

2 **Proximate composition, Functional and Sensory Properties of Pearl Millet, Soy flour**
3 **and Baobab Fruit Pulp Composite flour as a Complementary Food**

4
5 **Abstract**

6 **Aim:** to evaluate the proximate composition, functional and sensory properties of a
7 complementary food from pearl millet, soy flour and baobab fruit pulp composite flours.

8 **Study Design:** A complementary food was produced from Pearl millet, soy flour and baobab
9 fruit pulp powder of various proportions (10, 20, 25 and 30%). Proximate (protein, ash,
10 moisture, fibre, fat and carbohydrate and energy value) composition, functional (Bulk
11 density, gelation capacity, swelling index, water absorption capacity and oil absorption
12 capacity) properties and sensory (appearance, flavour, texture and overall acceptability)
13 attributes were determined.

14 **Results:** The results of proximate composition showed that Moisture content ranged from
15 10.09 – 10.98, Protein content ranged from 9.80 – 24.25, Fat content ranged from 4.94 –
16 16.65, Carbohydrate content ranged from 43.11 – 71.03, Fibre content ranged from 3.37 –
17 15.67, Ash content ranged from 2.59 – 2.87% and Energy value ranged from 367.78 – 423.69
18 Kcal. The functional properties showed that Water Absorption Capacity ranged from 2.70 –
19 2.91, Oil Absorption Capacity ranged from 1.90 – 2.72, Bulk Density ranged from 0.69 –
20 0.71, Swelling Index ranged from 0.68 – 1.04g/ml and Gelation Capacity ranged from 5 –
21 10% of the complementary food samples. The sensory attribute also revealed that the
22 complementary food samples proved to be of good quality but the controlled sample (A) was
23 most preferred by the panellist.

24 **Conclusion:** Complementary foods were produced from pearl millet and soybean
25 supplemented with baobab fruit pulp. Though the control sample (A) was the most preferred
26 sample. Samples with baobab fruit pulp were also accepted.

27 **Keywords:** Baobab Fruit Pulp (BFP), Pearl Millet, Soybean, Complementary Food

28
29 **INTRODUCTION**

30 Malnutrition is responsible, directly or indirectly, for over half of all childhood deaths.
31 Infants and young children are at increased risk of malnutrition from six months of age
32 onwards, when breast milk alone is no longer sufficient to meet all nutritional requirements
33 and complementary feeding needs to be started. Complementary foods are often of lesser
34 nutritional quality than breast milk. In addition, they are often given in insufficient amounts
35 and, if given too early or too frequently, they displace breast milk. Complementary foods are

36 food other than breast milk or infant formula such as solid, liquid and semi-solid food
37 materials which are introduced to infants to provide nourishment (Anigo *et al.*, 2010). Gastric
38 capacity limits the amount of food that a young child can consume during each meal.
39 Repeated infections reduce appetite and increase the risk of inadequate intakes. Infants and
40 young children need a caring adult or other responsible person who not only selects and
41 offers appropriate foods but assists and encourages them to consume these foods in sufficient
42 quantity (WHO, 2001). It is common knowledge that breast milk is the best food for infants
43 during their first six (6) months of life. Breast milk contains all the essential nutrients and
44 immunological factors an infant requires to maintain optimal health and growth. It also tends
45 to protect infants against upper respiratory infection and diarrhea which are the chief causes
46 of infant and child morbidity and mortality (Cristina *et al.*, 2004 and Solomon, 2005).
47 However, at an early age of six (6) months and above, the weight of the child is expected to
48 double which breast milk alone at this point may not be sufficient for the child's nutritional
49 and growth needs. The adoption of recommended breast feeding and complementary feeding
50 practice and access to the appropriate quality and amount of foods are essential component of
51 optimal nutrition for infant and young children (Anigo *et al.*, 2010). Several factors tends to
52 contribute to the vulnerability of children (infants) during the complementary feeding period.
53 These factors may include; low nutritional quality of complementary foods which most times
54 are provided in insufficient amount to the child (WHO, 2002; Anigo *et al.*, 2010). In recent
55 years, many important advances in breast feeding promotion have been made but
56 unfortunately the same may not be said for complementary feeding (PAHO/WHO, 2003).
57 This research therefore aims to improve the quality of complementary food through the
58 supplementation of Baobab Fruit Pulp to with other cereal e.g pearl millet and Legumes such
59 as soybean improve the nutritional quality of infant formula.

60

61 MATERIALS AND METHODS

62 Materials

63 The food commodities used for this research were pearl millet (*Pennisetum glaucum*),
64 soybean (*Glycine max. L*) and Baobab fruit pulp (*Adansonia digitata*). Soybean and pearl
65 millet were purchased from North Bank market Makurdi, were brought to the university of
66 Agriculture Makurdi seed research centre for identification. Baobab fruit pulp powder was
67 obtained from Lafia Market in Nasarawa state. Nigeria

68

69 Pearl Millet Flour Preparation

70 The process of flour preparation as shown in fig 1 consists of dry cleaning of the pearl millet
71 i.e winnowing etc. The kernels were thereafter dehulled after mild wetting using rice
72 dehuller. The grains were then washed and dried in a convection hot air laboratory oven
73 (MODEL TT-9053 (Techmel and Techmel) at 50⁰ C for 24 hrs to 14% moisture content. The
74 dried grain was milled using a single disk attrition mill and sieved through a 455µm screen
75 laboratory sieve (MODEL STMN 2-CO402 JAPAN) and the under flow was used for the
76 research (Filli, *et al* 2012).

77 Soy Flour Preparation from

78 The method of Filli *et al*, (2012) was adopted as shown in fig 2. Soybean seeds were steeped
79 in clean tap water at 28⁰C for 24hrs in a plastic bowl. The kernel was therefore dehulled using
80 the traditional pestle and mortar. The grains were then washed and the hulls removed. After
81 which it was dried in a convectional laboratory hot air oven (MODEL TT-9053 (Techmel) at
82 50⁰C for 24hrs to 14% moisture content and the mass was winnowed to remove the
83 remaining lighter material using trail. The dehulled soybeans kernels were ground in a
84 laboratory disc attrition mill to fine flour. The flour was sieved through a 455µm screen
85 laboratory sieve (MODEL STMN 2-CO402 JAPAN) and the under flow was used for further
86 use.

87 Baobab Fruit pulp Flour Preparation

88 Baobab pods were cracked using a hammer. The pulp and seeds were transferred into a
89 ceramic mortar and it was pounded using a pestle until all the pulp was separated from the
90 seed. The pulp was sieved through a 455µm screen laboratory sieve MODEL STMN 2-

91 CO402 JAPAN to remove the fibrous materials from the pulp and the under flow was used
92 for further use as shown in fig 3
93

UNDER PEER REVIEW

94

95

Pearl millet

96



97

Cleaning/washing

98



99

Oven drying (50⁰C for 24hrs)

100



101

Weighing

102



103

Toasting in microwaving (80⁰C for 15 min)

104



105

Cooling

106



107

Winnowing

108



109

Milling

110



111

Sieving (455 μ m)

112



113

Flour

114



115

Packaged and store

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Fig 1: Flow chart for the production of pearl millet flour.

117

Source: (Filli, 2012) with slight modification

118

119

Soybeans

120



121

Sorting

122



123

Cleaning

124



125

Blanching (60⁰C for 20 – 25 min)

126



127

Dehulling by hand rubbing

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129

Removal of hulls by floatation

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131

Oven drying (55⁰C for 24hrs)

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133

Toasting in microwaved (75⁰C)

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135

Milling

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137

Sieving (455µm)

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139

Flour

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141

Packaged and store

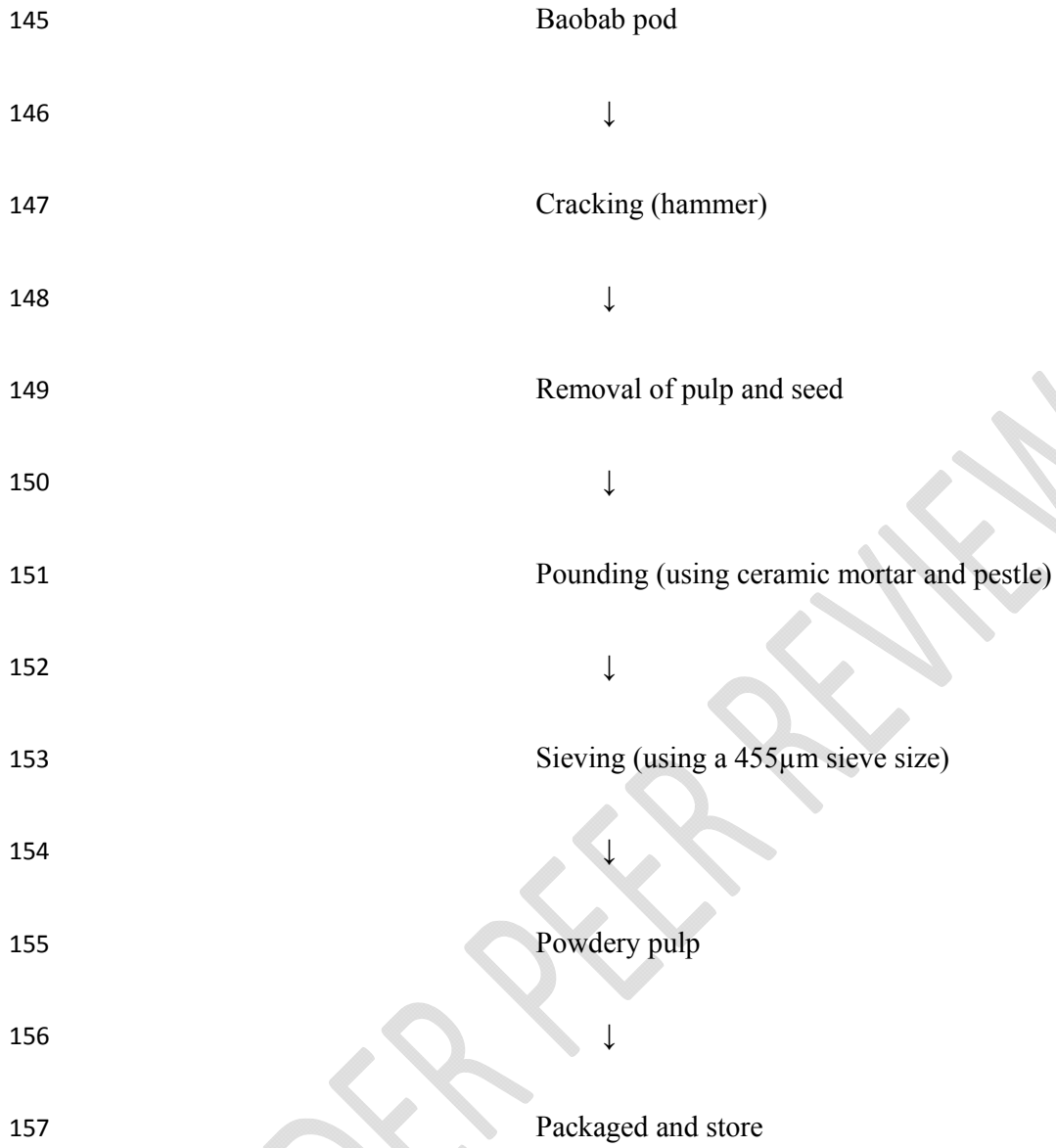
142

Fig 2: Flow chart for the production of soy flour.

143

Source: Ihekoronye, 1999) with slight modification

144



158 Fig 3: Flow chart for the production of baobab fruit pulp powder.

159 Source: (Chadre, 2009) with slight modifications.

160

161

162 .

163 **PROXIMATE ANALYSIS**

164 **Determination of Moisture Content**

165 Moisture content was determined by the air-oven method as described by AOAC (2005).
166 Two grams of the sample was weighed in duplicate into Petri dishes of known weight and
167 covered immediately. These were transferred into oven, uncovered and heated at $103^{\circ}\text{C} \pm 2$
168 for 3-5 hours. The samples were then removed from the oven and placed in the desiccator to
169 cool for 15 minutes before weighing. The process was repeated until constant weights were
170 recorded. The loss in weight from the original weight was reported as the moisture content.

$$171 \quad \% \text{ Moisture Content} = \frac{W_2 - W_3}{W_2 - W_1} \times 100 \quad (1)$$

172

173 **Determination of Crude Protein**

174 The Kjeldahl method was used for the determination of crude protein as described by AOAC
175 (2005). The samples (1.0g each) were first digested in Kjeldahl digesting system. The
176 digested samples were allowed to cool and then distilled into 2% boric acid solution
177 containing methyl orange indicator and diluted with water after the introduction of 40%
178 sodium hydroxide solution. The distilled samples were then titrated against 0.1 M HCL
179 solution. A blank titration was similarly carried out and the percentage content was estimated
180 as percentage Nitrogen $\times 6.25$ (1 ml of 0.1M HCL $\pm = 0.014$ g N)

$$181 \quad \%N = (b-a) \times 0.1N \text{ Hcl} \times 0.014 \times \text{dilution factor} \times 100 / \text{weight of sample} \quad (2)$$

$$182 \quad \% \text{ protein} = \% \text{ Nitrogen} \times 6.25 \quad (3)$$

183 **Determination of Crude Fat Content**

184 The Soxhlet solvent extraction method outlined in AOAC (2005) was used. Two gram
185 sample was weighed (A) into the extraction thimble and the thimble was plugged with cotton
186 wool. It was placed back in the Soxhlet apparatus fitted with a weighed flat bottom flask (B)
187 which was filled to about three quarter of its volume with petroleum ether of a boiling point
188 of $40\text{-}60^{\circ}\text{C}$. The extraction was carried for a period of 4-8 hours after which complete
189 extraction was made. The petroleum ether was removed by evaporation on the water bath and
190 the remaining portion in the flask was removed along with water by drying in the oven at 80
191 $^{\circ}\text{C}$ for 30 minutes and cooled in desiccators and weighed (C).

$$\% \text{ Fat Content} = \frac{W_4 - W_3}{W_2 - W_1} \times 100$$

192 where:

193 W_1 = weight of oven dried thimble,

194 W_2 = weight of sample used,

195 W_3 = weight of round bottom flask,

196 W_4 = weight of round bottom flask with fat residue.

197 **Determination of Crude Fibre Content**

198 Fibre content was determined following the procedure outlined in AOAC (2005) method as
199 reported by Onwuka (2005) Two grams portions of the samples were extracted using
200 petroleum spirit (boiling point 40-60°C.) This was digested in 1 liter flask using 200ml
201 concentrated Sulphuric acid and filtered through the California buchner system. The insoluble
202 matter was washed with boiling water until it was free from the acid. The residue was then
203 back into the flask with 200ml of 0.313M NaOH. The flask content was brought to boil for
204 30 minutes. The flask was allowed to stand for 1 minute and filtered immediately through a
205 filtering cloth. The insoluble material was transferred into 100ml beaker by means of boiling
206 water, washed with 1% HCl and again with boiling water to free it from acid. The insoluble
207 material was finally washed with alcohol twice and three times with diethyl ether. The
208 resulting residue was transferred to a dish (previously weighed) with boiling water. The dish
209 containing the residue was dried for 2 hours, at 100°C, cooled in desiccators and weighed
210 (W_1). The dried, cooled, and weighed residue was then transferred in a muffle furnace and
211 ignited at 600°C for 30 minutes, cooled and reweighed (W_2). The percent crude fibre content
212 was calculated as follows.

213

$$214 \quad \% \text{ Crude Fibre} = \frac{W_2 - W_3}{W_1} \times 100 \quad (5)$$

215 Where:

216 W_1 = weight of sample used,

217 W_2 = weight of crucible plus sample,

218 W_3 = weight of sample crucible + ash.

219

220 **Determination of Ash**

221 The ash content of the sample was determined by the method described by AOAC (2005) a
222 silica dish was heated to 600⁰C, cooled in desiccators and weighed. Then 5g of the sample
223 was weighed into the silica dish and transferred to the furnace. The temperature of the
224 furnace was allowed to reach 525⁰C before placing the dish in it. The temperature was
225 maintained until whitish grey colour was obtained indicating that all the organic matter
226 content of the sample had been destroyed. The dish was then brought out from the furnace
227 and placed in the desiccators, cooled and reweighed.

228

$$229 \quad \% \text{ Ash Content} = \frac{W_2 - W_1}{\text{Weight of sample}} \times 100 \quad (6)$$

230 Where:

231 W₂ = weight of crucible + ash,

232 W₁ = weight of empty crucible.

233 **Determination of Carbohydrates**

234 Carbohydrate was determined by difference as reported by Ihekoronye and Ngoddy, (1985).

$$235 \quad \% \text{ carbohydrate} = 100 - (\% \text{ moisture, protein, fibre, fat and ash}). \quad (7)$$

236 **FUNCTIONAL PROPERTIES OF SAMPLES**

237 **Determination of gelation capacity:**

238 The method described by Iwe *et al.* (2017) was used for the determination of the gelation
239 capacity. Suspensions of the samples in 5 ml of distilled water in test tubes were prepared
240 using 2 –20% (W/V) of the samples in test tubes. The sample test tubes were heated for 1
241 hour in a boiling water-bath followed by rapid cooling under running cold tap water. The test
242 tubes were further cooled for 2 hours at 40°C. Then, the gelation capacity was determined for
243 each sample as the least gelation concentration. That is, the concentration when the sample
244 from the inverted test tube will not slip

245 **Determination of Bulk Density**

246 The bulk density was determined as described by (Onwuka, 2005). A 10ml capacity
247 graduated measuring cylinder was weighed and 50g sample filled into it. The bottom of the

248 flask was tapped gently on the laboratory bench several times until there were no further
249 diminutions of the sample level after filling to 10ml mark.

$$250 \quad \text{Bulk Density (g/ml)} = \frac{\text{weight of sample}}{\text{volume of sample}} \quad (10)$$

251 **Determination of Swelling Index**

252 The method of Onwuka, (2005) was employed,. One gram of the flour samples was weighed
253 into 10ml graduated cylinder. Five (5ml) milliliters of distilled water was carefully added and
254 the volume occupied by the sample was recorded. The sample was allowed to stand
255 undisturbed in water for 1 hour and the volume occupies after swelling was recorded and
256 calculated as:

$$257 \quad \text{Swelling Index} = \frac{\text{vol.occupied by sample after swelling}}{\text{vol.occupied by sample after swelling}} \quad (11)$$

258 **Determination of Water Absorption Capacity**

259 The water absorption capacity of the flours was determine by the modified method of
260 Onwuka, (2005). One gram of sample was mixed with 10 mL distilled water and allow to
261 stand at ambient temperature (30 ± 2 °C) for 30 min, then centrifuged for 30 min at 3,000 rpm
262 or $2000 \times g$. Water absorption was examined as per cent water bound per gram flour.

263 **Determination of Oil Absorption Capacity**

264 The oil absorption capacity was also determined by the modified method of (Onwuka, 2005).
265 One gram of sample was mixed with 10 mL soybean oil (Sp. Gravity: 0.9092) and allow to
266 stand at ambient temperature (30 ± 2 °C) for 30 min, then centrifuged for 30 min at 300 rpm
267 or $2000 \times g$. Oil absorption was examined as percent water bound per gram flour.

268 **ENERGY VALUE**

269 This was calculated by multiplying the values of carbohydrate, fat and protein with the
270 Atwater Factor (4, 9, and 4) for carbohydrate, fat and protein respectively as described by
271 Onwuka, (2005).

272 **Sensory Evaluation**

273 Sensory evaluation based on the sensory attributes was conducted by using a standard 9-
274 points hedonic scales method (where 1 = dislike very much and 9 = like very much) as

275 described by Ihekoronye and Ngoddy, (1985). A total of 30 semi-trained panelists aged 18
276 years and above were involved in the evaluation of appearance, flavour, texture and overall
277 acceptability. The samples (100 g each) were coded randomly number using statistical
278 random Tables and served to the panellists with bottled water for rinsing their mouth after
279 every sample taste in a randomized order. The panellists were instructed to rate the attributes
280 indicating their degree of liking or disliking by putting a number as provided on the hedonic
281 scale according to their preference.

282

283 **Statistical Analyses**

284 All analyses were carried out in triplicate unless otherwise stated. Statistical significance was
285 established using one-way analysis of variance (ANOVA), and data were reported as the
286 mean standard deviation. Mean comparison and separation was done using Fisher's Least
287 Significant Difference test (LSD) at $p \leq 0.05$. Statistical analysis was carried out using the
288 SPSS 20 statistical package.

289

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UNDER PEER REVIEW

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292 **DISCUSSION**

293 **Proximate Composition**

294 The proximate composition of sample A was significantly ($P<0.05$) higher in protein content
295 (24.25%), fat content (16.65%) and Energy value (423.69 Kcal). According to Emmanuel *et al.*
296 *al.*, (2012), the addition of soybean flour to tiger-nut in the preparation of an infant diet
297 increases the protein, fat and energy values respectively. The Moisture content values for all
298 the samples tend to agree with the PAG (Protein Advisory Group – United Nations) which
299 reported moisture content of between 5-10% maximum. The range of moisture would have a
300 positive effect on the shelf life stability of the products (Bassey, 2004) and (Emmanuel *et al.*,
301 2012). The Ash content of the samples ranges from 2.59 – 2.87% with the highest value in
302 sample E (2.87%). The high Ash content of sample E could be due to the ratio of Millet Flour
303 and Baobab Fruit Pulp Powder in the sample since both are good sources of mineral
304 elements. Ash content of the samples was found to be less than the PAG standards which
305 reported 10% maximum ash content. The Protein content of the samples ranges from 9.80 –
306 24.25% with highest value in sample A (24.25%). These values are higher compared to PAG
307 standard (20%) respectively. This may be attributed to the protein content of soybean
308 addition (Emmanuel *et al.*, 2012). The fat content of the samples was found to range from
309 4.94 – 16.65% with sample A (16.65%) having the highest significance ($P<0.05$) value than
310 others. This is as a result of the high soy (50%) flour content in the sample. Though, the fat
311 contents of sample A and B met the PAG standard which is 10% and for weaning foods.
312 Sample D and E with low Fat content could be as a result of low amount of soy flour addition
313 and increased baobab fruit pulp addition which may have caused some dilution. High Fat
314 content is very important in infant diet because it contain essential Fatty Acids (soy flour)
315 which promote good health. It is also a carrier of fat soluble vitamins (A, D, E and K) and
316 promoting the absorption (Emmanuel *et al.*, 2012). The Fibre content of the samples on the
317 other hand ranges from 4.62 – 11.65% with samples E (15.67%) having the highest
318 significant ($P<0.05$) value. This could be due to increase in Baobab fruit pulp powder and
319 millet flour. An increase in the fibre content of weaning food has some beneficial effect on
320 the muscles of the large and small intestines. The values from the samples are higher than
321 those reported by PAG (5% Maximum). High fibre content was also reported to have adverse
322 effect on mineral element in the body (Emmanuel *et al.*, 2012) and (Bassey, 2004).

323 Carbohydrate content of the samples was found to range from 43.11-71.03% with sample E
324 having the highest significance ($P<0.05$) value. The high values of carbohydrate could be as a
325 result of millet flour and possibly baobab fruit pulp. Carbohydrate is required in infant diet
326 for Energy during growth. Energy values of the samples was found to range from 367.78-
327 423.69 Kcal with sample A (423.69 Kcal) having the highest significance ($P<0.05$) value.
328 The high Energy value of sample A is due to the high fat content of the sample. The Energy
329 value of the samples agrees with SON and PAG which reported 350-400Kcal respectively.
330 The Food and Agricultural Organisation reported that Home prepared weaning foods should
331 contain protein 15%, fat 11%, fibre 5% maximum, and for commercially prepared weaning
332 food for protein 15%, fat 6%, crude fibre 2% and moisture content 10% respectively.

333 **Functional Properties**

334 **Gelation concentration (GC)**

335 The least gelation concentration (LGC) which is defined as the lowest protein concentration
336 at which gel remained in the inverted tube was used as index of gelation capacity. The data
337 for LGC of different flours are given in Table 3. Composite (E) flours formed a gel at a
338 significantly higher concentration (10 g). Sample A and B flour formed gel quickly at very
339 lowest concentration (5 g). Wheat flours contain high protein and starch content and the
340 gelation capacity of flours is influenced by physical competition for water between protein
341 gelation and starch gelatinization (Kaushal *et al.* 2012). Suresh *et al.* (2015) reported that
342 protein gelation was significantly affected by exposed hydrophobicity and square of
343 sulfhydryls of proteins. As the percentage of incorporation of millet flour in wheat flour
344 (composite flour) increased, gelling properties decreased. The low gelation concentration of
345 A and B flour as composite flour may be added an asset for the formation of curd or as an
346 additive to other gel forming materials in food products. The variation in the gelling
347 properties may be ascribed to ratios of the different constituents such as protein,
348 carbohydrates and lipids in different flours, suggesting that interaction between such
349 components may also have a significant role in functional properties (Aremu *et al.* 2007). The
350 composite flours (E) would be useful in food system such as puddings, sauce and other foods
351 which require thickening and gelling (Suresh *et al.* 2015)

352 **Bulk density**

353 The bulk density (g/cm^3) of flour is the density measured without the influence of any
354 compression. The bulk densities of flours ranged from 0.69 g/cc to 0.71 g/cc. The highest
355 highest bulk density was observed A,B, C and D flour as shown in Table 3 and lowest was
356 sample E (0.69 g/cc). The present study revealed that bulk density depends on the particle
357 size and initial moisture content of flours. The obtained does not agree with those presented
358 by (Suresh *et al*, 2015), reported that Bulk density of composite flour increased with increase
359 in the incorporation of different flour. However, it is clear that decreased the proportion of
360 wheat flour increase the bulk density of composite flours. The high bulk density of flour
361 suggests their suitability for use in food preparations. On contrast, low bulk density would be
362 an advantage in the formulation of complementary foods (Suresh *et al*, 2015). Therefore, the
363 present study suggests that high bulk density of composite flour (A, B, C and D) suggests its
364 suitability to be used as thickener in food products and for use in food preparation since it
365 help to reduce paste thickness which is an important factor in convalescent and child feeding.

366 **Swelling capacity**

367 The swelling capacity of different flours ranged between 16.00 to 22.30 ml (Suresh *et al*,
368 2015). From Table 3, it is clear that lowest value of swelling capacity was observed in A
369 ($0.68 \pm 0.13 \text{ml}$) whereas the maximum in E ($1.04 \pm 0.13 \text{ml}$). The swelling capacity of flours
370 depends on size of particles, types of variety and types of processing methods and/or unit
371 operations. Suresh *et al*, (2015) reported that the flour of parboiled rice has more swelling
372 capacity as compared to raw rice. They also reported that the Swelling capacity of composite
373 flours increased with increase in the level of incorporation and decreased with level of wheat
374 flour addition. It is explicit that the swelling capacity of composite flours is highly affected
375 by the level of millet flour, because millet flour is rich in starch content.

376 **Water absorption capacity (WAC)**

377 The water absorption capacity for composite flours is given in Table 3. The WAC ranged
378 between 2.70 to 2.91 for all flours. The WAC was observed highest in C (2.91) and lowest in
379 D and E (2.70). The result suggests that addition of millet flour to wheat flour affected the
380 amount of water absorption. This could be due to molecular structure of millet starch which
381 inhibited water absorption, as could be seen from the lower values of WAC, with increase in

382 proportions of other flours to wheat flours. Similar observation was reported by Kaushal *et al.*
383 (2012). Suresh *et al.*, (2015) reported that lower WAC in some flours may be due to less
384 availability of polar amino acids in flours. The increase in WAC of blends after incorporating
385 millet flour may be due to increase in the amylose leaching and solubility and loss of starch
386 crystalline structure. High WAC of composite flours suggests that the flours can be used in
387 formulation of some foods such as sausage, dough and bakery products. The increase in the
388 WAC has always been associated with increase in the amylose leaching and solubility, and
389 loss of starch crystalline structure. The flour with high water absorption may have more
390 hydrophilic constituents such as polysaccharides. Protein has both hydrophilic and
391 hydrophobic nature and therefore they can interact with water in foods. The good WAC of
392 composite flour may prove useful in products where good viscosity is required such soups
393 and gravies. The observed variation in different flours may be due to different protein
394 concentration, their degree of interaction with water and conformational characteristics (Butt
395 and Batool, 2010).

396 **Oil absorption capacity (OAC)**

397 The composite flours (D and E) had highest OAC (2.72 and .44) and lowest for B (1.90). It is
398 clear that the OAC of composite flours increased with increase in the proportion of other
399 flours. The presence of high fat content in flours might have affected adversely the OAC of
400 the composite flours. The OAC was found to be insignificant to each other at $p \leq 0.05$ level of
401 significance. Therefore, the possible reason for increase in the OAC of composite flours after
402 incorporation of millet flour is the variations in the presence of non-polar side chain, which
403 might bind the hydrocarbon side chain of the oil among the flours. Similar findings were
404 observed by Kaushal *et al.* (2012). However, the flours in the present study are potentially
405 useful in structural interaction in food specially in flavor retention, improvement of
406 palatability and extension of shelf life particularly in bakery or meet products where fat
407 absorption is desired (Aremu *et al.* 2007). The major chemical component affecting OAC is
408 protein which is composed of both hydrophilic and hydrophobic parts. Non-polar amino acid
409 side chains which can form hydrophobic interaction with hydrocarbon chains of lipids
410 (Jitngarmkusol *et al.* 2008).

411

412

413 **Sensory Scores**

414 Table 4 shows the sensory scores of the samples tested. Appearance for sample A, B and C
415 was not significant ($P < 0.05$) difference level but was significant ($P < 0.05$) different level from
416 D and E. flavour shows that there are no significant ($P < 0.05$) difference level in all the
417 samples tested. In terms of texture, there are no significant ($P < 0.05$) difference level between
418 samples A, B and C and between samples B and C and also between sample C, D and D, E.
419 But there are significant ($P < 0.05$) difference level between sample A and E, B and E and C
420 and D. the general Acceptability indicates that there are no significant difference ($P < 0.05$)
421 between samples A, B, and C; samples B, C and D; samples C, D and E and between sample
422 D and E but there are significant difference ($P < 0.05$) between sample A and E, B and E. The
423 sensory scores and general acceptability shows that sample A (7.66) was the most preferred
424 amongst all the tested sample followed by sample B (7.47) and C respectively.

425 **CONCLUSION**

426 The addition of baobab fruit pulp (BFP) to pearl millet and soybean flour, in turn increases
427 the fibre, ash and carbohydrate contents of the complementary foods. The functional
428 properties also improved with addition of baobab fruit pulp levels. This improvement could
429 be noticed in water absorption capacity, oil absorption capacity, bulk density and swelling
430 index. The sensory attributes indicates that the baobab fruit pulp samples competes very well
431 with the control (A) sample. However, sample A was most preferred by the panellist.

432 **Acknowledgement**

433 We wish to acknowledge all the Authors who articles, books etc we used.

434

Table 1: Blend Formulation of Pearl Millet, Soybean flour and Baobab Fruit Pulp (%)

Composite Flour

SAMPLES	MAIZE	SOYBEAN	BAOBAB FRUIT PULP
A	50	50	0
B	50	40	10
C	60	20	20
D	65	10	25
E	65	5	30

435

436

Table 2: Effect of Baobab Fruit Pulp Addition on the Proximate Composition a Complementary Food Samples.

SAMPLES	MOISTURE	PROTEIN	FAT	CARBOHYDRATE	FIBRE	ASH	ENERGY Kcal
A	10.98 ^c ±0.40	24.25 ^a ± 0.23	16.65± 0.01	43.96 ^a ± 0.76	3.37 ^a ± 0.02	2.75 ^a ±0.00	423.69±0.00
B	10.50 ^a ±0.40	20.38 ^a ± 0.18	13.90± 0.08	43.11 ^a ± 0.34	7.68 ^b ±0.08	2.65 ^c ±0.03	379.06±0.01
C	10.27 ^a ±0.40	14.58 ^b ± 0.30	8.84± 0.00	62.00 ^b ±0.30	11.57 ^c ±0.08	2.68 ^a ±0.02	385.88±0.03
D	10.73 ^a ±0.40	11.51 ^b ± 0.93	5.62±0.01	67.91 ^b ±0.02	13.51 ^d ±0.06	2.59 ^b ±0.02	368.26±0.00
E	10.09 ^b ±0.40	9.80 ^c ± 0.62	4.94±0.02	71.03 ^c ±0.21	15.67 ^e ±0.05	2.87 ^a ±0.01	367.78±0.02
LSD	0.08	0.06	0.02	0.01	0.02	0.09	0.08
PAG	5 - 10	20	10	-	5	10	350 - 400

Values are means of standard deviation. Values in the same column with different superscript are significantly (P,0.05) different

437 Key:

438 A = Millet 50%, soybean 50%

439 B = Millet 50%, soybean 40% and Baobab fruit pulp 10%

440 C = Millet 60%, soybean 20% and Baobab fruit pulp 20%

441 D = Millet 65%, soybean 10%and Baobab fruit pulp 25%

442 E = Millet 65%, soybean 5% and Baobab fruit pulp 30%

443 LSD = Least significant difference

444 PAG = Protein Advisory Group

445

Table 3: Effect of Baobab Fruit Pulp addition on The Functional Properties of a Complementary Food from Pearl Millet and Soy flour

SAMPLES	GELATION (%)	BULK DENSITY(g/ml)	Swelling Index (g/vol)	WAC	OAC
A	5.00±1.22	0.71±0.009	0.68±0.13	2.83±0.084	2.11±0.31
B	5.00±1.22	0.71±0.009	0.87±0.13	2.84±0.084	1.90±0.31
C	8.00±1.22	0.71±0.009	0.79±0.13	2.91±0.084	2.21±0.31
D	8.00±1.22	0.71±0.009	0.79±0.13	2.70±0.084	2.72±0.31
E	10.00±1.22	0.69±0.009	1.04±0.13	2.70±0.084	2.44±0.31

Means in the same column with different superscript are significantly ($p<0.05$) different

447

448 **Key:**

449 A = Millet 50%, soybean 50%

450 B = Millet 50%, soybean 40% and Baobab fruit pulp 10%

451 C = Millet 60%, soybean 20% and Baobab fruit pulp 20%

452 D = Millet 65%, soybean 10% and Baobab fruit pulp 25%

453 E = Millet 65%, soybean 5% and Baobab fruit pulp 30%

Table 4: Effect of Baobab Fruit Pulp on The Sensory Attributes of a Complementary Food from Pearl Millet and Soy flour

SAMPLES	Appearance	Flavour	Texture	General Acceptability
A	7.26 ^a	6.60 ^a	6.53 ^a	7.66 ^a
B	7.20 ^{ab}	6.40 ^{ab}	6.33 ^{ab}	7.47 ^{ab}
C	7.13 ^{abc}	6.00 ^{abc}	6.07 ^{abc}	7.20 ^{abc}
D	6.53 ^{abcd}	5.73 ^{abcd}	5.40 ^{abd}	6.73 ^{abc}
E	5.80 ^d	5.27 ^{abcd}	4.67 ^d	5.33 ^{cd}
LSD	0.974	1.390	1.334	1.086

Means in the same column with different superscript are significantly ($p < 0.05$) different

455 **Key:**

456 A = Millet 50%, soybean 50%

457 B = Millet 50%, soybean 40% and Baobab fruit pulp 10%

458 C = Millet 60%, soybean 20% and Baobab fruit pulp 20%

459 D = Millet 65%, soybean 10% and Baobab fruit pulp 25%

460 E = Millet 65%, soybean 5% and Baobab fruit pulp 30%

461 **COMPETING INTERESTS**

462 Authors have declared that no competing interests exist.

463

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