

Original Research Article

Growth, Photosynthesis and Quality of Water Spinach (*Ipomoea aquatica*) as influenced by Magnetic nanoparticles (MNP) application

ABSTRACT

Aims: To characterize the growth, carbon assimilation and quality of *Ipomoea aquatica* as influenced by magnetic nanoparticles (MNP) application as well as to determine the best rates of iron oxide nanoparticles that give high growth, carbon assimilation and quality of *Ipomoea aquatica*.

Study design: *Ipomoea aquatica* plants were exposed to four different treatments of magnetic iron oxide nanoparticles (Fe_3O_4) (0, 50, 100 and 150 mg L⁻¹). The experiment was conducted in a randomized complete block design (RCBD) with 3 replications. One unit of experiment consisted of 8 plants and there were 96 plants used in the experiment.

Place and Duration of Study: Department of Biology, Faculty of Science, Universiti Putra Malaysia, between March 2018 and July 2018.

Methodology: The growth parameters measured included: plant height, basal diameter, total leaf number, leaf temperature, total chlorophyll content and plant biomass. The carbon assimilation parameters were measured using IRGA (Infrared Gas Analyzer, LICOR 6400 XT Portable Photosynthesis System). i.e. transpiration rate (E), stomatal conductance and water use efficiency (WUE). The chlorophyll fluorescence were measured by using Pocket PEA that measured maximum efficiency of photosystem ii, (fv/fm), maximum quantum yield of phytochemical and non-photochemical process in photosystem II (fv/fo), minimal fluorescence (fo), performance index (PI) and Density of Reaction Centers Per PSII Antenna Chlorophyll (RC/ABS). Total phenolics and flavonoids contents in leaves were measured using Folin-Ciocalteu method.

Results: It was observed that plant height, shoot length, plant temperature, total biomass, and total chlorophyll content were significantly influenced ($p \leq 0.05$) by the different concentrations of magnetic nanoparticles. The net photosynthetic rate (A), transpiration rate (E), stomatal conductance (gs), maximum efficiency of photosystem II (Fv/fm), maximum quantum yield of phytochemical and non-photochemical process in photosystem II (Fv/fo), performance index and the density of reaction centers per PSII antenna chlorophyll of *Ipomoea aquatica* were significantly reduced at higher concentration of magnetic nanoparticles. However, water use efficiency and minimal fluorescence value (Fo) of *Ipomoea aquatica* increased with increase of MNP concentration. In addition, the application of magnetic nanoparticles significantly influenced ($P \leq 0.05$) the total flavonoids and total phenolics content in water spinach. Both of these parameters were increased when higher concentration of magnetic nanoparticles was applied to *Ipomoea aquatica*. This study showed that MNP affected the growth, carbon assimilation and secondary metabolites production of *Ipomoea aquatica*.

Conclusion: In conclusion, the higher concentration of magnetic nanoparticles reduced the growth rate and carbon assimilation of water spinach and enhanced the production of secondary metabolites.

Keywords: [vegetables, nanoparticles, growth, photosynthesis, biometabolites]

14 1. INTRODUCTION

15

16 Based on the data from Department of Statistics Malaysia, agriculture sector was operated
17 at 11,628 establishments in the year 2015 with an annual growth of 5.7 percent [1] and
18 vegetables were contributing to agricultural commodities in which spinach (*Spinicia oleracia*),
19 long bean (*Vigna unguiculata*), mustard (*Brasica oleracia*), eggplant (*Solanum melongena*),
20 tomato (*Solanum lycopersicum*) and cucumber (*Cucumis sativus*) were selected by which
21 more than 100 percent were recorded in self-sufficiency ratio [2]. Malaysia population in
22 2018 is estimated at 32.4 million compared to 32.0 million in 2017 by which 1.1 per cent as
23 the growth rate [3]. As the population increases, the demand for food and commercial
24 energy is accelerating to fulfill the population requirements. According to Department of
25 Agriculture Malaysia, water spinach (*Ipomoea aquatica*) is one of the vegetables suggested
26 by Malaysia government to be consumed widely due to its low price and Asia had consumed
27 water spinach at the highest rate compared to other vegetables [4].

28

29 *Ipomoea aquatic*, also known as water spinach, is a popular vegetable in countries like
30 Malaysia, Hong Kong, Taiwan and other Asia countries. This edible vegetable is classified
31 into the family Convolvulaceae. Water spinach is an aquatic or semi-aquatic yearly herb
32 which usually creeps on moist soil or sand besides floating in water [5]. Countries like
33 Southern Asia, Bangladesh, China and India had been using *Ipomoea aquatica* in folk
34 medicine against different diseases including diabetes, malfunction of liver, constipation and
35 Arsenic poisoning as it is known for its high nutritive values and consumable leafy
36 vegetables [5]. Water spinach is also rich in minerals like flavonoids, phenolics and
37 carotenes. As water spinach is easily grown plant, has short time to harvest period and well
38 adapted to environment changes, this makes the plant favorable for cultivation. However,
39 water spinach could accumulate foreign minerals like Cadmium, Zinc and Copper which
40 enables it to be used as the sample plant for research [6].

41

42 Metals such as nickel, cobalt, and iron are used to demonstrate magnetic properties in which
43 their magnetic particle size are within nanoscale [7]. Currently, magnetic nanoparticles have
44 attracted researchers from different background like biotechnology, biomedicine and
45 agriculture because this material possess wide applications and easy to be produced
46 However, the application of magnetic nanoparticles relies on the particle's condition on its
47 steadiness [8]. The application of magnetic nanoparticles in the study of plant currently have
48 attracted interests of researchers for its ability to permit a particular localization to discharge
49 their load as conveyed to the plants. Recent studies being applied on pumpkin plants to test
50 on the specific localization, take-up, and translocation of magnetic nanoparticles (under 50
51 nm) [9]. Besides, gas exchange is also being influenced by the use of magnetic
52 nanoparticles in which they act on photosynthetic surface causing foliar warming and
53 changes physiological process and cell elements of plants as the leaves are faced with
54 stomatal obstacle [10].

55

56 The application of magnetic nanoparticles may enhance or retard the growth of *I. aquatica*
57 through: carbon nanotubes that increased the leaf gas trades properties of the plant. It was
58 seen in *Arabidopsis thaliana* that treatment with single wall carbon nanotubes (SWCNT) led
59 to higher plant photosynthetic, photoabsorption and higher electron transport rates
60 contrasted with the plant that was not treated with the materials. This was expected due to a
61 higher productivity of chloroplast when cooperated with the nanomaterials [11].

62

63 Secondary metabolites are also important factors that are influenced by the magnetic
64 nanoparticle application. Secondary metabolites are the natural products present in low
65 amounts and whose production rely on different species, genera and families. Secondary
66 metabolites are important in protecting plants from insects and pathogens, besides shaping

67 imperative UV-radiation absorbing compounds that eventually reduce the chances for the
68 plant to die [12]. The production of flavonoids and phenolics are the determinants for the
69 production of secondary metabolite in plant. It is often observed that plants undergoing
70 stress have higher total flavonoid and phenolics contents due to defensive mechanism in
71 them. The signal in plant to produce secondary metabolites is usually stimulated by the
72 accumulation of heavy metals like zinc, iron, and nickel which generates Reactive Oxygen
73 Species (ROS) and induces oxidative stress in plant. Consequently, the induction of
74 oxidative stress causes changes in signal transduction for the mechanism of gene coding
75 and enzyme [13]. Besides, the production of ROS may cause damages to cell membrane,
76 cell structure and photosynthetic site and thus the production of flavonoid at the generation
77 site act as defensive mechanism due to its high antioxidant properties. In addition, induced
78 phenolics are produced when the plant faces physical injury, infection or environmental
79 stresses due to heavy metal irradiation or temperature [14].
80

81 The low toxicity level of magnetic nanoparticles such as super paramagnetic nanoparticles
82 (SPION) has been the reason for growing studies on the application of magnetic
83 nanoparticles on organisms. A study shows that iron oxide nanoparticles is safe and non-
84 cytotoxic at the level of $100 \mu\text{g mL}^{-1}$ when being compared with few metal oxide
85 nanoparticles *in vitro* [15]. In one of the studies related to iron oxide nanoparticles exposure
86 on sunflower plant, *Helianthus annuus*, it was seen that at the concentration of $50\text{-}100 \text{ mg L}^{-1}$
87 the exposure resulted in reduction of root water pressure of the plant [16]. Besides, a study
88 by Liu, Zhang and Lal [17] showed that the iron oxide nanoparticles were less toxic, besides
89 stimulating root elongation during the germination of lettuce, *Lactuca sativa* at the
90 concentration of $5\text{-}20 \text{ mg L}^{-1}$ while inhibited root elongation at 50 mg L^{-1} [18].
91

92 Magnetic nanoparticles application is currently recognized in biomedicine to treat various
93 diseases but is still far behind in plant biology. Magnetic-based materials can be used in
94 production of certain chemicals to protect plant systematically from diseases besides
95 controlling externally the movement of nanocarriers in the plant by using high power external
96 magnet [19]. As water spinach is well known for its importance in culinary as vegetable and
97 in traditional medicine, this study would help the plant to be continuously used in studies.
98

99 The study of *I. aquatica* in the aspects of biochemical and physical towards magnetic
100 nanoparticles is still far behind in current research. Besides, the research on the impacts of
101 magnetic nanoparticles on growth, carbon assimilation and quality of plant in form of
102 secondary metabolites production is still few in science studies. Hence, this research was
103 conducted with few objectives which are to relate the growth, carbon assimilation and quality
104 of *I. aquatica* as influenced by magnetic nanoparticles application, to determine the best
105 concentration of magnetic nanoparticles that can promote the optimum growth and quality of
106 *I. aquatica* and to infer the relationship between growth and secondary metabolites of *I.*
107 *aquatica* as affected by magnetic nanoparticles application.
108
109

110 2. MATERIAL AND METHODS

111 2.1 Synthesis of Iron Oxide Nanoparticles (Fe_3O_4)

112 The iron oxide nanoparticles (Fe_3O_4) were synthesized using co-precipitation method [20].
113 Iron (III) chloride powder and Iron (II) chloride powder were mixed in the ratio of 2:1 in a 250
114 mL conical flask and were dissolved in 150 ml of deionized water. The solution was heated
115 at 45°C bubbled with nitrogen gas for 15 minutes. The solution was then added with 20 mL
116 of 25-30 % ammonia solution and stirred at 800 rpm for an hour. Then, the Iron (III) oxide
117 nanoparticles produced was collected by magnetic decantation. The Fe_3O_4 product was then
118
119

120 washed three times with acetone and centrifuged. The product was left to dry in furnace one
121 night and grinded after drying process to obtain the powder form. Fe₃O₄ produced were in
122 black colored fine powder.

123

124 **2.2 Plant Materials and Maintenance**

125

126 The experiment was conducted at the Vegetables Field plot for Teaching and Research,
127 Taman Pertanian Universiti, Universiti Putra Malaysia. The source of planting materials in
128 the study was the seeds of *I. aquatica*. Seeds of *I. aquatica* were propagated for 14 days in a
129 tray and transplant to polybags containing a mixture of top soil and sand (ratio 3:1). After 1
130 month, *I. aquatica* plants were exposed to four different treatments of Fe₃O₄ (0, 50, 100 and
131 150 mg L⁻¹). The magnetic nanoparticles were conveyed to the plants through watering. The
132 magnetic nanoparticles were diluted in distilled water before being applied to the plants.
133 Each plant was watered with 40 mL of magnetic nanoparticle solution. To maintain the plant
134 growth and avoid plant wilting or attack by any major plant disease that affect plants,
135 maintenance was done time to time.

136

137 **2.3 Experimental Design**

138

139 The polybags were arranged accordingly to Completely Randomized Block Design (RCBD)
140 with replication of 3 blocks. One unit of experiment consisted of 8 plants, and there were
141 total of 96 plants utilized in the experiment.

142

143 **2.4 Data Collection**

144

145 Data collection of plant growth were done on Week 3, 6, 9 and 12 while the destructive
146 analysis and leaf gas exchange measurement were conducted on the 12th week.

147

148 **2.4.1 Plant Growth Measurement**

149

150 The plant growth measurement was done to obtain the water spinach plant height, number
151 of leaf, diameter of stem, root to shoot ratio, plant temperature and the chlorophyll content.

152

153 **2.4.1.1 Plant height**

154

155 Measuring tape was used to measure the plant height which starting from stem at soil
156 surface up to the highest shoot growth or at its tip.

157

158 **2.4.1.2 Plant basal diameter**

159

160 The basal tips of the plant were measured by using vernier caliper.

161

162 **2.4.1.3 Plant leaves number**

163

164 The whole leaves of the plants were counted manually (one by one piece) and then recorded
165 in every 3 weeks.

166

167 **2.4.1.4 Plant leaf temperature determination**

168

169 Infrared (IR) Thermometer was used to measure plant leaf temperature.

170

171 **2.4.1.5 Chlorophyll content measurement**

171

172 Total chlorophyll content in the leaves was measured using SPAD 502 chlorophyll meter
173 (Spectrum Tech Inc; Aurora, IL; USA). The leaf was clipped under the chlorophyll meter
174 clipper to obtain the reading for every treatment in each of the replication. The data was
175 collected every week from week zero until week 12 of the experiment.

176

177 **2.4.1.6 Plant fresh weight measurement**

178

179 The plants were removed from the soil carefully and the dirt from the soil was rinsed with tap
180 water. The shoot, root and the leaf parts were separated and weighed using analytical
181 balance.

182

183 **2.4.1.7 Plant biomass (Dry weight) measurement**

184

185 All the plants were dried for 2 days in the oven at 60°C. The dry weight of root, stem and leaf
186 per seedling were recorded as plant biomass.

187

188 **2.4.1.8 Root to shoot ratio**

189

190 The root to shoot ratio was determined by dividing the dry weight of the root with the dry
191 weight of the shoot.

192

193 **2.4.2 Leaf gas exchange measurement**

194

195 A LI-6400XT (Li-COR Inc; Nebraska; USA) portable photosynthesis system gave the reading
196 of leaf gas exchange. The instrument was warmed and calibrated for 30 minutes with the
197 ZERO IRGA mode. The measurements used optimal conditions set at 400 $\mu\text{mol mol}^{-1} \text{CO}_2$,
198 30°C cuvette temperature, 60% relative humidity with air flow rate set at 500 $\text{cm}^3 \text{min}^{-1}$ and
199 modified cuvette condition of 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ photosynthetically photon flux density (PPFD).
200 The fully expanded young leaves were used to measure net photosynthesis (A),
201 transpiration rate (E) and stomata conductance (g_s) which also gave the measurement of
202 gas exchange. It was an automatic operation therefore the data was recorded in the LI-
203 6400XT console and further analyzed by Photosyn Assistant Software (Dundee Scientific,
204 Dundee, UK). Precautions step were considered while taking the measurements to avoid
205 errors.

206

207 **2.4.3 Chlorophyll fluorescence determination**

208

209 Measurements of chlorophyll fluorescence were taken from fully expanded second leaves.
210 Leaves were darkened for 15 min by attaching light-exclusion clips to the central region of
211 the leaf surface. Chlorophyll fluorescence was measured using a portable chlorophyll
212 fluorescence meter (Pocket PEA, Hansatech Instruments Ltd., Norwich, UK). Measurements
213 were taken at $>3,000 \mu\text{mol m}^{-2} \text{s}^{-1}$ and recorded for up to 5 seconds. The fluorescence
214 responses were induced by emitting diodes. Measurement of f_0 (initial fluorescence), f_m
215 (maximum fluorescence) and f_v (variable fluorescence) were obtained from this procedure.
216 f_v is derived as the difference between f_m and f_0 . Fluorescence values recorded, include the
217 initial/minimal fluorescence (f_0), the ratio of variable to maximum fluorescence (f_v/f_m) which
218 represents the maximum quantum yield of photosystem II (PS II), and the ratio of variable to
219 minimum fluorescence (f_v/f_0) which estimates the maximum primary yield of photochemistry
220 of PS II. The f_m is the maximal fluorescence value, and f_v is the variable fluorescence
221 calculated as $f_m - f_0$.

222

223 **2.4.4 Total phenolics and flavonoids quantification**

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

The total phenolics and flavonoid measurement follow methods from Ibrahim et al [21]. Plant tissue samples (0.1 g) that was initially ground were extracted with 80% ethanol (10 mL) on an orbital shaker at 50°C for 120 minutes. The mixture was filtered, and the filtrate used for the quantification of total flavanoids and total phenolics. Total phenolics content of the leaf samples was determined by using Follin-Ciocalteu reagent (Sigma Aldrich, Missouri, USA; diluted 10-fold). The absorbance measured at 725 nm. The results were expressed as mg g⁻¹ gallic acid equivalent (mg GAE g⁻¹ dry sample). For total flavonoids determination, a sample (1 mL) mixed with NaNO₃ (Sigma Aldrich, Missouri, USA; 0.3 mL) in a test tube which covered with aluminum foil, and left for 5 min. Then 10% AlCl₃ (Wako Pure Chemical Industries Ltd; Tokyo, Japan; 0.3 mL) added followed by addition of 1 M NaOH (Kanto Chemical Co. Inc.; Hokkaido, Japan; 2 mL). Later, the absorbance was measured at 510 nm using a spectrophotometer with rutin as a standard (results expressed as mg g⁻¹ rutin dry sample).

2.5 Statistical analysis

The recorded data was analyzed by using statistic software known as Statistical Package for Social Sciences (SPSS) with the version 24. A two-way ANOVA test was carried out for data analysis for all the parameters in the experiment. Data was significant if the *p*-value level ≤ 0.05.

3. RESULTS AND DISCUSSION

3.1 Plant height

Fig.1 showed that, different concentrations of magnetic nanoparticles application influenced significantly the plant height of *I. aquatica* at *P* ≤ 0.05 in week 3. The highest plant height was observed in 0 mg L⁻¹ (14.10 cm), followed by 50 mg L⁻¹ (12.2 cm), 10 mg L⁻¹ (11.2 cm) and lowest in 150 mg L⁻¹ (11.1 cm), these differences were significantly different. The current result indicates that the application of magnetic nanoparticles at higher concentration reduced the plant height of *I. aquatica*. Research had shown that higher concentration of Fe₃O₄ with pro-longed exposure may lead to iron toxicity in the plant. The plants that faced toxicity during growth stage may initiate stress and causes reduction in plant height due to effects of iron toxicity like production of free radicals, root break down and bronzing of leaves which lead to yield loss [22]. The application of Fe₃O₄ stimulate the production of Reactive Oxygen Species (ROS) under the condition of excess or deficiency of Fe₃O₄ that responsible of signaling molecule that stimulate or inhibit plant growth [23, 24].

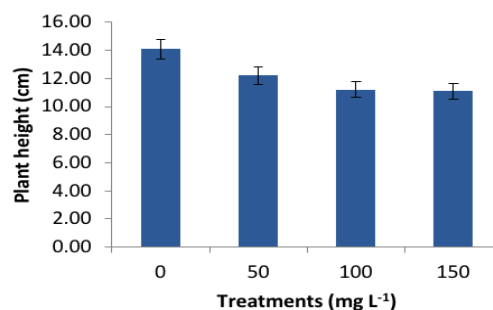
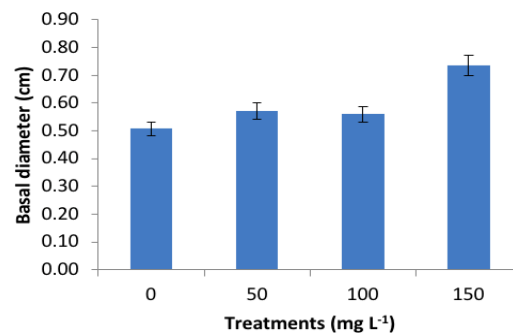


Fig. 1. The impact of magnetic nanoparticles on plant height of *Ipomoea aquatica*. Data are mean ± standard error of mean (SEM). N= 24.

271 **3.2 Basal diameter**

272 Different concentration of magnetic nanoparticles application has significantly influenced the
273 basal diameter of *I. aquatica* ($P \leq 0.05$; Fig. 3) in week 12. At 12th week, the application of
274 magnetic nanoparticles showed significant increase in basal diameter of *I. aquatica* at higher
275 concentration which is at 150 mg L⁻¹. The smallest basal diameter was observed in 0 mg L⁻¹
276 (0.51 cm), followed by 100 mg L⁻¹ (0.56 cm), 50 mg L⁻¹ (0.571 cm) and greatest in 150 mg L⁻¹
277 (0.74 cm). The current result indicates that the application of magnetic would promote the
278 stem diameter of *I. aquatica*. Based on research, aquaporins are found on tonoplast or also
279 known as vacuolar membrane which allows water to be freely moves across the cells in the
280 symplastic route. Therefore, the abundance of aquaporins found on the membrane depends
281 on the regulation of water flow. During water stress due to high concentration of
282 nanoparticles may lead to induction of turgor-responsive aquaporins to maintain turgor
283 pressure. This would cause the vacuole size to increase that brings to cell expansion and
284 increased of stem diameter [25]. Besides, a research done on radish cotyledon shows that
285 increase in turgor pressure increase the cell expansion and growth. However, the study
286 claimed that under unfavorable condition, cell would not divide [26]. Therefore, it is proposed
287 that higher concentration of Fe₃O₄ caused increase in stem diameter.
288

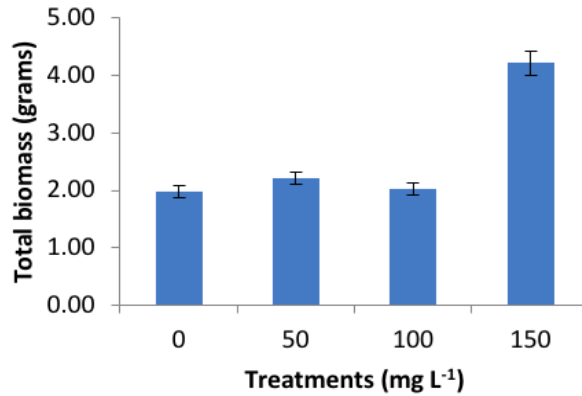


289
290
291
292
293
294
295
296
297
298
299
300
301 **Fig. 3. The impact of magnetic nanoparticles on basal diameter of *Ipomoea aquatica***
302 **Data are mean \pm standard error of mean (SEM). N= 24.**

303
304 **3.3 Total biomass**

305
306 Based on Fig. 4, different concentration of magnetic nanoparticles significantly influenced
307 the total biomass of *I. aquatica* ($p \leq 0.05$) in week 12. In week 12, a significant effect was
308 observed by which the application of Fe₃O₄ with the concentration of 150 mg L⁻¹ increased
309 the biomass of water spinach. The highest total biomass was observed in 150 mg L⁻¹ (4.21
310 gram), followed by 50 mg L⁻¹ (2.22 gram), 100 mg L⁻¹ (2.02 gram), and 0 mg L⁻¹ (1.98 gram).
311 The result obtained showed that iron oxide nanoparticles with highest concentration would
312 increase the total biomass of *I. aquatica*. This result was in agreement with the finding by
313 Sheykhbaglou and Sedghi [27]. From their research, it was found that the use of iron oxide
314 nanoparticles increased the dry weight of pod and peanut plant. They found that the
315 application of Fe₃O₄ helped in transferring iron and photosynthate particle in the leaves of
316 plants. Besides, the total biomass of *Vigna radiata* applied with Fe₃O₄ showed a positive
317 result compared to the application of ferum ions. This was d due to the increase of α -
318 amylase activity in the seeds exposed to Fe₃O₄. The increase of total biomass was also
319 observed in *Spinacea oleracea* applied with magnetic nanoparticles [18]. However, study by
320 Jeyasubramanian et al. [28] showed that higher concentration of Fe₃O₄ (200 mg L⁻¹) cause
321 decrease in both wet and dry weight of spinach [29].
322

323
324
325
326
327
328
329
330
331
332
333
334
335
336
337



338 **Fig. 4. The impact of magnetic nanoparticles on total biomass of *Ipomoea aquatica***
339 **Data are mean ± standard error of mean (SEM). N= 24.**

341 3.4 Leaf temperature

342

343 Fig. 5 showed that magnetic nanoparticle significantly influenced the plant temperature of *I.*
344 *aquatica* ($P \leq 0.05$) in week 12. The highest plant temperature was observed at 100 mg L⁻¹,
345 (32.0 °C) followed by 150 mg L⁻¹ (31.1 °C), 50 mg L⁻¹ (30.8 °C) and 0 mg L⁻¹ (28.7 °C) in
346 week 12. The result shows that a higher concentration of magnetic nanoparticles shows
347 higher leaf temperature of water spinach. Plant temperature is often related to transpiration
348 process. Transpiration is a cooling process taken by the plant to release water vapor from
349 the plant through stomata and cuticle. Through transpiration, thermal energy is balanced by
350 the loss of heat to its surrounding. Therefore, when the transpiration rate is decreased due to
351 the accumulation of nanoparticles at the root surface and inhibits the water intake capacity,
352 the plant temperature would increase. Besides, the stomata conductance also plays a major
353 role in maintaining the leaf plant temperature. The closing and opening of stomata do give
354 impact on the plant temperature [30]. Thus, higher concentration of Fe₃O₄ result higher plant
355 temperature due to its relationship with transpiration rate and stomata conductance.
356

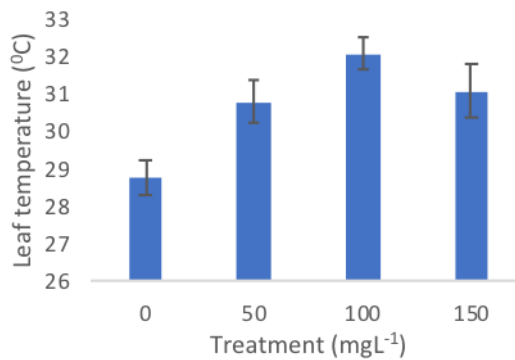
357

358

359

360

361



362

363

364 **Fig. 5. The impact of magnetic nanoparticles on plant temperature of *Ipomoea***
365 ***aquatica*. Data are mean ± standard error of mean (SEM). N= 24.**

366

367

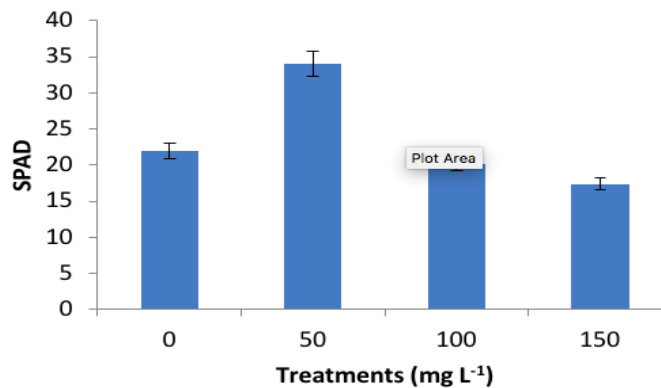
368

369

3.5 Total chlorophyll content

370
371
372
373
374
375
376
377
378
379
380
381
382
383
384
385
386
387
388
389
390
391
392
393

Fig. 6 shows the impact of magnetic nanoparticle on total chlorophyll content of water spinach. There was significant effect of the application of magnetic nanoparticles on the water spinach chlorophyll content at $P \leq 0.05$. After 12 weeks of treatment, the highest chlorophyll content was observed in *I. aquatica* treated with 50 mg/L Fe_3O_4 (34), followed by 0 mg L^{-1} (22) and 100 mg L^{-1} (20) and lowest in 150 mg L^{-1} (17). This result indicates that higher concentration of magnetic nanoparticles leads to decrease of chlorophyll content in water spinach which was supported by several studies. Based on research by Racuciu et al. [31], *Zea mays* plant treated with 20, 40, 60, 80, and 100 $\mu\text{l l}^{-1}$ of Fe_3O_4 suspension concentration showed decreased in chlorophyll content. The magnetic nanoparticles showed both chemical and magnetic influence on the water spinach enzymatic structure that eventually influence the photosynthetic system of the plant at higher concentration. Besides, the application of nanoparticles would induce oxidative stress in plant that reduced the chlorophyll content in plant leaf. For example, zinc oxide nanoparticles had proven to reduce the chlorophyll content in wheat plant due to the formation of free radical [32]. Another research done on watermelon showed that the application of magnetic nanoparticles at higher concentration loss the content of chlorophyll. Due to the toxic substance exposure to the plant, the Malondialdehyde (MDA) production could be observed as a result of lipid peroxidation. Low level of MDA is important in protecting the structure and function of cell membranes. As the increase of MDA in plant with the presence of Fe_3O_4 , the penetration of large particles into the cell is disturbed that result in less efficiency of cell. Thus, iron is claimed to be the cause for enzymatic activity inhibition that reduce chlorophyll synthesis [33].



394
395
396
397
398
399

400 **Fig. 6. The impact of magnetic nanoparticles on total chlorophyll content of *Ipomoea***
401 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 24.**

402 3.6 Net photosynthesis rate

403 Fig. 7 showed the net photosynthesis rate (A) of water spinach after 12 weeks of planting.
404 The results shown that the net photosynthesis rate was significantly influenced ($p \leq 0.05$) by
405 the application of Fe_3O_4 during the planting period. The higher concentration of magnetic
406 nanoparticles resulted in lower net photosynthesis rate of water spinach. The net
407 photosynthesis of control treatment (0 mg L^{-1}) was higher than other treatments: 50 mg L^{-1} ,
408 100 mg L^{-1} and 150 mg L^{-1} that recorded 6.9 $\mu\text{mol m}^{-2}\text{s}^{-1}$, 6.3 $\mu\text{mol m}^{-2}\text{s}^{-1}$, 3.4 $\mu\text{mol m}^{-2}\text{s}^{-1}$,
409 and 2.2 $\mu\text{mol m}^{-2}\text{s}^{-1}$ respectively. The photosynthesis rate could have been altered due to the
410 application of magnetic nanoparticles by which these nanoparticles could block the pathway
411 and caused stress to the water spinach. Although nanoparticles are in the size of nanometer
412 but their entry into the plant may cause some changes either by enhancing or inhibiting the
413 photosynthesis rate. In this case, the electron transport chain may be blocked by the

414 nanoparticles and enhance stress to the plant manipulating and changing the normality of
415 genes and enzymes like Rubisco. Rubisco is one of the enzymes that play an important role
416 in the conversion of carbon dioxide into biological substances [34]. Therefore, the changes in
417 production of Rubisco may lead to lower net photosynthesis rate with the application of
418 magnetic nanoparticles.

419

420

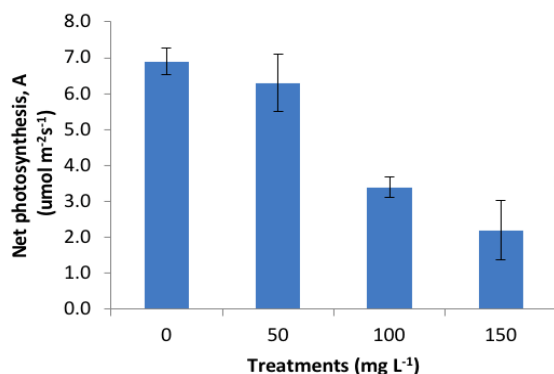
421

422

423

424

425



426 **Fig. 7. The impact of magnetic nanoparticles on net photosynthesis rate of *Ipomoea***
427 ***aquatica*. Data are mean ± standard error of mean (SEM). N= 24.**

428 3.7 Transpiration rate

429 Based on Fig. 8, the transpiration rate of *I. aquatica* was significantly influenced by the
430 application of magnetic nanoparticles after 3 months of experimental period as $P \leq 0.05$. The
431 transpiration rate was highest in 50 mg L⁻¹ treatment followed by the control treatment, 100
432 mg L⁻¹ and 150 mg L⁻¹ of Fe₃O₄ by 2.34 mmol H₂O m⁻²s⁻¹, 1.75 mmol H₂O m⁻²s⁻¹, 1.44 mmol
433 H₂O m⁻²s⁻¹, and 0.80 mmol H₂O m⁻²s⁻¹ respectively. This indicates that higher concentration
434 of Fe₃O₄ would decrease the transpiration rate while an optimum concentration did not inhibit
435 the transpiration rate in which in this case is 50 mg L⁻¹ of iron oxide nanoparticles.
436 Transpiration rate is linked up by photosynthesis rate by means if there is reduction in
437 photosynthesis rate, the transpiration rate would decrease too. Besides, nanoparticles
438 applied to the plant may cover the root surface of the plant and causes water stress in water
439 spinach. This is supported by a research using Titanium oxide nanoparticle in maize in which
440 the water transport capacity of the primary cell wall was reduced due to accumulation of
441 nanoparticles at the plant leaf [35]. Besides, the research done on *Citrullus lanatus* to study
442 the application of magnetic nanoparticles on the root activity showed that the Fe₃O₄
443 accumulated at the root surface that prevent the transmission of water and also other
444 nutritional components by the plant [33].

445

446

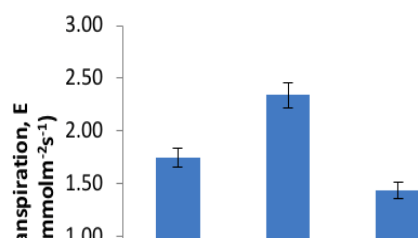
447

448

449

450

451



452

453

454

455

456

457

458

459 **Fig. 8. The impact of magnetic nanoparticles on transpiration rate of *Ipomoea***
460 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 24.**

461 **3.8 Stomata conductance**

462 The stomata conductance of water spinach was significantly affected by the application of
463 magnetic nanoparticles as showed in Fig. 9 as $P \leq 0.05$. After 12 weeks of treatment
464 application, the stomata conductance for 50 mg L⁻¹ Fe₃O₄ was the significantly highest
465 compared to 0 mg L⁻¹, 100 mg L⁻¹ and 150 mg L⁻¹ with 0.314 mol H₂O m⁻²s⁻¹, 0.246 mmol
466 H₂O m⁻²s⁻¹, 0.144 mol H₂O m⁻²s⁻¹, and 0.134 mol H₂O m⁻²s⁻¹ respectively. From the result, it
467 was observed that higher concentration of Fe₃O₄ would decrease the stomata conductance
468 of water spinach. Stomata conductance is determined by the degree of stomata aperture
469 which estimates the rate of gas exchange and transpiration rate. A greater conductance is
470 shown when the degree of stomata opening and its function in term of density and size is
471 greater. When the plant undergoes greater photosynthesis and transpiration rate, the
472 stomata conductance is greater. This statement is proven by the result of this experiment by
473 which when the photosynthesis and transpiration rate of water spinach is lower for higher
474 concentration of Fe₃O₄, the stomata conductance follows the same pattern. The application
475 of magnetic nanoparticles may cause leakage of electrolyte in the plant which alters the
476 mechanism of potassium pump that controls stomata opening. The accumulation of
477 magnetic nanoparticles may cause the stomata to be closed or partially closed which directly
478 reduce the stomata conductance [14].

479

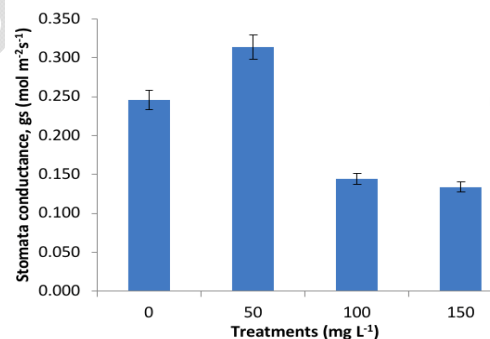
480

481

482

483

484



485 **Fig. 9. The impact of magnetic nanoparticles on stomatal conductance of *Ipomoea***
486 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 24.**

487 **3.9 Water use efficiency**

488 From the Fig. 10, the result showed that the water use efficiency of water spinach was
489 significantly affected by the application of Fe₃O₄ as $P \leq 0.05$ by which the highest water use
490 efficiency was recorded for 150 mg L⁻¹ treatment, followed by other treatments with 0 mg L⁻¹,
491 50 mg L⁻¹ and 100 mg L⁻¹ at 4.23, 2.86, 2.72, and 2.44 $\mu\text{molCO}_2\text{m}^{-2}\text{s}^{-1}$ respectively. From the
492 result, it was observed that the water spinach with the highest concentration of Fe₃O₄
493 showed greater water use efficiency. Water use efficiency is one of the important
494 determinants of plant growth under stress as the plant maximize the capturing of soil
495 moisture when there is limitation of water supply or lower stomata conductance in order to
496 increase yield production [36]. Water use efficiency is related with transpiration rate and
497 defined as the ratio of moles CO₂ assimilated per moles of water transpired [14]. Therefore,
498 with the application of magnetic nanoparticles at higher concentration increase the
499 transpiration rate of *I. aquatica*, the water use efficiency is also increased.

500

501

502

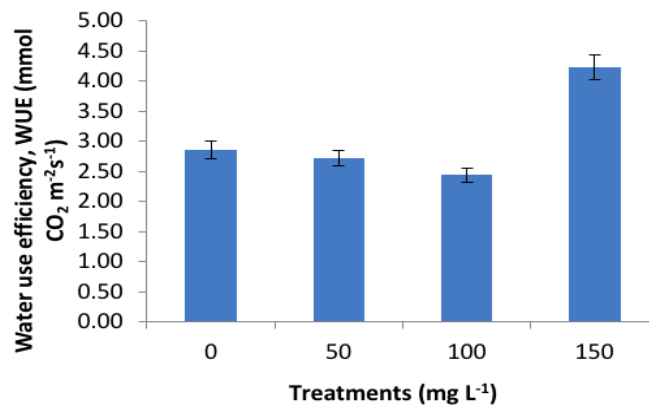
503

504

505

506

507



508 **Fig. 10. The impact of magnetic nanoparticles on water use efficiency of *Ipomoea***
509 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 24.**

510 **3.10 Maximum efficiency of Photosystem II**

511 Figure 11 shows the effect of different concentration of Fe₃O₄ on maximum efficiency of
512 photosystem II or known as fv/fm ratio. The result of the application on water spinach was
513 significant as $P \leq 0.05$ by which higher concentration of Fe₃O₄ showed lower Fv/Fm value.
514 The ratio of fv/fm for 150 mg L⁻¹ (0.65) was significantly lower than 100 mg L⁻¹ (0.69) and
515 followed by 50 mg L⁻¹ (0.74) and 0 mg L⁻¹ (0.76). The lowest Fv/Fm ratio showed by water
516 spinach treated with 150 mg/l clearly shows that higher concentration of Fe₃O₄ would reduce
517 the efficiency of photosystem II. Fv/Fm is the main parameter used to detect any injury in
518 photosystem II or photon inhibitory process as it is related to quantum yield of
519 photosynthesis. The induction of stress on photosynthetic surface in plant would limit the
520 transformation of light energy in photosystem II. Thus, the decrease in photosynthetic in
521 plant would reduce the reduction of phytochemical activity in photosystem II [37].

522

523

524

525

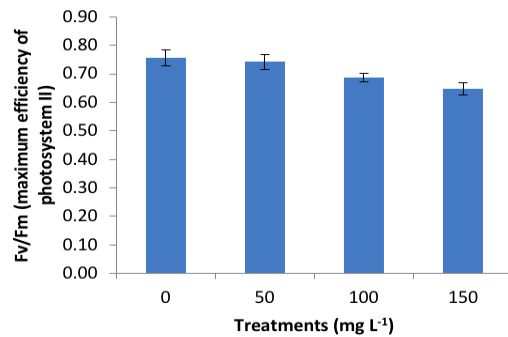
526

527

528

529

530



531 **Fig. 11. The impact of magnetic nanoparticles on fv/fm (maximum efficiency of**
532 **photosystem II) of *Ipomoea aquatica*. Data are mean ± standard error of mean (SEM).**
533 **N= 24.**

534 3.11 Maximum yield of photosystem II

535 Fig. 12 shows the effect of different concentration of Fe₃O₄ on maximum yield of
536 photosystem II or known as Fv/Fo ratio in 3 months planting periods. The result of the
537 application on water spinach was significant as $P \leq 0.05$ by which higher concentration of iron
538 oxide nanoparticles showed lower Fv/Fm value. The ratio of fv/fm for 0 mg L⁻¹ (3.15) and 50
539 mg L⁻¹ (3.14) was significantly higher than 100 mg L⁻¹ (2.40) and 150 mg L⁻¹ (2.35). As the
540 two parameters Fv/Fm ratio and Fv/Fo ratio showed the similar trend, it can be speculated
541 that both parameters are related to each other. The Fv/Fo is more sensitive towards
542 changes to efficiency of photosystem II compared to Fv/Fm as it shows quantum yield of
543 phytochemical and non-phytochemical process. This ratio indicates the state of photosystem
544 II on the energy absorbed and damaged occurred due to plant stress in leaf. This ratio is
545 also affected by the alteration of stomatal closure and carbon fixation process due to water
546 and temperature stress [38]. Besides, the accumulation of magnetic nanoparticles in the
547 photosystem I and II cause obstruction to photosynthesis in donor part of both photosystems
548 that reduce the fv/fo ratio [37].

549

550

551

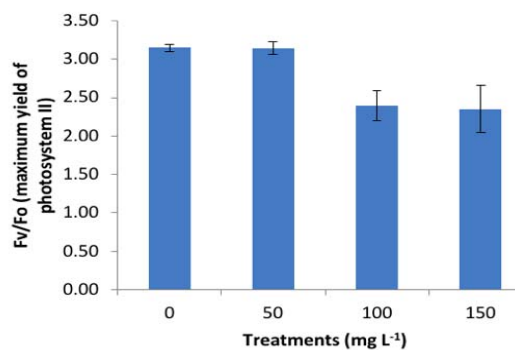
552

553

554

555

556



557 **Fig. 12. The impact of magnetic nanoparticles on fv/fo (maximum yield of**
558 **photosystem II) of *Ipomoea aquatica*. Data are mean ± standard error of mean (SEM).**
559 **N= 24.**

560 **3.12 Minimal fluorescence**

561 Based on Fig. 13, the application of Fe_3O_4 had significantly influenced the minimal
562 chlorophyll fluorescence yield of water spinach as $P \leq 0.05$. The F_o of water spinach treated
563 with 100 mg L^{-1} and 150 mg L^{-1} Fe_3O_4 were significantly higher compared to 0 mg L^{-1} and 50
564 mg L^{-1} , 722, 726, 653 and 607 respectively. From the result, it is shown that the F_o value
565 increased as the concentration of Fe_3O_4 increased. The application of nanoparticles like
566 titanium oxide showed that with the increase of its concentration, the minimal fluorescence
567 value decreased as titanium oxide nanoparticles do protect the photosynthetic structure of
568 the tomato plant under mild heat stress. However, the study by Gao et al. [39] showed that
569 at high light intensity the minimal fluorescence in *Ulmus elongata* seedlings decreased.
570 Therefore, the possible explanation for this would be the impact of magnetic nanoparticles
571 on water spinach is complex and it depends both on the concentration of Fe_3O_4 and the
572 environmental condition that would affect the activity of nanoparticles [40]. Besides, the
573 application of magnetic nanoparticles increased the heat dissipation in water spinach as the
574 absorbed light energy could not be used for photosynthesis [37].
575

576

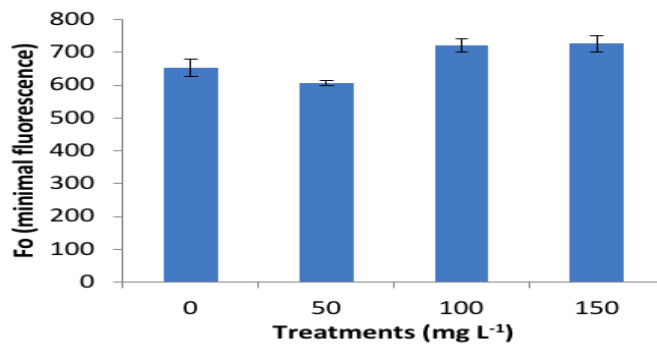
577

578

579

580

581



582 **Fig. 13. The impact of magnetic nanoparticles on f_o (minimal fluorescence) of**
583 ***Ipomoea aquatica*. Data are mean \pm standard error of mean (SEM). N= 24.**

584 **3.13 Performance index**

585 From the Fig. 14, the result showed that the performance index of water spinach was
586 significantly affected by the application of Fe_3O_4 as $P \leq 0.05$ by which the significantly highest
587 performance index was recorded for 50 mg/L treatment, followed by other treatments with 0
588 mg L^{-1} , 100 mg L^{-1} and 150 mg L^{-1} at 2.73, 2.51, 1.62, and 1.48 respectively. From the result,
589 it was observed that the water spinach with lower concentration of iron oxide nanoparticles
590 showed greater performance index. Performance index is the one of the parameters which is
591 sensitive to environmental conditions in a plant as it is used as a suitable tool to reflect water
592 deficit in a plant system. Based on research, performance index had been used as a
593 sensitive indicator of water stress in *Triticum aestivum*. Performance index is a combination
594 formula that takes into account the measurement of RC/ABS , maximal energy reflux that
595 reaches photosystem II and electron transport [41]. Therefore, the application of magnetic
596 nanoparticles that changes the net photosynthetic rate and transpiration rate due to the
597 accumulation of iron oxide nanoparticles at the root surface and the blockage of Electron
598 Transport Chain had affected the performance index of water spinach. Besides, the carbon
599 assimilation process and stomata conductance had correlated with the performance index of
600 water spinach which concluded that higher concentration of Fe_3O_4 showed lower
601 performance index in water spinach [42].

602

603

604

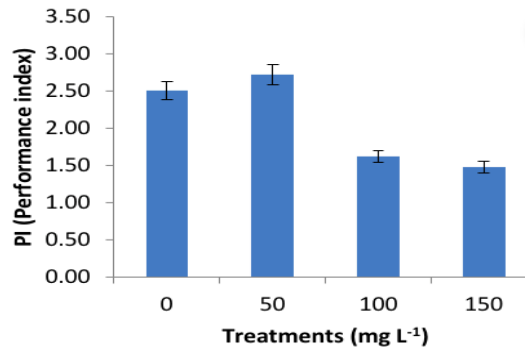
605

606

607

608

609



610 **Fig. 14. The impact of magnetic nanoparticles on PI (performance index) of *Ipomoea***
611 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 24.**

612 3.14 Density of reaction centers per PSII antenna chlorophyll

613 Fig. 15 shows the effect of different concentration of Fe₃O₄ on density of reaction centers per
614 Photosystem II antenna chlorophyll in three-month planting periods. The result of the
615 application on water spinach was significant as $P \leq 0.05$ by which higher concentration of
616 Fe₃O₄ showed lower density of reaction centers per Photosystem II antenna chlorophyll. The
617 density of reaction centers per Photosystem II antenna chlorophyll for 0 mgL⁻¹ and 50 mgL⁻¹
618 is 0.71 which was significantly higher than 100 mg L⁻¹ (0.56) and 150 mg L⁻¹ (0.60). RC
619 indicates the number of active reaction center in photosystem II while ABS shows the total
620 quantity of light absorbed by the antenna chlorophyll. Thus, RC/ABS indicate the total
621 number of active radiation per light absorption [43]. Due to the exposure of water spinach to
622 Fe₃O₄, free radicals production would induce stress to the plant and cause injury to the
623 photosystem II that limits the active reaction center and light absorption in photosystem II.
624 This parameter gives the same trend as the performance index as both shows the efficiency
625 of photosynthesis process in the plant. Therefore, the application of Fe₃O₄ at higher
626 concentration leads to plant stress and decrease the density of active reaction centers per
627 photosystem II antenna chlorophyll.

628

629

630

631

632

633

634

635

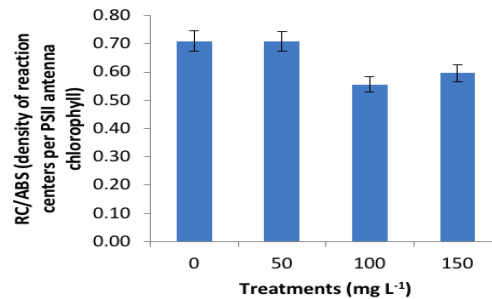
636

637

638

639

640



641 **Fig. 15. The impact of magnetic nanoparticles on RC/ABS (density of reaction centers**
642 **per PSII antenna chlorophyll) of *Ipomoea aquatica*.**

643 **3.15 Total phenolics content**

644 Based on Fig. 16, different concentration of Fe₃O₄ had influenced significantly on the
645 phenolics content of water spinach ($P \leq 0.05$). As the concentration of magnetic nanoparticles
646 increases, the total phenolics content in the plant increased too. The lowest total phenolics
647 content was observed in 0 mg L⁻¹ iron oxide nanoparticles at 1.38 mg GAE/g dry weight and
648 the highest for 150 mg L⁻¹ treatment at 2.29 mg GAE/g dry weight. The greater production of
649 phenolics at higher concentration of magnetic nanoparticles may be a sign of the plant under
650 stress. The production of secondary metabolites is an effort of the plant to defend itself from
651 the further damage of plant cell and ensure the survival of plant. The higher production of
652 phenolic stimulates antioxidant activity in the plant [44]. In order to the response of a plant
653 towards environmental stresses and protecting itself from damages in plant cell, the
654 production of phenolic compounds is essential to maintain the plant growth and reproduction
655 [45]. Based on the study by previous researchers, phenolics compounds do contain
656 antibiotic and anti-nutritional properties that help in the defense system of a plant. Phenolics
657 compound usually stored in the epidermal cells of leaves and shoots besides central vacuole
658 of guard cells. Phenolics content can be divided into two groups which are preformed
659 phenolics and induced phenolics by which preformed phenolics are synthesized by the plant
660 during development of plant tissues under normal condition. While, induced phenolics are
661 produced when the plant faces physical injury, infection or environmental stresses due to
662 heavy metal irradiation or temperature [14].

663

664

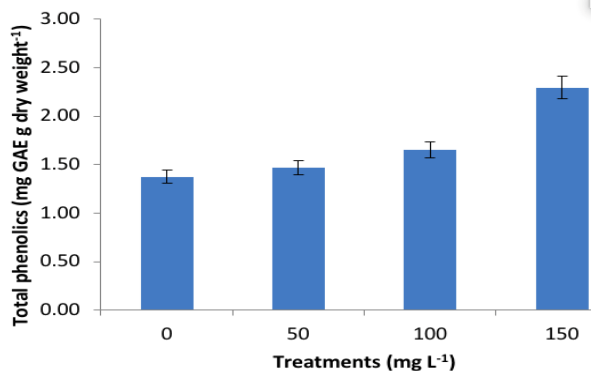
665

666

667

668

669

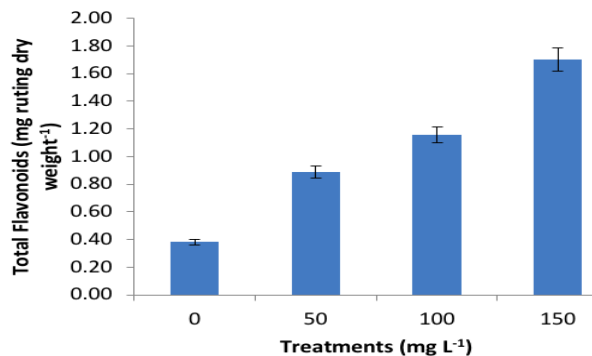


670 **Fig. 16. The impact of magnetic nanoparticles on total phenolics content of *Ipomoea***
671 ***aquatica*. Data are mean ± standard error of mean (SEM). N= 24.**

672 **3.16 Total flavonoids content**

673 Based on Fig. 17, the production of total flavonoid content was significantly influenced by the
674 application of magnetic nanoparticles at different concentrations of Fe₃O₄ ($P \leq 0.05$). At the
675 end of experiment of 12th week, the total flavonoid content in water spinach treated with 150
676 mg L⁻¹ Fe₃O₄ showed the highest reading at 1.70 mg rutin/g dry weight followed by 100 mg L⁻¹
677 treatment at 1.16 mg rutin/g dry weight, 50 mg L⁻¹ treatment at 0.89 mg rutin/g dry weight
678 and lastly 0 mg L⁻¹ treatment at 0.38 mg rutin/g dry weight. From the result, it was observed
679 that higher concentration of Fe₃O₄ increase the production of flavonoid in water spinach. The
680 production of flavonoid is one of the determinants for the production of secondary metabolite
681 in plant. It is oftenly observed that plant undergoing stress has higher total flavonoid content
682 due to defensive mechanism in plant. Signaling in plant to produce secondary metabolites is
683 usually stimulated by the accumulation of heavy metals like zinc, iron, and nickel which
684 generates Reactive Oxygen Species (ROS) and induces oxidative stress in plant.

685 Consequently, the induction of oxidative stress causes changes in signal transduction for the
686 mechanism of gene coding and enzyme [13]. Besides, the production of ROS may cause
687 damages to cell membrane, cell structure and photosynthetic site and thus the production of
688 flavonoid at the generation site act as defensive mechanism due to its high antioxidant
689 properties [14].



703 **Fig. 17. The impact of magnetic nanoparticles on total flavonoids content of *Ipomoea***
704 ***aquatica*. Data are mean \pm standard error of mean (SEM). N= 24.**

705

706 **4. CONCLUSION**

707

708 Overall, it was found that the application of magnetic nanoparticles which was in the form of
709 iron oxide nanoparticles had influence on growth of *Ipomoea aquatica*. The plant height,
710 plant temperature, total biomass and total chlorophyll content were significantly affected by
711 the application of magnetic nanoparticles at higher concentration. Besides that, the leaf gas
712 exchange characteristics were also influenced by the different concentrations of iron oxide
713 nanoparticles. The net photosynthesis rate, transpiration rate, stomata conductance,
714 maximum efficiency of photosystem II (Fv/fm), maximum quantum yield of phytochemical
715 and non-photochemical process in photosystem II (Fv/fo), performance index and the
716 density of reaction centers per PSII antenna chlorophyll of *I. aquatica* were significantly
717 reduced at higher concentration of magnetic nanoparticles. While, water use efficiency and
718 minimal fluorescence value of *I. aquatica* were increased with the increased of iron oxide
719 nanoparticles concentration. In addition, the application of magnetic nanoparticles had
720 significantly influenced the total flavonoids and total phenolics content in water spinach

721

722 **5. ACKNOWLEDGMENT**

723

724 The authors would like to thank the Ministry of Higher Education Malaysia for The
725 Fundamental Research Grant Scheme (FRGS) and the Research Management Centre,
726 Universiti Putra Malaysia (UPM) for sponsoring this work.

727

728

729 **REFERENCES**

730

- 1 Salaries, T. (2016). Department of Statistics Malaysia Press Release, (May), 1–4.
- 2 Department of Statistics Malaysia Press Release. (2017). Press Release Supply and Utilization Accounts Selected Agricultural Commodities, Malaysia 2012-2016, (December), 1–4.

- 3 Department of Statistics Malaysia Press Release. (2018), (February), 4–6. Retrieved from <https://www.dosm.gov.my/v1/index.php?r=column/pdfPrev&id=TkpmM05EK3NBV0JRU1pmOUJnS3RCQT09> on September 15, 2018.
- 4 Ibrahim, M. H., Yasmin, N., Rahman, A., Amalina, N., & Zain, M. (2018). Effect of Nitrogen Rates on Growth and Quality of Water Spinach (*Ipomea aquatica*), *Annual Research & Review in Biology*, 26(1), 1–12. <https://doi.org/10.9734/ARRB/2018/40352>
- 5 Dua, T. K., Dewanjee, S., Gangopadhyay, M., Khanra, R., Zia-Ul-Haq, M., & De Feo, V. (2015). Ameliorative effect of water spinach, *Ipomea aquatica* (Convolvulaceae), against experimentally induced arsenic toxicity. *Journal of Translational Medicine*, 13(1), 1–17. <https://doi.org/10.1186/s12967-015-0430-3>
- 6 Xiao, Q., Wong, M. H., Huang, L., & Ye, Z. (2015). Effects of cultivars and water management on cadmium accumulation in water spinach (*Ipomoea aquatica* Forsk.). *Plant and Soil*, 391(1–2), 33–49. <https://doi.org/10.1007/s11104-015-24095>
- 7 Markides, H., Rotherham, M., & El Haj, A. J. (2012). Biocompatibility and toxicity of magnetic nanoparticles in regenerative medicine. *Journal of Nanomaterials*, 13–15. <https://doi.org/10.1155/2012/614094>
- 8 Lu, A. H., Salabas, E. L., & Schüth, F. (2007). Magnetic nanoparticles: Synthesis, protection, functionalization, and application. *Angewandte Chemie - International Edition*, 46(8), 1222–1244, <https://doi.org/10.1002/anie.200602866>
- 9 Nair, R., Varghese, S. H., Nair, B. G., Maekawa, T., Yoshida, Y., & Kumar, D. S. (2010). Nanoparticulate material delivery to plants. *Plant Science*, 179(3), 154–163. <https://doi.org/10.1016/j.plantsci.2010.04.012>
- 10 Da Silva, L.C., Oliva, M.A., Azevedo, A.A., De Araujo, J.M. (2006). Responses of resting a plant species to pollution from an iron pelletization factory, *Water Air Soil Pollut.* 175, 241–256
- 11 Lin, C., Fugetsu, B., Su, Y., & Watari, F. (2009). Studies on toxicity of multi walled carbon nanotubes on Arabidopsis T87 suspension cells. *J. Hazard. Mat.* 170, 578-583.
- 12 Tiwari, D.K., Dasgupta-Schubert, N., Villasenor, L.M., Tripathi, D., & Villegas, J. (2013). Interaction of carbon nanotubes with mineral nutrients for the promotion of growth of tomato seedlings. *Nano Stud.* 7, 87–96.
- 13 Zhao, J., Davis, L. C., & Verpoorte, R. (2005). Elicitor signal transduction leading to production of plant secondary metabolites, *Biotechnology Advances*, 23, 283–333. <https://doi.org/10.1016/j.biotechadv.2005.01.003>
- 14 Izad, A. I., Ibrahim, M. H., Azurahaman, C., Abdullah, C., Amalina, N., & Zain, M. (2018). Growth , Leaf Gas Exchange and Secondary Metabolites of *Orthosiphon stamineus* as Affected by Multiwall Carbon Nanotubes Application, *Annual Research & Review in Biology*, 23, 1–13. <https://doi.org/10.9734/ARRB/2018/38113>
- 15 Karlsson, H.L., Cronholm, P., Gustafsson, J., & Möller, L. (2008). Copper oxide nanoparticles are highly toxic: a comparison between metal oxide nanoparticles and carbon nanotubes. *Chem Res Toxicol.* 21(9), 1726-32.
- 16 Martínez-Fernández, D., Barroso, D., & Komárek, M. (2016). Root water transport of *Helianthus annuus* L. under iron oxide nanoparticle exposure. *Environmental Science and Pollution Research*, 23(2), 1732–1741
- 17 Liu, R., Zhang, H., & Lal, R. (2016). Effects of stabilized nanoparticles of copper, zinc, manganese, and iron oxides in low concentrations on lettuce (*Lactuca sativa*) seed germination: nanotoxicants or nanonutrients?. *Water, Air, & Soil Pollution*, 227(1), 42.
- 18 Ruttkay-Nedecky, B., Krystofova, O., Nejdil, L., & Adam, V. (2017). Nanoparticles based on essential metals and their phytotoxicity. *Journal of Nanobiotechnology*, 15(1), 119. <https://doi.org/10.1186/s12951-017-0268-3>
- 19 Singh, S., Singh, B.K., Yadav, S.M., & Gupta, A.K. (2015). Applications of Nanotechnology in Agricultural and their Role in Disease Management. *Research Journal of Nanoscience and Nanotechnology*, 5(1), 1-5.
- 20 Mahdavi, M., Ahmad, M. B., Haron, M. J., Namvar, F., Nadi, B., Rahman, M. Z. A., &

- Amin, J. (2013). Synthesis, surface modification and characterisation of biocompatible magnetic iron oxide nanoparticles for biomedical applications. *Molecules*, 18(7), 7533–7548.
- 21 Ibrahim, M. H., Jaafar, H. Z., Rahmat, A., & Rahman, Z. A. (2011). The relationship between phenolics and flavonoids production with total non structural carbohydrate and photosynthetic rate in *Labisia pumila* Benth. under high CO₂ and nitrogen fertilization. *Molecules*, 16(1), 162–174.
 - 22 Becker, M., & Asch, F. (2005). Iron toxicity in rice conditions and management concepts, *Journal of Plant Nutrition Soil Science*, 168, 558–573. <https://doi.org/10.1002/jpln.200520504>
 - 23 Rui, M., Ma, C., Hao, Y., Guo, J., Rui, Y., Tang, X., & Zhu, S. (2016). Iron Oxide Nanoparticles as a Potential Iron Fertilizer for Peanut (*Arachis hypogaea*). *Frontiers in Plant Science*, 7(June), 1–10. <https://doi.org/10.3389/fpls.2016.00815>
 - 24 Dhoke, S. K., Mahajan, P., Kamble, R., & Khanna, A. (2013). Effect of nanoparticles suspension on the growth of mung (*Vigna radiata*) seedlings by foliar spray method. *Nanotechnology Development*, 3(1). <https://doi.org/10.4081/nd.2013.el>
 - 25 Chrispeels, M. J., & Agre, P. (1994). Aquaporins : water channel proteins of plant and animal cells, *Elsevier Science*, 8073–8077.
 - 26 Pressure, T., Kirkham, M. B., Gardner, W. R., & Gerloff, G. C. (1972). Regulation of Cell Division and Cell Enlargement, *Plant Physiology*, 49, 961–962.
 - 27 Sheykhbaglou, R., & Sedghi, M. (2010). Effects of Nano-Iron Oxide Particles on Agronomic Traits of Soybean, *Not Sci Biol*, 2(2), 112–113.
 - 28 Jeyasubramanian, K., Gopalakrishnan Thoppey, U. U., Hikku, G. S., Selvakumar, N. Subramania, A., & Krishnamoorthy, K. (2016). Enhancement in growth rate and productivity of spinach grown in hydroponics with iron oxide nanoparticles. *RSC Advances*, 6(19), 15451–15459. <https://doi.org/10.1039/C5RA23425E>
 - 29 Du, W., Tan, W., Peralta-vidua, J. R., & Gardea-torresdey, J. L. (2016). University of California Center for Environmental Implications of Nanotechnology (UC. *Plant Physiology and Biochemistry*. <https://doi.org/10.1016/j.plaphy.2016.04.024>
 - 30 Gates, D.M. (1968). Transpiration and leaf temperature. *Plant Physiology*, 19, 211–238.
 - 31 Racuciu M, Creanga D, Olteanu Z (2009). Water based magnetic fluid impact on young plants growing. *Rom. Rep. Phys.*, 61 (2): 259–268.
 - 32 Britt, D. W., Johnson, W. P., Boyanov, M. I., & Anderson, A. J. (2012). CuO and ZnO nanoparticles : phytotoxicity , metal speciation , and induction of oxidative stress in sand-grown wheat, *Journal of Nanoparticle Research*, 14, 1125. <https://doi.org/10.1007/s11051-012-1125-9>
 - 33 Li, J., Chang, P. R., Huang, J., Wang, Y., Yuan, H., & Ren, H. (2013). Physiological Effects of Magnetic Iron Oxide Nanoparticles Towards Watermelon, *Journal of Nanoscience and Nanotechnology*, 13(8), 5561–5567 <https://doi.org/10.1166/jnn.2013.7533>
 - 34 Shweta, Tripathi, D.K., Shweta, S., Swati, S., Dubey, N.K., & Chauhan, D.K. (2016). Impact of Nanoparticles on Photosynthesis: Challenges and Opportunities, *American Scientific Publishers*, 5(5), 404–411. <https://doi.org/10.1166/mat.2016.1327>
 - 35 Rizwan, M., Ali, S., Farooq, M., Sik, Y., Adrees, M., Ibrahim, M., & Abbas, F. (2017). Effect of metal and metal oxide nanoparticles on growth and physiology of globally important food crops: A critical review. *Journal of Hazardous Materials*, 322, 2–16. <https://doi.org/10.1016/j.jhazmat.2016.05.061>
 - 36 Blum, A. (2009). Field Crops Research Effective use of water (EUW) and not water-use efficiency (WUE) is the target of crop yield improvement under drought stress, *Field Crop Research*, 112, 119–123. <https://doi.org/10.1016/j.fcr.2009.03.009>
 - 37 Nurfarahin S. (2017). Growth, Leaf Gas Exchange and Secondary Metabolites of Water Spinach (*Ipomoea aquatica*) as Affected by Carbon Nanotubes Application, Universiti Putra Malaysia, Malaysia.
 - 38 Maxwell, K., & Johnson, G. N. (2000). Chlorophyll fluorescence -a practical guide, *Journal of Experimental Botany*, 51(345), 659–668.

- 39 Gao, J., Xu, G., Qian, H., Liu, P., Zhao, P., & Hu, Y. (2013). Effects of nano-TiO₂ on photosynthetic characteristics of *Ulmus elongata* seedlings. *Environmental pollution*, 176, 63-70.
- 40 Qi, M., Liu, Y., & Li, T. (2013). Nano-TiO₂ Improve the Photosynthesis of Tomato Leaves under Mild Heat Stress, *Biology Trace Element Research*, 156, 323–328. <https://doi.org/10.1007/s12011-013-9833-2>
- 41 Mehta, P., Jajoo, A., Mathur, S., & Bharti, S. (2010). Plant Physiology and Biochemistry Chlorophyll a fluorescence study revealing effects of high salt stress on Photosystem II in wheat leaves. *Plant Physiology et Biochemistry*, 48(1), 16–20. <https://doi.org/10.1016/j.plaphy.2009.10.006>
- 42 Živčák, M., Brestič, M., Olšovská, K., & Slamka, P. (2008). Performance index as a sensitive indicator of water stress in *Triticum aestivum* L., *Plant Soil Environment*, 54(4), 133–139.
- 43 Appenroth, K., & Sto, J. (2001). Multiple effects of chromate on the photosynthetic apparatus of *Spirodela polyrhiza* as probed by OJIP chlorophyll a fluorescence measurements, *Environmental pollution*, 115, 49-64
- 44 Kim, D., Weon, S., & Lee, C. Y. (2003). Antioxidant capacity of phenolic phytochemicals from various cultivars of plums, *Food Chemistry*, 81, 321–326.
- 45 Subbiah R., Veerapandian M., & Yun K.S. (2010). Nanoparticles: Functionalization and multifunctional applications in biomedical sciences. *Curr. Med. Chem*, 17, 4559–4577.

731
732
733
734
735
736
737