

Original Research Article

Growth, Photosynthesis and Quality of Water Spinach (*Ipomoea aquatica*) as influenced by Magnetic nanoparticles (MNP) application

ABSTRACT

Aims: To characterize the growth, carbon assimilation and quality of *Ipomoea aquatica* as influenced by magnetic nanoparticles (MNP) application as well as to determine the best rates of iron oxide nanoparticles (0, 50, 100, and 150 mg/L) that give high growth, carbon assimilation and quality of *Ipomoea aquatica*.

Study design: *Ipomoea aquatica* plants were exposed to four different treatments of magnetic iron oxide nanoparticles (Fe_3O_4) (0, 50, 100 and 150 mg/L). The experiment was conducted in a randomized complete block design (RCBD) with 3 replications. One unit of experiment consisted of 8 plants and there were 96 plants that have been utilized in the experiment.

Place and Duration of Study: Department of Biology, Faculty of Science, Universiti Putra Malaysia, between March 2018 and July 2018.

Methodology: The growth parameters measured include plant height, basal diameter, total leaf number, leaf temperature, total chlorophyll content and plant biomass. The carbon assimilation parameters were measured using LICOR 6400 XT Portable Photosynthesis System i.e. transpiration rate (E), stomatal conductance and water use efficiency (WUE). Total phenolics and flavonoids contents from the leaves extracts were measured using Folin-Ciocalteu reagents.

Results: It was observed that plant height, shoot length, plant temperature, total biomass, and total chlorophyll content were significantly influenced ($P \leq 0.05$) by the different concentration of magnetic nanoparticles. The net photosynthesis rate (A), transpiration rate (E), stomata conductance (gs), maximum efficiency of photosystem II (Fv/fm), maximum quantum yield of phytochemical and non-photochemical process in photosystem II (Fv/fo), performance index and the density of reaction centers per PSII antenna chlorophyll of *Ipomoea aquatica* were significantly reduced at higher concentration of magnetic nanoparticles. However, water use efficiency and minimal fluorescence value (Fo) of *Ipomoea aquatica* increased with the increased of MNP concentration. In addition, the application of magnetic nanoparticles had significantly influenced ($P \leq 0.05$) the total flavonoids and total phenolics content in water spinach. Both of these parameters were increased when higher concentration of magnetic nanoparticles was applied to *Ipomoea aquatica*. This study showed that application of MNP had affected the growth, carbon assimilation and secondary metabolites production of *Ipomoea aquatica*.

Conclusion: In conclusion, the higher concentration of magnetic nanoparticles can reduce the growth rate and carbon assimilation of water spinach and enhanced the production of secondary metabolites.

Keywords: [*Ipomoea aquatica*, magnetic nanoparticles, growth, carbon assimilation, secondary metabolites]

Comment [T11]: mg L⁻¹

Comment [T12]: mg L⁻¹

Comment [T13]: IRGA (Infrared Gas Analyser, LICOR 6400 XT)

Comment [T14]: Put all the parameters that were analyzed

Comment [T15]: Stomatal

1
2
3
4
5
6
8
9
10

11
12
13
14

15 1. INTRODUCTION

16
17 Based on the data from Department of Statistics Malaysia, agriculture sector was operated
18 at 11,628 establishments in the year 2015 with an annual growth of 5.7 percent [1] and
19 vegetables were contributing to agricultural commodities in which spinach, long bean,
20 mustard, brinjal, tomato and cucumber were selected by which more than 100 percent were
21 recorded in self-sufficiency ratio [2]. Malaysia population in 2018 is estimated at 32.4 million
22 compared to 32.0 million in 2017 by which 1.1 per cent as the growth rate [3]. As the
23 population increases, the demand for food and commercial energy is accelerating to fulfill
24 the population requirements. According to Department of Agriculture Malaysia, Kangkung is
25 one of the vegetables suggested by Malaysia government to be consumed widely due to its
26 low price and Asia had consumed water spinach at the highest rate compared to other
27 vegetables [4].
28

29 | *Ipomoea aquatica* which is also known as ~~kk~~kangkung is a popular vegetable in countries like
30 Malaysia, Hong Kong, Taiwan and other Asia countries. This edible vegetable is classified
31 into the family Convolvulaceae. Kangkung is an aquatic or semi-aquatic yearly herb which
32 usually creeps on moist soil or sand besides floating in water [5]. Countries like Southern
33 | Asia, Bangladesh, China and India had been using ~~Ipomoea aquatica~~ *Ipomoea aquatica* in folk medicine
34 against different diseases including diabetes, malfunction of liver, constipation and Arsenic
35 poisoning as it is known for its high nutritive values and consumable leafy vegetables [5].
36 Water spinach is also rich in minerals like flavonoids, phenolics and carotenes. As water
37 spinach is easily grown plant, has short time of harvesting period and well adapted with
38 environment changes, this enables the plant favorable to be cultivated. However, water
39 spinach could accumulate foreign minerals like Cadmium, Zinc and Copper which enables it
40 to be used as the sample plant for research [6].
41

42 Metals such as nickel, cobalt, and iron are used to demonstrate magnetic properties in which
43 their magnetic particle size are within nanoscale [7]. Currently, magnetic nanoparticles have
44 attracted researchers from different background like biotechnology, biomedicine, agriculture,
45 magnetic fluids and data storage in which different ways are used in synthesizing those
46 magnetic nanoparticles. However, the application of magnetic nanoparticles relies on the
47 particle's condition on its steadiness [8]. The application of magnetic nanoparticles in the
48 study of plant currently been interested by the researchers for its ability to permit a particular
49 localization to discharge their load as conveyed to the plants. Recent studies being applied
50 on pumpkin plants to test on the specific localization, take-up, and translocation of magnetic
51 nanoparticles (under 50 nm) [9]. Besides, gas exchange is also being influenced by the use
52 | of magnetic nanoparticles in which they act on ~~photo-synthetic~~photosynthetic surface
53 causing foliar warming and changes physiological process and cell elements of plants as the
54 leaves face with stomatal obstacle [10].
55

56 | The application of magnetic nanoparticles may enhance or retard the growth of ~~Ipomoea~~
57 *Ipomoea aquatica* somehow like carbon nanotubes that increased the leaf gas trades properties of the
58 plant. It was seen in *Arabidopsis thaliana* that treated with single wall carbon nanotubes
59 (SWCNT) the plant had higher photosynthetic, photoabsorption and higher electron transport
60 rates contrasted with the plant that not treated with the materials. This was expected to a
61 higher productivity of chloroplast when cooperated with the nanomaterials [11].
62

63 Secondary metabolites are also an important factor being influenced by the magnetic
64 nanoparticle application. Secondary metabolites are the natural products presence in low
65 amounts in which its production rely on different species, genera and families. Secondary
66 metabolites are important to protect plants from insects and pathogens besides shaping
67 imperative UV-radiation absorbing compounds that eventually reduce the chances for the

Comment [T16]: eggplant

Comment [T17]: Put scientific name after
common name

Comment [T18]: Use kangkung or water spinach.
Do not use both. Standardize throughout the text.

Comment [T19]: Confuse

68 plant to die [12]. The production of flavonoids and phenolics are the determinants for the
69 production of secondary metabolite in plant. It is often to observe that plant undergoing
70 stress has higher total flavonoid and phenolics content due to defensive mechanism in plant.
71 Biosynthesis in plant to produce secondary metabolites is usually stimulated by the
72 accumulation of heavy metals like zinc, iron, and nickel which generates Reactive Oxygen
73 Species (ROS) and induces oxidative stress in plant. Consequently, the induction of
74 oxidative stress causes changes in signal transduction for the mechanism of gene coding
75 and enzyme [13]. Besides, the production of ROS may cause damages to cell membrane,
76 cell structure and photosynthetic site and thus the production of flavonoid at the generation
77 site act as defensive mechanism due to its high antioxidant properties. In addition, induced
78 phenolics are produced when the plant faces physical injury, infection or environmental
79 stresses due to heavy metal irradiation or temperature [14].
80

81 The low toxicity level of magnetic nanoparticles such as super paramagnetic nanoparticles
82 (SPION) has been the reason for growing studies on the application of magnetic
83 nanoparticles on organisms. A study shows that iron oxide nanoparticles is safe and non-
84 cytotoxic at the level of $100 \mu\text{g mL}^{-1}$ when being compared with few metal oxide
85 nanoparticles *in vitro* [15]. In one of the studies related to iron oxide nanoparticles exposure
86 on sunflower plant, *Helianthus annuus*, it was seen that at the concentration of 50-100 mg L^{-1}
87 the exposure resulted in reduction of root water pressure of the plant [16]. Besides, a study
88 by Liu, Zhang and Lal [17] shown that the iron oxide nanoparticles was less toxic besides
89 stimulating root elongation during the germination of lettuce, *Lactuca sativa* at the
90 concentration of 5–20 mg L^{-1} while inhibited root elongation at 50 mg L^{-1} [18].
91

92 Magnetic nanoparticles application is currently recognized in biomedicine to treat various
93 diseases but still far behind in plant biology. Magnetic-based materials can be used in
94 production of certain chemical to protect plant systematically from diseases besides
95 controlling externally the movement of nanocarriers in the plant by using high power external
96 magnet [19]. As water spinach is well known for its importance in culinary as vegetable and
97 in traditional medicine, this study would help the plant to be continuously used in studies.
98

99 The study of *Ipomoea aquatica* in the aspects of biochemical and physical towards
100 magnetic nanoparticles is still far behind in current research. Besides, the research on the
101 impacts of magnetic nanoparticles on growth, carbon assimilation and quality of plant in form
102 of secondary metabolites production is still few in science studies. Hence, this research was
103 conducted with few objectives which are to relate the growth, carbon assimilation and quality
104 of *Ipomoea aquatica* as influenced by magnetic nanoparticles application, to determine the
105 best concentration of magnetic nanoparticles that can promote the optimum growth and
106 quality of *Ipomoea aquatica* and to infer the relationship between growth and secondary
107 metabolites of *Ipomoea aquatica* as affected by magnetic nanoparticles application. The
108 study hypothesized that the application of magnetic nanoparticles influence growth, carbon
109 assimilation and secondary metabolites of *Ipomoea aquatica* while the alternative
110 hypothesis stated that the application of magnetic nanoparticles does not influence growth,
111 carbon assimilation and secondary metabolites of *Ipomoea aquatica*.
112

113 114 2. MATERIAL AND METHODS

115 116 2.1 Synthesis of Iron Oxide Nanoparticles (Fe_3O_4)

117 The iron oxide nanoparticles (Fe_3O_4) were synthesized using co-precipitation method [20].
118 Iron (III) chloride powder and Iron (II) chloride powder were mixed with the ratio of 2:1 in a
119 250 mL conical flask and were dissolved in 150 mL of deionized water. The solution was
120

Formatted: Superscript

Formatted: Superscript

Formatted: Superscript

Formatted: Superscript

121 heated at 45 °C bubbled with nitrogen gas for 15 minutes. The solution was then added with
122 20 mL of 25-30-% ammonia solution and stirred at 800 rpm for an hour. Then, the Iron (III)
123 oxide nanoparticles produced was collected by magnetic decantation. The Fe₃O₄ product
124 was then washed 3 times with acetone and centrifuged. The product was left to dry in
125 furnace one night and grinded after drying process to obtain the powder form. Fe₃O₄
126 produced were in black colored fine powder.

128 2.2 Plant Materials and Maintenance

129
130 The experiment was conducted at the Vegetables Field plot for Teaching and Research,
131 Taman Pertanian Universiti, Universiti Putra Malaysia. The source of planting materials in
132 the study was the seeds of *Ipomoea aquatica*. Seeds of *Ipomoea aquatica* were propagated
133 for 14 days in a tray and transplant to polybags containing a mixture of top soil and sand
134 (ratio 3:1). After 1 month, *Ipomoea aquatica* plants were exposed to four different
135 treatments of Fe₃O₄ (0, 50, 100 and 150 mg L⁻¹). The magnetic nanoparticles were
136 conveyed to the plants through watering. The magnetic nanoparticles were diluted in distilled
137 water before being applied to the plants. Each plant was watered with 40 mL of magnetic
138 nanoparticle solution. To maintain the plant growth and avoid plant wilting or attacked by any
139 major plant disease that can make the plant die, maintenance was done time to time.

Formatted: Superscript

141 2.3 Experimental Design

142
143 The polybags were arranged accordingly to Completely Randomized Block Design (RCBD)
144 with replication of 3 blocks. One unit of experiment consisted of 8 plants, and there were
145 total of 96 plants utilized in the experiment.

147 2.4 Data Collection

148
149 Data collection of plant growth were collected on Week 3, 6, 9 and 12 while the destructive
150 analysis and leaf gas exchange measurement were conducted on the 12th week.

152 2.4.1 Plant Growth Measurement

153
154 The plant growth measurement was done to obtain the wWater sSpinach plant height,
155 number of leaf, diameter of stem, root to shoot ratio, plant temperature and the chlorophyll
156 content.

158 2.4.1.1 Plant height

159
160 Measuring tape was used to measure the plant height which starting from stem at soil
161 surface up to the highest shoot growth or at its tip.

163 2.4.1.2 Plant basal diameter

164 The basal tips of the plant were measured by using vernier caliper.

168 2.4.1.3 Plant leaves number

169
170 The whole leaves of the plants were counted manually (one by one piece) and then recorded
171 in every 3 weeks.

172 2.4.1.4 Plant leaf temperature determination

174
175 Infrared (IR) Thermometer was used to measure plant leaf temperature.
176

177 **2.4.1.5 Chlorophyll content measurement**

178 Total chlorophyll content in the leaves was measured using SPAD 502 chlorophyll meter
179 (Spectrum Tech Inc; Aurora, IL; USA). The leaf was clipped under the chlorophyll meter
180 clipper to obtain the reading for every treatment in each of the replication. The data was
181 collected every week from week zero until week 12 of the experiment.
182

183 **2.4.1.6 Plant fresh weight measurement**

184 The plants were removed from the soil carefully and the dirt from the soil was rinsed with tap
185 water. The shoot, root and the leaf parts were separated and weighed using analytical
186 balance.
187

188 **2.4.1.7 Plant biomass (dry weight) measurement**

189 All the plants were dried for 2 days in the oven at 60 °C. The dry weight of root, stem and
190 leaf per seedling were recorded as plant biomass.
191

192 **2.4.1.8 Root to shoot ratio**

193 The root to shoot ratio was determined by dividing the dry weight of the root with the dry
194 weight of the shoot.
195

196 **2.4.2 Leaf gas exchange measurement**

197 A LI-6400XT (Li-COR Inc; Nebraska; USA) portable photosynthesis system gave the reading
198 of leaf gas exchange. The instrument was warmed and calibrated for 30 minutes with the
199 ZERO IRGA mode. The measurements used optimal conditions set at 400 $\mu\text{mol mol}^{-1} \text{CO}_2$,
200 30 °C cuvette temperature, 60% relative humidity with air flow rate set at 500 $\text{cm}^3 \text{min}^{-1}$ and
201 modified cuvette condition of 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ photosynthetically photon flux density (PPFD).
202 The fully expanded young leaves were used to measure net photosynthesis (A),
203 transpiration rate (E) and stomata conductance (gs) which also gave the measurement of
204 gas exchange. It was an automatic operation therefore the data was recorded in the LI-
205 6400XT console and further analyzed by Photosyn Assistant Software (Dundee Scientific,
206 Dundee, UK). Precautions step were considered while taking the measurements to avoid
207 errors.
208

Formatted: Superscript

Formatted: Superscript

Formatted: Superscript

Formatted: Superscript

Formatted: Superscript

209 **2.4.3 Chlorophyll fluorescence determination**

210 Mature leaf tissue of water spinach was collected from plants which are grown in a
211 greenhouse at 20 °C with the supply of artificial light which provide a minimum photon flux
212 density of 550 $\mu\text{mol m}^{-2} \text{s}^{-1}$ for a 16 h photoperiod (Fi-totron H600, Fisons pic,
213 Loughborough, U.K.) and photosynthetically active radiation will be supplied at 250 $\mu\text{mol m}^{-2}$
214 s^{-1} during 16 h photoperiods.
215

Formatted: Superscript

Formatted: Superscript

216 **2.4.4 Total phenolics and flavonoids quantification**

217 The total phenolics and flavonoid measurement follow methods from Ibrahim, Jaafar,
218 Rahmat, and Zaharah [21]. Plant tissue samples (0.1 g) that initially grounded were
219 extracted with 80% ethanol (10 mL) on an orbital shaker at 50 °C for 120 minutes. The
220

227 mixture was filtered and the filtrate used for the quantification of total flavanoids and total
228 phenolics. Total phenolics content of the leaf samples was determined by using Follin-
229 Ciocalteu reagent (Sigma Aldrich, Missouri, USA; diluted 10-fold). The absorbance
230 measured at 725 nm. The results will be expressed as mg g⁻¹ gallic acid equivalent (mg
231 GAE g⁻¹ dry sample). For total flavonoids determination, a sample (1 mL) mixed with NaNO₃
232 (Sigma Aldrich, Missouri, USA; 0.3 mL) in a test tube which covered with aluminum foil, and
233 left for 5 min. Then 10% AlCl₃ (Wako Pure Chemical Industries Ltd; Tokyo, Japan; 0.3 mL)
234 added followed by addition of 1 M NaOH (Kanto Chemical Co. Inc.; Hokkaido, Japan; 2 mL).
235 Later, the absorbance was measured at 510 nm using a spectrophotometer with rutin as a
236 standard (results expressed as mg g⁻¹ rutin dry sample).

Comment [T110]: Flavonoids or flavanoids?

Formatted: Superscript

Formatted: Superscript

Formatted: Subscript

Formatted: Superscript

238 2.5 Statistical Analysis

239
240 The recorded data was analyzed by using statistic software known as Statistical Package for
241 Social Sciences (SPSS) with the version 24. A two-way ANOVA test was carried out for
242 data analysis for all the parameters in the experiment. Data was significant if the p -value
243 level ≤ 0.05 .

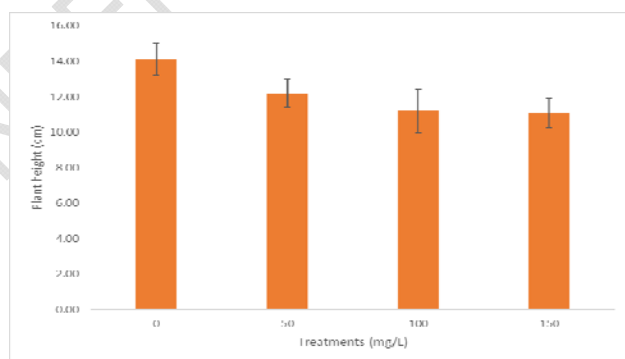
244 3. RESULTS AND DISCUSSION

246 3.1 Plant Height

247
248 Fig.1 showed that, different concentration of magnetic nanoparticles application influenced
249 significantly the plant height of *Ipomoea aquatica* as $P \leq 0.05$ in week 3. The highest plant
250 height was observed significantly in 0 mg/L (14.10 cm), followed by 50 mg/L (12.2 cm), 10
251 mg/L (11.2 cm) and lowest in 150 mg/L (11.1 cm). The current result indicates that the
252 application of magnetic nanoparticles at higher concentration would reduce the plant height
253 of *Ipomoea aquatica*. Research had shown that higher concentration of Fe₃O₄ with pro-long
254 exposure may lead to iron toxicity in the plant. The plants that faced toxicity during growing
255 stage may initiate stress and causes reduction in plant height due to consequences effects
256 of iron toxicity like production of free radicals, root break down and bronzing of leaves which
257 lead to yield loss [22]. The application of Fe₃O₄ stimulate the production of Reactive Oxygen
258 Species (ROS) under the condition of excess or deficiency of Fe₃O₄ that responsible of
259 signaling molecule that stimulate or inhibit plant growth [23].
260
261

Comment [T111]: Quantitative factors (with more than three levels) should be analyzed by regression analysis and not by means test. Repeat the analysis by placing the regression curves with their respective equations. When it is a quadratic effect, derive the equation and place the maximum/minimum point on the results. Redo the results according to the new statistical analysis.

Comment [T112]: The correct one is mg L⁻¹. Rectify in throughout the text

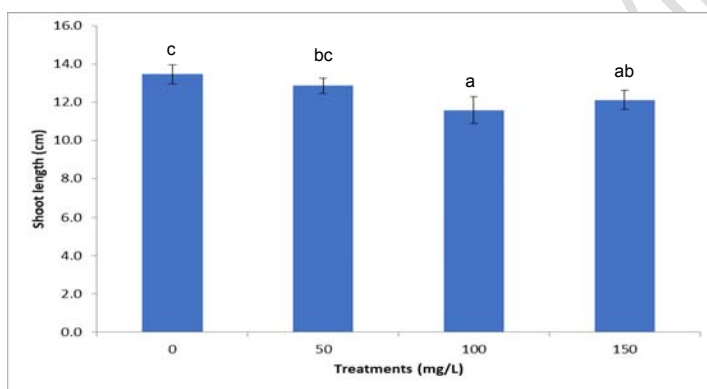


262

263 Fig. 1. The impact of magnetic nanoparticles on plant height of *Ipomoea aquatica*
264 Data are mean \pm standard error of mean (SEM). N= 24. Mean with different superscript
265 showed the significantly different at $P \leq 0.05$ using DMRT at 95% confidence levels.

3.2 Shoot Length

In this study, the shoot length of *Ipomoea aquatica* has been influenced significantly by the application of magnetic nanoparticle ($P \leq 0.05$; Fig. 2) in week 3. The longest shoot length was observed in 0 mg/L (13.5 cm) followed by 50 mg/L (12.9 cm), 150 mg/L (12.1 cm) and shortest for 100 mg/L (11.6 cm). From the result, we could observe that the shoot length of *Ipomoea aquatica* increased initially with the application of magnetic nanoparticles at both lower and higher concentration than 100 mg/L treatment. However, water spinach treated with 100 mg/L reduces the shoot elongation. Overall, the application of magnetic nanoparticles increased the shoot length of water spinach. From other researches, Fe_3O_4 between 100-200 mg kg^{-1} had shown positive effect on plant like *Spinacea oleracea* and no any effect at lower concentration which is lower than 100 mg/L on *Helianthus annus*. The stem length of *Spinacea oleracea* increased with the application of Fe_3O_4 while when there is no oxidative stress induced in the plant, no positive or negative effect is shown on shoot length like in *Helianthus annus* [18]. The shoot length of *Vigna radiata* applied with Fe_3O_4 had also shown positive effect [24].



283

284 **Fig. 2. The impact of magnetic nanoparticles on shoot length of *Ipomoea aquatica***
285 **Data are mean \pm standard error of mean (SEM). N= 24. Mean with different superscript**
286 **showed the significantly different at $P \leq 0.05$ using DMRT at 95% confidence levels.**
287

3.3 Basal Diameter

288

289

290

291

292

293

294

295

296

297

298

299

300

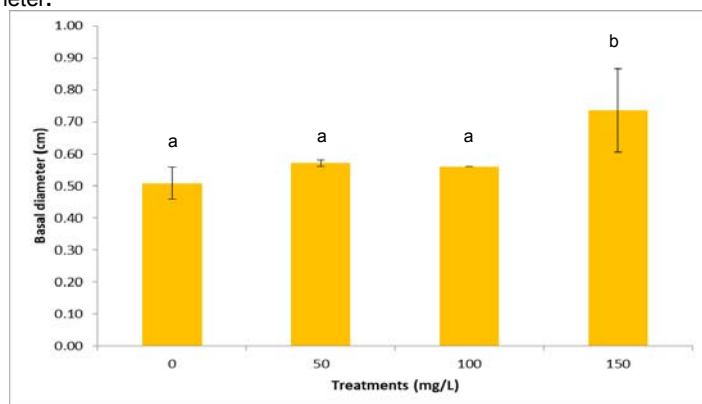
301

302

303

Different concentration of magnetic nanoparticles application has significantly influenced the basal diameter of *Ipomoea aquatica* ($P \leq 0.05$; Fig. 3) in week 12. At 12th week, the application of magnetic nanoparticles has shown to increase significantly the basal diameter of *Ipomoea aquatica* at higher concentration which is at 150 mg/L. The smallest basal diameter was observed in 0 mg/L (0.51 cm), followed by 100 mg/L (0.56 cm), 50 mg/L (0.571 cm) and greatest in 150 mg/L (0.74 cm). The current result indicates that the application of magnetic would promote the stem diameter of *Ipomoea aquatica*. Based on research, aquaporins are found on tonoplast or also known as vacuolar membrane which allows water to be freely moves across the cells in the symplastic route. Therefore, the abundance of aquaporins found on the membrane depends on the regulation of water flow. During water stress due to high concentration of nanoparticles may lead to induction of turgor-responsive aquaporins to maintain turgor pressure. This would cause the vacuole size to increase that brings to cell expansion and increased of stem diameter [25]. Besides, a research done on radish cotyledon shows that increase in turgor pressure increase the cell expansion and

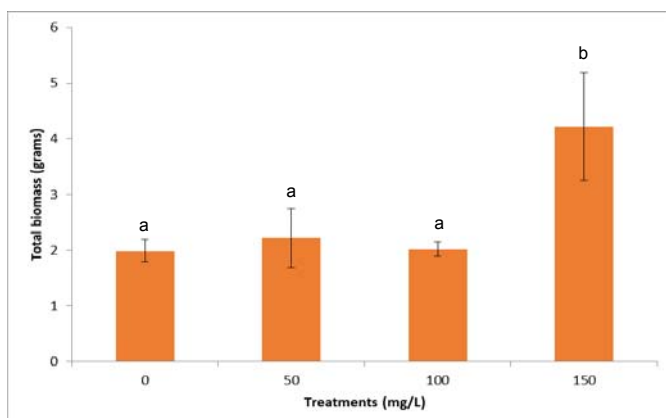
304 growth. However, the study claimed that under unfavorable condition, cell would not divide
305 but [26]. Therefore, we could conclude that higher concentration of Fe_3O_4 cause increase in
306 stem diameter.



307
308
309 **Fig. 3. The impact of magnetic nanoparticles on basal diameter of *Ipomoea aquatica***
310 **Data are mean \pm standard error of mean (SEM). N= 24. Mean with different superscript**
311 **showed the significantly different at $P \leq 0.05$ using DMRT at 95% confidence levels.**
312

313 3.4 Total Biomass

314
315 Based on Fig. 4, different concentration of magnetic nanoparticles had influenced
316 significantly the total biomass of *Ipomoea aquatica* as $P \leq 0.05$ in week 12. In week 12, a
317 significant effect was observed by which the application of Fe_3O_4 with the concentration of
318 150 mg/L increased significantly the biomass of water spinach. The highest total biomass
319 was observed in 150 mg/L (4.2179 gram), followed by 50 mg/L (2.2189 gram), 100 mg/L
320 (2.0213 gram), and 0 mg/L (1.9824 gram). The result obtained showed that iron oxide
321 nanoparticles with highest concentration would increase the total biomass of *Ipomoea*
322 *aquatica*. This result was in agreement with the finding by [27]. From their research, it was
323 found that the use of iron oxide nanoparticles increased the dry weight of pod and peanut
324 plant. They found that the application of Fe_3O_4 help in transferring iron and photosynthate
325 particle in the leaves of plants. Besides, the total biomass of *Vigna radiata* applied with
326 Fe_3O_4 showed a positive result compared to the application of ferum ions. This was claimed
327 due to the increase of α -amylase activity in the seeds exposed to Fe_3O_4 . The increase of
328 total biomass was also observed in *Spinacea oleracea* applied with magnetic nanoparticles
329 [18]. However, study by Jeyasubramanian et al. [28] showed that higher concentration of
330 Fe_3O_4 which is 200 mg/L cause decrease in both wet and dry weight of spinach [29].

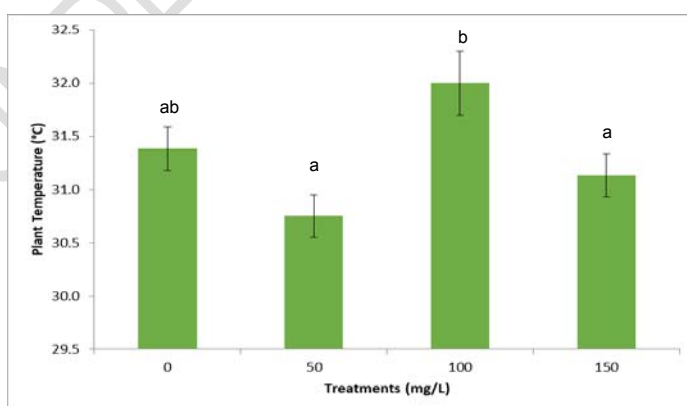


331 **Fig. 4. The impact of magnetic nanoparticles on total biomass of *Ipomoea aquatica***
 332 **Data are mean \pm standard error of mean (SEM). N= 24. Mean with different superscript showed**
 333 **the significantly different at $P \leq 0.05$ using DMRT at 95% confidence levels.**
 334
 335

3.5 Plant Leaf Temperature

336
 337
 338 Fig. 5 showed that magnetic nanoparticle had influenced significantly the plant temperature
 339 of *Ipomoea aquatica* as $P \leq 0.05$ in week 12. The highest plant temperature was observed for
 340 100 mg/L, 32.0 °C followed by 0 mg/L (31.4 °C), 150 mg/L (31.1 °C) and 50 mg/L (30.8 °C) in
 341 week 12. The result shows that a higher concentration of magnetic nanoparticles shows
 342 higher leaf temperature of water spinach. Plant temperature is often related to transpiration
 343 process. Transpiration is a cooling process taken by the plant to release water vapor from
 344 the plant through stomata and cuticle. Through transpiration, thermal energy is balanced by
 345 the loss of heat to its surrounding. Therefore, when the transpiration rate is decreased due to
 346 the accumulation of nanoparticles at the root surface and inhibits the water intake capacity,
 347 the plant temperature would increase. Besides, the stomata conductance also plays a major
 348 role in maintaining the leaf plant temperature. The closing and opening of stomata do give
 349 impact on the plant temperature [30]. Thus, higher concentration of Fe_3O_4 result higher plant
 350 temperature due to its relation with transpiration rate and stomata conductance.
 351

Comment [T113]: Review this statement (note the last level)



353 Fig. 5. The impact of magnetic nanoparticles on plant temperature of *Ipomoea*
354 *aquatica*. Data are mean \pm standard error of mean (SEM). N= 24. Mean with different
355 superscript showed the significantly different at $P\leq 0.05$ using DMRT at 95%
356 confidence levels.

357

3.6 Total Chlorophyll Content

358

359

360

361

362

363

364

365

366

367

368

369

370

371

372

373

374

375

376

377

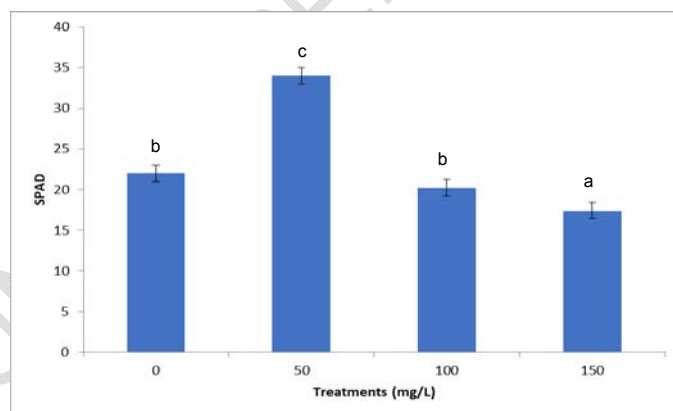
378

379

380

381

Fig. 6 shows the impact of magnetic nanoparticle on total chlorophyll content of water spinach. There was significant effect of the application of magnetic nanoparticles on the water spinach chlorophyll content at $P\leq 0.05$. After 12 weeks of treatment, the highest chlorophyll content was observed in *Ipomoea aquatica* treated with 50 mg/L Fe_3O_4 (34), followed by 0 mg/L (22) and 100 mg/L (20) and lowest in 150 mg/L (17). This result indicates that higher concentration of magnetic nanoparticles lead to decrease of chlorophyll content in water spinach which was supported by several studies. Based on research by Racuciu et al. [31], *Zea mays* plant treated with 20, 40, 60, 80, and 100 $\mu\text{l/l}$ of Fe_3O_4 suspension concentration showed decreased in chlorophyll content. The magnetic nanoparticles may showed both chemical and magnetic influence on the water spinach enzymatic structure that eventually influence the photosynthetic system of the plant at higher concentration. Besides, the application of nanoparticles would induce oxidative stress in plant that reduced the chlorophyll content in plant leaf. For example, zinc oxide nanoparticles had proven to reduce the chlorophyll content in wheat plant due to the formation of free radical [32]. Another research done on watermelon showed that the application of magnetic nanoparticles at higher concentration loss the content of chlorophyll. Due to the toxic substance exposure to the plant, the malondialdehyde (MDA) production could be observed as a result of lipid peroxidation. Low level of MDA is important in protecting the structure and function of cell membranes. As the increase of MDA in plant with the presence of Fe_3O_4 , the penetration of large particles into the cell is disturbed that result in less efficiency of cell. Thus, iron is claimed to be the cause for enzymatic activity inhibition that reduce chlorophyll synthesis [33].



382

383

384

385

386

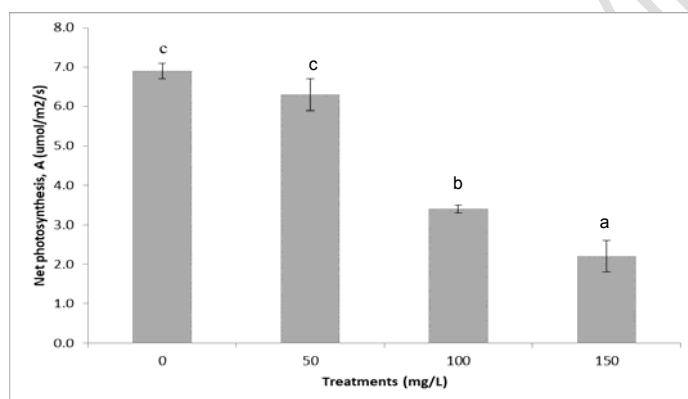
Fig. 6. The impact of magnetic nanoparticles on total chlorophyll content of *Ipomoea aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with different superscript showed the significantly different at $P\leq 0.05$ using DMRT at 95% confidence levels.

387

3.7 Net Photosynthesis Rate

388 | Fig. 7 showed the net photosynthesis rate (A) of water spinach after 12 weeks of planting.
 389 | The results shown that the net photosynthesis rate was influenced significantly ($P \leq 0.05$) by
 390 | the application of Fe_3O_4 during the planting period. The higher concentration of magnetic
 391 | nanoparticles resulted in lower net photosynthesis rate of water spinach. The net
 392 | photosynthesis of control treatment (0 mg/L) is higher than other treatments which are 50
 393 | mg/L, 100 mg/L and 150 mg/L that recorded 6.9 $\mu\text{mol}/\text{m}^2/\text{s}$, 6.3 $\mu\text{mol}/\text{m}^2/\text{s}$, 3.4 $\mu\text{mol}/\text{m}^2/\text{s}$,
 394 | and 2.2 $\mu\text{mol}/\text{m}^2/\text{s}$ respectively. The photosynthesis rate could be altered due to the
 395 | application of magnetic nanoparticles by which these nanoparticles could block the pathway
 396 | and causes stress to the water spinach. Although nanoparticles are in the size of nanometer
 397 | but their entry into the plant may cause some changes either by enhancing or inhibiting the
 398 | photosynthesis rate. In this case, the eElectron tTransport cChain may be blocked by the
 399 | nanoparticles and enhance stress to the plant manipulating and changing the normality of
 400 | genes and enzymes like Rubisco. Rubisco is one of the enzymes that play an important role
 401 | in the conversion of carbon dioxide into biological substances [34]. Therefore, the changes in
 402 | production of Rubisco may lead to lower net photosynthesis rate with the application of
 403 | magnetic nanoparticles.

Comment [TI114]: The correct one is $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$. Rectify in throughout the text



404

405 | **Fig. 7. The impact of magnetic nanoparticles on net photosynthesis rate of *Ipomoea***
 406 | ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with different**
 407 | **superscript showed the significantly different at $P \leq 0.05$ using DMRT at 95%**
 408 | **confidence levels.**

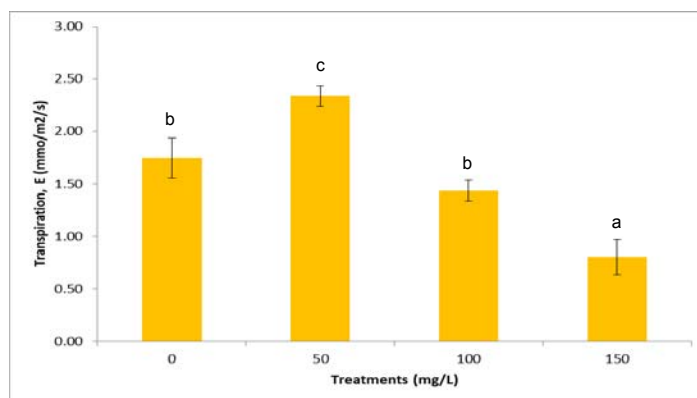
409 | 3.8 Transpiration Rate

410 | Based on Fig. 8, the transpiration rate of *Ipomoea aquatica* was significantly influenced by
 411 | the application of magnetic nanoparticles after 3 months of experimental period at $P \leq 0.05$.
 412 | The transpiration rate was highest observed in 50 mg/L treatment followed by the control
 413 | treatment, 100 mg/L and 150 mg/L of Fe_3O_4 by 2.34 $\text{mmol}/\text{m}^2/\text{s}$, 1.75 $\text{mmol}/\text{m}^2/\text{s}$, 1.44
 414 | $\text{mmol}/\text{m}^2/\text{s}$, and 0.80 $\text{mmol}/\text{m}^2/\text{s}$ respectively. This indicates that higher concentration of
 415 | Fe_3O_4 would decrease the transpiration rate while an optimum concentration could inhibit the
 416 | transpiration rate in which in this case is 50 mg/L of iron oxide nanoparticles. Transpiration
 417 | rate is linked up by photosynthesis rate by means if there is reduction in photosynthesis rate,
 418 | the transpiration rate would decrease too. Besides, nanoparticles applied to the plant may
 419 | cover the root surface of the plant and causes water stress in water spinach. This is
 420 | supported by a research using titanium oxide nanoparticle in maize in which the water
 421 | transport capacity of the primary cell wall was reduced due to accumulation of nanoparticles
 422 | at the plant leaf [35]. Besides, the research done on watermelon to study the application of

Comment [TI115]: $\text{mmol H}_2\text{O m}^{-2} \text{ s}^{-1}$

Comment [TI116]: Put scientific name

423 magnetic nanoparticles on the root activity showed that the Fe_3O_4 accumulated at the root
424 surface that prevent the transmission of water and also other nutritional components by the
425 plant [33].
426



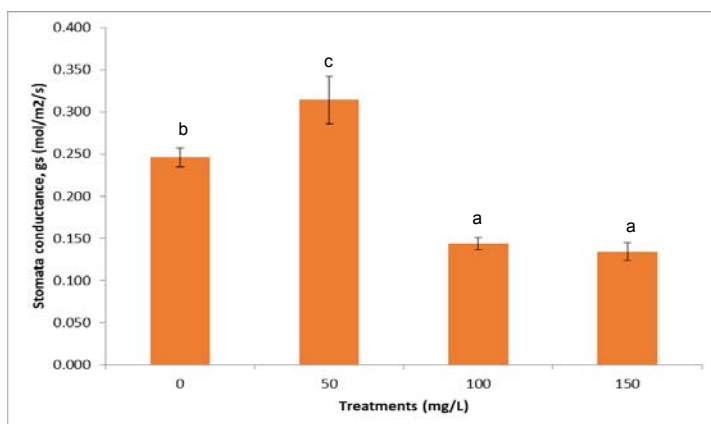
427

428 **Fig. 8. The impact of magnetic nanoparticles on transpiration rate of *Ipomoea***
429 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with different**
430 **superscript showed the significantly different at $P \leq 0.05$ using DMRT at 95%**
431 **confidence levels.**

432 3.9 Stomata Conductance

433 The stomata conductance of water spinach was significantly affected by the application of
434 magnetic nanoparticles as showed in Fig. 9 as $P \leq 0.05$. After 12 weeks of treatment
435 application, the stomata conductance for 50 mg/L Fe_3O_4 was the significantly highest
436 compared to 0 mg/L, 100 mg/L and 150 mg/L with 0.314 mol/m²/s, 0.246 mol/m²/s, 0.144
437 mol/m²/s, and 0.134 mol/m²/s respectively. From the result, we could observe that higher
438 concentration of Fe_3O_4 would decrease the stomata conductance of water spinach. Stomata
439 conductance is determined by the degree of stomata aperture which estimates the rate of
440 gas exchange and transpiration rate. A greater conductance is shown when the degree of
441 stomata opening and its function in term of density and size is greater. When the plant
442 undergoes greater photosynthesis and transpiration rate, the stomata conductance is
443 greater. This statement is proven by the result of this experiment by which when the
444 photosynthesis and transpiration rate of water spinach is lower for higher concentration of
445 Fe_3O_4 , the stomata conductance follows the same pattern. The application of magnetic
446 nanoparticles may cause leakage of electrolyte in the plant which alters the mechanism of
447 potassium pump that controls stomata opening. The accumulation of magnetic nanoparticles
448 may cause the stomata to be closed or partially closed which directly reduce the stomata
449 conductance [14].

Comment [T117]: mol H₂O m⁻² s⁻¹



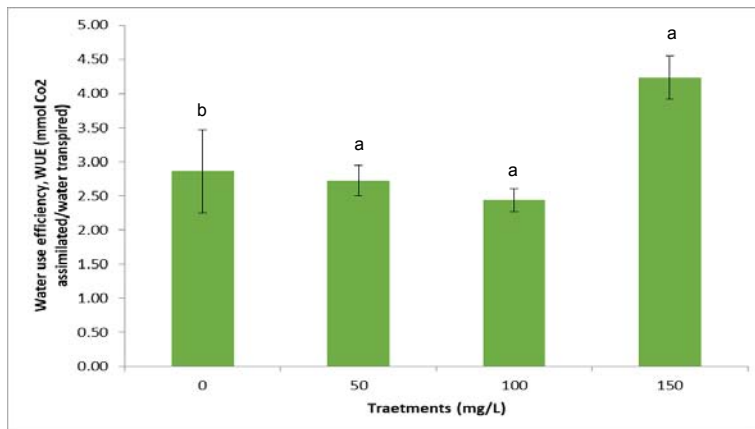
450

451 **Fig. 9. The impact of magnetic nanoparticles on stomatal conductance of *Ipomoea***
 452 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with different**
 453 **superscript showed the significantly different at $P \leq 0.05$ using DMRT at 95%**
 454 **confidence levels.**

455 3.10 Water Use Efficiency

456 From the Fig. 10, the result showed that the water use efficiency of water spinach was
 457 significantly affected by the application of Fe_3O_4 as $P \leq 0.05$ by which the significantly highest
 458 water use efficiency was recorded for 150 mg/L treatment, followed by other treatments with
 459 0 mg/L, 50 mg/L and 100 mg/L at 4.23, 2.86, 2.72, and 2.44 $\mu\text{mol}/\text{CO}_2/\text{mmol}/\text{H}_2\text{O}$
 460 respectively. From the result, we could observe that the water spinach with the highest
 461 concentration of Fe_3O_4 showed greater water use efficiency. Water use efficiency is one of
 462 the important determinants of plant which is under stress as the plant maximize the
 463 capturing of soil moisture when there is limitation of water supply or lower stomata
 464 conductance in order to increase yield production [36]. Water use efficiency is related with
 465 transpiration rate and defined as the ratio of moles CO_2 assimilated per moles of water
 466 transpired [14]. Therefore, with the application of magnetic nanoparticles at higher
 467 concentration increase the transpiration rate of *Ipomoea aquatica*, the water use efficiency
 468 is also increased.

Formatted: Highlight

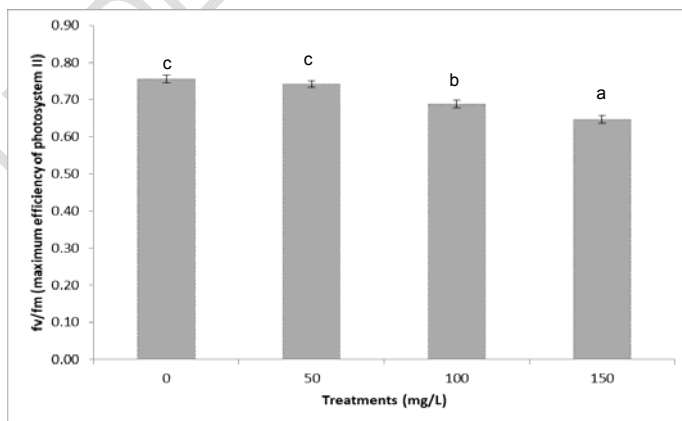


469

470 **Fig. 10. The impact of magnetic nanoparticles on water use efficiency of *Ipomoea***
 471 ***aquatica*. Data are mean ± standard error of mean (SEM). N= 5. Mean with different**
 472 **superscript showed the significantly different at P≤0.05 using DMRT at 95%**
 473 **confidence levels.**

474 **3.11 Fv/Fm (Maximum Efficiency of Photosystem II)**

475 Figure 11 shows the effect of different concentration of Fe₃O₄ on maximum efficiency of
 476 photosystem II or known as fv/fm ratio. The result of the application on water spinach was
 477 significant as P≤0.05 by which higher concentration of Fe₃O₄ showed lower Fv/Fm value.
 478 The ratio of fv/fm for 150 mg/L (0.65) was significantly lower than 100 mg/L (0.69) and
 479 followed by 50 mg/L (0.74) and 0 mg/L (0.76). The lowest Fv/Fm ratio shown by water
 480 spinach treated with 150 mg/l clearly shows that higher concentration of Fe₃O₄ would reduce
 481 the efficiency of photosystem II. Fv/fm is the main parameter used to detect any injury in
 482 photosystem II or photon inhibitory process as it is related to quantum yield of
 483 photosynthesis. The induction of stress on photosynthetic surface in plant would limit the
 484 transformation of light energy in photosystem II. Thus, the decrease in photosynthetic in
 485 plant would reduce the reduction of phytochemical activity in photosystem II [37].

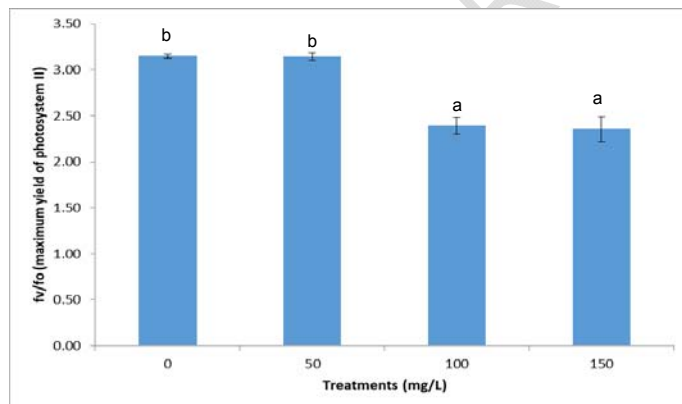


486

487 Fig. 11. The impact of magnetic nanoparticles on f_v/f_m (maximum efficiency of
488 photosystem II) of *Ipomoea aquatica*. Data are mean \pm standard error of mean (SEM).
489 N= 5. Mean with different superscript showed the significantly different at $P \leq 0.05$
490 using DMRT at 95% confidence levels.

491 3.12 Fv/Fo (Maximum Yield of Photosystem II)

492 Fig. 12 shows the effect of different concentration of Fe_3O_4 on maximum yield of
493 photosystem II or known as F_v/F_o ratio in 3 months planting periods. The result of the
494 application on water spinach was significant as $P \leq 0.05$ by which higher concentration of iron
495 oxide nanoparticles showed lower f_v/f_m value. The ratio of f_v/f_m for 0 mg/L (3.15) and 50
496 mg/L (3.14) was significantly higher than 100 mg/L (2.40) and 150 mg/L (2.35). As the two
497 parameters f_v/f_m ratio and F_v/F_o ratio showed the similar trend, we could say that both
498 parameters are related to each other. F_v/F_o is more sensitive towards changes to efficiency
499 of photosystem II compared to F_v/F_m as it shows quantum yield of phytochemical and non-
500 photochemical process. This ratio indicates the state of photosystem II on the energy
501 absorbed and damaged occurred due to plant stress in leaf. This ratio is also affected by
502 the alteration of stomatal closure and carbon fixation process due to water and temperature
503 stress [38]. Besides, the accumulation of magnetic nanoparticles in the photosystem I and II
504 cause obstruction to photosynthesis in donor part of both photosystems that reduce the
505 F_v/F_o ratio [37].
506



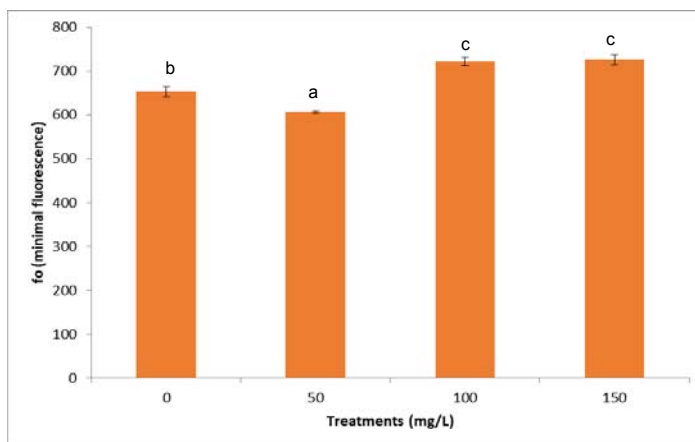
507

508 Fig. 12. The impact of magnetic nanoparticles on f_v/f_o (maximum yield of
509 photosystem II) of *Ipomoea aquatica*. Data are mean \pm standard error of mean (SEM).
510 N= 5. Mean with different superscript showed the significantly different at $P \leq 0.05$
511 using DMRT at 95% confidence levels.

512 3.13 Fo (Minimal Fluorescence)

513 Based on Fig. 13, the application of Fe_3O_4 had significantly influenced the minimal
514 chlorophyll fluorescence yield of water spinach as $P \leq 0.05$. F_o of water spinach treated with
515 100 mg/L and 150 mg/L Fe_3O_4 were significantly higher compared to 0 mg/L and 50 mg/L,
516 722, 726, 653 and 607 respectively. From the result, it is shown that the F_o value increase
517 as the concentration of Fe_3O_4 increase. The application of nanoparticles like titanium oxide
518 showed that with the increase of its concentration, the minimal fluorescence value
519 decreased as titanium oxide nanoparticles do protect the photosynthetic structure of the

520 tomato plant under mild heat stress. However, the study by Gao et al. [39] showed that at
521 high light intensity the minimal fluorescence in *Ulmus elongata* seedlings decreased.
522 Therefore, the best explanation for this would be the impact of magnetic nanoparticles on
523 water spinach is complex and it depends both on the concentration of Fe_3O_4 and the
524 environmental condition that would affect the activity of nanoparticles [40]. Besides, the
525 application of magnetic nanoparticles increased the heat dissipation in water spinach as the
526 absorbed light energy could not be used for photosynthesis [37].
527

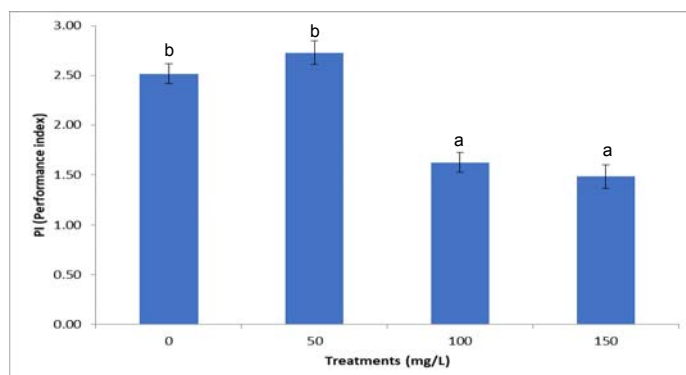


528

529 **Fig. 13. The impact of magnetic nanoparticles on fo (minimal fluorescence) of**
530 ***Ipomoea aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with**
531 **different superscript showed the significantly different at $P \leq 0.05$ using DMRT at 95%**
532 **confidence levels.**

533 3.14 PI (Performance Index)

534 From the Fig. 14, the result showed that the performance index of water spinach was
535 significantly affected by the application of Fe_3O_4 as $P \leq 0.05$ by which the significantly highest
536 performance index was recorded for 50 mg/L treatment, followed by other treatments with 0
537 mg/L, 100 mg/L and 150 mg/L at 2.73, 2.51, 1.62, and 1.48 respectively. From the result, we
538 could observe that the water spinach with lower concentration of iron oxide nanoparticles
539 showed greater performance index. Performance index is the one of the parameters which is
540 sensitive to environmental conditions in a plant as it is being used as a suitable tool to reflect
541 water deficit in a plant system. Based on research, performance index had been used as a
542 sensitive indicator of water stress in *Triticum aestivum*. Performance index is a combination
543 formula that takes into account the measurement of RC/ABS, maximal energy reflux that
544 reaches photosystem II and electron transport [41]. Therefore, the application of magnetic
545 nanoparticles that changes the net photosynthesis rate and transpiration rate due to the
546 accumulation of iron oxide nanoparticles at the root surface and the blockage of Electron
547 Transport Chain had affected the performance index of water spinach. Besides, the carbon
548 assimilation process and stomata conductance had correlated with the performance index of
549 water spinach which concluded that higher concentration of Fe_3O_4 showed lower
550 performance index in water spinach [42].



551

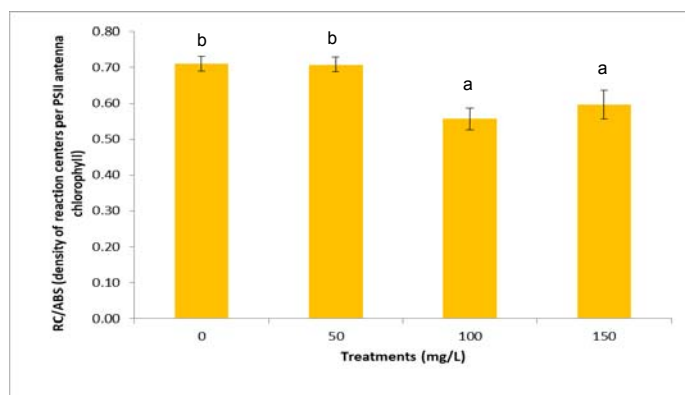
552 **Fig. 14. The impact of magnetic nanoparticles on PI (performance index) of *Ipomoea***
 553 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with different**
 554 **superscript showed the significantly different at $P \leq 0.05$ using DMRT at 95%**
 555 **confidence levels.**

556 3.15 RC/ABS (Density of Reaction Centers Per PSII Antenna Chlorophyll)

557 Fig. 15 shows the effect of different concentration of Fe_3O_4 on density of reaction centers per
 558 Photosystem II antenna chlorophyll in 3 month planting periods. The result of the application
 559 on water spinach was significant as $P \leq 0.05$ by which higher concentration of Fe_3O_4 showed
 560 lower density of reaction centers per Photosystem II antenna chlorophyll. The density of
 561 reaction centers per Photosystem II antenna chlorophyll for 0 mg/L and 50 mg/L is 0.71
 562 which was significantly higher than 100 mg/L (0.56) and 150 mg/L (0.60). RC indicates the
 563 number of active reaction center in photosystem II while ABS shows the quantity of light
 564 absorbed by the antenna chlorophyll. Thus, RC/ABS indicates the total number of active
 565 radiation per light absorption [43]. Due to the exposure of water spinach to Fe_3O_4 , free
 566 radicals production would induce stress to the plant and cause injury to the photosystem II
 567 that limits the active reaction center and light absorption in photosystem II. This parameter
 568 gives the same trend as the performance index as both shows the efficiency of
 569 photosynthesis process in the plant. Therefore, the application of Fe_3O_4 at higher
 570 concentration leads to plant stress and decrease the density of active reaction centers per
 571 photosystem II antenna chlorophyll.

572
 573

Comment [T118]: Put in the methodology how the procedures were performed to obtain this parameter.

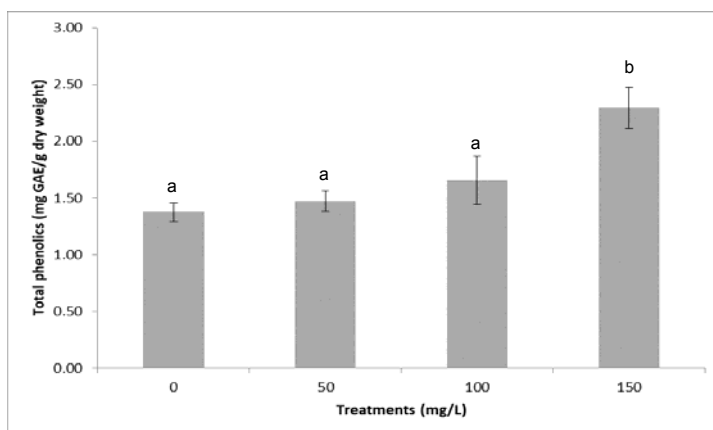


574

575 **Fig. 15. The impact of magnetic nanoparticles on RC/ABS (density of reaction centers**
 576 **per PSII antenna chlorophyll) of *Ipomoea aquatica*. Data are mean \pm standard error of**
 577 **mean (SEM). N= 5. Mean with different superscript showed the significantly different**
 578 **at $P \leq 0.05$ using DMRT at 95% confidence levels.**

579 3.16 Total Phenolics Content

580 Based on Fig. 16, different concentration of Fe_3O_4 had influenced significantly on the
 581 phenolics content of water spinach ($P \leq 0.05$). As the concentration of magnetic nanoparticles
 582 increases, the total phenolics content in the plant increases too. The lowest total phenolics
 583 content was observed in 0 mg/L iron oxide nanoparticles at 1.38 mg GAE/g dry weight and
 584 the highest for 150 mg/L treatment at 2.29 mg GAE/g dry weight. The greater production of
 585 phenolics at higher concentration of magnetic nanoparticles may a sign of the plant under
 586 stress. The production of secondary metabolites is an effort of the plant to defend itself from
 587 the further damage of plant cell and ensure the survival of plant. The higher production of
 588 phenolic stimulates antioxidant activity in the plant [44]. In order to the response of a plant
 589 towards environmental stresses and protecting itself from damages in plant cell, the
 590 production of phenolic compounds is essential to maintain the plant growth and reproduction
 591 [45]. Based on the study by researchers, phenolics compound do contain antibiotic and anti-
 592 nutritional properties that help in the defense system of a plant. Phenolics compound usually
 593 stored in the epidermal cells of leaves and shoots besides central vacuole of guard cells.
 594 Phenolics content can be divided into two groups which are preformed phenolics and
 595 induced phenolics by which preformed phenolics are being synthesized by the plant during
 596 development of plant tissues under normal condition. While, induced phenolics are produced
 597 when the plant faces physical injury, infection or environmental stresses due to heavy metal
 598 irradiation or temperature [14].

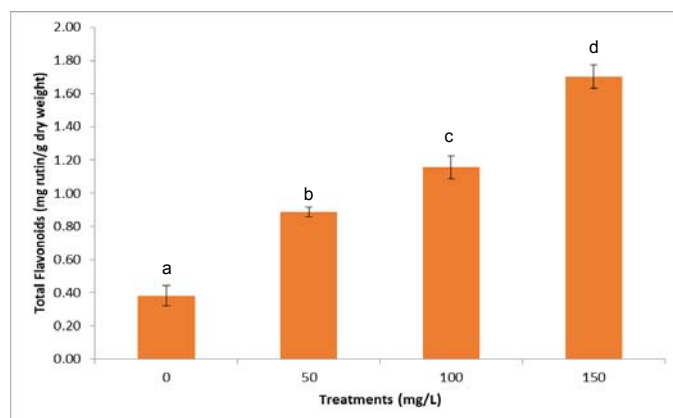


599

600 **Fig. 16. The impact of magnetic nanoparticles on total phenolics content of *Ipomoea***
 601 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with different**
 602 **superscript showed the significantly different at $P \leq 0.05$ using DMRT at 95%**
 603 **confidence levels.**

604 3.17 Total Flavonoids Content

605 Based on Fig. 17, the production of total flavonoid content had significantly influenced by the
 606 application of magnetic nanoparticles at different concentrations of Fe_3O_4 as ($P \leq 0.05$). At the
 607 end of experiment of 12th week, the total flavonoid content in water spinach treated with 150
 608 mg/L Fe_3O_4 showed the highest reading at 1.70 mg rutin/g dry weight followed by 100 mg/l
 609 treatment at 1.16 mg rutin/g dry weight, 50 mg/L treatment at 0.89 mg rutin/g dry weight and
 610 lastly 0 mg/L treatment at 0.38 mg rutin/g dry weight. From the result, we could observe that
 611 higher concentration of Fe_3O_4 increase the production of flavonoid in water spinach. The
 612 production of flavonoid is one of the determinants for the production of secondary metabolite
 613 in plant. It is often to observe that plant undergoing stress has higher total flavonoid content
 614 due to defensive mechanism in plant. Biosynthesis in plant to produce secondary
 615 metabolites is usually stimulated by the accumulation of heavy metals like zinc, iron, and
 616 nickel which generates Reactive Oxygen Species (ROS) and induces oxidative stress in
 617 plant. Consequently, the induction of oxidative stress causes changes in signal transduction
 618 for the mechanism of gene coding and enzyme [13]. Besides, the production of ROS may
 619 cause damages to cell membrane, cell structure and photosynthetic site and thus the
 620 production of flavonoid at the generation site act as defensive mechanism due to its high
 621 antioxidant properties [14].



622

623 **Fig. 17. The impact of magnetic nanoparticles on total flavonoids content of *Ipomoea***
 624 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 5. Mean with different**
 625 **superscript showed the significantly different at $P \leq 0.05$ using DMRT at 95%**
 626 **confidence levels.**

627 4. CONCLUSION

628

629 Overall, it was found that the application of magnetic nanoparticles which was in the form of
 630 iron oxide nanoparticles had influenced the growth of *Ipomoea aquatica*. The plant height,
 631 shoot elongation, plant temperature, total biomass and total chlorophyll content were
 632 significantly affected by the application of magnetic nanoparticles at higher concentration.
 633 Besides that, the leaf gas exchange characteristics were also influenced by the different
 634 concentrations of iron oxide nanoparticles as $P \leq 0.05$. The net photosynthesis rate,
 635 transpiration rate, stomata conductance, maximum efficiency of photosystem II (F_v/F_m),
 636 maximum quantum yield of phytochemical and non-photochemical process in photosystem II
 637 (F_v/F_o), performance index and the density of reaction centers per PSII antenna chlorophyll
 638 of *I. aquatica* were significantly reduced at higher concentration of magnetic nanoparticles.
 639 While, water use efficiency and minimal fluorescence value of *I. aquatica* were increased
 640 with the increased of iron oxide nanoparticles concentration. In addition, the application of
 641 magnetic nanoparticles had significantly influenced the total flavonoids and total phenolics
 642 content in water spinach.

643

644

645 REFERENCES

646

- 1 Salaries, T. (2016). Department of Statistics Malaysia Press Release, (May), 1–4.
- 2 Department of Statistics Malaysia Press Release. (2017). Press Release Supply and Utilization Accounts Selected Agricultural Commodities, Malaysia 2012-2016, (December), 1–4.
- 3 Department of Statistics Malaysia Press Release. (2018), (February), 4–6. Retrieved from <https://www.dosm.gov.my/v1/index.php?r=column/pdfPrev&id=TkpmM05EK3NBV0JRU1pmOUJnS3RCQT09> on September 15, 2018.
- 4 Ibrahim, M. H., Yasmin, N., Rahman, A., Amalina, N., & Zain, M. (2018). Effect of Nitrogen Rates on Growth and Quality of Water Spinach (*Ipomea aquatica*), *Annual Research & Review in Biology*, 26(1), 1–12. <https://doi.org/10.9734/ARRB/2018/40352>
- 5 Dua, T. K., Dewanjee, S., Gangopadhyay, M., Khanra, R., Zia-Ul-Haq, M., & De Feo, V. (2015). Ameliorative effect of water spinach, *Ipomea aquatica* (Convolvulaceae),

- against experimentally induced arsenic toxicity. *Journal of Translational Medicine*, 13(1), 1–17. <https://doi.org/10.1186/s12967-015-0430-3>
- 6 Xiao, Q., Wong, M. H., Huang, L., & Ye, Z. (2015). Effects of cultivars and water management on cadmium accumulation in water spinach (*Ipomoea aquatica* Forsk.). *Plant and Soil*, 391(1–2), 33–49. <https://doi.org/10.1007/s11104-015-24095>
 - 7 Markides, H., Rotherham, M., & El Haj, A. J. (2012). Biocompatibility and toxicity of magnetic nanoparticles in regenerative medicine. *Journal of Nanomaterials*, 13–15. <https://doi.org/10.1155/2012/614094>
 - 8 Lu, A. H., Salabas, E. L., & Schüth, F. (2007). Magnetic nanoparticles: Synthesis, protection, functionalization, and application. *Angewandte Chemie - International Edition*, 46(8), 1222–1244. <https://doi.org/10.1002/anie.200602866>
 - 9 Nair, R., Varghese, S. H., Nair, B. G., Maekawa, T., Yoshida, Y., & Kumar, D. S. (2010). Nanoparticulate material delivery to plants. *Plant Science*, 179(3), 154–163. <https://doi.org/10.1016/j.plantsci.2010.04.012>
 - 10 Da Silva, L.C., Oliva, M.A., Azevedo, A.A., De Araujo, J.M. (2006). Responses of resting a plant species to pollution from an iron pelletization factory, *Water Air Soil Pollut.* 175, 241–256
 - 11 Lin, C., Fugetsu, B., Su, Y., & Watari, F. (2009). Studies on toxicity of multi walled carbon nanotubes on Arabidopsis T87 suspension cells. *J. Hazard. Mat.* 170, 578–583.
 - 12 Tiwari, D.K., Dasgupta-Schubert, N., Villasenor, L.M., Tripathi, D., & Villegas, J. (2013). Interaction of carbon nanotubes with mineral nutrients for the promotion of growth of tomato seedlings. *Nano Stud.* 7, 87–96.
 - 13 Zhao, J., Davis, L. C., & Verpoorte, R. (2005). Elicitor signal transduction leading to production of plant secondary metabolites, *Biotechnology Advances*, 23, 283–333. <https://doi.org/10.1016/j.biotechadv.2005.01.003>
 - 14 Izad, A. I., Ibrahim, M. H., Azurahaman, C., Abdullah, C., Amalina, N., & Zain, M. (2018). Growth , Leaf Gas Exchange and Secondary Metabolites of *Orthosiphon stamineus* as Affected by Multiwall Carbon Nanotubes Application, *Annual Research & Review in Biology*, 23, 1–13. <https://doi.org/10.9734/ARRB/2018/38113>
 - 15 Karlsson, H.L., Cronholm, P., Gustafsson, J., & Möller, L. (2008). Copper oxide nanoparticles are highly toxic: a comparison between metal oxide nanoparticles and carbon nanotubes. *Chem Res Toxicol.* 21(9), 1726–32.
 - 16 Martínez-Fernández, D., Barroso, D., & Komárek, M. (2016). Root water transport of *Helianthus annuus* L. under iron oxide nanoparticle exposure. *Environmental Science and Pollution Research*, 23(2), 1732–1741
 - 17 Liu, R., Zhang, H., & Lal, R. (2016). Effects of stabilized nanoparticles of copper, zinc, manganese, and iron oxides in low concentrations on lettuce (*Lactuca sativa*) seed germination: nanotoxicants or nonnutrients?. *Water, Air, & Soil Pollution*, 227(1), 42.
 - 18 Ruttkay-Nedecky, B., Krystofova, O., Nejdil, L., & Adam, V. (2017). Nanoparticles based on essential metals and their phytotoxicity. *Journal of Nanobiotechnology*, 15(1), 119. <https://doi.org/10.1186/s12951-017-0268-3>
 - 19 Singh, S., Singh, B.K., Yadav, S.M., & Gupta, A.K. (2015). Applications of Nanotechnology in Agricultural and their Role in Disease Management. *Research Journal of Nanoscience and Nanotechnology*, 5(1), 1–5.
 - 20 Mahdavi, M., Ahmad, M. B., Haron, M. J., Namvar, F., Nadi, B., Rahman, M. Z. A., & Amin, J. (2013). Synthesis, surface modification and characterisation of biocompatible magnetic iron oxide nanoparticles for biomedical applications. *Molecules*, 18(7), 7533–7548.
 - 21 Ibrahim, M. H., Jaafar, H. Z., Rahmat, A., & Rahman, Z. A. (2011). The relationship between phenolics and flavonoids production with total non structural carbohydrate and photosynthetic rate in *Labisia pumila* Benth. under high CO₂ and nitrogen fertilization. *Molecules*, 16(1), 162–174.
 - 22 Becker, M., & Asch, F. (2005). Iron toxicity in rice conditions and management concepts, *Journal*

- of *Plant Nutrition Soil Science*, 168, 558–573. <https://doi.org/10.1002/jpln.200520504>
- 23 Rui, M., Ma, C., Hao, Y., Guo, J., Rui, Y., Tang, X., & Zhu, S. (2016). Iron Oxide Nanoparticles as a Potential Iron Fertilizer for Peanut (*Arachis hypogaea*). *Frontiers in Plant Science*, 7(June), 1–10. <https://doi.org/10.3389/fpls.2016.00815>
 - 24 Dhoke, S. K., Mahajan, P., Kamble, R., & Khanna, A. (2013). Effect of nanoparticles suspension on the growth of mung (*Vigna radiata*) seedlings by foliar spray method. *Nanotechnology Development*, 3(1). <https://doi.org/10.4081/nd.2013.el>
 - 25 Chrispeels, M. J., & Agre, P. (1994). Aquaporins : water channel proteins of plant and animal cells, *Elsevier Science*, 8073–8077.
 - 26 Pressure, T., Kirkham, M. B., Gardner, W. R., & Gerloff, G. C. (1972). Regulation of Cell Division and Cell Enlargement, *Plant Physiology*, 49, 961–962.
 - 27 Sheykhbaglou, R., & Sedghi, M. (2010). Effects of Nano-Iron Oxide Particles on Agronomic Traits of Soybean, *Not Sci Biol*, 2(2), 112–113.
 - 28 Jeyasubramanian, K., Gopalakrishnan Thoppey, U. U., Hikku, G. S., Selvakumar, N. Subramania, A., & Krishnamoorthy, K. (2016). Enhancement in growth rate and productivity of spinach grown in hydroponics with iron oxide nanoparticles. *RSC Advances*, 6(19), 15451–15459. <https://doi.org/10.1039/C5RA23425E>
 - 29 Du, W., Tan, W., Peralta-videa, J. R., & Gardea-torresdey, J. L. (2016). University of California Center for Environmental Implications of Nanotechnology (UC. *Plant Physiology and Biochemistry*. <https://doi.org/10.1016/j.plaphy.2016.04.024>
 - 30 Gates, D.M. (1968). Transpiration and leaf temperature. *Plant Physiology*, 19, 211-238.
 - 31 Racuciu M, Creanga D, Olteanu Z (2009). Water based magnetic fluid impact on young plants growing. *Rom. Rep. Phys.*, 61 (2): 259-268.
 - 32 Britt, D. W., Johnson, W. P., Boyanov, M. I., & Anderson, A. J. (2012). CuO and ZnO nanoparticles : phytotoxicity , metal speciation , and induction of oxidative stress in sand-grown wheat, *Journal of Nanoparticle Research*, 14, 1125. <https://doi.org/10.1007/s11051-012-1125-9>
 - 33 Li, J., Chang, P. R., Huang, J., Wang, Y., Yuan, H., & Ren, H. (2013). Physiological Effects of Magnetic Iron Oxide Nanoparticles Towards Watermelon, *Journal of Nanoscience and Nanotechnology*, 13(8), 5561–5567 <https://doi.org/10.1166/jnn.2013.7533>
 - 34 Shweta, Tripathi, D.K., Shweta, S., Swati, S., Dubey, N.K., & Chauhan, D.K. (2016). Impact of Nanoparticles on Photosynthesis: Challenges and Opportunities, *American Scientific Publishers*, 5(5), 404-411. <https://doi.org/10.1166/mat.2016.1327>
 - 35 Rizwan, M., Ali, S., Farooq, M., Sik, Y., Adrees, M., Ibrahim, M., & Abbas, F. (2017). Effect of metal and metal oxide nanoparticles on growth and physiology of globally important food crops: A critical review. *Journal of Hazardous Materials*, 322, 2 16. <https://doi.org/10.1016/j.jhazmat.2016.05.061>
 - 36 Blum, A. (2009). Field Crops Research Effective use of water (EUW) and not water-use efficiency (WUE) is the target of crop yield improvement under drought stress, *Field Crop Research*, 112, 119–123. <https://doi.org/10.1016/j.fcr.2009.03.009>
 - 37 Nurfarahin S. (2017). Growth, Leaf Gas Exchange and Secondary Metabolites of Water Spinach (*Ipomoea aquatica*) as Affected by Carbon Nanotubes Application, Universiti Putra Malaysia, Malaysia.
 - 38 Maxwell, K., & Johnson, G. N. (2000). Chlorophyll fluorescence -a practical guide, *Journal of Experimental Botany*, 51(345), 659–668.
 - 39 Gao, J., Xu, G., Qian, H., Liu, P., Zhao, P., & Hu, Y. (2013). Effects of nano-TiO₂ on photosynthetic characteristics of *Ulmus elongata* seedlings. *Environmental pollution*, 176, 63-70.
 - 40 Qi, M., Liu, Y., & Li, T. (2013). Nano-TiO₂ Improve the Photosynthesis of Tomato Leaves under Mild Heat Stress, *Biology Trace Element Research*, 156, 323–328. <https://doi.org/10.1007/s12011-013-9833-2>
 - 41 Mehta, P., Jajoo, A., Mathur, S., & Bharti, S. (2010). Plant Physiology and Biochemistry Chlorophyll a fluorescence study revealing effects of high salt stress on Photosystem II in

- wheat leaves. *Plant Physiology et Biochemistry*, 48(1), 16–20.
<https://doi.org/10.1016/j.plaphy.2009.10.006>
- 42 Živčák, M., Brestič, M., Olšovská, K., & Slamka, P. (2008). Performance index as a sensitive indicator of water stress in *Triticum aestivum* L., *Plant Soil Environment*, 54(4), 133–139.
 - 43 Appenroth, K., & Sto, J. (2001). Multiple effects of chromate on the photosynthetic apparatus of *Spirodela polyrhiza* as probed by OJIP chlorophyll a fluorescence measurements, *Environmental pollution*, 115, 49–64
 - 44 Kim, D., Weon, S., & Lee, C. Y. (2003). Antioxidant capacity of phenolic phytochemicals from various cultivars of plums, *Food Chemistry*, 81, 321–326.
 - 45 Subbiah R., Veerapandian M., & Yun K.S. (2010). Nanoparticles: Functionalization and multifunctional applications in biomedical sciences. *Curr. Med. Chem*, 17, 4559–4577.

647
648
649
650
651
652
653

654

UNDER PEER REVIEW