1	Original Research Article	
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3	Determination of Minerals, Total Phenolic	
4	Content, Flavonoids, Antioxidants and	
5	Antimicrobial Activities of Ethanolic Extract of	
6	sweet Lupinus Angustifolius of Palestine	
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11 12	ABSTRACT	
[Aims: To establish the most suitable extraction method for sweet lupine seeds and to	
	determine minerals, phenolic content, flavonoids, antioxidant activity and antimicrobial activities.	
	Study design: Known and standard experimental procedures are employed.	
	from January 2019 to March 2019.	
	Methodology: Seeds were ground and extracted by Soxhlet extractor using ethanol with	
	content were determined. Resistance to bacteria was performed against <i>Escherichia coli</i> and	
	Staphylococcus aureus, while antioxidant activity was determined by FRAP method. Two	
	types of flavonoids were measured: flavonones and dihydroflavonois via the reaction with 2.4-dinitrophenylhydrazine. Phenolics were determined by the Folin-Ciocalteu method.	
	Results: 50% ethanol resulted in the highest extract residue (18.6%) while 70% and 60%	
	showed the lowest content (10.0% for both). 80% ethanol extracted sample showed the bighest content for sodium (56.51 mg Na/g extract) while 60% and 50% ethanol extracts	
	showed the highest content of potassium (2.25 and 2.33mg K/g extract, respectively). The	
	maximum concentration of ferrous ion was obtained with 70% ethanol (6.854mg Fe^{+2}/g	
	extract). Similar results were obtained for total phenolic content and flavonoids: 24.60 mg	
	gallic acid/g extract for phenolics and 116.02 mg rutin/g extract for flavonoids. Extracts	
	Conclusion: 95% ethanol extracted samples showed the highest antioxidant activity and the	
	highest flavonoids and phenolic content. Sweet lupine extract did not perform any	
13	antimicrobial activity against both Gram positive and Gram negative bacteria.	
14	Keywords: Sweet lupine, Soxhlet extractor, Minerals, Total phenolics, Flavonoids,	
15 16	Antimicrobial Activity, Antioxidant Activity.	
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21 22	1. INTRODUCTION	
23	Sweet Lupinus angustifolius, also called "narrow-leafed lupine" is a member of the legume	
24 25	family (subfamily Papilionoideae) containing both herbaceous annual and shrubby perennial types with attractive long racemes of flowers [1]. There are twelve luning species within the	
	speet man analytic long recentle of netters [1]. There are menter upine species within the	

Lupinus genus, all of which are native to Europe and the Mediterranean regions. Sweet lupine is widely cultivated in Australia, the color of its flower varies from blue, to pink and white in demonstrated forms [2, 3]. Lupinus angustifolius is one of the four lupines that are widely known and fully domesticated for agriculture purposes (Lupinus albus, Lupinus angustifolius, Lupinus luteu and Lupinus mutablis).

31 For several years, lupine flour has been used in pasta, milk, soya substituents and diet products. Lupine seeds are also eaten as snacks in most regions of the world [4]. Lupine 32 seeds can contain toxicologically relevant bitter quinolizidine alkaloids, which cause 33 34 symptoms of poisoning of humans affecting the nervous, circulatory and digestive systems [5]. Typical symptoms of lupine alkaloid poisoning are dizziness, confusion, tachycardia, 35 nausea and dry mouth, loss of motor coordination and in high doses, cardiac arrest and 36 37 respiratory paralysis [5]. The levels of quinolizidine alkaloids in lupine seeds vary depending on the botanical and geographical origin of the lupine variety from which they derive. In 38 contrast to bitter lupine, sweet lupine has low level of toxic alkaloid and suitable for human 39 40 consumption even without debittering [6].

Lupine seeds, like other legumes are sources of vitamin, protein and fibers. Studies reported the pharmacological benefits of lupine alkaloids, with activity on circulatory system, metabolism against obesity and improving bowel health [7].

Due to the low concentration of biologically active materials in plants, it is necessary to use effective methods for extraction of these substances, specially using solvents that are environmentally friendly. Consequently, ethanol was the solvent of choice with different percentages to extract phenolics and flavonoids, which are responsible for the pharmacological properties such as antioxidants and antimicrobials. Therefore, a comprehensive determination of lupine properties is essential, not only because of its potential toxicity to humans, but also for its pharmacological properties.

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53 2. MATERIAL AND EXPERIMENTAL DETAILS

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55 2.1. Raw Materials and Equipment

56 Sweet lupine seeds were obtained from the local market, while reagents/chemicals were 57 purchased from Sigma-Aldrich. Deionized water was used in all preparations, and 58 commercial ethanol was used for extraction. An Analytik Jena Specord 40 UV-VIS 59 spectrophotometer was used for the determination of the antioxidant activity, phenolic 60 content and flavonoids. A model FP 640 flame photometer was used for the measurements 61 of sodium and potassium content. Bacteria strains were provided from Holy Family Hospital 62 in Bethlehem-Palestine.

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64 **2.2. Extraction of Seeds**

Lupine seeds were ground and extracted by Soxhlet extractor using different percentages of ethanol (50%, 60%, 70%, 80% and 95%) for three hours. The solvent was evaporated under vacuum and the residue was stored in refrigerator away from direct light.

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69 2.3. Stock Solution

Residue was dissolved in 50% ethanol (200 mg/100 mL) and this served as stock solution
 for the determination of sodium, potassium, ferrous ion, antioxidant activity, total phenolic
 content and flavonoids.

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74 **2.4. Determination of Sodium and Potassium**

Sodium and potassium were determined by flame photometry against reference standards
 for both elements. From the calibration curves, the concentration of the extracted samples
 was determined.

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79 2.5. Determination of Ferrous Ion (Fe⁺²)

80 Fe⁺² in sample extract was determined by a titrimetric method: redox titration of Fe⁺² with 81 potassium dichromate using sodium diphenylamine sulfonate, a pH independent redox 82 indicator. Endpoint was detected as the color turned to violet.

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84 **2.6. Determination of Antimicrobial Activity**

Antibacterial activity was studied on sweet lupine against *S. aureus* (Gram positive) and *E. coli* (Gram negative) bacteria. An "Agar Well" method was used to test the resistance of extract to bacteria [8]. In this method, three wells were created in the Agar plates of the Muller-Hinton broth [9]: the first of which was for negative control (H₂O), the second was for positive control (Amoxicillin), and the third one was for sample (the extract). High concentrations of extracts (1.2 g/100 mL) were used for the determination of antibacterial activity. Petri dishes were incubated at 37°C for 24-48 hours.

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93 **2.7. Antioxidant Activity**

The antioxidant activity was determined by the ferric reducing antioxidant power (FRAP) [10] method that relies on reduction by antioxidants of the complex ferric ion-TPTZ (2,4,6-tri (2pyridyl)-1,3,5-triazine). The binding of Fe^{+2} to the ligand makes a complex that gives the blue color intensity. The absorbance was measured to test the concentration of iron reduced, which is correlated with the concentration of antioxidant.

99 100 **2.7.1. Analysis**

For sample extract: 800 μ L of sample (Stock solution) was mixed with 1000 μ L FRAP, and for standard: 80 μ L of standard FeSO₄ (0.1–2.0 mM) was mixed with 1000 μ L H₂O and 1000 μ L FRAP. Solutions were incubated at 37 °C for 15 minutes and the absorbance of the colored product was measured at λ =593 nm against 50% ethanol as blank.

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106 **2.8. Total Phenolics Content**

The total concentration of phenolic compounds was determined using Folin-Ciocalteu
method [11, 12].

110 **2.8.1. Analysis**

For sample extract, 1.20 mL of 7.5% Na₂CO₃ was mixed with 100 μ L sample and 1.8 mL diluted Folin- Ciocalteu reagent (1:1). Standard preparation was done as the follows, 1.20 mL Na₂CO₃ was mixed with 40 μ L standard Gallic acid (90-900 ppm) and diluted Folin-Ciocalteu reagent (1:1). The mixtures were incubated for one hour at 30 °C where the sample was turned to greenish-blue, and absorbance was measured at λ =765 nm.

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117 2.9. Flavonoids

118 The colorimetric identification and quantification of the two types of flavonoids (flavonones 119 and dihydroflavonols) was based on their reaction with 2,4-dinitrophenylhydrazine (DNP) in 120 the presence of KOH in methanol [13,14].

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122 **2.9.1. Analysis**

For sample extract and standard (rutin, 5 – 100 ppm), 200 μ L of stock solution was mixed with 400 μ L 2,4-dinitrophenylhydrazine and placed in a water bath at 50 °C for 60 minutes. After cooling to room temperature, 800 μ L of a 10% KOH/methanol solution was added to the mixture, where after 350 μ L of the total mixture was diluted to 5.0 mL with 100% methanol. Absorbance was measured at λ =486 nm using a UV-VIS spectrophotometer.

128 **3. RESULTS AND DISCUSSION**

129 **3.1. Extraction**

Lupine seeds were extracted with different percentages of ethanol. Results are summarized in table 1. As shown, the highest percentage of extract was obtained when 50% ethanol was used (18.6%). On the other hand, the lowest percentage was obtained when 60% and 70% ethanol were used (10.0% for both).

134

135 Table 1. Percentages of residue obtained from sweet lupine seeds

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Solvent	Result	
95% EtOH	12.2%	
80% EtOH	10.9%	
70% EtOH	10.0%	
60% EtOH	10.0%	
50% EtOH	18.6%	

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138 **3.2. Determination of Sodium and Potassium**

Results of sodium and potassium are illustrated in table 2. The highest concentration of 139 sodium was obtained when 80% of ethanol was used while the lowest concentration was 140 141 obtained with 50% ethanol. This can be attributed to the fact that sodium is present in sweet lupine as organic salts that tends to dissolve in ethanol more than in water. In a previous 142 143 study on Lupinus albus seeds [15], the highest concentration of sodium was obtained with 144 50% ethanol suggesting that sodium is present as inorganic complexes in the seeds. The 145 highest concentration of potassium in sweet lupine was obtained when 50% and 60% 146 ethanol were used. This result is in agreement with results reported by Hanania et.al. (2018) 147 where bitter lupine seeds were extracted with 60% ethanol, which resulted in highest 148 potassium concentrations [15].

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150 **3.3. Determination of Ferrous lons**

Table 2 also shows that as the percentage of ethanol decreases, the ferrous content
increases until the 70% ethanol extraction, where the maximum content of ferrous was
extracted. However, below 70% ethanol, the ferrous content decreases.

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155 Table 2. Sodium, potassium and ferrous content of extracts (mg/g)

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Ethanol %	Sodium	Potassium	Ferrous	
95%	10.29	0.15	3.726	
80%	56.51	1.00	4.340	
70%	17.59	0.6	6.854	
60%	10.51	2.25	2.424	
50%	9.20	2.33	1.839	

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158 **3.4. Antimicrobial Activity**

Sweet lupine extract showed no inhibition against neither *E. coli* nor *S. aureus* bacteria. The negative results reported here are in agreement with previous studies in terms of *E. coli*, but it does not agree with the results of the study on *S. aureus*, where significant activity was observed [16, 17]. The extract of *Lupinus angustifolius* was weakly active on *E. coli*.

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164 **3.5. Determination of Antioxidant Activity and Total Phenolics Content**

As illustrated in table 3, the highest activity of antioxidants was obtained when 95% ethanol was used. Similar results were obtained for phenolics which is an important antioxidant as phytochemical in sweet lupine seeds. This result was expected since antioxidants such as phenolics are organic compounds that tend to dissolve in ethanol rather than water [18]. Our

- 169 results showed higher content of phenolics and similar antioxidant activity to those reported
- in literature [19, 20].

Ethanol was used in accordance with the literature data, to ensure optimum extraction of phenols, because the extraction efficiency of plant material using ethanolic-water is greater and environmentally friendly than methanolic-water extraction [21, 22]. Compared to bitter lupine, it was found that bitter seeds have a higher antioxidant activity since it contains a

175 higher content of phenols [15].

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177 Table 3. Antioxidant Activity and total Phenolics for sweet lupine extracts

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Ethanol %	mg FeSO₄/g extract	mg Gallic acid/g extract	_
95%	20.24	24.60	
80%	19.22	20.98	
70%	12.03	18.35	
60%	9.15	11.92	
50%	7.23	12.28	

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181 **3.6. Determination of Flavonoids Content**

Flavonones and dihydroflavones are the two types of flavonoids that were determined in sweet lupine. As illustrated in table 4, 95% ethanolic extract resulted in the highest concentrations of flavonoids i.e. 115.02 mg rutin/g extract. It is worth mentioning that the concentration of these bioactive chemicals depends on many factors including climate, precipitation and soil conditions [23].

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188 Table 4. Rutin (flavonoids) concentrations obtained from for different percentage

189 ethanol extraction

1	$\Omega \Omega$	
L	90	

Ethanol %	mg Rutin/g
	extract
95%	115.02
80%	11.77
70%	35.19
60%	22.56
50%	39.83

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193 4. CONCLUSION

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195 Based on the results, antioxidants present in sweet lupine are organic compounds and are more likely to dissolve in ethanol than in water. Sodium ion was shown to be present in high 196 197 percentages, especially in 80% ethanolic extract. Potassium, on the other hand, showed 198 high concentration when extracted with 60% ethanol. It was found that sweet lupine has 199 higher ferrous ion concentrationm than bitter lupine. Moreover, phenolics and flavonoids 200 have many biological properties in plant especially as antioxidants, while antibacterial agents 201 are absent from sweet lupine seeds. Although 50% ethanol was the highest percentage of 202 extracted content (residue), yet it may have inorganic compounds or compounds with no 203 biological effect to bacteria or oxidation reactions.

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205 COMPETING INTERESTS

206 Authors have declared that no competing interests exist.

207 208

209 CONSENT

210 Not applicable

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