Original Research Article

Nutritional and Phytochemical Characteristics of *Caesalpinioideae*Seeds

ABSTRACT

Aims: Caesalpiniaceae species have great medicinal and food values. In this work, six Caesalpiniaceae species:*Delonix regia, Entadagigas, Leucaena leucocephala, Mimosa pudica, Parkiajavanica*and *Senna siam*which grow abundantly in central India were selected for the chemical investigation.The objective of the present work is to describe phytochemical and mineralcomposition and bioaccumulation potentialities of six seeds derived from *Delonix regia, Entadagigas, Leucaena leucocephala, Mimosa pudica, Parkiajavanica*and *Senna siamea*.

Methodology: The spectrophotometric, enzymatic and X-ray fluorescence spectrophotometric technique were used for quantification of the polyphenol, starch and mineral, respectively.

Results: The sum of the total concentrations of17 macro- and micronutrients (P, S, Cl, K, Rb, Mg, Ca, Sr, Cr, Mn, Fe, Co, Cu, Zn, Se, Mo and Pb), oil and total starch in the six seeds were in the 20253-78489 mg/kg, 3.1-30.1% and 5.4–41.0% range, respectively. The highest concentrations of Fe, oil and phenolics were observed in *M. pudica* seeds. Both thermal and spectral characteristics allowed to differentiate *M. pudica* and *P. javanica* seeds (with the highest caloric contents) from the seeds from the other species.

Conclusion: Six Caesalpinioideae seeds i.e. *Delonix regia, Entadagigas, Leucaena leucocephala, Mimosa pudica, Parkiajavanica*and *Senna siamea*are potential sources of the nutrients (i.e. P, S, K, Mg, Ca and Fe) and polyphenols which are needed for biological metabolism and human health. The heavy metals are present under safe limits for their medicinal uses.

Keywords: Caesalpinioideae, starch, polyphenol, mineral, FTIR, thermal analysis.

1. INTRODUCTION

Caesalpinioideae is a subfamily of the Fabaceae family that includes 150 genera and 2500 species, which generally grow in tropical and sub-tropical regions (**1,2**). The seeds from six Caesalpinioideae species, common in central India, were selected for the study presented herein with a view to their valorization beyond their most frequent uses as sources of wood, of resin and gum, or of medicinal products.

Delonix regia (Bojer) Raf. (‘gulmohar’ or ‘flamboyant’) is a fast-growing tree that grows in most subtropical and tropical areas of the world and that is harvested for a range of uses, including medicines, timber, fuel and beads (**3-5**). Its seeds contain gum that is mainly used in the textile and food industries, but which is also being investigated for other applications (e.g., as a binder for the manufacture of tablets) (**6**).

Leucaena leucocephala (Lam.) de Wit (‘Subabul’ or ‘white Popinac’) is a perennial small tree mostly cultivated for fodder, as it is an excellent protein source (**7, 8**), but also as a bioenergy crop (**9, 10**). Its dried seeds can be also roasted and used as a coffee substitute due to emollient property.

Entadagigas(L.) Fawc. &Rendle (*Mimosa gigas* L., known as 'sea heart') is a perennial climbing shrub, known to be a rich source of saponins and commonly used for washing hair, clothes etc. (**11**). Its seeds and bark are astringent, and, together with its leaves, they have found numerous applications in Ayurvedic medicine.

Mimosa pudica L. is a creeping perennial herb, usually cultivated as a green manure and for soil stabilization, which is also used in folk medicine (**12-13**). Its applications as a source of bioactive products for pharmaceutical applications have been reviewed in (**14**).

*Parkiajavanica*Merr. ('tree bean' or 'khorial') isfound in most of South East Asian countries. Various parts of the plant are edible, and its bark and pods are used for treatment of various ailments, including intestinal disorders, bleeding piles, diarrhea and dysentery (**15, 16**).

Senna siamea(Lamarck) H.S.Irwin&Barneby (*Cassia siamea*Lam., 'black wood cassia') is a medium sized tree commonly planted in avenues and gardens, whose leaves can be used as manure and whose flowers are used as a vegetable. *Senna siamea* plays a key role in Jamu (Indonesia traditional medicine), as it possesses many medicinal properties (**17, 18**). Its chemical constituents and bioactivities have been reviewed in (**19**, **20**).

In this work, the nutritional, phytochemical, spectral and thermal characteristics of seeds from aforementioned six Caesalpinioideae species are described, together with an analysis of bioaccumulation factors and of the correlations found among the various constituents.

2. MATERIALS AND METHODS

2.1. Sample Collection

The seed legumes from plants: *D. regia* (DR), *L. leucocephala* (LL),*P. javanica*(PJ), *S. siamea* (SS), *E. gigas* (EG) and *M. pudica* (MP) were collected in April–May 2017 in Raipur area (21° 15′ 0″ N, 81° 37′ 48″ E), after botanical recognition using a standard monograph (**21**). The legumes (pods) were washed out with the de-ionized water and dried with the hot air. The surface layer of the soil on which the plants grew was also sampled. All samples were sundried for one week in a glass room. Size and mass of the seeds were measured using a Vernier scale and a Mettler-Toledo electronic balance, respectively.

Samples were then kept in an oven at 50 ^{0}C overnight for further dehydration, crushed with the help of mortar into fine powder (particle size ≤ 100 μm), and stored in glass bottles at -4 ^{0}C.

2.2. Characterization

The moisture content present in seeds was evaluated by drying the seeds at 105 ^{0}C in an air oven for 6 hr prior to the analysis, and mean values were computed. All characterization results were reported on a dry weight (dw) basis.

The infrared spectrum was characterized using a Thermo Scientific (Waltham, MA, USA) Nicolet iS50 Fourier-Transform Infrared (FTIR) spectrometer, equipped with an in-built diamond attenuated total reflection (ATR) system. The spectra were collected in the 400-4000 cm^{-1} spectral range with a 1 cm^{-1} spectral resolution and 64 scans.

Thermogravimetric/derivative thermogravimetric analyses (TG/DTG) and differential scanning calorimetry (DSC) analyses were conducted with a Perkin-Elmer (Waltham, MA, USA) STA6000 simultaneous thermal analyzer by heating the samples in a slow stream of N_2 (20 mL·min^{-1}) from room temperature up to 800 ^{0}C, with a heating rate of 20 ^{0}C·min^{-1}. Pyris v.11 software was used for data analysis.

AR grade sodium maleate (CAS 371-47-1) buffer, sodium acetate (CAS 127-09-3) buffer, potassium hydroxide (CAS 1310-58-3), amyl glucosidase (CAS 9032-08-0), pancreatic-α-amylase (MDL MFCD00081319), and glucose oxidase–peroxidase purchased from Megazyme International Ireland Ltd., and were used for color development for spectrophotometric determination.The soluble and resistant starch contents in the seeds were analyzed by the enzymatic method (**22**).

The oil content of the samples was analyzed by equilibrating a 5 g powdered sample with n-hexane (CAS 110-54-3, Sigma Aldrich) as prescribed by *Górnaś*et al. (**23**). The oil fraction was reported as a percentage on the basis of the dry weight (dw) of the seeds.

Sigma Aldrich analytical grade Folin-Ciocalteu reagent (MDL MFCD00132625), aluminum chloride (CAS 7446-70-0), tannic acid (CAS 1401-55-4), gallic acid (149-91-7) and quercetin (CAS 117-39-5) were used for the analysis of the phenols.For the determination of total polyphenol content (TPC) and flavonoid content (Fla), 100 mg of sample in powder form was equilibrated with 5 mL of an acetone:water mixture (70:30, v/v), and the solution was sonicated for 20 min at 20 °C in an ultrasonic bath, according to the procedure reported by *Bertaud*et al. (**24**).

The TPC of each extract was analyzed using Folin-Ciocalteu reagent, and expressed as tannic acid equivalents (TAE) (**25**). The Fla content was determined by the aluminum chloride method, and expressed as quercetin equivalents (QE) (**26**).

A Bruker Tracer 5i portable X-ray fluorescence (pXRF) spectrometer, equipped with a 4W rhodium anode and Xflash Silicon Drift Detector (SSD) with a typical resolution of 2028 channels, was used for the elemental analysis of the seed and soil samples. Two standard reference materials, brown and white cowpea (*Vigna unguiculata*(L.) Walp.) seeds, with reference values from ICP-OES and MS (As, Mo and Se in mg kg^{-1}) after *aqua regia* (HCl: HNO_3, 4:1) digestion were used for validation of the pXRF results. A standard soil sample (NCS DC 73382 CRM) was employed for the soil analyses. In soil analytical data, the confidence limit at p value of 0.05 was used.

Bioaccumulation factors were computed by dividing the seed analyte content by the soil one.

2.3. Statistics

Polyphenol, flavonoid, starch and mineral analyses of the seeds were carried out in triplicate. All values were reported as an average across three replicates with the STD. Correlation coefficients were calculated in IBM SPSS (Armonk, NY, USA).

3. RESULTS AND DISCUSSION

3.1. Physical Characteristics of Seeds

The Caesalpinioideae seeds were enclosed in a seed pod. Among those seed pods, that from *E. gigas* was the largest (1-2 m long and 10-12 cm wide). The number of seeds per seed pod were 20–25, 9–12, 15–20, 3–5, 10-15 and 15–20 for DR, EG, LL, MP, PJ and SS, respectively.All Caesalpinioideae seeds studied were brown colored, albeit with different shapes (elliptical, ovate or heart shaped, as depicted in **Figure 1**). The mass per seed varied from 21 to 23623 mg (**Table 1**): those from EG were exceptionally large (23623 mg), those from DR and PJ were of moderate size (304–510 mg), and the ones from LL, MP and SS were small (21–61 mg). The moisture content in the six seeds varied from 3.2 to 8.3%, with a fair correlation with mass size (r = 0.57).

Seed coats were found to range from thin to relatively thick: those of the seeds from MP and SS were found to be very thin, while the seed coat of other four seeds (DR, EG, LL and PS) were thicker, contributing from 37 to 69% of the mass of the whole seed. In particular, EG seed coat mass was 9449 mg per seed.

3.2. Polyphenol Content

Total polyphenols and flavonoid contents in the kernels/seeds varied from 1180 to 18840 mg/kg and from 2650 to 9100 mg/kg, respectively. The highest TPC values corresponded to MG and EG seed kernels. Remarkably high TPC and Fla concentrations were identified in the seed coats, ranging from 26900 to 32000 mg/kg and from 3900 to 12000 mg/kg, respectively.

3.3. Oil and Starch Content

The oil content in the seeds from the six species studied herein varied from 3.1 to 30.1%. Seeds from MP and PJ featured the highest oil contents (17.2 and 30.1%), comparable to those reported for other Caesalpinioideae seeds (**27, 28**).

Starch contents in the Caesalpinioideae seed kernels were in the 5.4 to 41.0% range. The highest starch content was detected in the EG seed kernel, for which the estimated amount of starch per seed was estimated to be 5811 mg. The content in resistant starch in the seed kernels from the studies species ranged from 0.5 to 1.2%.

The caloric value can be computed by multiplying by 9, 4 and 2 kcal for each gram of oil, protein and carbohydrate (**29**). Thus, the estimated calorie values of the DR, EG, LL, MP, PJ and SS seed kernels would be 365, 319, 388, 518, 442 and 390 kcal/100 g DW, respectively.

3.4. Mineral Content

The sum of the total concentrations of 17 elements (viz. P, S, Cl, K, Rb, Mg, Ca, Sr, Cr, Mn, Fe, Co, Cu, Zn, se, Mo and Pb) detected in the DR, EG, LL, MP, PJ and SS seed kernels was found to be 40324, 20253, 43769, 24606, 78489 and 42969 mg/kg of kernel (DW), **Table 2**. The very high mineral content of the PJ seed kernel would be due to its high content in sulphur (5.1%), due to the presence of thiol compounds in substantial amounts (**30**). Ten nutrients (P, S, K, Rb, Mg, Ca, Mn, Fe, Cu and Zn) were detected in the seed kernels from the six species, at concentrations (in mg/kg) in the following ranges: 2531-7298 (P), 3305–51438 (S), 5334–20198 (K), 4–24 (Rb), 1414–5916 (Mg), 1015-15236 (Ca), 9–233 (Mn), 54–507 (Fe), 13–29 (Cu) and 10–75 (Zn). Strontium was detected in all seed kernels except for those from EG, at concentrations ranging from 3 to 132 mg/kg. Cl, Cr and Se were only identified in the LL, DR and JS seed kernels, respectively. Mo and Pb were detected at low levels (1-3 mg/kg) in the seed kernels from LL, EG and SS; and from LL and MP, respectively. The maximum concentration of P, Rb, Ca Sr and Zn; Mn, Fe and Cu; S and Mg; and K were detected in the SS, MP, PR and DR seed kernels, respectively. Relatively low concentrations (11271 mg/kg) of elements (P, S, K, Rb, Mg, Ca, Sr, Mn, Fe, Co, Cu and Zn) were detected in EG seed coat.

3.5. Bioaccumulation

The pH value of the soil solutions was alkaline, ranging from 7.8 to 8.9, with a mean value of 8.2. The surface layer of the soil on which the plants grew was also analyzed by XRF. K, Mg, Ca, Mn and Fe were the main elements observed. Other elements were detected at moderate to low levels. The average concentrations of P, S, Cl, K, Rb, Mg, Ca, Sr, Mn, Fe, Cu and Zn found were found to be 160±10, 233±18, 135±10, 1387±127, 7±1, 1488±117, 5964±823, 49±4, 1187±94, 15673±1238, 48±2 and 29±2 mg/kg, respectively.

The bioaccumulation factor (BAF) was computed by dividing the elemental concentration in the seed by the soil mean values. The BAC values for P, S, K, Rb, Mg and Ca were in the 16–46, 14–221, 4–15, 0.6–3.4, 1.0–4.0 and 0.2–2.6 range, respectively. A strong bioaccumulation of P, S and K nutrients was observed. In the seeds from three of the species (DR, MP and SS), very high concentrations of P were accumulated, approximately twice those of S. In the other three species (EG, LL and PJ) the reverse trend was observed, with S concentrations approximately twice those of P. In particular, P was found to be strongly hyperaccumulated (BAF = 14-15) in the seeds from DR and LL. Both Mg and Ca were observed to be moderately hyperaccumulated (BAF = 2.3–4.0 and 1.7–2.6) in the seeds from SJ and SS.

3.6. Correlation Coefficients

Correlation coefficients of seed elements are summarized in **Table 3**. The seed oil, Fla, Mg, S and Se contents showed a good correlation with each other, either due to coordination with glycerides and/or accumulation of Mg as sulfur and selenium compounds. A good correlation of the starch content with the total phenolic content was observed, ascribed to intermolecular bond formation. Phosphorous and potassium had a fair correlation with each other and with heavy metals (Fe, Co, Cu, Zn and Mo), as the latter would be co-factor elements in the accumulation of the former. Rubidiumshowed a fair correlation with Mo, a co-factor element in its accumulation. Ca had a good correlation with Sr, probably because the latter would be a substituent element in Ca accumulation. Heavy metals (Cr, Fe, Cu, Zn, Mo and Pb) showed good correlations with each other.

3.7. Vibrational Characteristics

The ATR-FTIR spectra of the kernel samples are shown in **Figure 4**. The vibrations from the various functional groups in the molecular constituents of the seed kernels from the six Caesalpinioideae have been identified by their position (wavenumber) (**Table 3**). Such assignments, together the analysis of the intensity of the bands at 2923 cm^{-1}, 2853 cm^{-1} and 1744 cm^{-1}, allowed to differentiate the spectra of *Mimosapudica*and *Parkiajavanica*from the rest, and specially from those of *Leucaena leucocephala* and *Senna siamea.*

The band at 1710 cm^{-1} (conjugated C=O), 1515 cm^{-1} (aromatic skeletal) and 778 cm^{-1}, frequent in the biomass and seed spectra of plants, were missing in the analyzed spectra.

3.8. Thermal Characteristics

TG-DTG shape and DSC thermal effects were analyzed for all the studied samples (**Figure 3**). A small weight loss was recorded up to 100 ^{0}C (first DTG peak), mainly due to the evaporation of a fraction of free water contained in the seed kernel powder. Upon subsequent heating, a multiple DTG feature with peaks between 200 ^{0}C and 410 ^{0}C was observed, associated with an abrupt pattern of weight loss. Deconvolution of these features allowed to identify three peaks at 210 ^{0}C, 320 ^{0}C and 400 ^{0}C, which can be put in relationship with the final desorption of all bound water, the decomposition of the polysaccharide molecules with formation of low molecular weight volatiles, and the decomposition process of lignin, respectively.

The shape of the TG curves (**Figure 4**) and the temperature for DTG peaks and DSC effects (**Table 5**) evidence notable similitudes in the decomposition rate of the seed kernel samples from *D. regia*, *M. pudica* and *P. javanica*.

4. CONCLUSIONS

The seeds from *M. pudica* and *P. javanica*; *D. regia*, *L. leucocephala* and *S. siamea*; and *E. gigas* were found to be rich in oil, protein and starch, respectively, in good agreement with their vibrational spectra and thermal behavior. A strong bioaccumulation of P, S and K nutrients was observed in all seeds, with particularly high S and K contents in *P. javanica* seeds (51 g/kg) and *D. regia/L. leucocephala seeds (*20 mg/kg), respectively. *E. gigas*is a starchy seed. Whereas*M. pudica*and*P. javanica*are oily seeds. The *M. pudica* and *P. javanica* seeds featured the highest caloric values.

ETHICS APPROVAL

Not applicable.

REFERENCES

1.Lewis GP.Cassieae. Legumes of the World. Lewis GP, Schrire B, MacKinder B, Lock M. (eds). Royal Botanic Gardens, Kew, UK. 2005,111-125.

2.El-Nashar HAS,Eldahshan O,Singab AN. The Tribe Caesalpinieae (Fabaceae): An updated review on pharmacological aspects. Medicinal and Aromatic Plants.2015,4: 215. doi: 10.4172/2167-0412.1000215

3.Singh S, Sonia NK. A review introduction to genus delonix. World Journal **of** Pharmaceutical Sciences**.** 2014,3: 2042-2055

4.Shewale VD, Deshmukh TA, Patil LS, Patil VR. Anti-Inflammatory activity of *Delonix regia* (Boj. Ex. Hook),Advance in Pharmacological Sciences 2012, Article ID 789713.doi: 10.1155/2012/789713.

5.Suhane N, Shrivastava RR, Singh M. Gulmohar an ornamental plant with medicinal uses.Journal of Pharmacognosy and Phytochemistry. 2016,5(6): 245-248.

6.Adetogun GE,Alebiowu G. Influence of *Delonix regia* seeds gum on the compressional characteristics of paracetamol tablet formulations.Journal of DrugDelivery Science and Technology**.**2004,17(6): 443-445. doi: 10.1016/S1773-2247(07)50086-9.

7.Vandermeulen S, Ramírez-Restrepo C,Beckers Y,ClaessensH,Bindelle J. Agroforestry for ruminants: a review of trees and shrubs as fodder in silvopastoral temperate and tropical production systems. Animal Production Science, 2018, 58: 767–777. doi: 10.1071/AN16434.

8.Liu G, Bai C, Clements RJ, Peters M, Rao IM, Schultze-KraftR. Tropical forage legumes for environmental benefits: An overview. Trop Grasslands-ForrajesTropicales.2018, 6(1): 1-14. doi:10.17138/tgft(6)1-14.

9.Torbjorn ALM, Exotic drift seeds in Norway: vernacular names, beuefs, and uses.Journal of Ethnobiology.2003,23(2): 227-261.

10.Meena Devi VN, Ariharan VN, Nagendra Prasad P.Nutritive value and potential uses of *Leucaena leucocephala* as biofuel –a mini review.Research Journal of Pharmaceutical, Biological and Chemical Sciences.2013, 4(1):515-521.

11.Dawane JS,Pandit VA,Rajopadhye BD. Experimental evaluation of anti-inflammatory effect of topical application of *Entadaphaseoloides* seeds as paste and ointment, North American Journal of Medical Sciences.2011, 3(11): 513–517.doi: 10.4297/najms.2011.3513.

12.Ahmad H, Sehgal S, Mishra A, Gupta R. *Mimosa pudica* L. (Laajvanti): An overview, Pharmacognosy Review.2012,6(12): 115–124. doi: 10.4103/0973-7847.99945, PMCID: PMC3459453.

13.Muhammad G, Hussain, MA,JantanI, Bukhari SNA.*Mimosa pudica* L., a high-value medicinal plant as a source of bioactives for pharmaceuticals. ComprehensiveReviews inFood Science and Food Safety.2016,15: 303–315. doi:10.1111/1541-4337.12184.

14.Gunawardhana CB, Ranasinghe SJ, Waisundara VY. Review: Mimosa pudica Linn.: the garden weed with therapeutic properties, Israel Journal of Plant Science.2015, 62:4:234-241.doi: 10.1080/07929978.2015.1066997.

15.Chanu KV, Ali MA, Kataria M. Antioxidant activities of two medicinal vegetables: *Parkiajavanica* and *Phlogacanthusthyrsiflorus*.International Journal of Pharmacy and Pharmaceutical Sciences.2012,4(1): 102-6.

16.Angami T, Bhagawati R, Touthang L., Makdoh B., Nirmal, Lungmuana, Bharati KA, Silambarasan R, Ayyanar M. Traditional uses, phytochemistry and biological activities of *Parkiatimoriana* (DC.) Merr., an underutilized multipurpose tree bean: a review. Genetic Resources and Crop Evolution.2018,65: 679–692. Doi: 10.1007/s10722-017-0595-0.

17.Gutteridge RC.*Senna siamea* (Larnk) Irwin &Bameby. In: FaridahHanum, I. & van der Maesen, L.J.G. (Editors): Plant Resources of South-East Asia No. I 1. Auxiliary plants. Backhuys Publishers, 1977.

18.Ntandou GF,Banzouzi JT,MbatchiB, Elion-Itou RD, Etou-Ossibi A, Ramos S, Benoit-Vical F,Abena AA, Ouamba JM. (2010). Analgesic and anti-inflammatory effects of *Cassia siamea* Lam. stem bark extracts. Journal of Ethnopharmacology.2010,127 (1): 108-11.

19.Kumar D, Jain A, Verma A. Phytochemical and pharmacological investigation of Cassia siameaLamk: An insight.The Natural Products Journal**.**2017,7(4): 255-266. doi:10.2174/2210315507666170509125800.

20.Widyowati R,AgilM. Chemical constituents and bioactivities of several Indonesian plants typically used in Jamu. Chemical andPharmaceutical Bulletin.2018,66(5): 506-518. doi:10.1248/cpb.c17-00983.

21.Desikachary TV. Cyanophyta, Publisher: New Delhi, Indian Council of Agricultural Research, 1959.

22.AOAC Official Method, Resistant starch in starch and plant materials enzymatic digestion, First Action, 2002.

23. GórnaśP,Rudzińska M,Segliņa D.Lipophilic composition of eleven apple seed oils: a promising source of unconventional oil from industry by-products. Industrial Crops and Products**.**2014,60: 86-91.https://doi.org/10.1016/j.indcrop.2014.06.003.

24.Bertaud F,Tapin-Lingua S,Pizzi A, Navarrete P, Petit-Conil M. Characterisation of industrial barks for their tannin contents for further green-wood based adhesives applications. InTechFibres. 2010. COST FP0901-Hamburg.

25.Singleton VL, Orthofer R,Lamuela-Raventós RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Methods in Enzymology. 1999,299: 152-178.

26.Chang CC, Yang MH, Wen HM,Chern JC. Estimation of total flavonoid content in propolis by two complementary colorimetric methods. Journal of Food and Drug Analysis.2002, 10(3): 178-182.

27. Adewuyi A,Oderinde RA, Rao BVSK, Prasad RBN, Anjaneyulu B. Chemical component and fatty acid distribution of *Delonix regia* and *Peltophorumpterocarpum* seed oils. Food Science and Technology Research. 16. 565-570. 10.3136/fstr.16.565.

28.Nehdi IA,Sbihi H,Tan CP,Al-Resayes SI.*Leucaena leucocephala (Lam.)* de Wit seed oil: Characterization and uses,Industrial Crops and Products, 2014, 52:582-587.https://doi.org/10.1016/j.indcrop.2013.11.021.

29.Meiners CR,Derise NL, Lau HC, Crews MG, Ritchey SJ, Murphy EW. Proximate composition and yield of raw and cooked mature dry legumes.Journal of Agricultural and Food Chemistry. 1976,24(6): 1122-1126.doi: 10.1021/jf60208a035.

30.GmelinR, Susilo R, Fenwick GR. Cyclic polysulphides from *Parkiaspeciosa*. Phytochemistry.1981, 20(11):2521-3.https://doi.org/10.1016/0031-9422(81)83085-3.

Table 1. Physico-chemical characteristics of Caesalpinioideae seeds.

Parameter	Delonix Regia	EntadaGigas	LeucaneaLecocephala	Mimosa Pudica	ParkiaJavanica	Senna Siamea
Color	Glossy brown	dark-red brown	Glossy brown	Brown	Brown	Glossy brown
Shape	Oblong	Heart-shaped	Ovate	Oval to orbicular	Elliptic	Flat ovate
Seed mass, mg	510±11	23623±	61±2	21±1	304±7	22±1
Seed coat, %	69	40	47	-	37	-
Moisture, %	7.5±0.2	8.3±0.3	6.5±0.1	3.2±0.1	6.5±0.2	3.6±0.1
Oil,%	3.1±0.1	6.1±0.2	5.9±0.2	30.1±0.8	17.2±0.0.6	3.1±0.1
Total starch, %	10.3±0.4	41.0±1.1	7.5±0.3	7.6±0.4	6.7±0.2	5.4±0.1
Resistant starch, %	0.50±0.02	1.00±0.0.03	0.60±0.03	1.20±0.04	1.10±0.04	0.6±0.01
TPh (Kernel), mg/kg	1820±32	18840±360	12430±251	18460±375	4880±98	1180±23
Fla (Kernel), mg/kg	2850±	2650±48	3200±57	6250±128	9100±176	3050±61
TPh (seed coat), mg/kg	28200±540	26900±522	30100±580	-	32000±625	-

Fla (seed coat), mg/kg	4100±81	3900±79	4600±90	-	12000±232	-

TPh = Total polyphenol, Fla = Flavonoid

Table 2. Mineral characteristics of Caesalpinioideae seeds.

Element	Delonix Regia	EntadaGigas	LeucaneaLecocephala	Mimosa Pudica	ParkiaJavanica	Senna Siamea
P	6812	2531	5564	5678	3005	7298
S	4475	5268	10833	3305	51438	3936
Cl	ND	ND	360	ND	ND	ND
K	20198	9899	19543	5334	7742	12640
Rb	22	24	8	4	10	24
Mg	2781	1414	3429	2358	5916	3463
Ca	5524	1015	3788	7119	10162	15236
Sr	3	ND	8	27	32	132
Cr	6	ND	ND	ND	ND	ND
Mn	139	9	15	233	30	22
Fe	262	54	156	507	99	133
Co	1	ND	1	ND	1	1
Cu	21	13	18	29	13	15
Zn	75	25	46	10	39	67
Se	ND	ND	ND	ND	2	0
Mo	3	1	ND	ND	ND	2
Pb	2	ND	ND	2	ND	ND

ND = Not detectable

Table 3. **Correlation coefficient matrix of seed elements.**

	Oil	Sta	TPh	Fla	P	S	Cl	K	Rb	Mg	Ca	Sr	Cr	Mn	Fe	Co	Cu	Zn	Se	Mo	Pb
Oil	1																				
Sta	-0.12	1.00																			
TPh	0.02	0.80	1.00																		
Fla	0.96	-0.34	-0.24	1.00																	
P	-0.68	-0.63	-0.68	-0.48	1.00																
S	0.98	-0.30	-0.16	0.99	-0.54	1.00															
Cl	-0.11	-0.25	0.34	-0.20	0.13	-0.12	1.00														
K	-0.65	-0.33	-0.17	-0.59	0.69	-0.57	0.55	1.00													
Rb	-0.62	0.46	-0.08	-0.58	0.24	-0.64	-0.68	-0.07	1.00												
Mg	0.78	-0.71	-0.52	0.89	-0.08	0.87	0.01	-0.31	-0.67	1.00											
Ca	0.11	-0.68	-0.80	0.33	0.47	0.24	-0.33	-0.26	0.08	0.58	1.00										
Sr	-0.17	-0.44	-0.56	0.00	0.50	-0.09	-0.27	-0.25	0.30	0.24	0.91	1.00									
Cr	-0.38	-0.14	-0.44	-0.27	0.45	-0.29	-0.25	0.61	0.31	-0.21	-0.16	-0.32	1.00								
Mn	-0.29	-0.25	-0.55	-0.15	0.47	-0.18	-0.29	0.56	0.25	-0.08	-0.05	-0.26	0.99	1.00							
Fe	-0.44	-0.54	-0.61	-0.28	0.75	-0.30	0.11	0.83	0.04	-0.02	0.05	-0.11	0.87	0.88	1.00						

Co	0.09	-0.99	-0.81	0.31	0.64	0.27	0.25	0.41	-0.45	0.68	0.61	0.35	0.25	0.35	0.62	1.00					
Cu	-0.56	-0.39	-0.36	-0.46	0.71	-0.46	0.32	0.95	0.00	-0.21	-0.17	-0.26	0.81	0.78	0.95	0.48	1.00				
Zn	-0.52	-0.65	-0.84	-0.28	0.93	-0.35	-0.12	0.61	0.28	0.06	0.50	0.41	0.67	0.71	0.86	0.69	0.73	1.00			
Se	0.97	-0.28	-0.22	1.00	-0.52	0.99	-0.25	-0.62	-0.54	0.86	0.30	-0.03	-0.25	-0.13	-0.30	0.25	-0.48	-0.31	1.00		
Mo	-0.69	-0.03	-0.51	-0.54	0.64	-0.61	-0.51	0.40	0.78	-0.44	0.18	0.22	0.77	0.75	0.64	0.09	0.55	0.76	-0.51	1.00	
Pb	-0.38	-0.14	-0.44	-0.27	0.45	-0.29	-0.25	0.61	0.31	-0.21	-0.16	-0.32	1.00	0.99	0.87	0.25	0.81	0.67	-0.25	0.77	1.00

Table 4.Main absorption bands in the ATR-FTIR spectra of the *Fabaceae* kernel samples under study (all wavenumbers are expressed in cm^{-1}).

Delonix kernel	*Entada* kernel	*Leucaena* kernel	*Mimosa* kernel	*Parkia* kernel	*Senna* kernel	Assignments
3281	3285	3291	3286	3288	3285	O-H stretching (cellulose)
2923	2923	2923	2923	2922	2923	$–CH_2$ stretch. (cutine, wax, pectin)
2854	2953	2853	2854	2853	2853	$–CH_2$ stretching (cutine and wax)
1745	1744	1744	1744	1742	1744	C=O stretching (hemicellulose)
1651			1654		1651	C=C (cellulose)/COO^- sym. Stret
	1640		1648			uronic acids?
1634		1634		1635	1634	C=O stretching (hemicellulose)
	1547		1541	1540	1539	COO^- symmetric
1538		1537	1535			stretching
1455	1455	1454	1457	1456	1455	$O-CH_3$ stretching
					1445	
1398						CH rocking
	1377	1378	1377	1378		$-CH_3$ symmetric deformation (hemicellulose)
1315		1301	1316		1316	C-H (cellulose)
1238		1231	1238	1235	1237	C-C-O asym stret.,
	1218					acetylated glucomannan

1139	1146	1136	1157	1159		C-O-C in bridge, asymmetric (cellulose)
			1095	1097		C-O-C stretching in the pyranose
1046	1073 1012	1047	1037	1043	1046	C-O stretching (cellulose)
997	997					C-H wags, vinyl
		921	916			
			895			O-C=O in-plane deformation or a CH_2 rocking deformation
	840 830	831 804		857		aromatic C-H out-of-plane binding or C-O-C deform
			695			β-glycosidic linkage (cellulose)
	575	572		584	572	saccharide moities
525	525	525		529	526	

Table 5.DTG peak temperatures for the kernel samples.

Species	1st step	2nd step	3rd step	4th step
D. regia	82⁰C	210⁰C / 242⁰C	323⁰C	411⁰C
E. gigas	86⁰C	210⁰C	309⁰C	396⁰C
L. leucocephala	87⁰C	200⁰C	319⁰C	-
M. pudica	82⁰C	-	310⁰C / 332⁰C	404⁰C
P. javanica	98⁰C	253⁰C	-	379⁰C
S. siamea	88⁰C	-	318⁰C	-

F

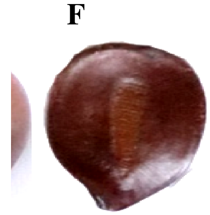


Figure 1. **Imageofseeds: Delonix regia (A), Entada gigas (B), Leucanealecocephala (C), Mimosa pudica (D), Parkiajavanica (E), Senna siamea (F).**

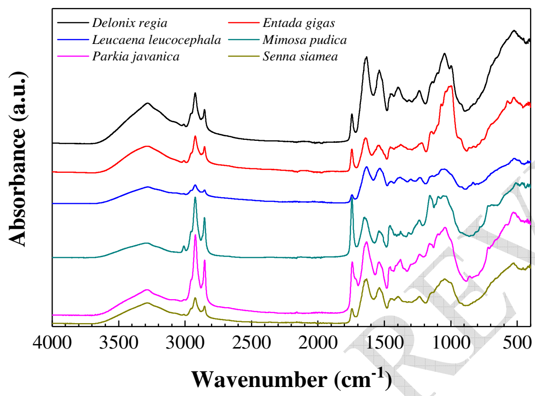
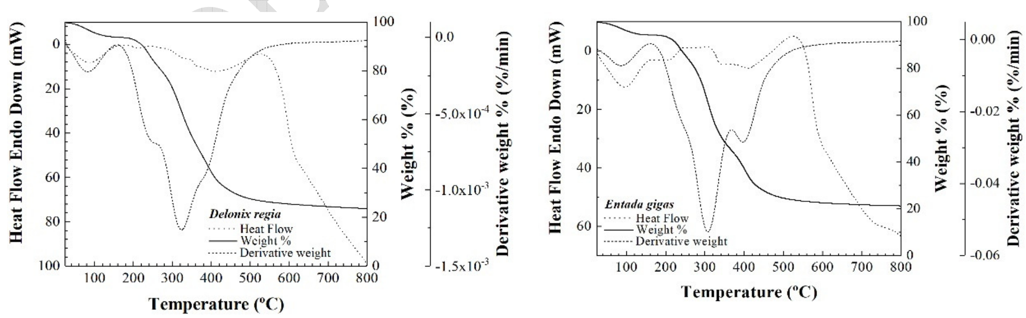


Figure 2.ATR-FTIR spectra of kernel samples from species of *Fabaceae* family, *Caesalpinioideae*subfamily. Some offset has been added in the *y* axis for clarity purposes.



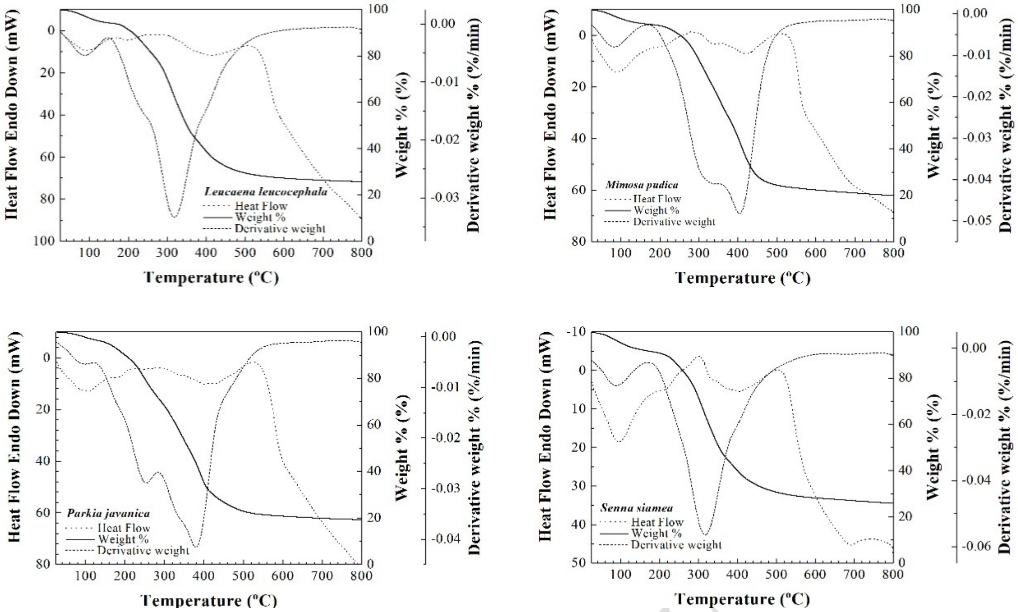


Figure 1. TG (*solid line*), DTG (*dashed line*) and DSC (*dotted line*) curves for the *Caesalpinioideae*seedsamples.

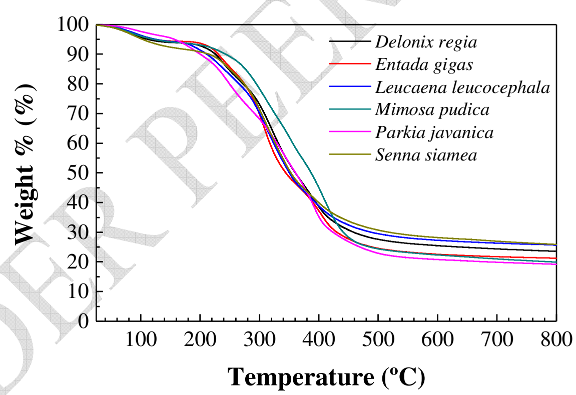


Figure 4. Comparison of the TG curves for the different *Caesalpinioideae* seed samples.