptyline group (g2). Findings of this study showed significant elevation in serum alkaline phosphatase in rat groups intoxicated by amitriptyline (g2).

In this study showed the elevation levels of serum aspartate aminotransferase (ast) under the effect of amitriptyline as a marker cytotoxicity in groups under study which supported by the result estimated by afify et al., [29]. The results of this study showed the effect of amitriptyline poisoning on the blood, different parameters of blood parameters. Red blood cell indices showed a significant reduction in the amitriptyline group (g2). In addition, the results showed that amitriptyline could cause thrombocytopenia and a decrease in white blood cell count in amitriptyline group (g2), unlike control group (g1). The results were consistent with tousson et al. [12]. The higher dose of amitriptyline increased concentrations of Na, k levels in serum this is consistent with previous studies thorstrand et al. [30].

5. CONCLUSION

Amitriptyline toxicity is life-threatening and can cause acute myocarditis in addition to the known cardiotoxic profile of tricyclic anti-depressant medications. Physicians should be aware of this rare entity as a differential diagnosis for myocarditis with an unknown etiology.

COMPETING INTERESTS

There is no conflict of interest

ETHICAL APPROVAL

"All authors hereby declare that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985) were followed, as well as specific national laws where applicable. All experiments have been examined and approved by the appropriate ethics committee"

REFERENCES

1. Tousson E, Hafez E, Massoud A, Sweef O , Atta N. Protective role of folic acid in thyroxine-induced cardiac hypertrophy in hyperthyroid rat. Biomedicine & Aging Pathology. 2013; 3:89–95.
2. Tousson E, Hafez E, Zaki S, Gad A. The cardioprotective effects of L-carnitine on rat cardiac injury, apoptosis, and oxidative stress caused by amethopterin. Environ Sci Pollut Res. 2016; 23:20600–20608.
3. Tousson E, Elgharabawy RM, Elmasry TA. Grape seed proanthocyanidin ameliorates cardiac toxicity induced by boldenone undecylenate through inhibition of nadph oxidase and reduction in the expression of NOX2 and NOX4. Oxidative Medicine and Cellular Longevity. 2018; Article ID 9434385, 12 pages. <https://doi.org/10.1155/2018/9434385>
4. Massoud A, El-Atrash A, Tousson E, Ibrahim W and Abou-Harga H. Light and ultrastructural study in the propylthiouracil - induced hypothyroid rat heart ventricles and the ameliorating role of folic acid. Toxicology and Industrial Health. 2012; 28(3):262–270.
5. Hafez E, Tousson E. Thyroxine-induced cardiac hypertrophy: Role of ascorbic acid in treatment. Biomedicine & Aging Pathology. 2014; 4(2):161-7.

157 6. Glassman AH. Cardiovascular effects of tricyclic antidepressants. *Annu Rev Med.* 1984; 35:503–11.

158 7. Gur A, Oktayoglu P. Central nervous system abnormalities in fibromyalgia and chronic fatigue syndrome: new concepts in
159 treatment. *Current pharmaceutical design.* 2008; 14(13):1274-1294.

160 8. Magalhães E, Menezes C, Cardeal M, Melo A. Botulinum toxin type A versus amitriptyline for the treatment of chronic daily
161 migraine. *Clinical neurology and neurosurgery.* 2010; 112(6):463-466.

162 9. Graud G, Lantri-Minet M, Lucas C, Valade D. French Society for the Study of Migraine Headache (SFEMC, French guidelines for
163 the diagnosis and management of migraine in adults and children. *Clinical therapeutics.* 2004; 26 (8):1305-1318.

164 10. Stahl SM. Stahl's essential psychopharmacology: neuroscientific basis and practical applications. Cambridge university press.
165 2013.

166 11. Thamer K, Toufik M, Amandeep R, Amjad K, Ahmad Q, Mohammad S, et al. A Case of Amitriptyline-induced Myocarditis.
167 *Cureus.* 2018; 10(6).

168 12. Tousson E, Zaki S, Hafez E, AG. Biochemical and immunocytochemical studies of the testicular alteration caused by
169 Amitriptyline in adult male rat. *Journal of Bioscience and Applied Research.* 2018 b; 4 (4):418-424.

170 13. Basuony M, Hafez E, Tousson E, Massoud A, Elsomkhraty S, Eldakamawy S. Beneficial role of Panax ginseng root aqueous
171 extract against Cisplatin induced blood toxicity in rats. *Am J Biol Chem.* 2015; 3 (1):1-7.

172 14. Whitaker J. A general colorimetric procedure for the estimation of enzymes which are linked to the NADH/ NAD⁺ system.
173 *Clinica Chimica Acta.* 1969; 24(1):23–37.

174 15. Zilva J, Pannall P. Clinical chemistry in diagnosis and treatment. Lloyd-Luke, London. 1985.

175 16. Bishop C, Chu T, Shihabi Z. Single stable reagent for creatine kinase assay. *Clinical chemistry.* 1971; 17(6):548-550.

176 17. Cummins P, Young A, Auckland M, Michie C, Stone P, Shepstone B. Comparison of serum cardiac specific troponin-I with
177 creatine kinase, creatine kinase-MB isoenzyme, tropomyosin, myoglobin and C-reactive protein release in marathon runners: cardiac
178 or skeletal muscle trauma?. *Eur J Clin Invest.* 1987;17:317–24.

179 18. Reitman S, Frankel S. Determination of glutamate-pyruvate transaminase (ALT) and aspartate aminotransfrase (AST). *J Clin*
180 *Pathol.* 1957; 28:1-56.

181 19. Belfield A, Goldberg D. Normal ranges and diagnostic value of serum 5' nucleotidase and alkaline phosphatase activities in
182 infancy. *Archives of disease in childhood.* 1971; 46(250):842-846.

183 20. Bowers L, Wong E. Kinetic serum creatinine assays. II. A critical evaluation and review. *Clinical chemistry.* 1980; 26 (5):555-
184 561.

185 21. Abd Eldaim MA, Tousson E, El Sayed IE, Awd WM. Ameliorative effects of Saussurea lappa root aqueous extract against
186 Ethephon-induced reproductive toxicity in male rats. *Environmental toxicology.* 2019; 34(2):150-9.

187 22. Aldubayan M, Elgharabawy R, Ahmed A, Tousson E. Antineoplastic Activity and Curative Role of Avenanthramides against
188 the Growth of Ehrlich Solid Tumors in Mice. *Oxidative medicine and cellular longevity.* 2019. <https://doi.org/10.1155/2019/5162687>

189 23. Dworkin R, Backonja M, Rowbotham M, Allen R, Argoff C, Bennett G, et al. Advances in neuropathic pain: diagnosis,
190 mechanisms and treatment recommendations. *Archives of neurology.* 2003; 60(11):1524-1534.

191 24. Prahlow J, Landrum J. Amitriptyline abuse and misuse. *The American journal of forensic medicine and pathology.* 2005;
192 26(1):86-88.

193 25. Boles R, Lovett-Barr M, Preston A, Li B, Adams K. Treatment of cyclic vomiting syndrome with co-enzyme Q10 and
194 amitriptyline, a retrospective study. *BMC neurology.* 2010; 10(1):10.

195 26. Acosta D, Ramos K. Cardiotoxicity of tricyclic antidepressants in primary cultures of rat myocardial cells. *Journal of Toxicology*
196 and *Environmental Health*. 1984; 14(2-3):137-143.

197 27. Gurer G, Sendur O, Ay C. Serum lipid profile in fibromyalgia women. *Clinical rheumatology*. 2006; 25(3):300-303.

198 28. Kaur H, Hota D, Bhansali A, Dutta P, Bansal D, Chakrabarti A. A comparative evaluation of amitriptyline and duloxetine in
199 painful diabetic neuropathy: a randomized, double-blind, cross-over clinical trial. *Diabetes care*. 2011; 34(4):818-822.

200 29. Afify M, Abd Elmaksoud M, Mosa T, Elshaer M, Kotb N. Differential effects of amitriptyline treatment on testicular and liver
201 functions in adult male rats. *New York Science Journal*. 2010; 3(3):10-18.

202 30. Thorstrand C, Bergström J, Castenfors J. Cardiac effects of amitriptyline in rats. *Scandinavian journal of clinical and laboratory*
203 *investigation*. 1976; 36(1):7-15.

204

205 31. Tousson E, Ali EM, Ibrahim W and Ashraf RM (2012): Histopathological and immunohistochemical alterations in rat heart after
206 thyroidectomy and the role of hemin and ketoconazole in treatment. *Biomedicine & Pharmacotherapy*. 66 (2012) 627–632.