Original Research Article

Validated stability indicating HPTLC, UHPLC and UV-spectrophotometric techniques for the determination of bepotastine besilate in presence of its oxidative degradate

Abstract

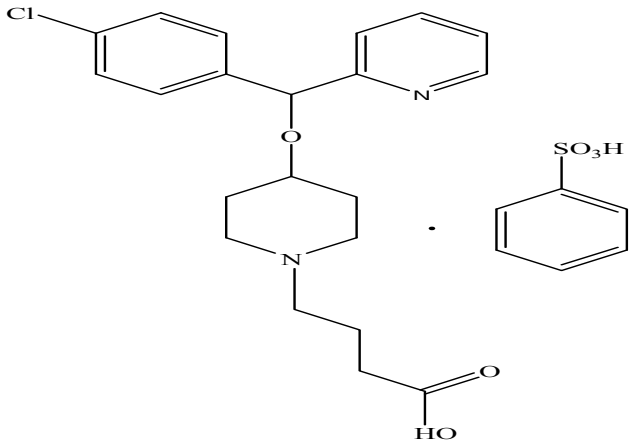
The study is aimed at developing methods which have a complete validation as stipulated in the ICH guidelines and to be applied for the determination of Bepotastine besilate (BB) in pure form and in pharmaceutical formulations in the presence of its oxidative degradation product. High performance thin layer chromatography (HPTLC), Ultra high performance liquid chromatography (UHPLC) and different spectrophotometric methods (first derivative, first derivative of ratio spectra and ratio difference are developed for simultaneous determination of bepotastine besilate in laboratory-prepared mixtures of bepotastine besilate with its oxidative degradate and in pharmaceutical formulations were used in the study design. Firstly, HPTLC was performed and separation occurred on silica gel 60 F254 plates, with butanol: ammonia (8:2, v/v) as a developing system. UHPLC in which separation occurred on a Kinetex C 18 column using methanol- 0.1% O-phosphoric acid - acetonitrile (70:20:10, by volume) as mobile phase, followed. And lastly was UV/Vis spectrophotometry which included first derivative determination of the drug at 252.6 nm, first derivative of ratio of peak amplitudes at 233.4, 250 and 275.6 nm and the ratio difference with the amplitude difference between (240 nm and 260 nm). Result showed that HPTLC method was applicable over the concentration range of 0.5-5 µg / band, while UHPLC method was linear over the concentration 2- 12 µg/mL and spectrophotometric methods were linear over the concentration range 20-120 µg/mL for bepotastine besilate. The proposed three techniques are quite accurate and precise. They can be used for routine analysis of bepotastine besilate in pharmaceutical formulation and stability indicating methods.

Keywords: Bepotastine besilate; UHPLC; HPLC; UV-spectrophotometry; oxidative degradate; Stability

1. Introduction

Bepotastine besilate (Bepotastine-B) (Structure 1) is second generation antihistamine and is chemically benzenesulfonic acid; 4-[4-[(S)-(4-chlorophenyl)- pyridin-2-ylmethoxy]piperidin-1-yl] butanoic acid[1]. It is soluble in acetonitrile and methanol and was approved in Japan for use in the treatment of allergic rhinitis and urticaria/pruritus. It is a direct H1 receptor

antagonist that inhibits the release of histamine from mast cells[2]. Literature survey revealed that few methods have been reported for its determination in pharmaceutical formulations and biological samples including RP-HPLC techniques[3,4] and LC-MS/MS [5,6]. LC-MS/MS method was used for determination of the drug in human plasma and urine where the sample was prepared by solid phase extraction using mobile phase of acetonitrile: water: 200mmole/L ammonium formate (75:20:5) at flow rate 0.3 mL/min, 30°C and electrospray ionization mass spectrometry (MRM) detection. Additionally stability indicating HPTLC[6] was used in the determination of Bepotastine-B in presence of its acid degradate using chloroform: methanol (5: 5) as mobile phase where Bepotastine-B and acid degradate were obtained at Rf 0.50 and 0.78, respectively with UV detection at 225 nm. The acid degradate was undescribed in details. None of these methods is concerned with the identification and elucidation of the structure of the resulting oxidative degradation product, thus the objective of the present study was to develop simple and accurate stability -indicating methods for selective determination of Bepotastine-B in the presence of its oxidative degradation product.



Structure (1): Chemical structure of bepotastine besilate

2. Experimental

2.1 Instrumentation

The UHPLC system used was an Agilent 1100 UPLC with binary pump and UV detector. Analysis was performed on a Kinetex C 18 column (100 mm, 4.6 mm i.d., 2.6 µm). Torrance, USA. Merck TLC plates 20 x 20 cm precoated with silicagel 60 F 254 (Fluka, Chemie, Switzerland) were used. A camag Linomate 5 sample applicator equipped with a 100 µL syringe (Hamilton,Germany) 20 x 20 cm twin through glass chamber (Camag). The

plates were scanned with a camag TLC scanner 3 with WINCATS computer software (Switzerland) using UV lamp with short wavelength (254 nm) (Desega- Germany). Shimadzu UV/Vis spectrophotometer (PC – 1601, Tokyo, Japan), using 1.0 cm quartz cells. Scans were carried out in the range of 200–400 nm at 0.5 nm intervals. The spectra were automatically obtained by Shimadzu UV-Probe 2.32 system software.

2.2. Chemicals

Standard Bepotastine-B(99.80%, purity) was kindly supplied by RAMEDA Co., Giza, Egypt, Bepogra® tablets market samples; B.N. (J701271) labelled, to contain 10 mg Bepotastine -B (Lupin LTD, Jammu, India). Solvents: hydrogen peroxide (30%), Butanol and ammonia were obtained from El-Nasr Co., Qalyubia, Egypt, methanol and O-phosphoric acid were obtained from Sigma Aldrich (Schnelldorf, Germany) while acetonitrile HPLC grade was obtained from Fisher (Loughborough, UK).

2.2.1 Preparation of Standard solutions

Stock solution of Bepotastine-B (1 mg/mL) was prepared in methanol. Working solutions were freshly prepared by suitable dilution of stock solution with methanol to obtain a concentration of 0.1 mg/mL drug.

2.2.2 Preparation of degradation product

Accurately weighed 100.0 mg of intact Bepotastine-B was dissolved in 25 mL 10% H_2O_2 and kept at room temperature for two days then allowed to dry at room temperature. Residue was extracted with methanol, applied as a band on preparative TLC plate and developed in the mobile phase. The band corresponding to oxidation product was scratched, dissolved in methanol and evaporated. The isolated oxidative degradate solid was used to prepare the stock solution and identified by mass spectral analysis.

2.3. Validation procedures

2.3.1. Linearity

HPTLC method: Different volumes of stock standard solution (1 mg/mL) containing 0.5-5.0 mg Bepotastine-B were introduced into a series of 10- mL volumetric flasks, and then volume was filled to mark with methanol. Ten (10) µL from each solution was applied to the TLC plates of silica gel 60 F254 plates. The plates were washed with the mobile phase consisting of butanol: 33% ammonia (8:2, v/v). The pre-washed activated plates, of 6-mm bands, 6 mm apart, were sprayed by means of a Camag Linomat IV automated spray-on band applicator equipped with a 100-µL syringe. The plates were developed with the mobile phase in a Camag twin-trough chamber previously saturated with mobile phase vapour for 20 min.

After development, the plates were removed and air dried. Densitometry was performed at 266 nm in reflectance mode with slit dimensions of 6.00 mm × 0.3 mm and scanning speed of 20 mm/s. Peak area of each concentration was then plotted against its corresponding drug concentration and regression equation was computed.

UHPLC method: Aliquots of working standard drug solution (0.1 mg /mL in methanol) containing 0.02-0.12mg drug were introduced into series of 10- ml volumetric flasks and adjusted to the volume with methanol. Triplicate 10µL injections were made for each concentration on a C18 column followed by elution with a mobile phase of methanol- 0.1% O-phosphoric acid - acetonitrile (70:20:10, by volume) at a flow rate of 1 mL/ min and UV wave length of 260 nm was used for detection. The Peak area of each concentration was then plotted against the corresponding drug concentration and regression equation was obtained.

Spectrophotometric methods: Aliquots equivalent to 0.2-1.20 mg/mL Bepotastine-B were accurately transferred from its standard solution (0.1 mg/mL) into a series of 10 -mL volumetric flasks then completed to volume with methanol. The spectra of the prepared solutions were scanned from 200 - 400 nm and stored in the computer.

First derivative (1D) method: 1D spectra of both Bepotastine-B and its oxidative degradate were recorded against methanol as blank, where the zero crossing wavelengths at 252.6 nm was selected for the analysis of Bepotastine-B in presence of its oxidative degradate. The calibration curve was constructed and the regression equation was calculated.

Derivative ratio (1DR) method: The stored spectra of Bepotastine-B was divided by the spectrum of 40µg/mL of its oxidative degradate. Then the first derivative of the ratio spectra (1DR) with scaling factor 10 is obtained and smoothed with $\Delta\lambda$ = 8 nm. 1DR signals at 233.4nm, 250nm and 275.6 were measured and then plotted against their corresponding concentration from which regression equations were determined.

Ratio difference (RD) method: The above procedure detailed under derivative ratio (1DR) method was followed and after the division of the stored spectra of Bepotastine-B by the spectrum of 40µg/mL oxidative degradate, the amplitude difference between 240 nm and 260 nm for spectrum was plotted against its corresponding drug concentration and the regression equation was evaluated.

2.3.2. Application to laboratory prepared mixtures

HPTLC method: Aliquots equivalent to 4.5-0.5 mg of Bepotastine-B from its stock standard solution (1 mg/mL) were transferred into series of 10 ml volumetric flasks containing

aliquots of its oxidative degradate derived from 0.5-4.5 mg intact drug and then diluted to the volume with methanol.

UHPLC method: Aliquots equivalent to 0.09-0.01 mg of Bepotastine-B from its working standard solution (0.1 mg /mL) were transferred into series of 10 mL volumetric flasks containing aliquots of its oxidative degradate derived from 0.01-0.09 mg intact drug and then diluted to the volume mark with methanol.

Spectrophotometric method: Aliquots equivalent to 0.9-0.1 mg of Bepotastine-B from its working standard solution (0.1 mg /mL) were transferred into series of 10 ml volumetric flasks containing aliquots of its oxidative degradate derived from 0.1-0.9 mg intact drug of degraded solution and then diluted to the volume mark with methanol.

2.3.3. Application to pharmaceutical formulation

Ten Bepogra® tablets were accurately weighed and finely powdered. A weight equivalent to 100mg of Bepotastine-B was sonicated with about 30 mL of methanol in 100 ml volumetric flask for 20 minutes then the volume was completed with methanol to obtain a solution labeled to contain 1mg/mL Bepotastine-B. The solution was filtered and used to analyze the drug by HPTLC as detailed under linearity. The filtrate was diluted with methanol to obtain solution labeled to contain 0.1 mg/mL Bepotastine-B to be analyzed by UHPLC and UV-spectrophotometry as described under linearity.

3. Results and Discussion

3.1. Forced degradation

Stability of Bepotastine-B was studied under different stressed conditions. It was found that it was stable to acidic and alkaline hydrolysis upon refluxing with 5N HCL and 5N NaOH for 6 h while it is liable to oxidative degradation upon keeping with 10% H_2O_2 at room temperature for two days. The obtained oxidative degradation product was separated on preparative TLC plates using butanol: 33% ammonia (8:2, v/v) as a developing system and its structure was identified by mass spectroscopy. Where its oxidative degradate showed main molecular ion peak at 581.45 m/z, whereas intact Bepotastine-B molecular ion at 549.45m/z, which are equivalent to their molecular weights this indicate increasing in molecular ion peak equal 32 unit, Fig. 1. This suggested the addition of two oxygen to each nitrogen in both piperdine and pyridine rings. Thus, a proposed pathway for this oxidative degradation was

illustrated in Scheme (1). Fragmentation of intact Bepotastine-B was also illustrated in Scheme (2).

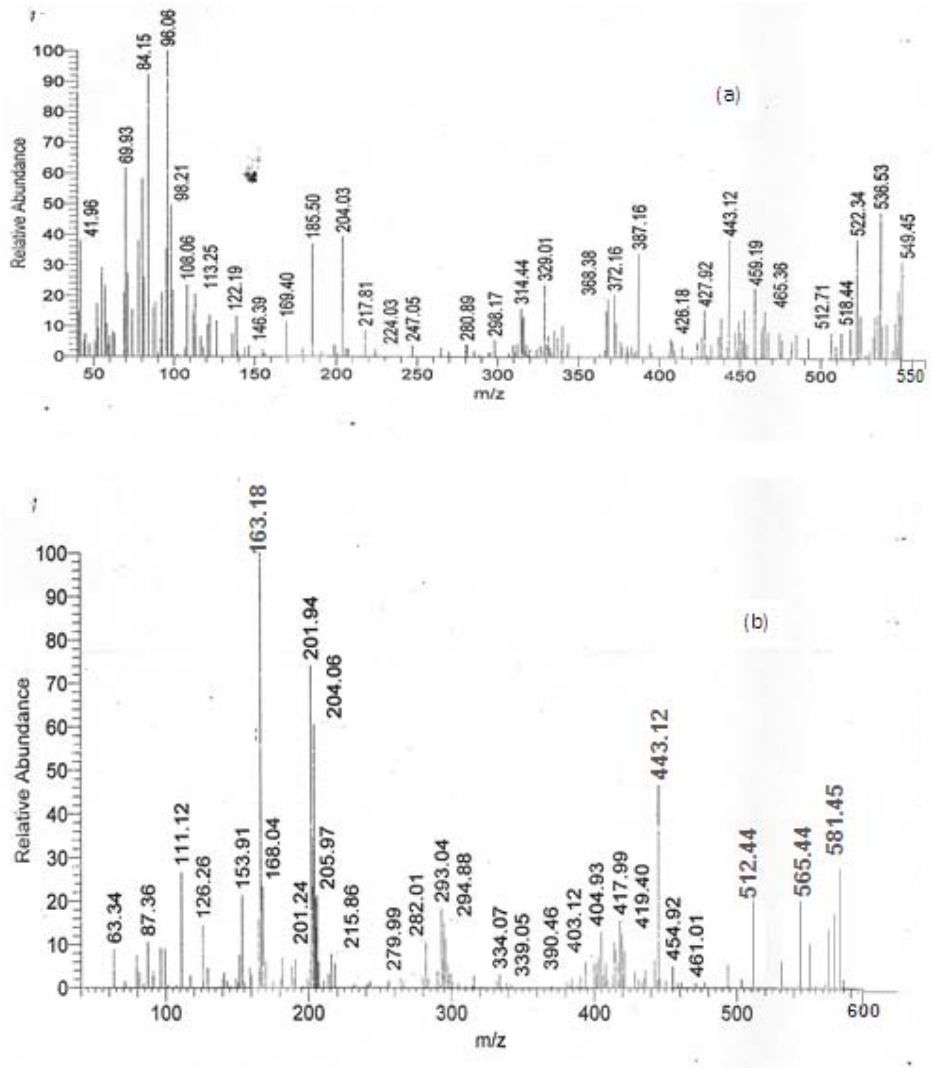
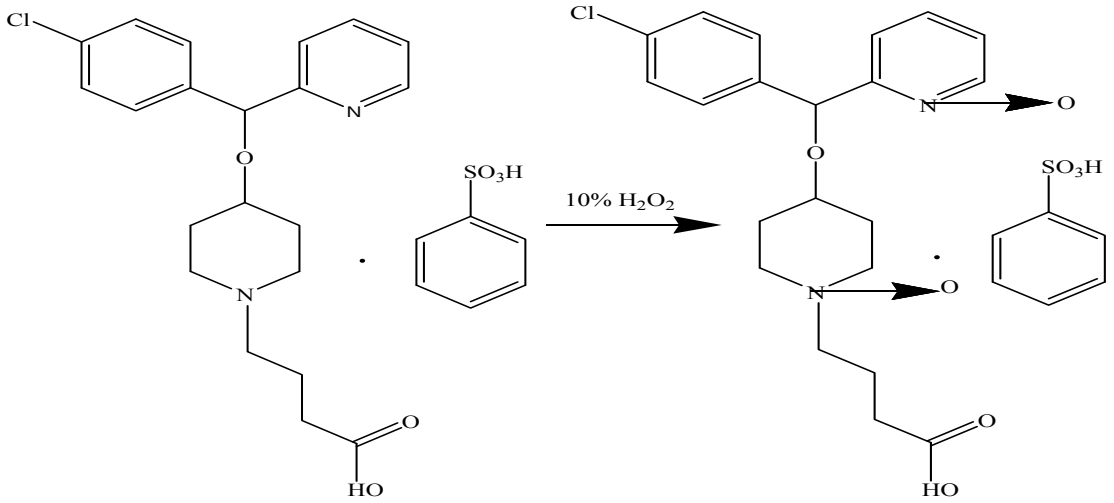
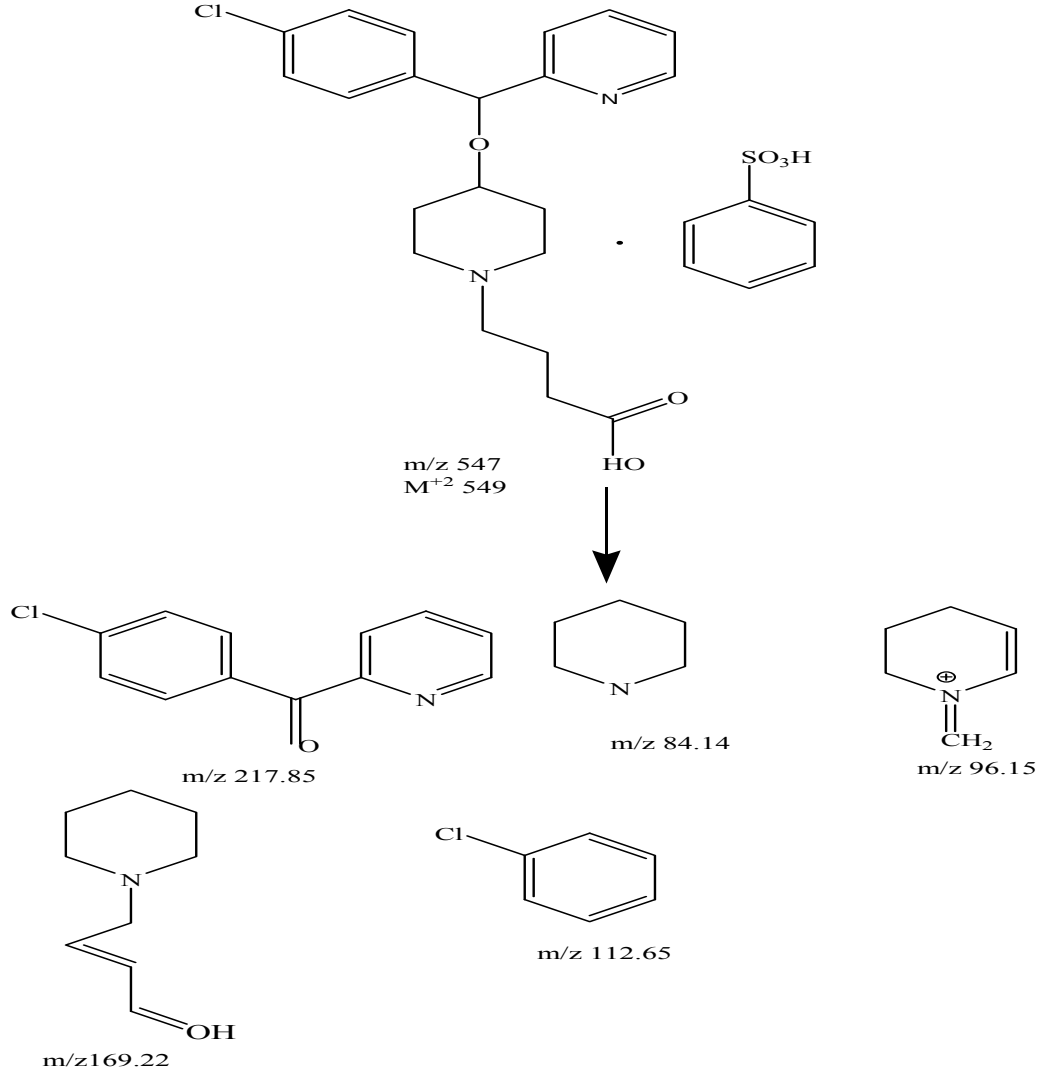


Fig. (1): Mass spectra of (a) intact Bepotastin besilate and (b) its oxidative degradate**.**

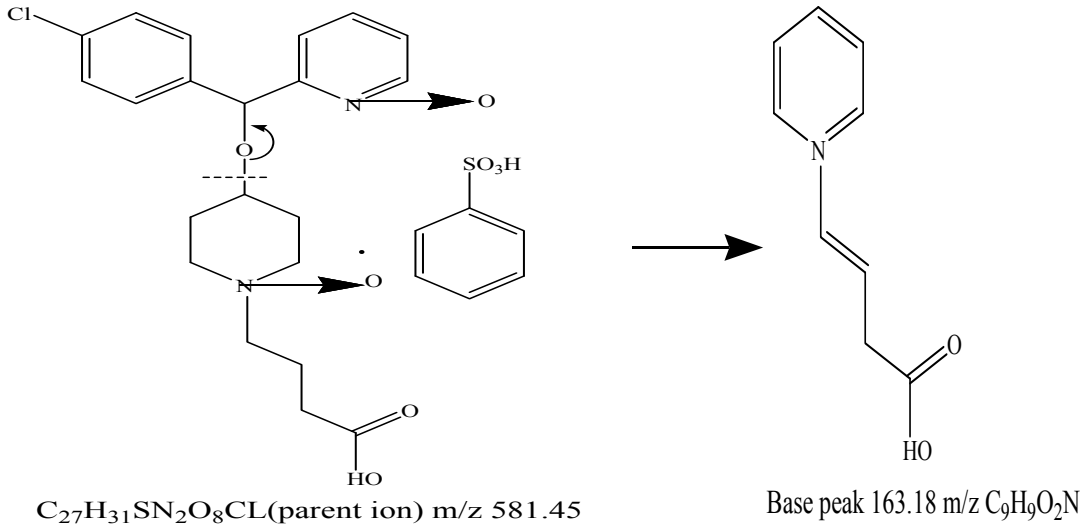


Scheme(1): Suggested oxidative degradation pathway of Bepotastin besilate



Scheme(2): Fragmentation of intact Bepotastin besilate

The drug does not degradate upon treatment with H_2O_2 but it undergoes oxidation of both nitrogen atoms due to presence of molecular ion peak (parent ion) at 581.45 m/z corresponding to its molecular weight. However when the vaporized drug passes into ionization champer of mass spectrum it is bombarded by a stream of electrons which break it to smaller fragments. The base peak 163.18 m/z may be due to fragmentation of the parent ion to give the most stable ion at 163.18 m/z which has molecular formula $C_9H_9O_2N$. This fragmentation was illustrated in Scheme (3) where piperidine ring stabilize itself to more stable pyridine ring.



$C_{27}H_{31}SN_2O_8CL$(parent ion) m/z 581.45

Base peak 163.18 m/z $C_9H_9O_2N$

Scheme(3): Fragmentation of Bepotastin besilate oxidative degradate

3.2. HPTLC method

Different mobile phases in different ratios and at different λmax for detection were tried. It was found that butanol: ammonia 33% (8:2, v/v) as a developing system followed by densitometric determination at 266 nm, Fig.(2). offered the best separation and resolution where R_f were 0.35 and 0.26 for Bepotastine-B and its oxidative degradate, were obtained respectively.

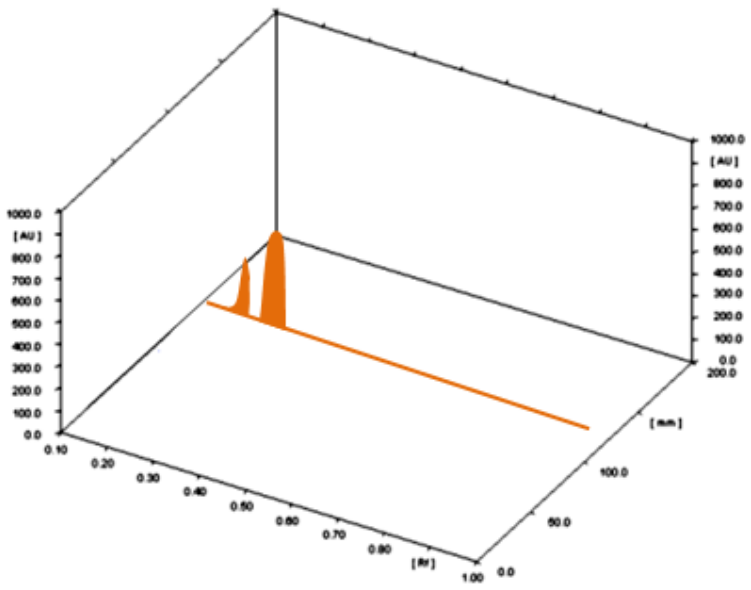


Fig. 2: Densitogram of bepotastine-besilate (2.5 µg/ mL) and its oxidative degraded derived from intact drug (2.5 µg/ mL).

3.3. UHPLC method

Ultra Performance Liquid Chromatography (UPLC) is a new category of separation technique based upon well-established principles of liquid chromatography, which utilizes sub-2µm particles for stationary phase. These particles operate at elevated mobile phase linear velocities to affect dramatic increase in resolution, sensitivity and speed of analysis. Owing to its speed and sensitivity, this technique is gaining considerable attention in recent years for pharmaceutical and biomedical analysis[7]. Chromatographic separation of Bepotastine-B and its oxidative degradate using UHPLC were optimized. Different mobile phases in different ratios were studied, where best peak shape and adequate separation was obtained by using methanol- 0.1% O-phosphoric acid – acetonitrile (70:20:10, by volume). Different flow rates and wavelengths were tried; good resolution with sensitive detector response was obtained at 260 nm using a flow rate of 1 mL min^{-1} to obtain a retention time 1.518 min for Bepotastine-B and 2.773 min for its oxidative degradate, respectively, Fig.(3).

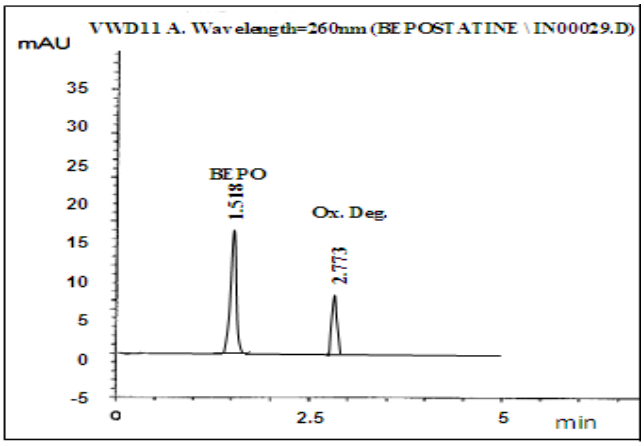


Fig. 3: UHPLC chromatogram of bepotastine-besilate (6 µg/ mL) and its oxidative degradate derived from intact drug (4 µg/ mL).

3.4. Spectrophotometric method

The zero-order absorption spectra of Bepotastine-B and its oxidative degradate showed severe overlapping over the entire spectrum of the intact drug; Fig. (4a). Therefore, the use of direct absorbance measurements for assaying Bepotastine-B in the presence of its degradation product was not possible.

First-derivative (1D) method: It was found that the above overlapping were somewhat resolved in 1D spectra and Bepotastine-B can be determined at 252.6 nm, at which its degradate has no contribution (zero crossing); Fig. (4b).

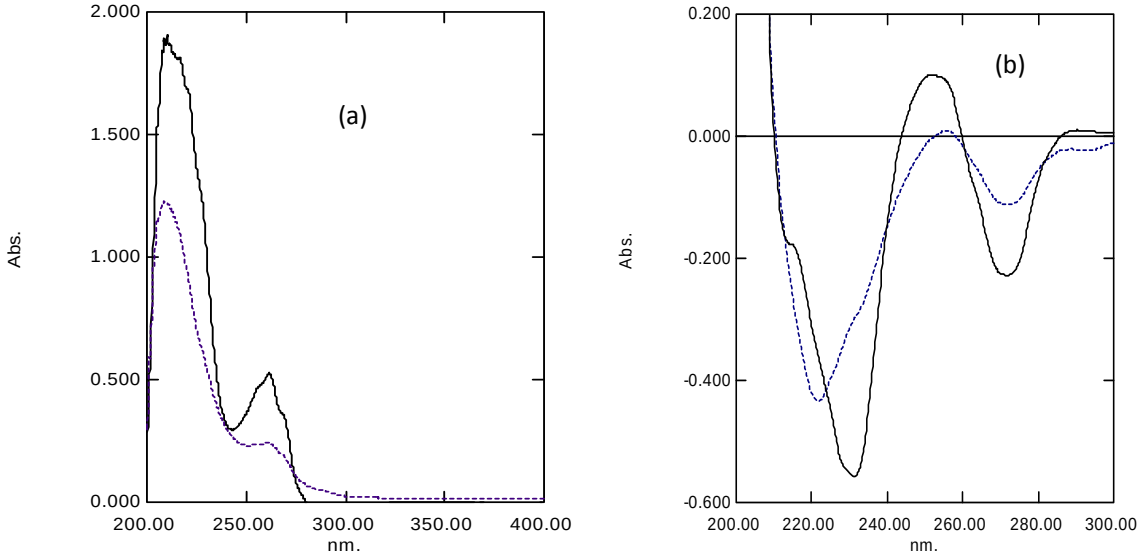


Fig. 4:(a) zero order spectra and (b) first order derivative spectra of 40 ug mL^{-1} Bebotastine besilate (–) and its oxidative degradate derived from 40 ug mL^{-1} intact drug (- - -).

Derivative ratio (1 DR) method- In this method the absorption spectrum of Bepotastine-B is divided by the absorption spectrum of oxidative degradate and the first derivative of the ratio spectrum is obtained. Different concentrations of oxidative degradate were tried as a divisor (20-120 µg/ml) and a divisor concentration of oxidative degradate derived from 40 µg/ml Bepotastine-B was the best regarding average % recovery. At wavelengths 233.4, 250 and 275.6 nm using $\Delta\lambda$ = 8 nm and scaling factor 10, good linearity and recovery percent were observed; Fig. (5b). Consequently, the peaks amplitudes of 1DR were then recorded at 233.4, 250 and 275.6 nm.

Ratio difference (RD) method- This method comprises two critical steps. The first is the choice of the divisor and the selected divisor should compromise between minimal noise and maximum sensitivity. The second is the choice of the wavelengths at which measurements are recorded [8]. Linear correlation was obtained between the differences in amplitudes between 240 and 260 nm against the corresponding concentration of Bepotastine-B using 40 µg/ml of oxidative degradate as divisor; Fig.(5a).

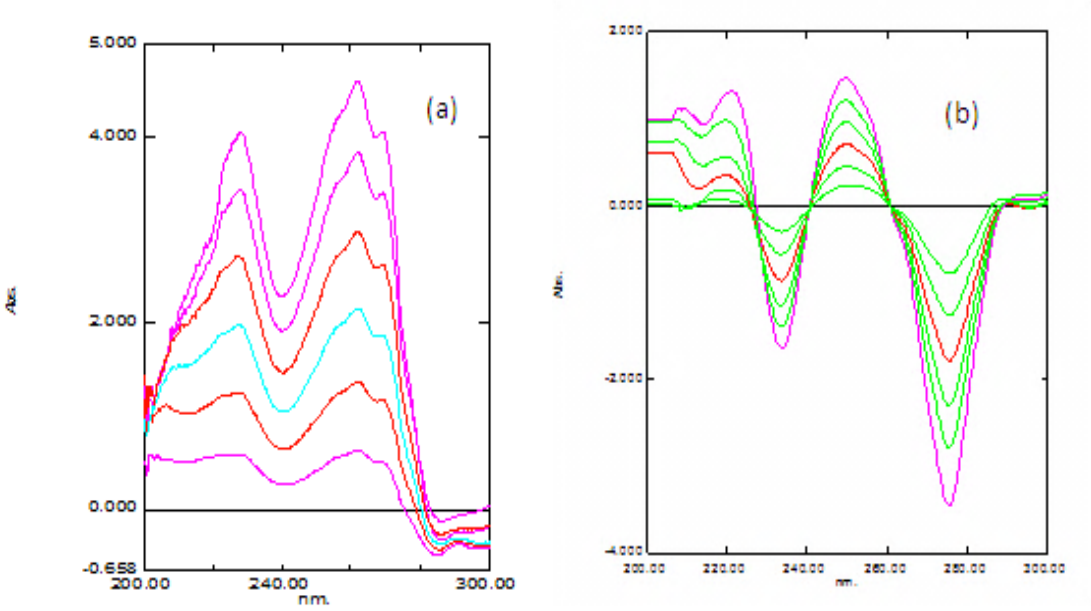


Fig. 5: (a) Ratio spectra and (b) First derivative of smoothed ratio spectra of Bebotastine besilate (20-120µg/mL) using 40 µg/mL oxidative degradate as divisor

1.1. Method Validation

System suitability: System suitability test was performed in accordance with USP[9] to ensure system performance before or during the drug analysis. Results shown in Table 1 indicated adequate resolution**.**

Table 1: System suitability results of the UPLC method

Parameter	Bepotastine-B	Reference value
Number of theoretical plates (N)	6711	The higher the value, the more efficient the column is
Resolution factor	17.19	>2
Capacity factor (K)	3.52	1–10
Selectivity factor	8.20	≥1

Linearity: Under the described experimental conditions, linear calibration curves between peak areas to respective drug concentrations were obtained through the concentration ranges of 0.5 -5 µg/ spot and 2-12 µg/ mL and 20-120 µg/mL of Bepotastine-B using HPTLC, UHPLC and spectrophotometric methods, respectively. Regression parameters were computed, where values of r^2 ranged between 0.9993 and 0.9999 indicating good linearity.

Accuracy and precision: Accuracy calculated as (R%) ranged from 99.67 to 100.58% for Bepotastine-B. Intraday precision RSD % ranged from 0.15 to 1.84%, while inter day precision ranged from 0.19 to 1.93%; indicating good repeatability and reproducibility of the proposed methods, Tables 2**.**

Selectivity: The selectivity of the proposed methods was assured by application to laboratory prepared mixtures of the intact drug together with its oxidative degradate. They were successfully applied for the determination of Bepotastine-B in the presence of up to 70% of its oxidative degradate for UHPLC and HPTLC methods and up to 60% for spectrophotometric methods with mean % recovery of 99.52±0.65 to 101.48±1.63 , Table 3.

1.2. Application to pharmaceutical formulation

The proposed techniques were successfully applied for analysis of Bepotastine-B in the Bepogra® Tablets with mean % recovery of 99.11±0.57 to 100.90±1.34. The validity of the proposed method was further assessed by applying the standard addition technique. The results obtained were reproducible with acceptable SD, Table (4). Statistical analysis of the results obtained by the proposed methods compared with a reported HPTLC method[6]showed that the calculated t and F values were less than the tabulated ones indicating no significant difference between them confirming accuracy and precision at 95% confidence limit, Tables (5) where the theoretical t- and f- values at p= 0.05 are 2.31 and 6.39, respectively and

reported method[6] involved stability indicating HPTLC method for estimation of Bepotastine-B. separation was carried out on Merck TLC aluminium sheets precoated with silica gel 60F254 using mobile phase as chloroform : methanol. (5:5 v/ v) at 225nm.

The two chromatographic methods were more sensitive, less time and solvent consuming, UV-spectrophotometric methods are cheap and easily available instrument. Therefore, should be cost-effective for routine analysis in the pharmaceutical industry.

Table 2: Regression and validation parameters for the determination of Bepotastine-Besilate by the HPTLC and UHPLC methods

	HPTLC	**UHPLC**
$\lambda_{max\ (nm)}$	266	260
Linearity range	0.5-5 µg/ spot	2-12µg/ mL
Regression parameters		
Slope (b)± SD	3004.5±12.476	25.784±0.0004
Intercept (a)± SD	115.73±37.858	0.004±0.003
Correlation coefficient (r^2)	0.9999	0.9999
Accuracy (R %)(n=9)	99.86	99.95
Precision (RSD %)		
Intra day	0.31-0.99	0.15-0.33
Inter day (n=9)	0.57-1.47	0.19-0.62

Table 3: Regression and validation parameters for the determination of Bepotastine-Besilate by the UV- spectrophotometric methods

	(1D) First-derivative method	**1DR method**			**RD method**
$\lambda_{max\ (nm)}$	252.6	233.4	250	275.6	240-260
Linearity range			20-120µg/ mL		
Regression parameters					
Slope (b)± SD	0.003±2.67E-05	0.014±0.0002	0.012±0.0001	0.026±0.024	0.018±0.0003
Intercept (a)± SD	-0.004±0.002	0.027±0.013	-0.033±0.011	0.208±0.0003	-0.03±0.02
Correlation coefficient (r^2)	0.9996	0.9994	0.9995	0.9995	0.9993
Accuracy (R %)(n=9)	99.69	99.85	100.15	100.58	99.67
Precision (RSD %)					
Intra day	0.95-1.84	0.44-0.92	0.93-1.56	0.32-0.64	0.27-0.49
Inter day (n=9)	1.42-1.93	0.74-1.78	1.25-1.84	0.60-1.30	0.65-1.19

Table 3: Determination of Bepotastine-Besilate in laboratory prepared mixtures with its oxidative degradate by the proposed methods.

Method		Intact µg mL-1	Degradate µg mL-1	%Degradation	Recovery % of intact	Average recovery (mean ± SD)
HPTLC		4.5	0.5	10	101.21	100.09±0.97
		4	1	20	100.97	
		3.5	1.5	30	100.65	
		3	2	40	100.12	
		2.5	2.5	50	99.67	
		2	3	60	99.62	
		1.5	3.5	70	98.39	
UHPLC		9	1	10	100.24	99.40±1.00
		8	2	20	100.23	
		7	3	30	100.08	
		6	4	40	99.59	
		5	5	50	99.22	
		4	6	60	99.03	
		3	7	70	97.41	
(1D) First-derivative method		90	10	10	99.30	101.19±1.22
		80	20	20	100.14	
		70	30	30	101.75	
		60	40	40	101.42	
		50	50	50	102.44	
		40	60	60	102.13	
1DR method	At 233.4 nm	90	10	10	100.90	101.38±0.67
		80	20	20	100.93	
		70	30	30	100.96	
		60	40	40	101.12	
		50	50	50	101.78	
		40	60	60	102.59	
	At 250 nm	90	10	10	99.39	100.84±1.63
		80	20	20	99.44	
		70	30	30	99.63	
		60	40	40	101.45	
		50	50	50	101.74	
		40	60	60	103.39	
	At 275.6 nm	90	10	10	100.37	101.70±1.11
		80	20	20	100.51	
		70	30	30	101.67	
		60	40	40	101.83	
		50	50	50	102.80	
		40	60	60	103.02	
RD method		90	10	10	99.74	100.69±0.91
		80	20	20	100.33	
		70	30	30	100.13	
		60	40	40	100.61	
		50	50	50	101.06	
		40	60	60	102.28	

Table 4: Application of standard addition technique for the determination of Bepotastine-Besilate in its pharmaceutical formulation by the proposed methods.

Method	Pharmaceutical formulations	Taken (ug mL^{-1})	Added (ug mL^{-1})	% recovery	Mean%±SD
HPTLC	99.57±0.57	2	0.5 1 2 3	98.21 100.00 101.60 101.06	100.22±1.49
UHPLC	99.91±0.69	4	2 4 6 8	101.51 100.50 100.38 100.47	100.72±0.53
(1D) First-derivative method	99.11±0.92	40	20 40 60 80	100.00 99.08 100.00 100.46	99.89±0.58
1DR method At 233.4 nm	100.52±1.34	40	20 40 60 80	101.10 102.03 100.86 99.55	100.89±1.02
1DR method At 250 nm	100.90±0.99	40	20 40 60 80	100.38 98.58 99.86 101.51	100.08±1.22
1DR method At 275.6 nm	100.02±1.09	40	20 40 60 80	98.30 98.38 98.22 101.15	99.01±1.43
RD method	100.19±0.82	40	20 40 60 80	100.81 100.54 101.45 101.77	101.14±0.57

Table 5: Statistical analysis of the proposed analytical methods and reported method for the determination of Bepotastine-Besilate in Bepogra® tablets

Parameter	HPTLC	UHPLC	(1D) First-derivative method	1DR method 233.4 (nm)	1DR method 250 (nm)	1DR method 275.6 (nm)	RD method	Reported method
Linearity	0.5-5 (µg/spot)	2-12 (µg/mL)	20-120 (µg/ mL)					5-25 (µg/ spot)
N	5	5	5	5	5	5	5	5
Mean%	99.57	99.91	99.11	100.52	100.90	100.02	100.19	99.90
±SD	±0.57	±0.69	±0.92	±1.34	±0.99	±1.09	±0.82	±0.57
Variance	0.32	0.48	0.85	1.80	0.98	1.19	0.67	0.32
t-	0.92	0.03	1.63	0.96	1.96	0.22	0.65	-
F-	1.00	1.47	2.61	5.53	3.02	3.66	2.07	-

4. Conclusion

The proposed three techniques are rapid, accurate and precise and can be used for the analysis of Bepotastine-B in pure form and in pharmaceutical formulations in the presence of its oxidative degradation product. The sample recovery for all five methods was in good agreement with their respective label claims which suggested no interference of additives and excipients.

Competing interests

Authors have declared that no competing interests exist.

References

1- O' Neil J.M. The Merck Index, 14th Ed., Merck Research Laboratories, Merck and Co. Inc, Rahway, USA, 2006, p.189.

2- Godse K. and Kumari N.; Bepotastine Besilate: A Novel Anti-histamine, Indian J Drugs Dermatol 2017; 3: 64-8.

3- Kotla NR, Nagaraju Ch. VS.; Stability indicating HPLC method for the quantification of bepotastine besilate and its related substances, Der pharma chemical,2014; 6(2): 343-351.

4- Gupta KR, Askarkar SS; Simultaneous estimation of bepotastine besilate and benzalkonium chloride in ophthalmic formulation by RP-HPLC method, Int.J. Pharm. Pharm.Sci., 2016; 8(6): 225-233.

5- SHI Ai-Xin, LI Ke-Xin, HU Xin; Study on determination of bepotastine in human plasma and urine by HPLC-MS/MS and pharmacokinetics , Chinese Journal of Pharmaceutical Analysis, 2014; 34(6): 2139-2144.

6- Damle M. C. and Ghode Sh. P.; Stability indicating chromatographic method for estimation of bebotastin besilate, IAJPS, 2018; 05 (06): 5222-5228.

7- Rashed N.S., Abdallah O. M. and Said N. S.; Validated stability indicating methods for determination of oseltamivir phosphate, BJPR, 2017; 16(1): 1-9.

8- Abdel-Razeq S. A., Abdel-Salam K.M., Fouad M. M. and Mohamed T.F.; Stability indicating spectrophotometric methods for the determination of pramipexole dihydrochloride, WJPPS. 2015; 3(12): 1444-1457.

9- The United State Pharmacopoeia 35, NF 30, Asian Ed. Rand Mc Nally, USA, 2012.