

1
2
3
4
5
6
7
8
9

Original Research Article
**Reduction of pathogens in palisade grass
seeds by contact with fertilizer**

ABSTRACT

The objective of this study was to evaluate the effects of the contact of *Brachiaria* seeds with the fertilizer 05-25-15 on the sanitary quality of the seeds. Experiment was carried out in a completely randomized design, in a 2x5 factorial, with four replications. Treatments consisted of the contact times (0, 24, 48, 72 and 96 hours) of the fertilizer with seeds of ruziziensis grass and marandu grass. After the contact times with the fertilizer with *Brachiaria* seeds, the seeds were separated from each species and the analyzes were carried out: water content, germination, sanitation ("Blotter Test"), and an accelerated, with later germination test and sanitary analysis of seeds. Data were analyzed for variance and regression analysis at the significance level of 5%, and the descriptive analysis for the results of the sanitary analysis. Contact time reduces the incidence of pathogens in seeds of ruziziensis grass and marandu grass treated with fertilizer due to salinity and acid pH from the fertilizer. The fungi related to the reduction in germination were *Aspergillus* sp. and *Fusarium* sp. Disinfestation process increased the incidence of *Fusarium* sp., under high internal infestation of this fungus in palisade grass seeds.

10
11
12

Keywords: Aspergillus sp.; Fusarium sp.; Brachiaria sp.; accelerated aging; blotter test; germination.

13 **1. INTRODUCTION**

14
15
16
17
18
19

The crop-livestock integration system is an alternative to promote socioeconomic and sustainable development before a growing food demand and the need to reduce the deforestation [1], increasing the production system efficiency [2]. In addition to the financial benefits [3,1], there is synergism between pastures and annual crops, such as: improvement of soil physical, chemical and biological properties, control of diseases, pests and weeds [4].

20
21
22
23
24

This system can be used by farmers with the succession of grain crop and annual forage, in rural properties where agriculture with annual summer crops predominates, mainly soybeans and maize. Besides, it can be used as an alternative to soil cover in the form of straw for the no-till system and as an income source in the off-season (Santa Fe System), or in properties where the main activity is the cattle raising [4].

25
26
27
28
29
30

Based on the implantation premises of the Santa Fe system, where the grain crop is intercropped annually with a forage crop, we highlight the corn and *Brachiaria* (Syn. *Urochloa*) *ruziziensis* cv. Kennedy (Ruziziensis palisade grass). To a deployment of the Barreirão system, which a grain crop is intercropped and / or rotated with a forage crop that acts in the area as a perennial culture, the most used species are corn and *Brachiaria* (Syn. *Urochloa*) *brizantha* cv. Marandu (Marandu palisade grass) [5].

31
32

The corn culture is an alternative for the second crop in the Cerrado (Brazilian Savanna) due to its adaptability to the region edaphoclimatic conditions, with several commercial cultivars

33 for the production of grains or silage [5]. In addition, it stands out in integrated systems
34 because it has competitiveness in intercropping, suppresses weeds, presents selective
35 herbicides, increases the surface soil residue input and maximizes nutrient cycling, being a
36 good economic-environmental option of production [6]. For the corn crop implantation, in
37 MatoGrosso state, it is commonly used the fertilizer 05-25-15 (N-P₂O₅-K₂O).

38 Tropical forages of the genus *Brachiaria* (Syn. *Urochloa*) are known for their adaptation to
39 tropical climate and soil conditions, with high potential for dry matter production under
40 adequate temperature and soil moisture conditions. In addition, they present a broad range
41 of grazing height, greater regrowth and soil cover capacity, as well as lower clumps
42 formation and greater ease of desiccation [7]. The forage plants desiccation promotes root
43 death, increasing soil porosity, due to the formation of canaliculi, and the organic matter
44 content, nutrients source for soil microorganisms [8].

45 Marandu palisade grass use expanded in the central region of Brazil due to adaptation to
46 edaphoclimatic conditions, dry mass production with medium soil fertility, and resistance to
47 spittlebug [9], a major problem in the region. In contrast, ruziziensis palisade grass stands
48 out for its rapid soil cover, chemical composition, palatability, excellent nutrient recycling,
49 ease of desiccation and uniform seed production, since it only blooms once [10].

50 To minimize the problems arising from the forage sowing in integrated systems, such as the
51 lack of uniformity in the initial stand, due to the reduced size and low weight of the seeds,
52 simultaneous sowing of the grain crop seeds with the forage seeds is commonly done,
53 associating them to the fertilizer applied at the grain-producing crop seeding [11].

54 However, the contact of the seeds with the fertilizer can influence the seeds sanitary quality,
55 altering the incidence of pathogens associated to the seeds due to the fertilizer salinity and
56 pH. The goal was to evaluate the contact time effect of fertilizer 05-25-15 (N-P₂O₅-K₂O). on
57 the sanitary quality of palisade grass seeds.

58 2. MATERIAL AND METHODS

59
60 The trial was carried out in a completely randomized design, with four replications.
61 Treatments consisted of a 5x2 factorial, with five contact times (0, 24, 48, 72 and 96 hours)
62 of the fertilizer 05-25-15 (N-P₂O₅-K₂O) to the *Brachiaria* (Syn. *Urochloa*) *ruziziensis* cv.
63 Kennedy (ruziziensis palisade grass) and to the *Brachiaria* (Syn. *Urochloa*) *brizantha* cv.
64 Marandu (Marandu palisade grass). The fertilizer consisted of monoammonium phosphate,
65 single superphosphate, triple superphosphate and potassium chloride, with a saline index of
66 70.59% and pH 4.83.

67 For each grass species, the associations of the seeds with the fertilizer were carried out,
68 which were transferred to closed plastic packages and stored until the pre-established
69 contact times. Then, the fertilizer was removed from the mixtures and the grasses seeds
70 were submitted to the analyses of water content, germination, sanitation ("Blotter Test" with
71 salt stress), and accelerated aging, with later germination test and sanitary conditions
72 analysis of seeds.

73 To determine the water content, three 4.0 g samples were placed in a drying oven for 24
74 hours at a temperature of 105 ± 1 °C, for each species and treatment. After the drying
75 process, the samples were placed in desiccators to promote the cooling and then the
76 weighing was carried out in analytical scale (0.0001 g). The results were expressed as
77 percentage [12].

78 The methodology used for the standard germination test was the one described in the Rules
79 of Seed Analysis [12], in which four sub-samples of 50 seeds were used for each species
80 and treatment. The seeds were placed equidistantly in plastic box (gerbox) on two sheets of
81 blotter paper as substrate, moistening them with distilled water in the ratio of two and a half
82 times the dry mass of the paper.

83 Subsequently, the boxes were sealed with film paper, to reduce moisture loss, and taken to
84 the BOD (Biochemical Oxygen Demand) chamber with photoperiod and temperature
85 regulation (12 hours of light at 35 °C and 12 hours in the absence of light at 20 °C). The
86 moisture inside the gerbox was maintained with the addition of distilled water. At 21 days the
87 germinated seeds were evaluated, considering germinated the seeds that had emitted 2 mm
88 of root. The results were expressed as percentage.

89 Seed sanitary analysis was performed according to the "Blotter Test" method [13] modified
90 with water restriction [14]. For each treatment and species of grass, 100 pure and viable
91 seeds and 100 pure, viable and disinfested seeds were used. The disinfestation process
92 was performed in a laminar flow chamber by immersing the seeds in a 1% sodium
93 hypochlorite solution for three minutes, and then the seeds were washed with sterile distilled
94 water [13].

95 Then, the seeds were distributed equidistantly in a gerbox containing three sheets of filter
96 paper, previously moistened with sterile sodium chloride solution (-0.6 MPa) [14], in a
97 proportion equivalent to two and a half times the dry mass of the substrate. During the
98 analyses period the moisture of the substrate was maintained by the addition of sterile
99 sodium chloride solution (-0.6 MPa) to restrict the seeds germination and ensure the
100 accurate evaluation of the incident microorganisms.

101 The seeds were incubated in a BOD chamber under a constant temperature of 20 °C and a
102 12-hour photoperiod [13]. After seven days, the seeds individual evaluation was carried out
103 with the aid of stereoscopic and biological microscopes. The fungi was identified by the
104 morphological structures observation and with the aid of specialized literature. The results
105 were expressed as percentage of fungi incidence [13].

106 The accelerated aging test was realized by the methodology proposed by the author cited in
107 reference [15], in which the seeds of each treatment were distributed on aluminium screen
108 attached to the gerbox with 40 mL of distilled water in the recipient. The boxes were then
109 capped, forming a wet chamber, and placed in a BOD chamber for a period of 36 hours at a
110 temperature of 42 °C. After the accelerated aging, the germination and the sanitation test
111 (Blotter Test adapted) with seed evaluation were carried out at seven days.

112 Data were submitted to analysis of variance and regression analysis at a significance level of
113 5% of probability ($P < 0.05$) and descriptive analysis for the results of the sanitary analysis.

114 **3. RESULTS**

115 In the water content analysis, there was no effect of the contact time and neither interaction
116 between forages and contact times. An average water content of 7.91% was observed.

117 In the germination standard test there was interaction effect of contact time x forage species.
118 The contact time to the fertilizer did not affect the germination of the ruziziensis palisade
119 grass seeds, with an average value of 75.10%. However, the evaluation of the marandu
120 palisade grass seeds showed a reduction in germination percentage as the seeds contact
121 time to the fertilizer was prolonged (Fig. 1).

Ruziensiis	6,36	2,96	78,66	73,33	71,08	5,40	63,42	35,20
Marandu	5,05	2,05	81,33	62,33	21,21	3,80	63,72	40,23

149 ¹WOD: without disinfestation; ²WD: with disinfestation.

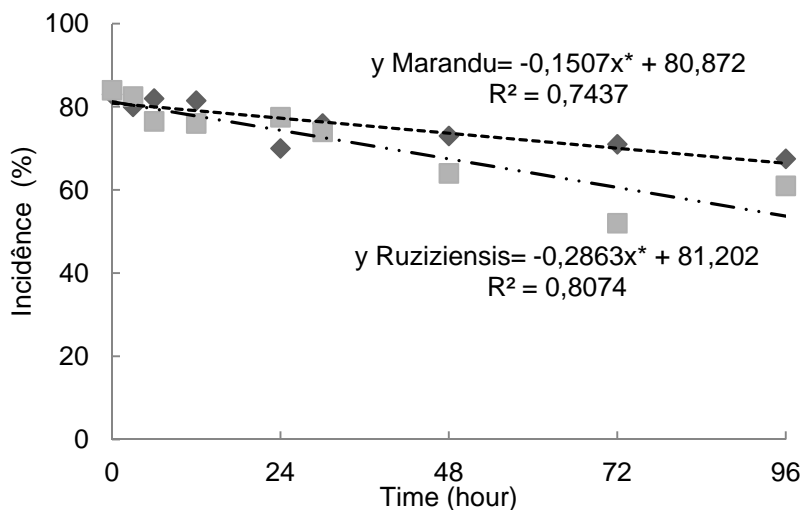
150 There was a high percentage of *Fusarium* sp. in seeds treated with fertilizer (78.66 and
151 81.33% in ruziensiis palisade grass and marandu palisade grass, respectively), and in
152 seeds treated with fertilizer exposed to accelerated aging (63.42 and 63.72%, in ruziensiis
153 palisade grass and marandu palisade grass, respectively) (Table 1).

154 The incidence of *Fusarium* sp. in seeds remained high in fertilized and disinfested seeds
155 (73.33 and 62.33% in ruziensiis and marandu palisade grass seeds, respectively), and in
156 seeds treated with fertilizer and exposed to accelerated and disinfested aging (35.20 and
157 40.23% in ruziensiis and marandu palisade grass seeds, respectively) (Table 1).

158 It was observed that the percentage of *Fusarium* sp. was higher in seeds treated with
159 fertilizer than in seeds treated with fertilizer and subjected to accelerated aging, independent
160 of the disinfestation process (Table 1).

161 The increase in contact time of the marandu and ruziensiis palisade grass seeds with the
162 fertilizer decreased the incidence of fungi (Fig. 2). When comparing the incidence of fungi
163 in the absence of contact with fertilizer (time zero) with the maximum time studied (96 h), it was
164 observed a reduction of 18.67 and 27.38% in the seeds of ruziensiis palisade grass and
165 marandu palisade grass, respectively.

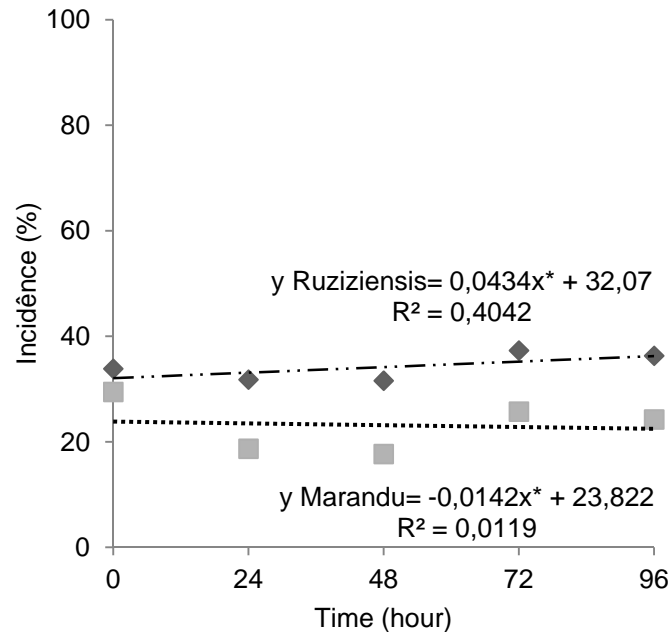
166 In relation to the seeds of ruziensiis and marandu palisade grass treated with fertilizer and
167 exposed to accelerated aging, when comparing the incidence of fungi in the absence of
168 contact with fertilizer (time zero) with the maximum time studied (96 h), it was observed
169 increase of 13.00% and reduction of 5.42% in the seeds of ruziensiis palisade grass and
170 marandu palisade grass, respectively.



171

172 **Fig. 2. Percentage of fungi incidence in ruziensiis and marandu palisade grass seeds**
173 **as a function of the contact time with the fertilizer 05-25-15, after the**
174 **treatments. *Significant at 5% probability ($P = .05$).**

175 When analysing the accelerated aging test, the germination percentage of the ruzizensis
176 palisade grass seeds was reduced up to 42 hours of contact to the fertilizer, possibly due to
177 the interference of the external pathogens intense sporulation after accelerated aging,
178 confirmed by the sanitary analysis (Fig. 3).



179

180 **Fig. 3. Percentage of fungi incidence in aged seeds of ruzizensis and marandu**
181 **palisade grass as a function of the contact time with the fertilizer 05-25-15.* Significant**
182 **at 5% probability ($P = .05$).**

183 4. DISCUSSION

184 In the analysis of water content, the seeds contact time to the fertilizer does not interfere in
185 the seeds water content, although the used fertilizer consists of single and triple
186 superphosphate and potassium chloride, besides having a high saline index (70.59%) and
187 acidic pH (4.83).

188 Differently, the authors cited in the reference [16] studying ruzizensis palisade grass seeds
189 submitted to contact with urea, obtained positive linear effect for the water content, as a
190 consequence of the fertilizer high hygroscopicity. The authors cited in the reference [17, 18,
191 19, 20] observed an increase in the water content of marandu palisade grass seeds during
192 the contact time to fertilizers due to the high urea hygroscopicity, the acid phosphate
193 fertilizers obtainment, and the high saline index of potassium chloride.

194 In the evaluation of the ruzizensis palisade grass seeds germination percentage, it was
195 verified that there was no effect of the seeds contact time to the fertilizer on the germination.
196 However, a reduction was observed for the marandu palisade grass seeds as the contact
197 time to the fertilizer increased (Fig. 1), due to tegument rupture and extravasation of
198 electrolytes by saline effect [21].

199 The authors cited in the reference [17,18, 19] verified a reduction in the germination
200 percentage of marandu palisade grass as the seeds contact time to the fertilizers was
201 prolonged, corroborating with the present study.

202 The same phenomenon was not observed for the ruziense palisade grass seeds, probably
203 due to the tegument being less susceptible to damage by intrinsic factors to the fertilizer,
204 such as acid pH and saline effect. Further studies on the tegument constitution of the
205 species used in the present work are necessary.

206 There was no effect of the seeds contact time to the fertilizer on the germination of the
207 marandu palisade grass seeds submitted to accelerated aging. The phenomenon may be
208 related to reduced sporulation of external pathogens, as shown in Fig. 3, derived from the
209 sanitary quality of the seeds.

210 Based on this, it is concluded on the importance of seed health, since the aging test predicts
211 the behaviour of the seeds stored, and it was verified in this study the interaction between
212 the germination decrease of aged seeds and the low sanitary quality of the seeds.

213 From the seeds sanitary analysis, it was verified that the results of fungi incidence obtained
214 by the authors cited in the reference [22, 23, 24, 25] corroborate with those found in the
215 present study (*Alternaria* sp., *Aspergillus* sp., *Bipolaris* sp., *Cladosporium* sp., *Cercospora*
216 sp., *Fusarium* sp., *Nigrospora* sp., *Penicillium* sp., *Rhizoctonia* sp. e *Rhizopus* sp.).

217 The authors cited in the reference [22] have similarly identified the fungi *Alternaria* sp.,
218 *Aspergillus* sp., *Cladosporium* sp., *Fusarium* sp., *Penicillium* sp. and *Rhizopus* sp. in
219 *Brachiaria* (Syn. *Urochloa*) *brizantha* seeds. Whereas, other authors cited in the reference
220 [23] reported a similar incidence of *Bipolaris* sp. and *Cladosporium* sp. in the marandu
221 palisade grass seeds produced in Mato Grosso, besides *Alternaria* sp. and *Fusarium* sp. In
222 seeds of *Brachiaria* (Syn. *Urochloa*) sp. and *Panicum maximum*.

223 In studies carried out by the authors cited in the reference [24, 25] it was verified the
224 presence of fungi *Aspergillus niger*, *Bipolaris* sp., *Fusarium* sp., *Penicillium* sp. e *Rhizopus* sp.
225 in *Brachiaria* (Syn. *Urochloa*) *brizantha* cv. BRS Piatã (piatã palisade grass). In addition, the
226 authors cited in the reference [25] observed the presence of *Alternaria* sp., *Aspergillus flavus*,
227 *Aspergillus ochraceus* and *Cladosporium* sp.

228 The main fungi evidenced in this work were *Aspergillus* sp. and *Fusarium* sp., in both forage
229 species, and may be related to the reduction in the seeds germination [25], causing damage
230 to the quality and establishment of forage plants [26, 24].

231 In addition to the physiological damage caused to *Brachiaria* seeds, both fungi can produce
232 mycotoxins under low humidity conditions, which can lead to intoxication, cancer and death if
233 ingested by animals [27].

234 The fungi that can be transmitted by seeds, with infestation / infection capacity during
235 storage and in the physiological maturity point, interfere in seed quality, reducing
236 germination and forage production by compromising the establishment, mainly under
237 development favourable conditions and inefficient control method [25, 28, 26]. In the
238 transmission of seed pathogens to seedlings, the authors cited in the reference [25] reported
239 the occurrence of pathogens *Aspergillus* sp. and *Fusarium* sp.

240 It is observed that the use of low quality sanitary seed results in unsuccessful pasture
241 formation and seed lots commercialization, due to the presence of fungi and nematodes [23].

242 Based on the seeds treated with fertilizer and subjected to accelerated aging (Table 1), a
243 high incidence of *Aspergillus* sp. on the surface of the *Brachiaria* seeds can occur, due to the
244 high incidence of this pathogen after the process of disinfestation and accelerated aging on
245 the seeds.

246 In accelerated aging, the seeds are submitted to high temperature and humidity, which are
247 favourable conditions for sporulation and development of pathogens, mainly of the seeds
248 outer layer. In this sense, it was observed that the percentage of *Aspergillus* sp. was higher
249 in *Brachiaria* seeds treated with fertilizer and subjected to accelerated aging than in
250 *Brachiaria* seeds treated with fertilizer, regardless of the disinfestation process (Table 1).

251 The contamination of the seeds by *Aspergillus* sp. can cause damage to the physiological
252 quality of seed, reducing germination and vigor[27], causing a reduction of the planting
253 stand, as well as being a inoculum source for the development of diseases and introducing
254 pathogens in unaffected regions [29], such as the disease of burned grains in corn ears [30].

255 The incidence of *Fusarium* sp. in *Brachiaria* seeds (treated with fertilizer and treated with
256 fertilizer and exposed to accelerated aging) remained high after the disinfestation process,
257 evidencing the possibility of high incidence in the interior of the seeds (Table 1).

258 In addition, the incidence of *Fusarium* sp. was higher in *Brachiaria* seeds treated with
259 fertilizer than in seeds treated with fertilizer and submitted to accelerated aging, regardless
260 the disinfestation process (Table 1), confirming the high incidence of the pathogen inside the
261 *Brachiaria* seeds, once that the accelerated aging test provides the proper conditions (high
262 temperature and humidity) to the development and sporulation of fungi in the outer layer.

263 As a consequence of the fungi, incidence there may be a reduction in the viability
264 percentage of the lots or death of the *Brachiaria* seeds, once *Fusarium* sp. is a soil fungus
265 that can be associated with the seeds [30, 31].

266 By having alternative hosts (corn, sorghum, sugarcane, grass, among others), crop rotation
267 is not a very efficient control practice in these cases. Among them, *Fusariummoniliforme* and
268 *Fusariumgraminearum*, stand out for causing stalk and root rot to infected plants (Costa et
269 al., 2009), and *Fusariumclamydosporium* for causing wilt symptom followed by death in
270 forage plants such as *Stylosanthes* sp. [31].

271 It is observed that the contact of the marandu and ruziensi palisade grass seeds to the
272 fertilizer occasioned improves the seeds sanitization, since the contact time prolongation to
273 the fertilizer decreased the fungi incidence.

274 The effect of dormancy overcoming due to the accelerated aging process was excluded,
275 since the germination percentage of ruziensi palisade grass seeds submitted to
276 accelerated aging in the absence of contact (time zero) to the fertilizer was lower (74.90%)
277 than the germination percentage obtained in the standard germination test (75.10%) (Fig. 1).
278 The authors cited in the reference [32] observed a reduction in the germination percentage
279 of marandu palisade grass seeds after 24 hours of aging at 43°C.

280 The increase in the contact time to the fertilizer of the *Brachiaria* seeds exposed to
281 accelerated aging leads to an increase in the fungi incidence on ruziensi palisade grass
282 seeds and reduces the fungi incidence on marandu palisade grass seeds (Fig. 3). This effect
283 can be attributed to the accelerated aging process of the seeds treated with fertilizer, which
284 promotes the proliferation and development of fungi due to the optimal conditions provided
285 (Fig. 3).

286 **4. CONCLUSION**

287

288 It is concluded that there is presence of the fungi *Bipolaris* sp., *Fusarium* sp., *Rhizoctonia*
289 sp., *Cercospora* sp., *Alternaria* sp., *Aspergillus* sp., *Cladosporium* sp., *Penicillium*
290 sp., *Rhizopus* sp., *Nigrospora* sp. in the seeds of ruziziensis and marandu palisade grass,
291 treated with fertilizer and, treated with fertilizer and exposed to accelerated aging.

292 The fungi related to the reduction in the germination of *Brachiaria* seeds are *Aspergillus* sp.
293 and *Fusarium* sp., with a high internal infestation of *Fusarium* sp.

294 It is verified that the increase in the contact time to the fertilizer of the *Brachiaria* seeds
295 treated with fertilizer reduces the incidence of fungi, improving the sanitary quality of the
296 seeds.

297

298 **COMPETING INTERESTS**

299

300 We declare that no competing interests exist.

301

302

303

304

305 **REFERENCES**

306

307 1. Vilela LMartha Junior GB, Macedo MCM, Marchão R L, Guimarães Júnior R., Pulrolnik, K
308 et al. Integration systems for livestock farming in the Cerrado region. Agricultural Research.
309 2011; 46 (10): 1127-1138. English. DOI: 10.1590 / S0100-204X2011001000003.

310

311 2. Balbino LC, Barcellos AO, Stone, LF. Reference frame: crop-livestock-forest
312 integration. Brasília: Embrapa; 2011. English.

313

314 3. Macedo MCM. Crop-livestock integration: the state of the art and technological
315 innovations. Revista Brasileira de Zootecnia. 2009; 38: 133-146. Portuguese. DOI: 10.1590 /
316 S1516-35982009001300015.

317

318 4. Leonel FP. Integration of crop-livestock: production and quality of fodder. In: Zervoudakis
319 JT, Cabral LS, editors. Nutrition and production of debovine cuttings. Page 2 221-
320 233. Portuguese.

321

322 5. Freitas FCL, Ferreira LR, Ferreira FA, Santos MV, Agnes EL, Cardoso AA et al. Pasture
323 formation via *Brachiaria* consortium with corn for silage in no - tillage system. Weed
324 plant. 2005; 23: (1) 49- 58. Portuguese. DOI: 0.1590 / S0100-83582005000100007.

325

326 6. Cobucci T, Wruck FJ, Kluthcouski J, Muniz LC, Martha Junior GB, Carnevalli RA et al.
327 Options for crop-livestock integration and some of its economic aspects. Agricultural Report.
328 2007; 28: (240) 64-79. Portuguese.

329

330 7. Almeida RG, Barbosa, RA, Zimmer, AH, Kichel, NA. Forages in systems of cattle
331 production in integration. In: Bungenstab DJ, editor. Systems of crop-livestock-forest
332 integration: sustainable production. Field of study: Embrapa, 2011; 25-36. Portuguese.

333

- 334 8. Mellow LMM. Integration of crop-livestock in no-tillage system. In: Brazilian Congress of
335 Soil Science. Vicosa: Brazilian Society of Soil Science, 2003. Annual ... Portuguese.
336
- 337 2. Balbino LC, Barcellos AO, Stone, LF. Reference frame: crop-livestock-forest
338 integration. Brasília: Embrapa; 2011. English.
339
- 340 9. Valle CB, Euclides VPB, Pereira JM, Valério JR, Pagliarini MS, Macedo MCM et al. Xaraés
341 grass (*Brachiaria brizantha* cv. Xaraés) in the diversification of pastures of *Brachiaria*. Campo
342 Grande: Embrapa, 2004. 36 p. Portuguese.
343
- 344 10. Trecenti R. Consortium techniques help in the formation of straw for no-till. Direct Plant
345 Review. 2005; 86. Portuguese.
346
- 347 11. Physiological quality of millet seeds (*Panicum dichotomiflorum* Mix) as a function of
348 mixing time with triple superphosphate. Agronomic Culture. 2000; 9 (1): 177-
349 189. Portuguese.
350
- 351 12. Brazil Ministry of Agriculture, Livestock and Food Supply. Rules for seed analysis.
352 Brasília DF. MAP / ACS. 2009a. Portuguese.
353
- 354 13. Brazil Ministry of Agriculture, Livestock and Food Supply. Rules for sanitary seed
355 analysis. Brasília DF. MAP / ACS. 2009b. English.
356
- 357 14. Machado AQ, Machado JC, Vieira MDGGC, Cassetari Neto D, Souza MV. Potential of
358 the use of water restriction in sanitary tests of cotton seeds. Brazilian Phytopathology. 2007;
359 32: (5) 408-414. Portuguese. DOI: 10.1590 / S0100-41582007000500006.
360
- 361 15. Marcos Filho J. Accelerated aging test. In: Krzyzanowski FC, Vieira RD, France Neto JB.
362 Seed vigor: concepts and tests. Londrina: Abrates; 1999. English.
363
- 364 16. Dan HA, Dan LGM, Barroso ALL, Braccini AL, Puccinin GG. Mixture of seeds of
365 *Brachiaria ruziziensis* G. et E with urea aiming at the implantation of the crop-livestock
366 integration system. Caatinga Magazine. 2011; 24 (4): 68-73. English.
367
- 368 17. Lima EDV, Tavares JC, Silva EC, Leitão-Lima P. Triple superphosphate as route of
369 distribution of *Brachiaria brizantha* seeds for renewal of pastures in the Amazon. Revista
370 Brasileira de Zootecnia. 2009; 38 (5): 796-800. English. DOI: 10.1590 / S1516-
371 35982009000500003.
372
- 373 18. Lima EDV, Tavares JC, Azevedo VR, Leitão-Lima PS. Mixture of *Brachiaria brizantha*
374 seeds with NPK fertilizer. Rural Science. 2010; 40 (2): 471-474. English. DOI: 10.1590 /
375 S0103-84782010005000003.
376
- 377 19. Lima EV, Tavares JCS, Leitão-Lima PS, Pinheiro DP. Contact periods of the KCl
378 fertilizer on the physiological quality of *Brachiaria brizantha* Stapf seeds. Amazon: Science &
379 Development. 2013; 38 (5): 53-64. English.
380
- 381 20. Peres AR, Vazquez GH, Cardoso RD. Physiological potential of *Brachiaria brizantha* cv.
382 Marandú seeds kept in contact with phosphatic fertilizers. Brazilian Journal of Seeds. 2012; 34
383 (3): 424-432. Portuguese. DOI: 10.1590 / S0101-31222012000300009.
384

- 385 21. Mateus GP, Borghi E, Marques RR, Bôas RLV, Crusciol CAC. Fertilizer sources and
386 contact periods and seed germination of *Brachiariabrizantha*. Brazilian Journal of Soil
387 Science. 2007; 31 (1): 177-183. English. DOI: 10.1590 / S0100-06832007000100018.
388
- 389 22. Days DCFS, Toledo FF. Germination and incidence of fungi in tests with
390 *Brachiariabrizantha* (Hochst.exA.Rich) seeds Stapf. Agricultural Science. 1993; 1 (3): 68-76.
391 DOI: 10.1590 / S0101-31222008000300019.
392
- 393 23. Mallmann G, Verzignassi JR, Fernandes CD, Santos JM, Vechiato MH, Inácio CA,
394 Batista MV, Queiroz CA. Fungi and nematodes associated with tropical forage seeds.
395 SummaPhytopathol. 2016; 39 (3) 201-203.Portuguese.
396
- 397 24. Santos GR et al. Sanity analysis, transmission and pathogenicity off fungi associated with
398 forage plant seeds in tropical regions of Brazil. Journal of Seed Science. 2014;(1)36:54-
399 62.DOI:10.1590/S2317-15372014000100007.
400
- 401 25. Sbalcheiro CC, Jose SCBR, Barbosa JCRCM. Physiological and sanitary quality, and
402 transmission of fungi associated with *Brachiariabrizantha*(Hochst. ex. A. Rich.) Stapf seeds
403 submitted to thermal and chemical treatments. Journal of Seed Science.2014;36(4)443-
404 450.Portuguese. DOI: 10.1590/2317-1545v36n41032.
405
- 406 26. Marchi CE, Fernandes CD, Bueno ML, Batista MV, Fabris LR. Fungi propagated by
407 commercial seeds of *Brachiaria*. Archive Biological Institute. 2010; 77: (1) 65-73.Portuguese.
408
- 409 27. Vechiato MH, Aparecido CC, Fernandes CD. Frequency of fungi in commercial seed lots
410 of *Brachiaria* and *Panicum*. Campo Grande: Embrapa. 2010. English.
411
- 412 28. Marchi CE, Fernandes CD, Borges CT, Santos JM, Jerba VF, Trentin RA, Guimarães,
413 L.R.A. Nematofaunapitopathogenic of tropical forage commercial seeds. Pesquisa
414 Agropecuária Brasileira. 2007; 42: 655-660.Portuguese. DOI: 10.1590 / S0100-
415 204X2007000500007.
416
- 417 29. Soave J, Whetzel MMVS. Pathology of seeds. Campinas: Cargill Foundation. 1987.
418 English.
419
- 420 30. Costa RV, Casela CR, Cota LV. Corn Crops: Diseases. In: Cruz JP, Magalhães PC,
421 Pereira Filho IA, Moreira JAA, editors. Sete Lagoas, Embrapa. 2009; 138-169.
422
- 423 31. Verzignassi JR, Fernandes CD. Diseases in forages. Campo Grande, Embrapa. 2001.
424
- 425 32. Meschede DK, Sales JGC, Braccini ADL, Scapim CA, Schuab SRP. Treatments to
426 overcome the dormancy of the seeds of *Brachiaria* grass marandu cultivar. Brazilian Journal
427 of Seeds. 2004; 26 (2) 76-81.Portuguese.