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Determination of Minerals, Total Phenolic Content, Flavonoids, Antioxidants and Antimicrobial Activities of Ethanolic Extracts of sweet Lupinus Angustifolius of Palestine **Ethanolic Extract in Palestine**

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ABSTRACT

Aims: To establish finthe most suitable dout the best extraction method for of-sweet lupine seeds and to determine minerals, phenolic content, flavonoids, antioxidant activity and antimicrobial activities.

Study design: Known and standard experimental procedures are employed.

Place and Duration of Study: Department of Chemistry, Bethlehem University, from January 2019 to March 2019.

Methodology: Seeds were ground grinded and extracted by Soxhlet extractor using ethanol with different percentages (50%, 60%, 70%, 80% and 95%). Sodium, potassium and ferrous ion_ion content were determined. As for pharmacological properties, Resistance to bacteria was performed against Escherichia coli and Staphylococcus aureus, while aAntioxidant activity was determined by FRAP method. Two types of flavonoids were measured: in this work: f-lavonones and dihydroflavonols via the reaction with 2,4-dinitrophenylhydrazine. Phenolics were determined by the Folin-Ciocalteu method.

Results: 50% ethanol resulted in produced the highest content of extract (18.6%) while 70% and 60% showed the lowest content (10.0% for both). 80% ethanol extracted sample showed the highest content for sodium (56.51 mg Na/g extract), while 60% and 50% ethanol extracts showed the highest content percentage of potassium (2.25 and 2.33 mg K/g extract, respectively). The maximum concentration of ferrous ion was obtained with 70% ethanol (6.854 mg Fe⁺²/g extract). 95% ethanolic extract showed the highest antioxidant activity (20.24 mg FeSO₄/g extract). Similar The same results wereas obtained for total phenolic content and flavonoids: 24.60 mg gGallic acid/g extract for pPhenolics and 116.02 mg rRutin/g extract for flavonoids. Extracts showed no bacterial activity against both types of bacteria used.

Conclusion: 95% ethanol extracted samples showed the highestbest antioxidant activity and the highest flavonoids and phenolic content. Sweet lupine extract did not perform any antimicrobial activity against both Gram positive and Gram negative bacteria.

Keywords: Sweet lupine, Soxhlet extractor, Minerals, Total phenolics, Flavonoids, Antimicrobial Activity, Antioxidant Activity.

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1. INTRODUCTION

Sweet Lupinus <u>a</u>Angustifolius, also called "narrow-leafed lupine" is a member of the legume family (subfamily Papilioniodeae) containing both herbaceous annual and shrubby perennial types with attractive long racemes of flowers [1]. There are twelve lupine species within the <u>Lupinus</u> genus, all of which are native to Europe and the Mediterranean regions. Sweet lupine is widely cultivated in Australia, the color of its flower varies from blue, to pink and white in demonstrated forms [2,3]. Lupinus angustifolius is one of the four lupines that are widely known and fully domesticated for agriculture <u>purposes</u> (Lupinus albus, Lupinus angustifolius, Lupinus luteu L. and Lupinus mutablis).

For several years, lupine flour has been used in pasta, milk, soya substituents and diet products. Lupine seeds are also eaten as snacks in most regions of the world [4]. Lupine seeds can contain toxicologically relevant bitter quinolizidine alkaloids whichalkaloids, which cause symptoms of poisoning of humans affecting the nervous, circulatory and digestive systems [References needed]. Typical symptoms of lupine alkaloid poisoning are dizziness, confusion, tachycardia, nausea and, dry mouth, loss of motoric coordination and in high doses, cardiac arrest and respiratory paralysis [5]. The levels of quinolizidine alkaloids in lupine seeds vary depending on the botanical and geographical origin of the lupine variety from which they derive. In contrast to bitter lupine, sweet lupine has low level of toxic poisoning alkaloid and suitable for human consumption even without debittering [6].

Lupine seeds, like other legumes are <u>good</u> sources of vitamin, protein and fibers. Studies reported the pharmacological benefits of lupine alkaloids, with activity on circulatory system, metabolism against obesity and improving bowel health [7].

Due to the low concentration of biologically active materials in plants, it is necessary to use effective methods for extraction of these substances, specially using solvents that are environmentally friendly. Consequently, ethanol was the solvent of choice with different percentages to extract polyphenols and flavoncids whichflavoncids, which are responsible for the pharmacological properties such as antioxidants and antimicrobials. Therefore, a complete comprehensive determination of lupine properties is essential, not only because of its potential toxicity to humans but also for its pharmacological properties.

2. MATERIAL AND EXPERIMENTAL DETAILS

2.1. Raw Materials and Equipment

Sweet lupine seeds were obtained from the local market, while all other fine reagents/chemicals were purchased from Sigma Aldrich Company. Dejonized water was used in all preparations, and commercial alcohol was used for extraction. An Analytik Jena Specord 40 UV-VIS spectrophotometer was used for the determination of the antioxidant activityeapacity, phenolic content and flavonoids. A mModel FP 640 flame photometer was used for the measurements of sodium and potassium content. Bacteria strains were provided from Holy Family Hospital in Bethlehem, Palestine.

2.2. Extraction of Seeds

Lupine seeds were groundgrinded and extracted by Soxhlet extractor using different percentages of ethanol (50%, 60%, 70%, 80% and 95%) for three hours. The sSolvent was then evaporated under vacuum and the residue was stored away from direct light.

2.3. Preparation of Samples (Stock Ssolution)

Residue was dissolved in 50% ethanol (200 mg/100 mL) and this served as <u>a stock</u> solution for the determination of sodium, potassium, ferrous <u>ion</u> iron, antioxidant activity, total phenolic content and flavonoids.

2.4. Determination of Ssodium and Ppotassium

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Sodium and potassium were determined by flame photometry against reference standards for both elements. From the calibration <u>curvescurves</u>, the concentration of the extracted samples was determined.

2.5. Determination of Ferrous Lion (Fe⁺²)

Fe⁺² in sample extract was determined by <u>a</u>titrimetric method: redox titration of Fe⁺² with potassium dichromate using sodium diphenylamine sulfonate, a pH independent redox indicator. Endpoint was detected as the color turned to violet.

2.6. Determination of Antimicrobial Activity

Antibacterial activity was studied on sweet lupine against S. aureus (Gram positive) and E. coli (Gram negative) bacteria. "An "Agar Well" method was used to test the resistance of extract to bacteria [8]. In this method, three wells were created in the Agar plates of the Muller-Hinton broth [9]: the first of which was for negative control (H_2O), the second was for positive control (Amoxicillin), and the third one was is for sample (the extract). High concentrations of extracts (1.2 g/100 mL) were used for the determination of antibacterial activity. Petri dishes were incubated at 37°C for 24-48 hours.

2.7. Antioxidant Activity

The antioxidant activity <u>or capacity</u> was <u>determined</u> by the ferric <u>reducing antioxidant power</u> (FRAP) [10] method that relies on reduction by antioxidants of the complex ferric ion-TPTZ (2,4,6-tri (2-pyridyl)-1,3,5-triazine). The binding of Fe^{+2} to the ligand makes a complex that gives the blue color intensity. The absorbance was measured to test the <u>concentrationamount</u> of iron <u>reduced which reduced</u>, <u>which</u> is correlated with the <u>concentration amount</u> of antioxidant.

2.7.1. Analysis

For sample extract: 800 μ L of sample (Stock solution) was mixed with 1000 μ L FRAP, and for standard: 80 μ L of standard FeSO₄ (0.1–2.0 mM) was mixed with 1000 μ L H₂O and 1000 μ L FRAP. Solutions were incubated at 37 °C for 15 minutes and the absorbance of the colored product was measured at λ =593 nm against 50% ethanol as blank.

2.8. Total Phenolics Content

The total <u>concentration</u>amount of phenolic compounds was determined using Folin-Ciocalteu method [11,12].

2.8.1. Analysis

For sample extract, 1.20 mL of 7.5% Na_2CO_3 was mixed with 100 μ L sample and 1.8 mL diluted Folin- Ciocalteu reagent (1:1). Standard preparation was done as the followsing, 1.20 mL Na_2CO_3 was mixed with 40 μ L standard Gallic acid (90-900 ppm) and diluted Folin-Ciocalteu reagent (1:1). The mixtures were incubated for one hour at 30 °C where the sample turned to color was turned to greenish-blue, and absorbance was measured at λ =765 nm.

2.9. Flavonoids

The colorimetric identification and quantification of the two types of flavonoids (flavonones and dihydroflavonols) was based on their reaction with 2,4-dinitrophenylhydrazine (DNP) in the presence of KOH in methanol [13,14].

2.9.1. Analysis

For sample extract and standard (r-Rutin, -(5 – 100 ppm), -200 µL of ssample (Stock solution) was mixed with 400 µL 2,4-dinitrophenylhydrazine and placed in a water bath at 50 °C for 60 minutes. After cooling to room temperature, 800 µL of a 10% KOH/methanol

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3. RESULTS AND DISCUSSION

3.1. Extraction. Extraction

Lupine seeds were extracted with different percentages of ethanol. R and results are summarized in table 1. As shown, the highest percentage of extract was obtained when 50% of ethanol was used (18.6%). On the other hand, the lowest percentage was obtained was when 60% and 70% of ethanol were used (10.0% for both).

Table 1. Percentages of residue obtained from sweet lupine seeds

Solvent	Result	
95% EtOH	12.2%	
80% EtOH	10.9%	
70% EtOH	10.0%	
60% EtOH	10.0%	
50% EtOH	18.6%	

3.2. Determination of Ssodium and Ppotassium

Results of sodium and potassium are illustrated in table 2. As table 2 shows. The highest concentrationamount of sodium was obtained when 80% of ethanol was used while the lowest concentrationamount was obtained with 50% ethanol. This can be attributed to the fact that sodium is present in sweet lupine as an organic salts that tends to dissolve in ethanol more than in water. In a previous study on bitter—Lupinus albus seeds [15], the highest concentrationamount of sodium wase obtained withas with 50% ethanol, suggesting that sodium is present as in—inorganic complexes in the bitter—seeds. The highest concentrationamount of potassium in sweet lupine was obtained was—when 50% and 60% ethanol were used. This result is in agreement with results reported by Hanania et al. (2018) where of-bitter lupine seeds where extracted with 60% ethanol—ethanol, which resulted in produced the highest potassium concentrationsamount [15].

Table 2. Sodium, potassium and ferrous content of extracts (mg/g)

Ethanol %	Sodium	Potassium	Ferrous	
95%	10.29	0.15	3.726	
80%	56.51	1.00	4.340	
70%	17.59	0.6	6.854	
60%	10.51	2.25	2.424	
50%	9.20	2.33	1.839	

3.3. Determination of Ferrous Ions

Table 2 <u>also</u> shows <u>that</u> as the percentage of ethanol decreases, the ferrous content increases until <u>it reaches-the</u> 70% ethanol <u>extraction</u>, where the maximum content <u>of ferrous</u> <u>was extractedis observed.</u> However, below 70% alcohol, the ferrous content decreases.

3.4. Antimicrobial Aactivity

Sweet lupine extract showed no inhibition against neither *E. coli* nor *S. aureus*. The Our negative results reported here are in agreement with previous studies in terms of *E. coli* [16]. The extract of *Lupinus* Angustifolius was weakly active on *E. coli*.

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201 4. CONCLUSION 202

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3.5. Determination of Antioxidant Activity and Total Phenolics Content

As illustrated in table 3, the highest activity of antioxidants was obtained when using 95% ethanol was used. Similar The same results wereas obtained for phenolics which that is an important play a significant antioxidant role as phytochemical in sweet lupine seeds. This result was expected since antioxidants such as phenolics are organic compounds that tend to dissolve in ethanol rather more than water [17]. Ethanol was used in accordance with the literature data, to ensure optimum extraction of phenols, because since the extraction efficiency of <u>plant material using</u> ethanol<u>-water</u> extraction from plant material is greater and environmentally friendly when using ethanol-water system than methanol-water extraction [18, 19]. Compared to bitter lupine, it was found that bitter lupine? seeds have a higher antioxidant activity since it contains a higher content of phenois-[15]. Since ethanol ethanolic extract contains other compounds in addition to phenols. The, Folin-Ciocalteu method was used because of due to its low sensitivity.

Table 3. Antioxidant aActivity and total pPhenolics for sweet lupine extracts

Ethanol %	mg FeSO₄/g extract	mg Gallic acid/g extract
95%	20.24	24.60
80%	19.22	20.98
70%	12.03	18.35
60%	9.15	11.92
50%	7.23	12.28

3.6. Determination of Flavonoids Content

Flavonones and dihydroflavones are the two types of flavonoids that were determined in sweet lupine. As illustrated in table 4, 95% ethanolic? extract resulted in the showed highest concentrations of significant amount of flavonoids, i.e. (115.02 mg rRutin/g extract). It is worth mentioning that the concentration of these bioactive chemicals depends on many factors including climate, precipitation and soil conditions [20].

Table 4. Rutin (flavonoids) concentrations Results obtained from or different percentage ethanol extraction flavonoids content

Ethanol %	mg <u>R</u> Rutin/g extract
95%	115.02
80%	11.77
70%	35.19
60%	22.56
50%	39.83

Based on the above mentioned results, antioxidants present existing in sweet lupine are organic compounds and are more likely to dissolve in ethanol than in water. Moreover, polyphenols (phenolcs?) and flavonoids have many biological properties in plant especially as antioxidants, while antibacterial agents are absent from sweet lupine seeds. Although Comment [PM9]: Sweet/bitter lupine

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COMPETING INTERESTS

Authors have declared that no competing interests exist.

CONSENT

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