

## **Original Research Article**

### **Molecular detection of *Coxiella burnetii* in stray dogs in Klang Valley, Malaysia**

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#### **Abstract**

**Aims:** *Coxiella burnetii* is a tick-borne pathogen that causes clinical disease in a number of animals including dogs. *C. burnetii* has garnered attention as a zoonotic agent. With the increasing contact between dogs and humans through ownership of pets and the growing stray dog population, it is alarming how there is still a paucity of information concerning this pathogen in stray dogs in Klang Valley, Malaysia.

**Methods:** In order to determine the rate of infection of Q fever, blood and ticks were collected from stray dogs from animal shelters and dog pounds and screened for *htpAB*-associated repetitive element of *Coxiella burnetii* using nested PCR. Chi-square test was used to establish the association or differences among/between proportions of the categories (age, breed and sex) at the significance level of  $\alpha=0.05$ .

**Results:** The method was initially applied on blood samples of which 62 (33%) of dogs were detected out of 188 total population then ticks as it is one of the transmission means but, non was found positive. However, none of the sampled ticks were found to be positive. Comparison based on age showed young dogs to have a high prevalence of 35.7% (95%CI, 0.07-0.64) than adults (32.8%, 95%CI, 0.26-0.40). According to breed, the local breed had a higher infection rate (33.2%, 95%CI, 0.26-0.40) than pedigree dogs (25.0%, 95%CI, -0.55-1.05) and based on sex, male dogs had a higher infection rate (34.3%, 95%CI, 0.21-0.42) than their female counterparts (31.3%, 95%CI, 0.25-0.44). However, there was no significant difference  $P > 0.05$  between all the three factors observed for *Coxiella burnetii*.

**Comment [s1]:** Restructure the sentence. Not very clear

26 **Conclusion:** This study found *Coxiella burnetii* present in Klang Valley, therefore  
27 necessitate action against stray dogs population to avoid shedding or contamination of the  
28 organism in environments and transmission to native animals and humans.

29 **Keywords:** blood, *Coxiella burnetii*, nested PCR, Klang Valley, stray dogs and ticks.

30

### 31 **Introduction**

32 *Coxiella burnetii* is an obligate intracellular bacterium that parasitizes eukaryotic cells and is  
33 the aetiological agent of Q fever [1]. It is an infectious zoonotic agent that has several modes  
34 of transmission and has the potential to cause significant disease in both animals and humans.  
35 It is extremely contagious and even a few organisms can lead to infection in man [2]. *C.*  
36 *burnetii* is hardy and can survive in the external environment and resist disruptions from  
37 physical and chemical substances [3]. There are a wide range of reservoirs of *C. burnetii* that  
38 include many wild and domestic mammals, birds and ticks [3]. Rodents, birds and rabbits  
39 play an important role as reservoirs [2, 4]. However, cattle, sheep and goats are the common  
40 reservoirs that pass infection to humans. Cats and less frequently dogs have also been  
41 implicated in human infections [5]. Therefore, extra care is recommended during parturition  
42 because, reactivation of infection may occur and placentas become heavily infected during  
43 pregnancy [2]. Consequently, veterinarians, animal handlers and owners are expected to  
44 proceed with precaution when handling possible reservoirs of the agent.

45 In Malaysia, there are few reports of Q fever in humans [6, 7], ruminants and farm workers  
46 [8] and pet dogs [9]. Despite increase in pet ownership in Malaysia coupled with the  
47 transmission mode of the disease, it is alarming how there is still a paucity of scientific  
48 information regarding the agent of Q fever in Malaysia. Nested PCR appeared to be highly  
49 sensitive in *C. burnetii* diagnosis [10] and was suggested to be more superior than the

50 Immuno fluorescence Assay (IFA) serological test for the primary diagnosis of Q fever  
51 because greater sensitivity achieved and being less time consuming [11]. In light of the  
52 aforementioned research information, a cross sectional study was designed focusing on stray  
53 dogs due to their closeness with human/animal environments, constant contact with ticks and  
54 lack of documented history of diagnosis and treatment of Q fever infection.

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## 56 **Methods**

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### 58 **Aim, design and area of the study**

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60 In order to detect *C. burnetii*, blood and ticks collected from stray dogs from animal shelters  
61 and dog pounds were screened for *htpAB*-associated repetitive element of *Coxiella burnetii*  
62 using nested PCR. Fisher exact test was used to establish the association or differences  
63 among/between proportions of the categories (age, breed and sex) at the significance level of  
64  $\alpha=0.05$ . A total of 188 blood and 438 ticks' samples were collected from shelters and dog  
65 pounds from different locations in Klang Valley for detection of *C. burnetii*. Of 188 sampled  
66 population, 159 dogs were sampled from Dewan Bandaraya Kuala Lumpur (DBKL), dog  
67 pounds and 29 from Society for Prevention of Cruelty in Animals (SPCA), animal shelters  
68 (Ampang and Paws Subang Jaya . Of the 438 ticks collected; 212 were males, 171 females  
69 and 55 nymphs. On average, 3-9 ticks were collected per dog and placed into properly  
70 labeled and sterile microcentrifuge tubes for further processing. Age, breed and sex of the  
71 dogs from which blood and ticks samples were collected were recorded.

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73 This study was carried out from 2013- 2014 on stray dogs in and around Klang Valley. Klang  
74 Valley is an area in Peninsular Malaysia that comprises of Kuala Lumpur and its outer

75 reaches and neighbouring cities and towns in the state of Selangor. It is situated within 3° 6'  
76 Northern latitude and 101° 39' Eastern latitude. A heartland of industry and commerce  
77 covering important areas including Kuala Lumpur International Airport [12].

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#### 79 **DNA extraction from blood and ticks**

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81 DNA from canine blood was extracted from 200 µl of whole blood using QIAamp animal  
82 Dneasy blood and tissue kit (QIAGEN® GmbH, Hilden, Germany). The protocol was  
83 followed based on manufacturer's instruction and the extracted canine DNA was stored at -  
84 20°C until further analysis. Ticks were initially identified under stereomicroscope using  
85 Kohls taxonomic classification guideline [13] and Soulsby morphological keys [14]. After  
86 identification, the ticks were washed twice with 95% ethanol followed by a final washing  
87 step with sterile distilled water. They were then crushed with certified RNase, DNase and  
88 DNA free pestle and transferred to 1.5 ml microcentrifuge tubes mixed and some were cut  
89 into pieces with a scalpel blade on glass slides then transferred to 2 ml microcentrifuge tubes.  
90 DNA was then extracted from the ticks using blackPREP Tick DNA / RNA Kit, Analytik  
91 Jena AG (Life Science, Konrad-Zuse-Strasse, Jena, Germany) and QIAamp animal Dneasy  
92 blood and tissue kit (QIAGEN® GmbH, Hilden, Germany) using manufacturer's instruction  
93 and stored at -20°C until further use.

94

#### 95 **Amplification of *Coxiella burnetii* DNA**

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97 Screening PCR for detection of *C. burnetii* in canine blood was then carried out. The first  
98 PCR amplification was performed using primers; IS111F1 and IS111R1 (Fournier and  
99 Raoult, 2003) which flank a 484-500 bp fragment of the *htpAB*-associated repetitive element

100 (Table 1). A nested PCR was carried out with primers IS111F2 and IS111R2 flanking a 260-  
101 300 bp fragment. The amplification was performed in a reagent mixture of 4.0 µl of 5x Green  
102 Go Taq buffer, 1.2 µl of 25mM Mgcl, 0.8 µl of 10mM dNTP, 1.0 µl of 20 pmol of both  
103 forward and reverse primers, 0.3 µl of 5U/µl of Taq polymerase and 5.0 µl of DNA template  
104 set up to 20 µl by adding 6.7 µl of sterile distilled water. The amplification protocol for both  
105 screening and nested PCR were the same except that the first product was diluted ten times  
106 and used as template for nested PCR. The positive control was *C. burnetii* Nine Mile strain  
107 DNA isolated from cell culture obtained from Japan. The first PCR cycling conditions  
108 consisted of an initial denaturation step at 95°C for 2 min followed by 40 cycles at 95°C for  
109 30 s, 52°C for 30 s, 72°C for 1 min and a final extension step at 72°C for 5 min. Cycling  
110 conditions for the nested PCR was carried out using the same parameters except that the  
111 number of cycles was reduced to 35 and an annealing temperature was set to 48°C. The  
112 resulting amplified products were electrophoresed on a 2.0% agarose gel at 100 V for 30 min,  
113 stained for 30-40min in ethidium bromide and subsequently visualized under a UV  
114 transilluminator.

115  
116 In order to determine whether tick DNA was successfully extracted, PCR was conducted to  
117 amplify the 28S rRNA gene of ticks. The primers (Inokuma *et al.*, 2003) (Table 1). PCR  
118 amplification was performed in a final volume of 20 µl containing 5.0 µl of DNA template,  
119 0.8 µl of 10 mM of each dNTP, 25 mM of Mgcl, 1µl of 20pmol of each primer, 0.3µl of  
120 5U/µl of GoTaq DNA polymerase in 5X Green GoTaq reaction buffer (Promega Corporation,  
121 Woods Hollow, Madison, WI, USA). The cycling conditions consisted of an initial  
122 denaturation step at 95°C for 2 min followed by 35 cycles at 95°C for 30 s, 55°C for 30 s,  
123 72°C for 30 s and a final extension step at 72°C for 5 min. The resulting amplified products

124 were electrophoresed on a 2.0% agarose gel at 100 V for 30 min, stained for 30-40 min in  
125 ethidium bromide and subsequently visualized under a UV transilluminator.

126

127 PCR was performed with tick DNA using the same reaction volume, IS111F1 and IS111R1  
128 primers for the first PCR and IS111F2 and IS111R2 set for nested PCR as described for  
129 screening of *C. burnetii* in canine blood

130 **Table 1: Sequences and target regions of oligonucleotide primers used for detection of**

131 ***Coxiella burnetii* in canine blood and tick DNA**

Primer target regions	Primer sequences 5'-3'	Amplicon sizes (bp)	References
<i>C. burnetii</i> - <i>htpAB</i> ; associated repetitive element	IS111F1-TACTGGGTGTTGATATTGC	485-500	[15]
	IS111R1-CCGTTTCATCCGCGGTG		
	IS111F2-GTAAAGTGATCTACACGA	260-300	
	IS111R2-TTAACAGCGCTTGAACGT		
Ticks -28S rRNA	28SF-GACTCTAGTCTGACTCTGTG 28SR-GCCACAAGCCAGTTATCCC	484-500	[16]

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133

134 For sequencing, PCR products prepared in total volumes of 75 µl were initially confirmed for  
135 successful amplification. After the verification, the products were then purified using  
136 Wizard® SV Gel and PCR clean-up system (Promega, USA) according to the manufacturer's  
137 instructions before sending for standard sequencing to NHKBioscience (Korean). Sequences  
138 were aligned and blasted to compare with other sequences stored in the GenBank.

139 **Data analysis**

140

141 The data collected during this study were recorded and stored in Microsoft Excel for  
142 windows. Frequency tables were used to calculate the prevalence of *C. burnetii* based on  
143 age, breed and sex of the stray dogs at the estimated confidence interval of 95%. The data  
144 were then subjected to statistical analysis using statistical package IBM SPSS statistics  
145 (version 20.0, SPSS, Inc., Chicago, IL, USA). Fisher exact test was used to establish the  
146 association or differences among/between proportions of the categories at a significance level  
147 of  $\alpha=0.05$ .

148

149 The sampled population was categorized according to age; young and adult, according to  
150 breed; local and pedigree and according to sex; male and female. Stratification of the dogs  
151 based on district or region was not possible because some of the information on these  
152 variables were not clearly recorded or could not be obtained.

153

154 **Results**

155

156 **Descriptive statistics and prevalence of *Coxiella burnetii* in stray dogs**

157

158 Of the total 188 dogs sampled, 105 were males while 83 were females (Table 2).  
159 In terms of sources, a total 159 dogs were sampled from Dewan Bandaraya Kuala Lumpur  
160 (DBKL) dog pound and 29 dogs were sampled from Animal shelters (SPCA, Ampang and  
161 Paws, Subang Jaya). *C. burnetii* was detected in 62 / 188 (33.0%) of the dogs sampled with  
162 GenBank, accession number KU215930. Comparison based on age showed young dogs had a  
163 relatively higher prevalence of 35.7% than adults (32.8%). According to breed, the local

164 breed had a relatively higher infection rate (33.2%) and based on sex, male dogs had a  
 165 relatively higher infection rate (34.3%) than their female counterparts (31.3%). However, it  
 166 should be noted that differences between all the three factors age, breed and sex of dogs for  
 167 the presence of *C. burnetii* were not statistically significant  $P > 0.05$  ( Table 3).

168  
 169  
 170 **Table 2. Proportion of the total dogs sampled and that of tick infested dogs that had**  
 171 ***Coxiella burnetii* infection according to age, breed and sex**

Variables	Category	Sample Proportion n=188 (%)	Tick positive dogs n= 141 (%)	Coxiella positive tick infested dogs. n=141(%)
Age	Young	14 (7.4)	10 (7.1)	1 (0.1)
	Adult	174 (92.6)	131 (92.9)	42 (29.9)
Breed	Local	184 (97.9)	139 (98.6)	43 (30.0)
	Pedigree/mixed	4 (2.1)	1 (1.4)	0 (0.0)
Sex	Male	105 (55.9)	82 (58.2)	26 (18.0)
	Female	83 (44.1)	59 (41.8)	17 (12.0)

173  
 174  
 175  
 176 **Table 3: Prevalence of *Coxiella burnetii* amongst the total dog population sampled**  
 177 **according to age, breed and sex**

Variables	Category	Prevalence (%)	Standard dev. ±	95% CI	P-value
Age	Young	35.7	0.50	0.07- 0.64	0.777
	Adult	32.8	0.47	0.26- 0.40	-
Sex	Male	34.3	0.47	0.21-0.42	0.755
	Female	31.3	0.48	0.25- 0.44	-



Breed	Local	33.2	0.47	0.26-0.40	1.000
	Pedigree	25.0	0.50	-0.55-1.05	-

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179

180 **Discussion**

181

182 This study to the best of our knowledge is the first to report *C. burnetii* in canine blood in  
183 Malaysia with a prevalence of 33.0% (62/188) in stray dogs in Klang Valley. There are  
184 previous reports of Q fever in humans [6, 17] as well as a recent report in ruminants and farm  
185 workers [8] in Malaysia, but no reports thus far on stray dogs. A most recent report detected a  
186 Coxiella-like bacteria (89%) in ticks recovered from wild boars, single porcupine and goats  
187 (Khoo et al., 2016). Another high prevalence (59.0%) of the pathogens in ticks was also  
188 reported from pet dogs visiting University Veterinary Hospital (UVH), University Putra  
189 Malaysia (UPM) [9]. This poses a risk of sharing of infection between dogs and owners [9].  
190 The implication of our findings is that stray dogs may play a greater role in dissemination of  
191 the pathogen and may serve as a source of infection to humans and other naive dogs. The  
192 recent report of the pathogen detection in pets [9], together with the present study on stray  
193 dogs reiterates the existence and persistence of *C. burnetii* in dogs in Klang Valley, Malaysia.

194

195 Since the data used in this study is limited, information on prevalence of *C. burnetii* in the  
196 study area in terms of the spatial and temporal distribution of the pathogen cannot be  
197 explored as some variables were too scanty and most samples were obtained from Dewan  
198 Bandaraya Kuala Lumpur (DBKL), thus the limitations of this study.

199

200 The primers used for amplification of the *htpAB*-associated repetitive element was reported  
201 sensitive and specific for detection of *Coxiella* DNA [15]. It should be noted that the first  
202 PCR with the outer primers; IS111F1 and IS111R1 could not amplify the *C. burnetii* DNA in  
203 blood, however when a nested PCR was carried out with the inner primers, IS111F2 and  
204 IS111R2, positive results were obtained from the canine blood. Despite the reported  
205 sensitivity of the primers, nested PCR on all the tick DNA was negative for *C. burnetii* and  
206 screening test was carried after running the success of tick DNA extraction. This nested  
207 approach is adequate for detection because it is highly conserved with a unique component  
208 common to a variety of *C. burnetii* strains, thus able to amplify a small bacterial load in blood  
209 and milk [9].

210  
211 Adult 57/62 (91.9%) stray dogs in Klang Valley had a relatively higher infection ratio  
212 compared to the younger dogs 5/62 (8.1%) as shown in Table 2. This may result from  
213 increase in the risk of abortion, still birth or death of puppies during pregnancy or after  
214 whelping related to *C. burnetii* infection in some bitches [5]. However, adult dogs were  
215 over represented and the difference was not statistically significant  $P > 0.05$ . Comparison  
216 within breed showed the local breed had a relatively higher infection rate (33.2%) than  
217 pedigree (25%) even though the difference was not statistically significant. The greater part  
218 of the local canine population in the country consists of stray dogs and owned [18] local  
219 breeds, thus explaining the results. According to sex, male dogs appeared to have a high  
220 prevalence (34.3%) compared to the females (31.3%), but the difference was not statistically  
221 significant (Table 3). As the difference between all variables was not statistically significant,  
222 it can be assumed that age, sex and breed do not contribute or affect *C. burnetii* infection  
223 rates.

224

225 **Conclusion**

226 In conclusion *C. burnetii*, the aetiological agent of Q-fever was found to be present in blood  
227 samples obtained from stray dogs in Klang Valley, Malaysia. Ticks collected from dogs were  
228 all negative suggestive that ticks may not necessarily be the source of infection of *C. burnetii*  
229 to the stray dogs sampled in the study. Rather, the dogs may likely be the source of infection  
230 to ticks during blood meal up-take. The relatively high prevalence rate of *C. burnetii* in dogs  
231 sampled is worrisome in light of the public health risk and therefore warrants further  
232 investigation and control as most of the infected dogs were sampled from Dewan Bandaraya  
233 Kuala Lumpur and are mainly dogs that are caught roaming around the city.

234

235 **Abbreviations**

236 IFA= Immuno Fluorescence Assay

237 UPM= Universiti Putra Malaysia

238 UVH= Universiti Veterinary Hospital

239 DBKL= Dewan Bandaraya Kuala Lumpur

240 SPCA= Society for Prevention of Cruelty in Animals

241

242 **Declarations**

243

244 **Ethics approval** : This research was conducted following approval from the Animal Care  
245 and Use Committee, Faculty of Veterinary Medicine, Universiti Putra Malaysia (UPM),  
246 which was an adopted version of the Australian Code of Practice for the Care and Use of  
247 Animals for Scientific Purposes. (The research grant No. 01-01-09-0662RU. Informed

248 consent and approval was obtained from the International care and use committee, UPM  
249 (approval code RD74/2013)).

250

251 **Consent of publication:** Not applicable.

252

253 **Availability of data and material:** GenBank, accession number is KU215930.

254

255 **Competing interests:** All authors approved and declared that they have no competing  
256 interest.

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