1	Original Research Article
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2	EFFECT OF DIFFERENT COMBINATIONS OF NPK AND BIOFERTILIZERS
3	IN <mark>ON</mark> ZINNIA (Zinnia elegans J.)
4	Abstract
5	The experiment was carried out to study the effect of different combinations of NPK
6	and biofertilizers in on zinnia (Zinnia elegans J.). The results reviled that treatment
7	combination of (NPK 21:12:7.5 g/m^2 + Azotobacter + PSB + KSB) recorded maximum
8	plant height (125.32 cm), number of primary branches (9.73), plant spread (66.55 cm ²),
9	minimum days taken to anthesis (48.88 days), maximum flowering duration (42.42
10	days), seed yield per plant (21.19 g) respectively compared to control.
11	Keywords: Anthesis, Azotobacter, Biofertilizers, Flowering, Zinnia
12	Introduction
13	Zinnia is a genus of plants of the sunflower tribe (Asteraceae) within the daisy family
14	(Linnaeus, 1759). They are native to scrub and dry grassland in an area stretching from
15	the South-western to South America, with a centre of diversity in Mexico. Members of
16	the genus are notable for their solitary long-stemmed flowers that come in a variety of
17	bright colours. The genus name honours German master botanist Johann Gottfried Zinn.
18	Zinnia elegans, known as youth-and-age, zinnia is a popular garden flower, usually
19	grown from seed and preferably in fertile, humus-rich and well-drained soil, in an area
20	with full sun. Zinnias flower are champion of season among summer annual flowers.
21	Zinnia is originated from Mexico; the Spanish referred it as "mal de ojos" (meaning
22	sickness of the eyes). Modern Zinnia has been developed from species Zinnia elegans
23	Jacq. Zinnia range in height from 15-100 cm. zinnia leaves are sandpapery in texture,
24	contrary, generally stalk less (sessile), pale to middle green in colour and having
25	different forms (linear and ovate). Zinnias may be used as cut flowers, in beds, container,
26	border and background or as cottage; garden plants attracts birds, butterflies and other
27	humming birds <u>hummingbirds</u> .
28	Zinnia requires appropriate nutrition for its proper growth and development to be
29	sufficiently green, vigorous and produce abundant flowers of adequate size and color
30	intensity with good lasting qualities (Joiner and Gruis, 1961). Though the chemical

fertilizers are an important source of nutrients, they are not only costly but growing 31 32 concerns of environmental pollution and limitation of non-renewable resources may introduce additional constraints. The use of chemical fertilizer also poses a major threat 33 to sustain soil health and crop productivity. At present we are not in a position to 34 abandon the use of chemical fertilizers completely, so the best option available is to use 35 36 these fertilizers in lesser amount along with other nutrients sources. To minimise the use 37 of these inputs without effecting the overall production and the ecosystem, it is necessary to use eco-friendly, economical and easily available biofertilizers for the development of 38 more efficient fertility management programme. These are cost effective and renewable 39 source of plant nutrients to supplement the chemical fertilizers for sustainable 40 floriculture. Incorporation of biofertilizers in combination with chemical fertilizers can 41 42 completely prevent the detrimental effect of current practice (Maurya and Beniwal, 2003). 43

44 Materials and methods

The present investigation was carried out at floriculture research farm, division of 45 Floriculture and Landscape Architecture. Sher-e-Kashmir University of Agriculture Science 46 47 and Technology, Shalimar Srinagar, during year 2017-2018 / 2018-2019. The experimental farm is located between 34^o 05' N latitude and 74^o 98' E longitude at an altitude of 1587 48 meters above mean sea level. The climate is temperate-cum-mediterranean and continental 49 type characterized by hot summer and severe winters. The average annual precipitation is 50 944.6 mm, and more than 80% precipitation received from western disturbances. Three levels 51 of chemical fertilizers (NPK @ 28:16:10, 21:12:7.5, and 14:8:5 g/m²) along with different 52 53 combinations of biofertilizers (Azotobacter, PSB, KSB). Seedlings were treated by dipping root portion of seedlings in solution prepared by mixing biofertilizers in 1000ml water for 30 54 minutes before transplanting. Treated seedlings were plant by maintaining spacing of 30×40 55 cm thus accommodating nine plants. Five plants are randomly selected from each unit plot 56 57 for collecting data and the mean value of all the parameters were analysed by Duncan's multiple range test (DMRT) AT 5% level of probability. He experiment comprises of 18 58 59 different treatment combinations laid out in Randomized Block Design (RBD) replicated 60 thrice.

61 **Results** and discussion

62 Plant height at harvest (cm)

The result of analysis for plant height is presented in table 1. Among different treatment 63 combination (NPK 21:12:7.5 g/m² + Azotobacter + PSB + KSB) recorded maximum plant 64 height (125.32 cm) which was statistically superior to other combination of NPK and 65 biofertilizers. The possible reason for increase in plant height is that combined application of 66 biofertilizers with (NPK 21:12:7.5 g/m^2) resulted in better nutrition which leads to increased 67 photosynthetic activity, enhanced cell division and enlargement as nitrogen is important 68 69 constituent of nucleic acid and it might have increased the synthesis of carbohydrates, amino acids etc. From which phytohormones like auxins, gibberellins and cytokines have been 70 71 synthesized and phosphorous being an essential component of protoplasm and chlorophyll, 72 cause conversion of photosynthates into phospholipids resulting in adequate vegetative 73 growth thus increased plant height at harvest. Biofertilizers produce several growth 74 promoting hormones (auxins, cytokinins and gibberellins etc.) in addition to increasing the 75 availability of nitrogen, phosphorous and potash to the plants resulting in better plant growth. 76 Similar results of increase in plant height at harvest due to combined application of 77 biofertilizers with reduced dose of NPK have been reported by Chaitra and Patil (2007), Patil 78 and Agasimani (2013) and Kiran et al., (2014) in China Aster; Verma et al., (2011) in 79 chrysanthemum and Airadevi (2012) in annual chrysanthemum.

80 NUMBER OF PRIMARY BRANCHES PER PLANT

The perusal of pooled data presented in Table 1 clearly shows difference in number of 81 primary branches per plant due to different combination of NPK and biofertilizers. Among 82 different treatments T_{12} (NPK 21:12:7.5 g/m² + Azotobacter + PSB + KSB) recorded 83 maximum number of primary branches per plant (9.73). He increase in number of primary 84 85 branches per plant with treat might be due to formation of nitrogenous compounds such as proteins, amino acids, nucleic acids, various enzymes and coenzymes which were responsible 86 87 for cell division and cell enlargement and the role of phosphorous in structural component as in phospholipid and in translocation of food material this results might be due to role of 88 89 Azotobacter in nitrogen fixation and production of growth promoting substances such as IAA 90 and gibberellins which lead to more no of primary branches per plant. Similar result with increase in number of primary branches with inoculation of... has been reported by Gupta et 91 92 al (1999), Panchal et al (2010)

93 PLANT SPREAD (cm²)

perusal of pooled data presented in Table 2 clearly shows that (NPK 21:12:7.5 g/m^2 + 94 Azotobacter + PSB + KSB) recorded maximum plant spread (66.55 cm²) Result clearly 95 showed that the combined application of Azotobacter PSB and KSB along with... NPK 96 97 proved to be beneficial for robust growth of plant as compared to other treatments this may be attributed to the possible role of nitrogen in improving structural parameters as it is an 98 99 important constituent of protein and the role of phosphorous in structural component as in 100 phospholipid and in absorbing and in translocation of food material. Moreover, biofertilizers 101 viz. Azotobacter, PSB and KSB proved to be beneficial as they fix atmospheric nitrogen in 102 soil and also secrete growth promoting substances like auxins which stimulate the plant 103 metabolic activity and photosynthetic efficiency leading to better growth of plant, above 104 result are in conformity with the findings of Panchal et al (2010).

105 DAYS TAKEN TO ANTHESIS

Perusal of pooled data presented in Table 2 clearly shows that (NPK 21:12:7.5 $g/m^2 +$ 106 Azotobacter + PSB + KSB) recorded minimum days taken to anthesis (48.88 days). The 107 reason for earliness of flowering can be proper uptake of nutrient and production of growth 108 promoting substances like auxins, gibberellins, vitamins and organic acids by the 109 110 biofertilizers further phosphorous is an important element and essential for initiation of 111 flowering and PSB is known to increase the availability of phosphorous resulting in early flowering similar result for early flowering by combined application of NPK along with 112 biofertilizers are reported by Kiran et al 2014, Chitra and Patil (2007) 113

114 FLOWERINF DURATION (days)

The perusal of pooled data presented in table 3 reviled that (NPK 21:12:7.5 g/m² + Azotobacter + PSB + KSB) recorded maximum flowering duration (42.42 days). It is because nitrogen, phosphorous and potassium leads to increase in growth parameters and translocation and accumulation of photosynthates might be the reason for increase in flowering duration result got support from Airadevi (2012)

120 SEED YIELD PER PLANT (g)

121 (NPK 21:12:7.5 g/m² + Azotobacter + PSB + KSB) recorded maximum seeds yield 122 per plant (21.19 g), as shown in table 3, Increase in Phosphorous availability due to 123 application of PSB result in significant increase in seed yield per plant this might be due to 124 more dry matter production by the plant which exhibit superior vegetative growth, results are in concordance with the findings of Sehrawat *et al* (2003). Whereas potassium has many different roles in plant: in photosynthesis, potassium regulates the opening and closing of stomata, and therefore regulate CO2 uptake. Potassium trigger activation of enzymes and essential for production of Adenosine Triphosphate (ATP).

129 CONCLUSION

130 Integrated nutrient management enhance the availability of applied as well as native soil 131 nutrients. Application of NPK @ 21:12:7.5 g/m² along with biofertilizers (Azotobacter + PSB 132 + KSB) significantly improves vegetative, flowering and seed parameters compared to 133 control.

149 Table 1. Effect of different combinations of NPK and biofertilizers on plant height

150 (cm) and number of primary branches per plant in zinnia (Zinnia elegans j.)

Details of treatment		nt height (o	em)	Number of primary		
				brar	iches per	plant
	2017	2018	Pooled	2017	2018	Pooled
NPK 28:16:10 g/m ²	118.14	118.64	118.39	6.83	6.83	6.83
NPK 28:16:10 g/m ² + Azotobacter	119.53	121.02	120.28	7.76	7.90	7.83
NPK 28:16:10 g/m ² + PSB	118.53	119.26	118.90	7.33	7.40	7.36
NPK 28:16:10 g/m ² + KSB	117.60	118.66	118.13	7.33	7.46	7.40
NPK 28:16:10 g/m ² + Azotobacter + PSB	121.08	122.57	121.82	7.86	8.00	7.93
NPK 28:16:10 g/m ² + Azotobacter + PSB + KSB	122.87	124.05	123.46	9.46	9.50	9.48
NPK 21:12:7.5 g/m ²	117.23	118.09	117.66	6.93	7.03	6.98
NPK 21:12:7.5 g/m^2 + Azotobacter	121.03	122.97	122.00	8.96	9.16	9.06
NPK 21:12:7.5 $g/m^2 + PSB$	120.36	122.13	121.25	8.90	9.03	8.96
NPK 21:12:7.5 g/m ² + KSB	120.16	121.04	120.60	7.66	7.83	7.75
NPK 21:12:7.5 g/m ² + Azotobacter + PSB	122.95	124.49	123.72	9.00	9.13	9.06
NPK 21:12:7.5 g/m^2 + Azotobacter + PSB + KSB	124.43	126.21	125.32	9.66	9.80	9.73
NPK 14:8:5 g/m ²	116.46	117.32	116.89	5.66	5.70	5.68
NPK 14:8:5 g/m ² + Azotobacter	118.13	120.21	119.17	7.03	7.20	7.11
NPK 14:8:5 $g/m^2 + PSB$	117.66	118.23	117.95	7.00	7.20	7.10
NPK 14:8:5 $g/m^2 + KSB$	116.86	117.53	117.20	6.33	6.46	6.40
NPK 14:8:5 g/m^2 + Azotobacter + PSB	119.88	121.08	120.48	8.00	8.13	8.06
NPK 14:8:5 g/m^2 + Azotobacter + PSB + KSB	120.96	122.90	121.93	8.43	8.63	8.53
C.D _(p≤0.05)	1.662	2.21	1.78	1.15	1.10	1.11

156 Table 2. Effect of different combinations of NPK and biofertilizers on plant spread

157 (cm ²) and days taken to anthesis in zinnia (<i>Zinn</i>	<i>ia elegans</i> i.)

Details of treatment	Plant spread (cm ²)		² m ²)	Days taken to a		nthesis
	2017	2018	Pooled	2017	2018	Pooled
NPK 28:16:10 g/m ²	58.88	59.21	59.04	52.00	51.59	51.79
NPK 28:16:10 g/m ² + Azotobacter	59.03	59.37	59.20	51.67	51.30	51.48
NPK 28:16:10 g/m ² + PSB	61.10	61.77	61.43	51.00	50.84	50.92
NPK 28:16:10 g/m ² + KSB	59.43	59.97	59.70	51.33	51.06	51.20
NPK 28:16:10 g/m ² + Azotobacter + PSB	60.13	61.43	60.78	50.67	50.26	50.46
NPK 28:16:10 g/m ² + Azotobacter + PSB + KSB	64.10	64.17	64.13	49.67	49.52	49.59
NPK 21:12:7.5 g/m ²	55.13	55.40	55.27	53.00	52.66	52.83
NPK 21:12:7.5 g/m ² + Azotobacter	61.77	62.13	61.95	51.00	50.73	50.87
NPK 21:12:7.5 g/m ² + PSB	63.30	63.67	63.48	50.33	50.54	50.44
NPK 21:12:7.5 g/m ² + KSB	61.27	61.60	61.43	50.67	50.46	50.56
NPK 21:12:7.5 g/m ² + Azotobacter + PSB	65.30	65.83	65.57	50.00	49.77	49.89
NPK 21:12:7.5 g/m^2 + Azotobacter + PSB + KSB	66.30	66.80	66.55	49.00	48.76	48.88
NPK 14:8:5 g/m ²	52.43	53.63	53.03	54.33	53.98	54.16
NPK 14:8:5 g/m ² + Azotobacter	54.20	54.57	54.38	52.67	53.17	52.92
NPK 14:8:5 g/m ² + PSB	55.00	56.57	55.78	51.00	50.44	50.72
NPK 14:8:5 g/m ² + KSB	53.67	54.23	53.95	52.67	52.40	52.53
NPK 14:8:5 g/m ² + Azotobacter + PSB	57.13	57.47	57.30	51.33	51.00	51.16
NPK 14:8:5 g/m^2 + Azotobacter + PSB + KSB	59.77	60.50	60.13	50.67	50.39	50.53
C.D _(p≤0.05)	5.20	4.86	4.91	1.30	1.40	1.27

165 Table 3. Effect of different combinations of NPK and biofertilizers on flowering

duration (days) and seed yield per plant in zinnia (Zinnia elegans j.)

Details of treatment	Flowerin	g duration	ı (days)	Seed y	ield per pl	ant (g)
-	2017	2018	Pooled	2017	2018	Pooled
NPK 28:16:10 g/m ²	38.33	38.37	38.35	13.89	14.07	13.98
NPK 28:16:10 g/m ² + Azotobacter	37.67	37.80	37.74	15.68	16.42	16.05
NPK 28:16:10 g/m ² + PSB	39.67	39.87	39.77	17.30	18.15	17.72
NPK 28:16:10 g/m ² + KSB	38.33	38.50	38.42	16.93	17.84	17.38
NPK 28:16:10 g/m ² + Azotobacter + PSB	40.67	40.87	40.77	18.12	19.01	18.56
NPK 28:16:10 g/m ² + Azotobacter + PSB + KSB	41.67	41.83	41.75	19.60	20.39	19.99
NPK 21:12:7.5 g/m ²	37.67	37.87	37.77	12.34	12.11	12.22
NPK 21:12:7.5 g/m ² + Azotobacter	39.00	39.23	39.12	17.27	17.90	17.59
NPK 21:12:7.5 g/m ² + PSB	40.33	40.60	40.47	18.78	19.57	19.17
NPK 21:12:7.5 g/m ² + KSB	39.67	39.80	39.73	18.21	19.18	18.69
NPK 21:12:7.5 g/m ² + Azotobacter + PSB	41.67	42.00	41.83	20.12	21.03	20.58
NPK 21:12:7.5 g/m^2 + Azotobacter + PSB + KSB	42.33	42.50	42.42	20.86	21.51	21.19
NPK 14:8:5 g/m ²	35.00	35.00	35.00	10.56	11.04	10.80
NPK 14:8:5 g/m ² + Azotobacter	36.33	36.53	36.43	12.73	13.35	13.04
NPK 14:8:5 g/m ² + PSB	37.67	37.90	37.78	13.87	14.47	14.17
NPK 14:8:5 g/m ² + KSB	37.00	37.20	37.10	13.55	14.29	13.92
NPK 14:8:5 g/m ² + Azotobacter + PSB	38.67	38.90	38.78	14.94	15.87	15.40
NPK 14:8:5 g/m^2 + Azotobacter + PSB + KSB	39.00	39.17	39.08	16.79	17.70	17.24
C.D _(p≤0.05)	1.12	1.15	1.12	1.37	1.38	1.31

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