Allelopathic activity of some medicinal plants against plant pathogenic bacteria (*Erwinia carotovora*)

## Abstract:

The in depth use of chemicals in plant protection has given rise to concerns about pesticide residues in the environment. Allelopathic plants provide better alternative for this purpose due to being cost-saving, simple to use, eco-friendly, economical and safe. investigation on plant pathogens has not been given an adequate attention. The present investigation was meted out to guage the allelopathic effect of three medicative plants to seek out a suitable alternative to artificial antibiotics for the management of plant diseases caused by bacteria.

Aqueous extracts of three plant species (Artemisia herba alba, Pistacia atlantica and Juniperus phoenicea which from Al-Jabal Al-Akhdar region (Libya), were screened against plant pathogenic bacteria (Erwinia carotovora subsp. Carotovora) in comparison to the antibiotic Streptomycin (positive control) under laboratory conditions. Antibacterial activities was evaluated for different concentrations of different extracts for each plant studied.

Results showed varying in levels of the inhibition were observed in response to the tested extracts at the concentrations of 20, 40, 80 and 100 %. The differences in their inhibition depend on to the type of plant and concentration of extract.

The effectiveness of the extracts against tested bacteria could be arranged by the following order; *Artemisia herba alba >Pistacia atlantica > Juniperus phoenicea* .the study gave encouraging results for the use of these plant extracts in the control of tested bacteria.

The results indicate that the rate of inhibition increases with increased concentration in all transactions, and that it increases the diameter of the inhibition of bacteria by increasing the concentration of the extract and that the highest inhibition of *Erwinia carotovora* bacteria was obtained using the *Artemisia herba alba* extract.

Plant extracts resulted in antibacterial activity is potential to use in the management of plant diseases as an alternative to antibiotics. Further phytochemical analysis is required to identify the bioactive compounds responsible for antibacterial activity.

**Key words:** Allelopathy, plant pathogenic bacteria, Medicinal plants, Inhibition zone, Disc diffusion Method, Aqueous extract.

#### :[1U]Comment

Too many formatting errors throughout the text ... need deep fixes !!

Formatted: Font: Not Italic

Formatted: Font: Not Bold, Not Italic

## :[2U]Comment

For the corrections made ... a careful correction must be made by standardizing the text and placing in the norms of the magazine ...

## Introduction-:

Biological management of plant pathogens and plant diseases is critical in agriculture. There is great incentive to find biologically active natural products from higher plants to act as bactericides, insecticides ,fungicides and herbicides that are better than artificial agrochemicals, and more safe environmentally and a healthily. Allelopathy offers beautiful environmentally friendly different to pesticides in agricultural pest management (Inderjit and Mukerji, 2006). Allelopathy has been defined is a physiological process with ecological implications (Reigosa *et al.*, 2006).

The International Allelopathy Society defined allelopathy as any process it involves secondary metabolites produced by plants, microorganisms, viruses and fungi-which effect on the growth and development of agricultural and biological systems (Cheng and Cheng, 2015).

It may also include the substances effect on growth and development of nearby plants through both inhibitory and stimulatory biochemical interactions. (Rice,1974; Rice,1984; Tang,1986). These chemicals vary among plant species and organs and are released into the environment by different processes; root exudation, volatilization, leaching, and tissue decomposition in soil (Parvez et al., 2004).

Allelopathy as an ecological phenomenon, has been defined as any direct or indirect effects of one plant, including micro organisms, on another through the release of phytochemical compounds (allelochemicals) into the environment, (Rice, 1984; Weir et al., 2004; Inderjit, et al., 2006).

these chemicals vary among plant species and organs and are released into the environment by different processes; root exudation, volatilization, leaching, and tissue decomposition in soil (Parvez et al., 2004).

Allelochemicals are produced by any organ of the plant and are mainly secondary metabolites.

plant secondary metabolites are usually associated with plant defense against herbivores and pathogens, these distinctive compounds may be linked to wide range of ecological functions. (Hussain and Reigosa, 2011).

Phenolic compounds are one in every of the biggest group of secondary metabolites, consisting of four main groups divided in keeping with the number of phenol rings and the structural parts that bind those rings, including flavonoids, phenolic acids, tannins, saponins, cinnamic acid coumarins, terpenoids, quinones, and lignans (Balasundram *et al* 2006). some secondary metabolites are considered as natural pesticides against pathogens, bacteria, fungi, insects, and weeds. (Soltoft *et al* 2008), merge allelopathy into agricultural management systems could scale back the employment of pesticides, reduction environment/soil pollution, and reducing the risk of auto toxicity dangers, increase crop growth and enhancing productivity (Sodaeizadeh and Hosseini, 2012).

## :[3U]Comment

Redo Introduction ... many parts of the text are redundant ...

redundant :[4U]Comment

very subjective :[5U]Comment

#### :[6U]Comment

add the paragraphs of the text ... will improve the aesthetics of the manuscript ... And finish the topics mentioned!!

 ${\sf redundant:} \textbf{[7U]} \textbf{Comment}$ 

redundant :[8U]Comment

redundant : [9U]Comment

Among the massive diversity of plant pathogens, that include viruses, bacteria, fungi, nematodes, and insects, nearly a hundred and fifty are bacterial species cause diseases to plants that limit the crop production and caused crop losses the use of agrochemicals to manage bacterial diseases should be minimized due to the increased risk of its harmful residues (Kannan and Bastas, 2015).

The current and future trend towards the use of cheap and safe insecticides from natural plant extracts instead of harmful and expensive industrial chemicals has become the current control of these lesions at present time. (Danahap and Wonang, 2016). Some medicinal plants have the ability to eliminate antibiotic- resistance bacterial species, because they effect on a large number of microbes as well as safe and low cost. (Hassawi and Kharma, 2006).

Al-Jabal Al-Akhdar region (Libya) has highest species diversity and having distinct environmental characteristics associated with evergreen forest and it has environment similar to other region in Southern Europe (Al-Idrissi et al., 1996).

The number of plant species reach up of 1100 species from the total of plant species in Libya (2000 species) with about 75 species of plants that grow only in AL-Jabal AL Akhdar and have been served for as basis of traditional medicinal systems for thousands of years. (Al-Idrissi *et al.*, 1996 and El-Barasi *et al.*, 2013).

In addition, such plants produce a remarkable diverse array of over low known and high high molecular mass natural products which are 5,00,000 as secondary metabolites, which can be used as an alternative from of health care as well as screening for active compound that have significant effects against human and plant pathogens. (Hegazy et al., 2011; Attitalla et al., 2010 and Fatope et al., 2000).

In the current study, three of these plants with medicinal properties used in folk medicine were selected in Libya they are *Artemisia herba alba ,Pistacia atlantica* and *Juniperus phoenicea*. To test its inhibitory activity for the growth of plant pathogenic bacteria.

The objective of the present work was to evaluate the allelopathic potential of three Libyan folk medicinal plants aqueous extracts against plant pathogenic bacteria <a href="Erwinia carotovora">Erwinia carotovora</a>.

#### Materials and methods:

## **Plant Material Collection**

Plant materials of species belonging to 3 botanical families included in this study were collected from Al-Jabal Al-Akhadar of Libya, during October 2018.

redundant :[10U]Comment

Table 1. Plant species screened for Allelopathic activity.

Plant species	Family	Local name	Part used
Artemisi aherba alba	Asteraceae	Sheah	Leaves &steams
Juniperus phoenicea	Cupressaceae	Araar	Leaves &steams
Pistacia atlantica	Anacardiaceae	Battom	Leaves &steams

Artemisia herba alba-e\_(-Asteraceae-) from Sosa region (125 m) which considered as littoral zone near to the Mediterranean Sea. while Juniperus phoenicea, (Cupressaceae) and Pistacia atlantica (Anacardiaceae). from Shahat region which represents (450 m) above the sea level.

The plants were classified and authenticated according to Jafri and EL- ghadi (1978). as well as herbarium at Department of Botany, Faculty of Sciences, Omar EL-Mukhtar University, Elbyda, Libya.

**Tested microorganisms:** plant pathogenic bacteria (*Erwinia carotovora subsp. Carotovora*) (Family: Enterobacteriaceae), The bacterium infects a variety of vegetables and plants including carrots, potatoes, cucumbers, onions, tomatoes, lettuce and ornamental plants like iris.. It was obtained from Laboratory of bacterial plant diseases Department of prevention, Faculty of Agriculture Omar EL-Mukhtar University, Elbyda, Libya. The bacterial strains were maintained on nutrient agar (NA: Peptone 5g, Beef extract 3g, NaCl 8g).

# **Plant Material Preparation**

After the collection of the plants, they were dried in a shady place at room temperature for ten days.

## Preparing the aqueous extracts of plants

the dried aerial parts (-leaves and stems) were ground into powdered form then 50 g of the powder were diluted into 500 ml of distilled water (-Bajalan et al., 2013-). Next to that, this mixture was left on a Shaker for 24 hrs in room temperature at speed of 120 rpm (Ghorbani et al., 2008).

four-folded cotton fabric was used as a filter to separate rough solid particles from solution, The contents were then filtered with Whatman No. 1 filter paper and then it was centrifuged with the speed of 2000 rpm for 15 minutes (Bajalan et al., 2013). Three concentrations of solutions were prepared based on volume/volume percent (v/v. %), except for the basic solution 100 % which was weight was a weight/ volume percent (w/v. %) (Elshatshat, 2010). Four concentrations were prepaid (20, 40, 80 and 100%) in addition to the antibiotic Streptomycin as a positive control.

## **Bactericidal activity**

The Bactericidal activity of plant aqueous extracts was evaluated by disc diffusion Technique developed by Bauer et al. (1966).

Formatted: Font: Not Bold
Formatted: Font: Not Bold
Formatted: Font: Not Bold

Formatted: Font: Not Italic

Formatted: Font: Not Bold

Formatted: Font: Not Bold

Formatted: Font: Not Bold, Not Italic

Formatted: Font: Not Italic

Briefly, this method is based on the diffusion of water extract from a filter paper disc through the solidified culture medium (nutrient agar) of Petri dishes.

Whatman filter paper (NO<sub>e2</sub>) was used to prepare small discs (5 mm in diameter), then sterilized at 121 °C for 15 min in an autoclave.

then it was poured the culture medium (nutrient agar) into the Petri dishes (three dishes /concentration). Then processing a bacterial suspension from colony (24 hours old) ( $10^8$  cfu /ml) and performed three dilutions (1000 time), taking from the last dilution  $250\mu$ l by plastic pipette and distributed regularly in Petri dishes.

The discs of Whatman filter paper were separately impregnated with extract (serial of dilution of aqueous extract ranging from 20,40,80 and 100%) for 5 minutes and left to dry for 1-2 hrs at room temperature, four discs of each plant extracts were placed on the surface of the Petri dishes which Containing the bacterial suspense.

In the same way, tablets were immersed in sterile distilled water as a test and placed on the surface of the dishes.

and under the same laboratory conditions, an antibiotic experiment (streptomycin) was performed with the concentration (0.004) % for comparison.

all dishes are placed in the incubator at a temperature of 28 °  $C \pm 2$  ° C for 24 hours, (three replicated plates were used for each treatment).

The antimicrobial plant extracts activity was detected as a result of clear inhibition zone, The diameter of inhibition zone created by each disc was measured (in mm) using a ruler.

Antibacterial activities of different concentrations of different extracts for each plant studied were assessed, and was recorded the lowest concentration of the sample that inhibits visible growth giving clear zones of inhibition after 24 h incubation.

### **Statistical Analysis:**

Statistical analysis was performed using a computer run program (Minitab software). One way ANOVA followed by Tukey,s HSD test was performed to show the statistical significance among the means of the groups. Results were expressed as mean  $\pm$  Standard Division (SD). P-value below 0.05 was considered to be statistically significant.

# Results and discussion-:

In the present study, the tested bacteria were to be sensitive to tested plant extracts. The preliminary screening of selected plant extracts against the phytopathogen bacteria was done using disc diffusion method.

It was found that inhibition zone (IZ) with a diameter greater than 5mm indicated there Antibacterial activity-. (Palombo and Semble 2001).

Our results showed that , all the aqueous extracts screened against plant bacteria shows significant allelopathic potency ,the in vitro antimicrobial activity of *Artemisia herba-alba*, *Pistacia atlantica* and *Juniperus phoenicea* against phytopathogen bacteria (*-Erwinia carotovora*) was assessed by the diameter of inhibition zone around the discs, with varying degrees of inhibition of different concentrations of different extracts for each plant studied, from 20% to\_100% among all the aqueous extracts obtained from the three studied plants.

These results are consistent with previous research on the containment of aqueous extracts of a number of plants on biologically effective compounds against bacteria

Formatted: Font color: Auto

Formatted: Font color: Auto

Formatted: Font color: Auto, Subscript

Formatted: Font: Not Bold, Font color: Auto

Formatted: Font color: Auto

Formatted: Font: Not Italia

and pathogenic fungi, where (Saleem *et al.*, 2010) proved that aqueous extracts could contain anthocyanins, tannins, saponins, terpenoids, polypeptides, and lectins.

Terpenoids derived from plants have antibacterial activity, (Drewes *et al.*, 2005; Mathabe *et al.*, 2008)—. Tannins are water soluble polyphenols found in almost all plants. It is known to possess various biological effects with antifungal and antibacterial effects. (Lanchoti Fiori *et al.*, 2013; Scalbert, 1991).

In a study carried out by Mohammed and Abdu EL-Salam, (2013) on the extracts of some medicinal plants developing in al-Jabal al-Akhdar area of Libya (Rosmarinus fficinalis, Datura alba and Capparis spinose-) which showed allelopathic activity against bacteria and pathogenic fungi of plant. The effect of the allelopathic was examined using the disc diffusion method. which proved to be that the inhibitory effect increased by increasing the concentration of plant extracts, and diameters of the inhibition zone of bacteria increases by increasing the concentration of the extract. This is similar to the results obtained in our current study, as shown in table (2) which explained that the rate of the inhibition increased with increased concentration in all transactions and that it increases the diameter of the inhibition of bacteria by increasing the concentration of the extract and that the highest inhibition of Erwinia carotovora bacteria was obtained by using the A. herba alba extract, the diameters of the inhibition zone was 8.33-mm, 10.00-mm, 12-mmand, 13.33 mm, followed by P. atlantica, with 7.67-mm, 10.00-mm, 10.67 and mm, 12 mm, then J. phoenicea extract with a diameters of 7.00-mm, 9.00-mm, 10.67 and mm, 11.67 mm, for concentrations of 20, 40, 80, 100% respectively, compared to 15 mm, when using the antibiotic at concentrations of 0.004, as indicated in the table (2).

Increasing the efficiency with increasing the concentration is may be due to the effect of the extract on the membrane permeability of the cell membrane and the work of enzymes transferring (Transferase) of cell bacteria-

The effectiveness of the extracts of the selected plants is attributed to the presence of phenolic compounds that have a high inhibitory effect on the bacteria. Mounir Tilaoui, et al. (2015). Also found that tannins are toxic to fungi, yeast and bacteria through their association with the wall of these microorganisms, preventing their growth and the effectiveness of protease this is accordance with (Jones, G. A, et al 1994).

Formatted: Font: Not Italic

**Table 2**. Allelopathic activity of aqueous extracts of the studied plants against plant pathogenic bacteria using agar disc diffusion method, Streptomycin was used as positive control.

 $IZ (mm \pm SD) in mm$ 

The plant	streptomycin	20	40	80	100
	Mean± SD	Mean ± SD	Mean ±SD	Mean ±SD	Mean± SD
A. herba alba	$15^{a} \pm 0$	$8.33^{\circ} \pm 0.47$	$10^{c} \pm 0.82$	$12^{\rm b}\pm0$	$13.33^{ab} \pm 0.94$
J. phoenicea	15 <sup>a</sup> ± 0	$7^{\rm d} \pm 0.82$	9° ± 0	$10.67^{bc} \pm 0.94$	$11.67^{\rm b} \pm 0.47$
P. atlantica,	$15^a \pm 0$	$7.67^{\rm d} \pm 0.47$	$10^{c} \pm 0.82$	$10.67^{bc} \pm 0.47$	12 <sup>b</sup> ± 0

Data are expressed as mean  $\pm$  SD of three replicate. Within each row, means with different superscript (a, b, c or d) were significantly different at p<0.05. Where means superscripts with the same letters mean that there is no significant difference (p>0.05).

Our results for the allelopathic activity of *Artemisia hrba alba* are consistent with many previous studies including those referred to by(Abdel-Rahman et al., 2017) conducted a study to assess the allelopathic effect of three desert plants (*Artemisia* judaica, *Asphodelus microcarpus* and *Solanum nigrum*) against plant pathogenic bacteria (*Erwinia carotovora*, *Xanthomonas campestris* and *Ralestonia solanacearum*), using leaf extracts of each plant with different concentrations.

Artemisia judaica extract showed the highest level of antibacterial activities against all the tested bacterial strains also recorded the best result of minimum inhibitory concentrations.

Dababneh (2008).which reported that *Artemisia hrba alba* possesses great antibacterial effect of pathogenic bacteria using disc diffusion method.

Hassawi and Kharma (2006) stated that Artemisia sp. extract is anti-diabetic, antitumor, antifungal against *Candida albicans* and bactericidal.

It was pointed out by (Halbuda et al., 2014) that the water extract of Artemisia herba alba that growing in Libya has an inhibitory effect against bacteria\_-

Chemical studies of Artemisia species indicate that all classes of compounds compounds, especially terpenes and flavonoids are present in these species According to Wright (2002). (Sellami *et al.*2010) confirmed the presence of alkaloids, sesquiterpene, coumarins and acetylenes.

Our results showed the presence of antimicrobial effects of the aqueous extract from the leaf of the *Pistacia atlantica* and this corresponds to some of the previous research (Panahi *et al.*, 2013 and Roozegar *et al.*, 2016).

The antibacterial activity of the extracts of *Pistacia atlantica* may be due to the presence of certain antimicrobial secondary metabolite, the leaf extract is rich in phenol compounds and antibacterial properties. (Farzangi *et al* 2013).

Leaf extracts of *P. atlantica* proved to be very efficient against plant pathogenic fungi and bacteria. (Rhouma *et al.*, 2009).

*P. atlantica* extracts contain polyphenols, which have a high level of antimicrobial activity against plant pathogens. Several studies have suggested that polyphenols inhibit the growth of microorganisms. (Mila *et al.*, 1996).

Formatted: Font: (Default) Times New Roman

Formatted: Font: (Default) Times New Roman

Formatted: Font: (Default) Times New Roman, 12 pt

Formatted: HTML Preformatted, Adjust space between Latin and Asian text, Adjust space between Asian text and numbers, Pattern: Clear (White)

Formatted: Font: Not Bold

Our results agrees with that obtained by Ekweny and Elegalam (2005) who reported that juniper phoenicea inhibit growth E. coli.

Also, Malu *et al.*, (2009) reported that *juniper phoenicea* extracts have antibacterial properties and could be used for the treatment of bacterial infections.

Bouzouita et al. (2008) found that J. phoenicea oil shows inhibition effect against Klebsiella oxytoca, Lactobacillus plantarum, Saccharomyces cereviseae and Geotrichum candidum.

terpenoids have been isolated from the genus Juniperus according to (Barrero et al 2006 and Mansouri et al 2010)

and flavonoids according to (Nakanishi, 2005). The species of Juniperus important medicinal plants used extensively in traditional medicine as Juniperus leaves have anti-inflammatory activity. (Mansouri *et al* 2010).

There are many reasons that may lead to varying efficacy of the plant extracts and their allelopathic effect. The difference in efficacy may be due to the stage of plant sample collection, soil nature, other environmental factors, storage conditions, plant part used, extraction method, Extraction, and different sensitivity of strain test. (Barbour *et al.*, 2004 and Entezari *et al.*, 2009).

The presence of various secondary metabolites such as alkaloids, quaternary alkaloids, coumarins, flavanoids, steroids/ terpenoids, phenols etc. have been reported in the various plants extracts (Aswal *et al.*, 1984; Abraham et al., 1986; Chopra et al., 1992)

which may be responsible for the antibacterial properties of the plants.

leaves contain several types of water-soluble polyphenols which have a high antioxidant and antimicrobial potential (Barotto *et al.*, 2003; Ljubuncic *et al.*, 2005).

Based on these results, the antibacterial activities seem attributable to flavonoids, these compounds have been associated with antimicrobial effects in various studies using plant extracts (Nwaogu *et al.*, 2007; Newze *et al.*, 2004).

### **Conclusion:**

In conclusion, the obtained results showed that the aquatic extracts of some medicinal plants that grow wild in the eastern region of Libya showed allelopathic activities, which proved effective in reducing the growth of bacteria responsible for changes in some vegetables. These results obtained from laboratory experiments can be

supplemented by other, more comprehensive, open-field studies to assess the practical use of these extracts within an integrated pest management system. To date, this type of studies has been conducted only a limited number of them to test commercially grown commercial plant compounds available to the agro-industry. While some of these compounds have a role in activating the mechanisms of plant protection. Therefore, we need to deepen our knowledge about the mechanisms of plant water extracts against microorganisms.

Allelopathy offers safe solutions in pest management; in the long run it would be a luminous direction to proceed in order to develop bactericidal by using the allelochemicals.

## **REFERENCE:**

Weir TL, Park SW, Vivanco JM (2004) Biochemical and physiological mechanisms mediated by allelochemicals. *Current Opinion in Plant Biology*. 7:472-479.

Inderjit, Callaway RM, Vivanco JM (2006) Can plant biochemistry contribute to understanding of invasion ecology? *Trends Plant Sciences* 11:574-580.

Parvez SS, Parvez MM, Fujii Y, Gemma H (2004)Differential allelopathic expression of bark and seed of *Tamarindus indica* L. Plant Growth Regulation. 42:245-252.

Abdel-Rahman, A. G.; Hashem, Hanan, A.; Kassem, Hala, A. and Abdel Aziz, Nehad, F(2017) .Allelopathic activity of some desert plants against plant pathogenic bacteria and nematodes. *Journal of Environmental Sciences*, 37(2):15-35

Mohammed, N.A and Abdu EL-Salam, F.M (2013) The inhibitory effect of extracts of *Rosmarinus officinalis*, *Datura alba* and *Capparis spinosa* against some bacteria and pathogenic fungi of plants. *Al-Mukhtar Journal of Science* 28(1):70-81.

Inderjit, H.K. and Mukerji, K.G. (eds.) (2006): Allelochemicals: biological control of plant pathogens and diseases, Vol. 2, 208.

Cheng, F. and Cheng, Z. (2015): Research progress on the use of plant allelopathy in agriculture and the physiological and ecological mechanisms of allelopathy. *Front plant science*, 6: 1-16.

Jafri , S. and El-Gadi , A. (1978) : Asteraceae . In flora of Libya , 107. Al-Fateh University Press, Tripoli, Libya .

Rice, E.L. (1974) Allelopathy:1st edition Academic Press.pp366.

Rice, E.L. (1984) Allelopathy: 2nd edition Academic Press. pp. 422.

Tang, C.S. (1986). Allelopathy: State of the science. In: The Science of Allelopathy (Ed., A.R. Putnam and C.S Tang), pp. 1-19, Wiley, New York

Hussain I.M and Reigosa J.M (2011) Allelochemical stress inhibits growth, leaf water relations, PSII photochemistry, non-photochemical fluorescence quenching and heat energy dissipation in three C3 perennial species. J Exp Bot 624533-4545.

Balasundram N, Sundram K, Samman S (2006) Phenolic compounds in plants and agri-industrial by products: antioxidant activity, occurrence and potential uses. Food Chem 99: 191-203

Soltoft M, Joergensen LN, Svensmark B, Fomsgaard IS (2008) Benzoxazinoid concentrations show correlation with Fusarium Head Blight resistance in Danish wheat varieties. Biochemical Systematics

and Ecology 36: 245-259.

Sodaeizadeh, H. and Hosseini, Z. (2012): Allelopathy an environmentally

friendly method for weed control. *International Conference on Applied Life Sciences*, Turkey.387-392.

Kannan, V.R. and Bastas, K.K. (2015): Sustainable approaches to controlling plant pathogenic bacteria. International Standard Book. CRC press. New York.

Danahap, L.S. and Wonang, D.L. (2016): Antinematicidal efficacy of root exudates of some crotalaria species on *Meloidogyne incognita*(root-knot nematode) (kofoid and white) chitwood isolated from infected

lycopersicumesculentum (Tomato) plant. International Journal of Scientific & Technology Research, 5: 79-84

Hassawi D. and Kharma A., 2006; Antimicrobial activity of some medicinal plants against *Candida albicans*. *Journal of Biological Sciences* . 6 (1): 109-114.

Al-Idrissi, M., Sbeita, A., Jebriel, A., Zintani, A., Shreidi, A., Ghawawi, H. and Tazi, M. 1996. Libya: Country report to the FAO international technical conference on plant genetic resources. Leipzig, Germany. Tripoli, Libya; 1996. p. 1–29.

Hegazy A, Boulos L, Kabiel H, and Sharashy O .( 2011 )Vegetation and species altitudinal distribution in Al-Jabal Al-Akhdar landscape, Libya. *Pakistan Journal of Botany*. 43:1885–1898.

El-Barasi, Y.M.M, and Saaed, M.W.B.(2013). Threats to plant diversity in the north eastern part of Libya (Al-Jabal Al-Akahdar and marmarica plateau ) *Journal of Environmental Science and Engineering* A.; 2:41

Attitalla, I.H., A.M. Alhasin, M.A. Nasib, A.H. Ghazali, L. Zakaria, H.M. Jais, I.A.A. Balal, and B. Salleh ,(2010). Occurrence and microbiological characteristics of azospirillum strains associated with leguminous and non-leguminous plants in Al-

Jabal Al- Akhdar region, Libya. *American-Eurasian Journal of Agricultural and Environmental Science*. 8(6):617–625.

Fatope MO, Adoum OA, Takerda Y.(2000) Oleanene palmitate from ximenia Americana. *Pharmaceutical Biology* . 38(5):391–393

Bajalan I., Zand M., and Rezaee S.(2013). Allelopathic effects of aqueous extract from *Salvia officinalis* L. on seed germination of barley and

purslane. International Journal of Agricultural Crop Sciences., 5(7): 802-805.

Ghorbani M., Bakhshi-Khaniki G., Shojaei A.A. (2008). Examination of the effects of allelopathy of *Artemisia sieberi* Besser subsp. sieberi on seed germination and *Avena lodoviciana* and *Amaranthus retroflexus* 

seedlings growth. Pajouhesh Sazandegi 79:129–134.

Elshatshat S. A. (2010). Allelopathic Effects of *Artemisia Herba-Alba* Aqueous Extracts on Germination of Tomato and Wheat Seeds. *Journal* 

of Science and Its Applications, 4(1): 1-6

Bauer, A.W., Kirby, M.M., Sherris, J.C. and Turck, M. (1966) Antibiotic susceptibility testing by a standardized single disc method. *American* 

Journal of Clinical Pathology, 45, 493-496.

Palombo, E.A and Semple, S.J 2001. Antibacterial activity of traditional Australian medicinal plants. *Journal of Ethanopharmacology* 77:151-157

Saleem M, Nazir M, Shaiq Ali M, Hussain H, Lee YS, Riaz N, Abdul Jabbar. 2010. Antimicrobial natural products an update on future antibiotic drug candidates. Natural Product Reports., 27(2): 238–254.

Drewes SE, Khan F, Van Vuuren SF, Viljoen AM. 2005; Simple 1, 4- benzoquinones with antibacterial activity from stems and leaves of *Gunnera perpense Phytochemistry*. 66:1812-1816.

Mathabe MC, Hussein AA, Nikolova RV, Basson AE, Meyer JJ, Lall N.2008; Antibacterial activities and cytotoxicity of terpenoids isolated from

Spirostachys Africana. Journal of Ethnopharmacology. 116:194-197.

Lanchoti-Fiori G, Fachin A, Correa V, Bertoni B, Giuliatti S, Amui S,2013 . Antimicrobial activity and rates of tannins in Stryphnodendron adstringens Mart. Accessions collected in the Brazilian Cerrado. *American Journal of Plant Sciences*. 4(11):2193-2198.

Scalbert A.1991 . Antimicrobial properties of tannins. *Phytochemistry*. 30(12):3875-3883.

Formatted: English (United States)

Dababneh, B.F. (2008): Antimicrobial activity of selected Jordanian medicinal plant extracts against pathogenic microorganisms. *Journal Food Agriculture And Environment*. 6(2): 134-139.

Halbuda M.B., Indbha A.M., and Karaza A.E., 2014. The antimicrobial effect of the *Artemisia herba alba* and *Eoridum glycobhyllum* against pathogenic bacteria. *Arts Journal University of Benghazi Education Libya* 9:1-16

Wright, C.W. 2002. Artemisia. First edition. London, New York: Taylor & Francis. 344 pp.

Sellami S., Mezrket A., Dahmane T. ., 2010. Activité nématicide de quelques huiles essentielles contre *Meloidogyne incognita*. Nematologia . Mediterranea 38: 195-201.

Panahi J, Havasiyan MR, Gheitasi S, Pakzad I, Jaliliyan A, Hoshmandfar R, Havasiyan M. 2013. The in Vitro Inhibitory Effects of the Aqueous Extracts of Summer Onion on Candida Albicans. *Scientific Journal of Ilam University of Medical Sciences*; 21(1):54-59.

Roozegar M.A., Jalilian F.A., Havasian M.R., Panahi J., Pakazed I. 2016. *Bioinformation* 12(1):19-21.

Farzanegi, P., Mousavi, M., Ghanbari-Niaki, A. 2013. Effect of *Pistacia atlantica* Extract on Glutathione Peroxidase Tissue Levels and Total Oxidative Capacity of Liver and Plasma Lipid Profile of Rabbit. Zahedan

*Journal of Research in Medical Sciences* 15(11): 59-63.

Rhouma A., Ben Daoud H., Ghanmi S., Ben Salah H., Romdhane M. and Demak M., 2009. Antimicrobial activities of leaf extracts of *Pistacia* 

and *Schinus* species against some plant pathogenic fungi and bacteria. *Journal of Plant Pathology* 91(2):339-345

Mila I., Scalbert A., Expert D., 1996. Iron withholding by plant polyphenols and resistance to pathogens and rots. *Phytochemistry* 42: 1551-1555

Malu S, Obochi G, Tawo E, Nyong B (2009) Antibacterial activity and medicinal properties of *juniperus phoenicea*. *Global Journal of Pure and Applied Sciences* 15: 365-368.

Ekweny U, Elegalam N (2005) Antibacterial activity of *juniperus phoenicea* extracts on *Escherichia coli* and *Salmonella typhyi*. *International Journal of Molecular Medicine and Advance Sciences* 1: 411-417.

Bouzouita, N., Kachouri, F., Ben Halima, M., & Chaabouni, M. M. (2008). Composition chimique et activités antioxydante, antimicrobienne et insecticide de

Formatted: Portuguese (Brazil)

l'huile essentielle de *juniperus phoenicea*. *Journal de la Société Chimique de Tunisie*, 10, 119-125.

Barrero A, Herrador M, Arteaga P, Quilez F, Sanchez E (2006) Chemical composition of the essential oil from the leaves of *Juniperus phoenicea*. L from North Africa. *Journal Essential Oil Research* 18: 168–169.

Mansouri N, Satrani B, Ghanmi M, El Ghadraoui A, Aafi A (2010) Valorisation des huiles essentielles de *Juniperus thurifera* et de *Juniperus oxycedrus* du Maroc. *Phytothérapie* 8: 166–170.

Nakanishi T (2005) Pair of new atropisomeric cupressuflavone glucosides isolated from *Juniperus communis* var. depressa. *Tetrahedron Letters* 46: 6533–6535.

Barbour EK, Al Sharif M, Sagherian VK, Habre AN, Talhouk RS, Talhouk SN. 2004. Screening of selected indigenous plants of Lebanon for antimicrobial activity. *Journal of Ethnopharmacology* 93(1): 1-7

Entezari M, Hashemi M, Ashki M, Ebrahimian S, Bayat M, Azizi Saraji AR, Rohani SR. 2009. Studying the effect *Echinophora platyloba* extract on bacteria (*Staphylococcus aureus* and *Pseudomonas aeruginosa*) and fungi *Candida albicans*, *Aspergillus flavus* and *Aspergillus niger* in vitro. *World Journal of Medical Sciences* 4(2): 89-92.

Abraham, Z., D.S. Bhakunl, H.S. Garg, A.K. Goel, B.N. Mehrotra and G.K. Patnalk. 1986. Screening of Indian plants for biological activity: Part XII. *Indian Journal of Experimental Biology* 24:48-68.

Aswal, B.S., Bhakuni. D. S., Goel, A. K., Kar, K. and Mehrotra, B. N., 1984. Antimicrobial activity of plants. *Indian Journal of Experimental Biology* 22: 487 – 504.

Chopra, R.N., Nayer, S.L., Chopra, I.C., 1992. Glossary of Indian medicinal plants: (3rd End) *Council of Scientific and Industrial Research*, New Delhi, pp. 7-246.

Barotto M.C., Tattini M., Galardi C., Pinelli P., Romani A,. Visioli F., Basosi R., Pogni R., 2003. Antioxidant activity of galloyl quinic derivatives isolated from *P. lentiscus* leaves. *Free Radical Research* 37: 405-412.

Ljubuncic P.H., Songa U., Coganb H., Azaizehc, Bomzon A., 2005. The effect of aqueous extracts prepared from the leaves of *Pistacia lentiscus* in experimental liver disease. *Journal of Ethnopharmacology* 100: 198-204.

Nwaogu L.A., Alisi C.S., Ibegbulem C.O., Igwe C.U., 2007. Phytochemical and antimicrobial activity of ethanolic extract of *Landolphia owariensis* leaf. *African Journal of Biotechnology* 6: 890-893.

Nweze E.I., Okafor J.I., Njoku O., 2004. Antimicrobial activities of methanolic extract of *Trume guineesis* (Schumm and Thorn) and *Morinda lucinda* Benth. used in Nigerian herbal medicinal. *Journal of Biology and Research Biotechnology* 2: 34-46.

Reigosa, J., N. Pedrol and L. Gonzalez. 2006. Allelopathy: A Physiological Process with Ecological Implications. Springer, pp:639.

Mounir Tilaoui, Hassan Ait Mouse, Abdeslam Jaafari, Abdelmajid Zyad, (2015). Comparative Phytochemical Analysis of Essential Oils from Different Biological Parts of Artemisia herba alba and Their Cytotoxic Effect on Cancer Cells . Plos One | DOI:10.1371/journal.pone.0131799 1-15

Jones, G. A.; Macalliser, T. A.; Muri, A. D. and Cheng K. J. (1994).

Effects of sainfion (onobrychis and Proteolysis by four of ruminal bacteria. Applied Environmental Microbiology. 60: 1374- 1378.