

## Original Research Article

### **Phytoremediation of Heavy Metals from Water of Yamuna River by *Tagetes patula*, *Bassica scoparia*, *Portulaca grandiflora***

#### **Abstract**

Heavy metal contamination is a worldwide problem, causing many serious diseases and the levels of contamination varied from place to place. Heavy metals like cadmium (Cd), mercury (Hg), zinc (Zn), chromium (Cr), and lead (Pb) etc. are very injurious even at low concentration and are present in Yamuna river water. Phytoremediation has great potential as an efficient cleanup technology for contaminated soils, groundwater, and wastewater. It is a cheap and very efficient technique for metal removal. A study had been carried out to detect the efficiency of phytoremediation technique for removal of heavy toxic metals from water of Yamuna river. This study also focused on the phytoremediation capacity of all of three selected plants: *Tagetes patula*, *Bassica scoparia*, and *Portulaca grandiflora*. Bioaccumulation of heavy metals in various parts of plants has also been checked.

**Keywords**–: Yamuna river, *Tagetes patula*, *Bassica scoparia*, *Portulaca grandiflora*, Heavy metals, Phytoremediation.

#### **1. INTRODUCTION**

Yamuna river originates from Yamunotri glaciers of Himalayas. It is the largest tributary of river Ganga. It is around 1370 kilometers in length. It flows across the states of Haryana, Delhi, Uttar Pradesh. It merges into Ganga river in Allahabad. Big cities like Mathura, Agra, Delhi lie on the

28 Yamuna river bank. It is classified into five segments like Delhi segment, Upper segment,  
29 Himalayan segment, Eutrophicated segment, Diluted segment depend on the basis of ecological  
30 and hydrological conditions. The quality of water, river water in Himalayan segment is very  
31 good and also meets all the standards within this segment. Yamuna river water is trapped by a  
32 Wazirabad barrage for the purpose of domestic supply of water in Delhi. The Okhla barrage of  
33 Delhi receives the water of seventeen drain sewage, Najafgarh drain. It is the most polluted  
34 segment of river Yamuna. Today it has become the most polluted and dirtiest river of the country  
35 and was once described as the lifeline of Delhi city.

36 It has been given the grade "E" by the Central Pollution Control Board (CPCB), which means it  
37 is only good for recreation and industrial cooling. No underwater life found in this segment of  
38 the river. The domestic discharges from Delhi, Faridabad, Noida, Ghaziabad, Mathura, Agra,  
39 Haryana, has rendered the river unfit for any use.

40 Even taking a dip in river water can cause various health and skin regarding issues. One of the  
41 major contaminants present in river water is toxic heavy metals. Presence of toxic heavy metals  
42 is an issue of major concern because of bio-accumulative nature of metals. These metals have  
43 geological origin, but entering into the river water can be by erosion, weathering and  
44 anthropogenic activities of human beings like agricultural runoff, industrial processing, sewage  
45 disposal etc. Environmental related exposure of these heavy metals are like lead paint, household  
46 dust, silver foil in food, surface soil, batteries, peeling paints, sewage wastes, plumbing system  
47 etc. Use of fertilizers and pesticides is also a great source of heavy metals like Cd, As. Some of  
48 these metals are essential for human beings, but in very low concentration, such as Ca, Cu, Fe,  
49 Cr, Mg, K, Zn, Ni, Mn, Co and Na are essential for normal growth of plants and living  
50 organisms. Cd, Ag, Al, Pb are some non essential metals and are very toxic.

51 High uptake and slow elimination of Heavy metals cause harm to the aquatic life. As the heavy  
52 metals get settled down in the sediment and uptake by the plants or aquatic organisms, drink by  
53 the animal and this will ultimately harm the life of organisms (Ghosh et al, 2015; 2016; 2017).  
54 Human by many ways are highly exposed to heavy metals as they are also the part of the food

55 chain. Table 1 shows the permissible limit of heavy metals (Ad, Zn, Cr, Pb, Hg) prescribed by  
56 WHO.

57

58 **Table 1: Maximum permissible value of heavy metals by WHO**

Metals	Water (L/kg <sup>-1</sup> )	Sediment (µg/kg <sup>-1</sup> )
Cadmium	0.003	6
Zinc	3	123
Chromium	0.05	25
Lead	0.01	----
Mercury	1.3	0.3

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60 High uptake of lead causes changes in the gill, kidney and liver of fish. Intestine and gills are the  
61 major site of metal accumulation in fishes. It causes variation in the lipids of aquatic organisms.

62 Lead cause swelling in the gills and jaws of fishes. Nausea, anemia and vomiting, etc problems  
63 are the side effects of lead exposure in humans.

64 Zinc accumulates in the gills of fish, this indicate a depressive effect in tissue respiration cause  
65 hypoxia or death of the fish. Zinc also causes a decrease in total white blood cells. Zinc cause  
66 changes in heart physiology and also cause toxic changes in ventilatory System. Headache,  
67 fever, vomiting, chest tightness, aches, chills, metallic taste in the mouth And cough are the side  
68 effects of acute exposure to zinc. Chronic exposure causes problems like cancer, kidney and lung  
69 failure.

70 Cadmium mostly accumulates in the gills, intestine and stomach of fishes. It causes changes in  
71 enzymatic activities in marine animals and also changes in oxygen consumption. High  
72 concentration of cadmium also affects the osmotic-regulation activity in fishes. Cadmium also  
73 causes reduction in red blood cells in the fishes. Exposure to heavy metals causes various serious  
74 diseases in human beings. Cadmium exposure cause lung inflammation and lung cancer as  
75 | cigarette smoking is the largest source of cadmium [In-in](#) humans. Osteomalacia -and proteinuria

76 are the kind of problems occur in humans due to cadmium.

77 Chromium cause acute and chronic effects on fishes. High chromium Uptake causes changes in  
78 metallo-enzymatic activity. Chromium gets accumulated in the gills of aquatic biota. High  
79 chromium concentration cause altered blood chemistry, osmoregulatory changes, behavioral  
80 modifications and in severe conditions hypoxia. Acute renal failure, hemolysis and  
81 gastrointestinal ~~hemorrhage~~ ~~hamorrhage~~ are the problems occur in humans at acute exposure to  
82 chromium. At chronic exposure to Chromium lungs cancer and pulmonary fibrosis diseases will  
83 take place.

84 Mercury is highly toxic to aquatic animals. It shows variable effects on oxygen consumption,  
85 osmoregulation, and enzyme activity of marine life. It also shows several effects on blood  
86 circulation system and causes a reduction in RBC count. ~~Diarrhea~~ ~~Diarrheoa~~, fever and vomiting  
87 are the side effects of acute mercury exposure. Nausea, nephrotic syndrome, pink disease,  
88 stomatitis, neurotic disorders and tremor diseases are the side effects of cadmium at chronic  
89 exposure as mercury is highly toxic.

90 Various techniques are available for remediation of contaminants. **Which are chemical, physical**  
91 **and biological methods.** The chemical method involves the use of several harsh chemicals like  
92 leaching of metals by chelating agents and chemical wash. Physical methods are very expensive  
93 and cause labour demand. That's why researchers have developed highly efficient, cost effective,  
94 eco-friendly remediation techniques, in which organic waste are biologically degraded into an  
95 innocuous state.

96 Removal of heavy metals with the help of microorganisms is a very efficient method, but it is  
97 confined to water system only. Some other remediation methods are bio augmentation, land  
98 farming, bio leaching, rhizofiltration, biostimulation, composting, bioreactor, and  
99 phytoremediation. Phytoremediation is a technique that uses plants for degradation or  
100 accumulation of toxic contaminants present in environment (Ali et al., 2013; Mahar et al., 2016;  
101 Ullah et al., 2015; Mani and Kumar, 2014; Tauqeer et al., 2016; Sarwar et al., 2017). It involves  
102 the use of living organisms, especially plants and microorganisms to eliminate the effects of  
103 contaminants present in air, water, soil (Ganjo and Khwakaram, 2010; Tiwari et al., 2008;  
104 Bhardwaj et al., 2017; Ahuja et al., 2011; Saha et al., 2017; Mojiri et al., 2013; Farraji, 2014).

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105 Phytoextraction of heavy metals by the hyperaccumulator plants from both soil and water is also  
106 a key area of search. This study was also focused on the phytoremediation capacity of all of three  
107 selected plants *Tagetes patula*, *Bassica scoparia*, *Portulaca grandiflora*.

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## 109 1.2 OBJECTIVES

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- 111 i. Determination of heavy metal content in Yamuna river water sample
- 112 ii. Removal of contaminants from river water sample with the help of Hyper - accumulator  
113 plants
- 114 iii. Evaluation of Bio-accumulation capacity of all of three selected plants

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## 116 2. Material and Methods

117 **2.1 Waste water collection:** Water sample was collected from Yamuna river enrooted Delhi-  
118 Agra via Haryana, near Palwal District. Water sample was preserved in a can at freezing  
119 temperature (6-8°C). This water was further used for phytoremediation study.

120 **2.2 Plants used:-** Three different plants (*Tagetes patula*, *Bassica scoparia*, *Portulaca*  
121 *grandiflora*) were used for the study. The seeds of the plants were collected from a local  
122 nursery at Delhi-NCR. The plant classifications have been listed in Table 2.

123 Table 2:

Classification	<i>Tagetes patula</i>	<i>Bassica scoparia</i>	<i>Portulaca grandiflora</i>
Kingdom	Plantae	Plantae	Plantae
Order	Asterales	Caryophyllales	Caryophyllales
Family	Asteraceae	Amaeanthaceae	Portulacaceae
Genus	Tagetes	Bassia	Portulaca
Species	<i>T. patula</i>	<i>B. scoparia</i>	<i>P. grandiflora</i>

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125 *T.patula* grown and harvested annually and flowers are yellow and red in colour, reaching 0.3 m

126 to 0.5 m in size. The plant size varies from 0.1 to 2.2 m tall. They have fibrous roots. In India it  
127 grows from October to April. The plants common name is called “Marygold”. The leaves of the  
128 plants include oil glands and the oils are pungent. It can grow in any sort of soil. *T. patula* is  
129 widely cultivated in India it also have various uses in medicines.

130 The main reason for selecting this plant for phytoremediation is its ability of resisting adverse  
131 condition like pests, salinity, drought etc. *T. patula* is good for phytoextraction of heavy metals  
132 like arsenic, Mercury etc.

133 It is a small but fast growing annual plant as it has grown 30 cm tall. The leaves of the plant are  
134 thick and fleshy, up to 2.5 cm long arranged in a cluster like structure. the flowers are 2.5-3 cm  
135 diameter with five petals. The colour of flowers varied -from red, pink, white, orange and yellow.  
136 In India it is called “9 o’clock” flower -because it blooms at 9 a.m. It generally requires no  
137 attention as it gets spread very easily by itself. This plant can easily grow in adverse conditions  
138 like pesticides, high heavy metal concentration, chemicals etc. This plant consumption known to  
139 reduce the risk of cancer and heart diseases (Tangahu et al., 2011; Ahmad, 2015; Moubasher et al.,  
140 2015; Vijayaraghavan et al., 2017; Purakayastha et al, 2008)

141 It is a large annual herb. The plant is helpful in controlling soil erosion. This plant is suggested  
142 as an agent for phytoremediation technique because it is hyperaccumulator of cadmium, zinc,  
143 mercury, chromium. It is an evergreen foliage plant. The seeds of the plant help in regulation of  
144 hypertension and obesity etc.

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## 146 2.3 Procedure

### 147 2.3.1 Model set up:

- 148 i. Six -plastic boxes were taken.
- 149 ii. Two -boxes for each plant.
- 150 iii. For setting up the model, one plastic box was placed on another.
- 151 iv. Small holes were induced in the centre of each plastic box for the passage of plant -roots  
152 as shown by the pictures below in figure a, b, c, d, e.
- 153 v. After germination of seeds in soil, small plants were transplanted. From the soil in the  
154 upper plastic box which was already filled with garden soil.

- 155 vi. Roots of the plants were allowed to reach the lower plastic box. Already filled with  
156 contaminated water sample of Yamuna river through induced wholes.



157 **Figure a: Set up for plant**

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160 **Figure b: Set up of different plants**

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**Figure c: Set up for *P. grandiflora***



**Figure d: Picture of *B. scoparia***

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Figure e: Picture of *T. patula*

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#### 170 **2.3.2 Growth period:**

- 171 i. Plants were allowed to grow in that setup for eight weeks.
- 172 ii. During these eight weeks, generally called “Growth period”, proper attention to the plants  
173 was given just to make sure. That none of the plant will die.
- 174 iii. Fertilizers such as cow dung [wasere](#) mixed into the soil.
- 175 iv. Plants were placed beneath a tree, because much, sunlight exposure can cause browning  
176 of plants.

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178 | **2.3.3 Change in size parameters:** Growth in the length of the plants was measured. [A](#)fter  
179 completion of fourth and eighth week by a centimetre scale.

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181 **2.3.4 Lab work:** After 8 weeks, all of the three plants were harvested and the water samples  
182 **(initial untreated and final treated)** from all the three plants were taken and stored in three  
183 different plastic bottles with proper labelling.

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**2.3.5 Acid Digestion:** Acid digestion method was used for preparing the water and tissue samples. It is done by adding a considerable amount of acids and heating, until the solution gets completely decompose and release metals.

**a. For acid digestion of water samples,** the water samples were autoclaved and added in the glass beakers.

As nitric acid can never use alone, so it was combined with sulphuric acid. To the water samples, first added 5 ml of concentrated  $\text{HNO}_3$  and 10 ml of concentrated  $\text{H}_2\text{SO}_4$ , boil on a hot plate at  $90^\circ\text{C}$  for evaporation, until dense fumes of dense  $\text{SO}_3$  appears.

After clearing of the solution, no brownish fume appears, then distilled water was added to make solution dilute and heated.

Then the solution was centrifuged at 3000 rpm for 25 min and the pellet was discarded, supernatant was taken and stored in test tubes with proper labeling.

**b. For acid digestion of plant tissues**

Plants were first wiped with 0.01N HCl followed by rinsing with distilled water, then the plants were separated into different parts viz. roots, stems, leaves. And let them dry in oven for 15 min or less. All the parts were ground into grinder and 2 g of sample were taken in the glass beaker after weighing. For digestion,  $\text{HNO}_3$  And  $\text{HClO}_4$  acids was used. To the sample first 5 ml of  $\text{HNO}_3$  -added and heated on a Hot plate at temperature  $100^\circ\text{C}$  for 30 to 35 min, then 2.5 ml of  $\text{HClO}_4$  added to the mixture and boiled, white fumes appeared, later 5 ml of dilute water added to the mixture and again boiled until the fumes were totally released.

Detection of heavy metals present in all the samples was done by AAS technique.

**3. Results and Discussion:** Final growth in the length of plants is given in the table below

211 and also shown in the picture given below.

212 **Table 4: Change in length (cm) of the Plants**

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Plants	Zero day	After four weeks	After eight weeks
<i>T. patula</i>	5 cm	9.5 cm	19 cm
<i>B. scoparia</i>	6 cm	8.5 cm	11.5 cm
<i>P. grandiflora</i>	3.5 cm	7 cm	13 cm

214

215 The amounts of heavy metals present in the water sample and in the plant tissue sample were  
216 analyzed by a technique called “Atomic absorption spectrometry”. The amount of heavy metals  
217 such as Cd, Hg, Zn, Cr, Pb in the initial untreated water sample and also in final treated Water  
218 samples are given in the table below.

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220 **Table 5: Presence of heavy metals (mg/L) in water sample**

Metals	Initial water sample	<i>Tagetes patula</i>	<i>Portulaca grandiflora</i>	<i>Bassia scoparia</i>
<b>Cd</b>	0.715	0.489	0.315	0
<b>Cr</b>	0.513	0.269	0.418	0.379
<b>Zn</b>	0.948	0.533	0.697	0.705
<b>Hg</b>	1.079	0.782	0.969	0.783
<b>Pb</b>	1.098	0.055	0.079	0.069

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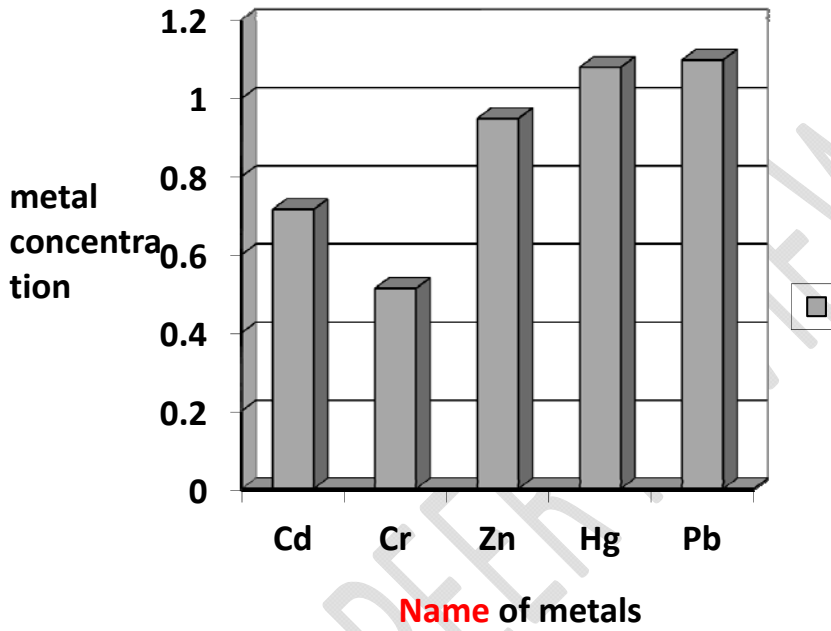
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223 In the present study, cadmium was undetectable in the water sample of *B. scoparia* and *T. patula*

224 absorbed greater amount of Cd as compared to *T. patula*. The chromium concentration found  
225 very less in the treated water sample by *T. patula* and it was highest in *P. grandiflora*. Zinc level  
226 highest in *P. grandiflora* and lowest in *T. patula*. The Hg concentration found highest in *P.*  
227 *grandiflora* and there is approximately no difference in the results of *T. patula* and *B. scoparia*.  
228 Pb concentration has been found in this decreasing order *P. grandiflora* > *B. scoparia* > *T. patula*.  
229 ~~So~~ according to this result *T. patula* is good for treatment of chromium, zinc, mercury, lead  
230 from wastewater-. *B. scoparia* is good for the removal of mercury most as compared to other  
231 heavy metals from waste water and *P. grandiflora* is proved to be a good remediation agent for  
232 cadmium etc mostly as compared to other heavy metals from contaminated water.

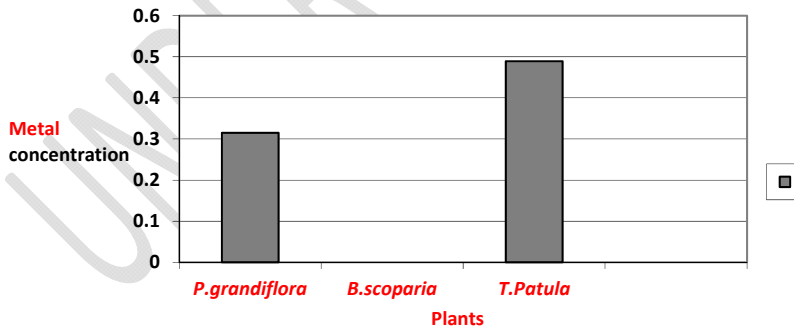
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234 **Graph3. 1: Graphically representation of concentration of heavy metals in untreated initial**  
235 **water sample**

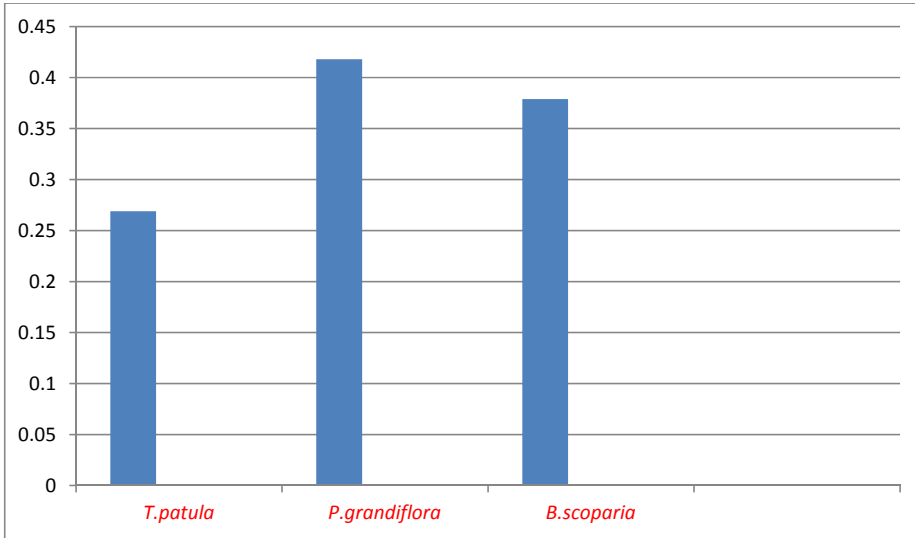


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237 Graph 3.2: Cadmium concentration left in treated water sample after eight weeks



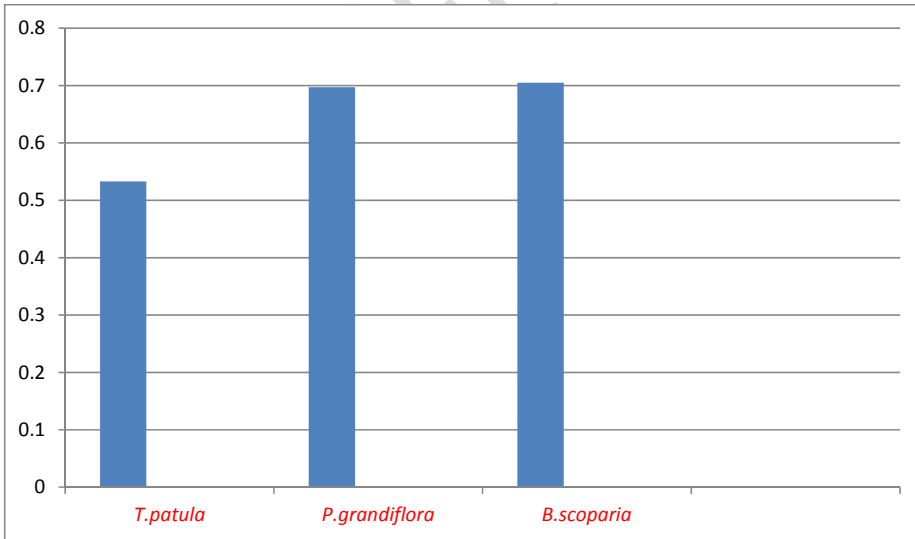
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240 **Graph 3.3: Chromium concentration left in water samples after eight weeks**

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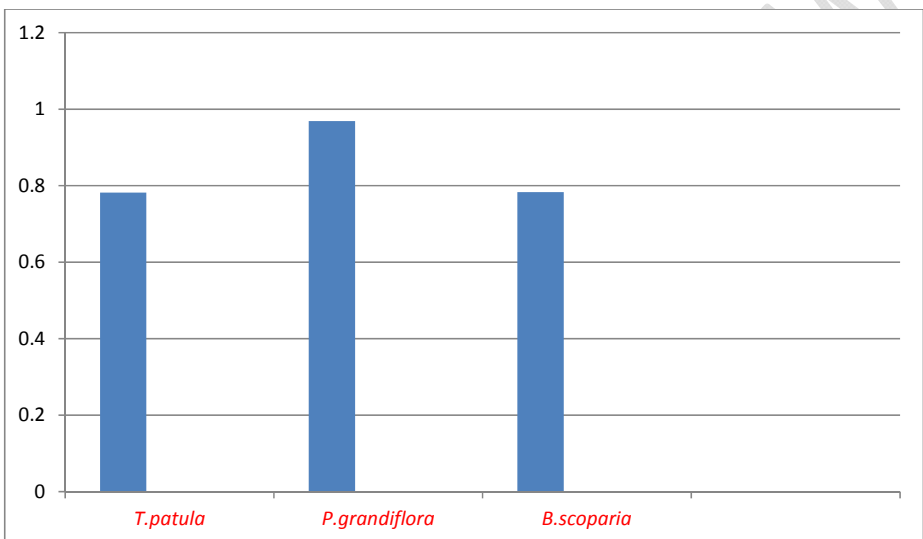
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**Graph 3.4 :Zinc concentration left in water sample after eight weeks**

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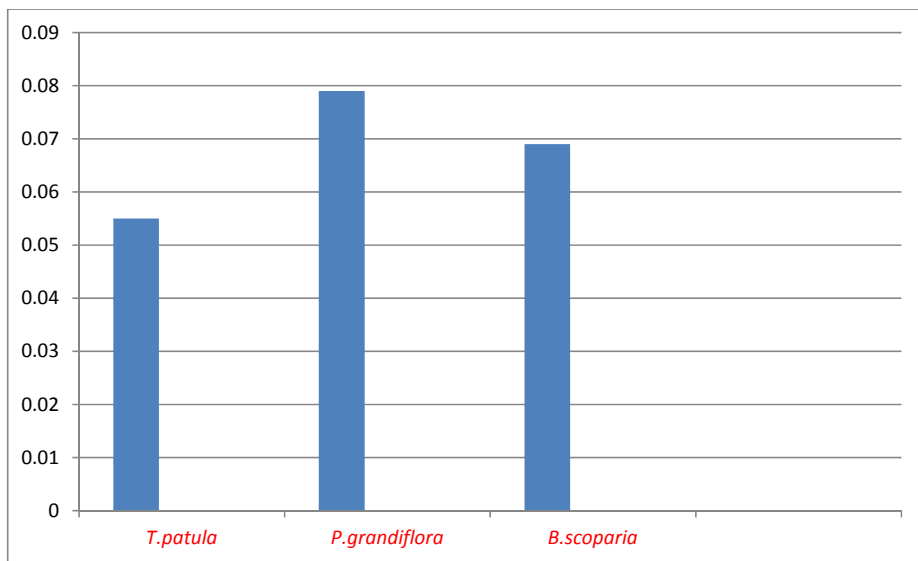


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**Graph3. 5: Mercury concentration left in water after eight weeks**





**Graph3. 6: Lead concentration left in water sample after eight weeks**

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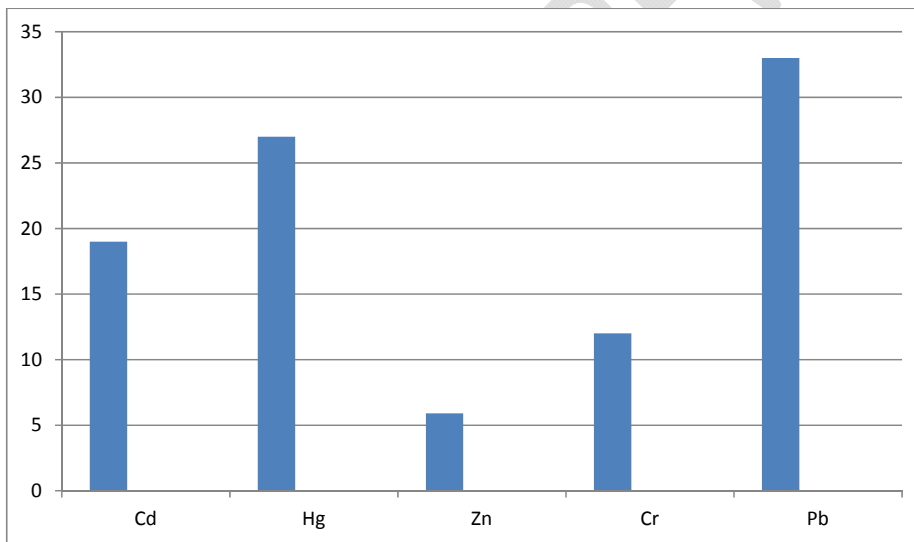
Bioaccumulation of heavy metals by plants: Plants also have the ability to accumulate the metals, were checked with the help of AAS technique, after the acid digestion process of samples. The results of AAs are given in the table below.

**Table 6: Presence of heavy metals in the Roots (mg/kg<sup>-1</sup>) of plants**

Metals	<i>Tagetes patula</i>	<i>Portulaca grandiflora</i>	<i>Bassia scoparia</i>
<b>Cd</b>	19	22	8
<b>Cr</b>	12	7.9	6.3
<b>Zn</b>	5.9	4	2.6
<b>Hg</b>	27	25.2	15.9
<b>Pb</b>	33	38	13.7

267 According to the above result, accumulation of zinc, mercury and chromium was highest in the  
 268 roots of *T. patula*. Lead and cadmium accumulation was highest in the roots of *P. grandiflora*.

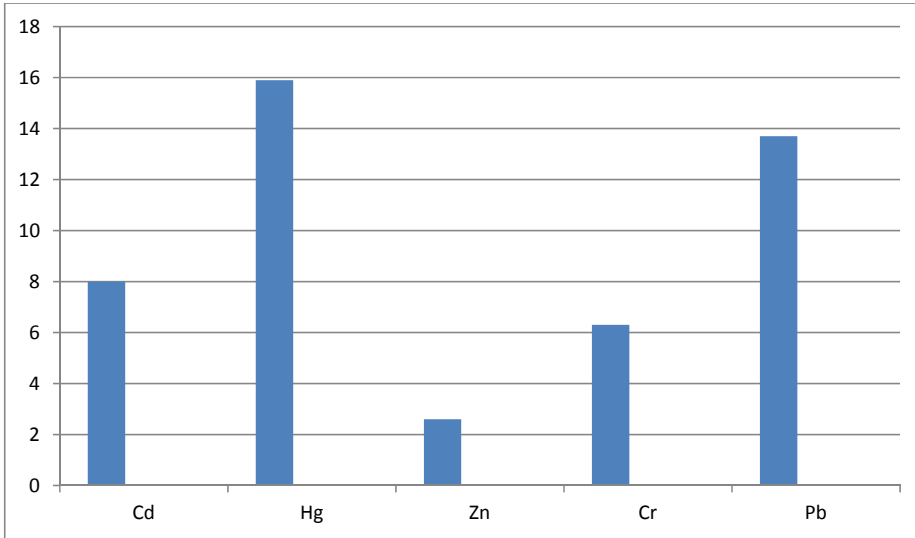
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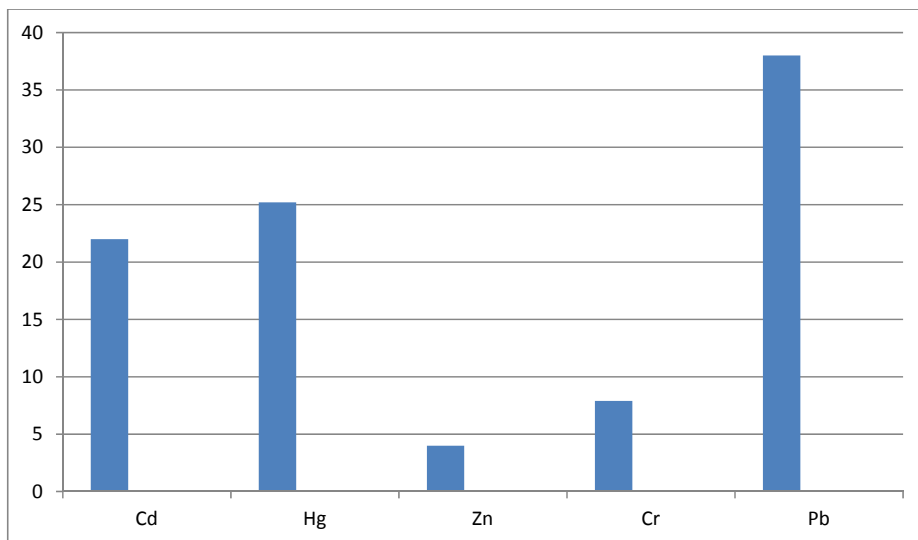
**Graph 3.7: Heavy metal concentration in roots of *T. patula***



**Graph 3.8: Heavy metal concentration in roots of *B. scoparia***

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**Graph 3.9: Heavy metal concentration in roots of *P. grandiflora***

**Table 7: Presence of heavy metals in the Stems (mg/kg<sup>-1</sup>) of plants**

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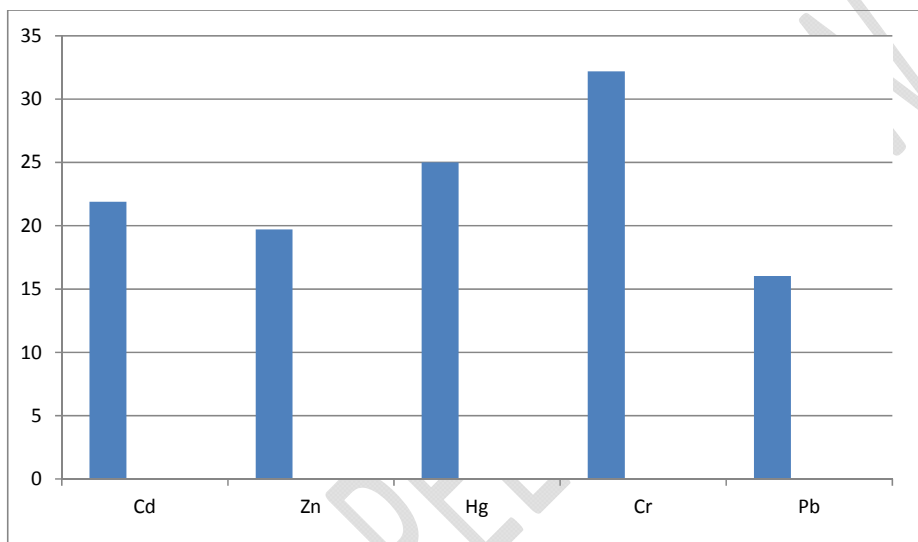
Metals	<i>Tagetes patula</i>	<i>Portulaca grandiflora</i>	<i>Bassia scoparia</i>
<b>Cd</b>	21.9	18.8	6.9
<b>Cr</b>	32.2	30.1	4
<b>Zn</b>	19.7	17	3.1
<b>Hg</b>	25	8.6	21
<b>Pb</b>	16.02	11.7	7

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292 According to the result given in [above-Table \(7\)](#), stems of *T. patula* has the highest efficiency for  
293 accumulating all the above heavy metals, even *P. grandiflora* and *T. patula* shows approximately  
294 the same results for accumulation of heavy metals in their stems.

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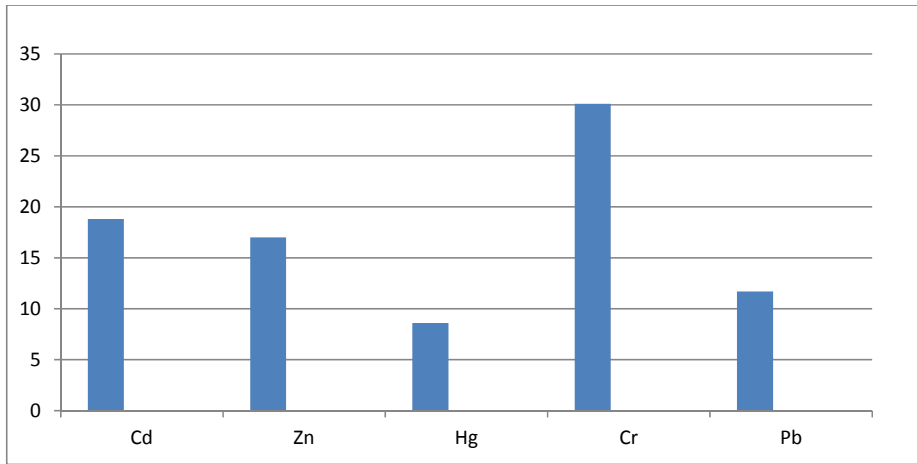
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**Graph 3.10: Heavy metal concentration in Stems of *T. patula***

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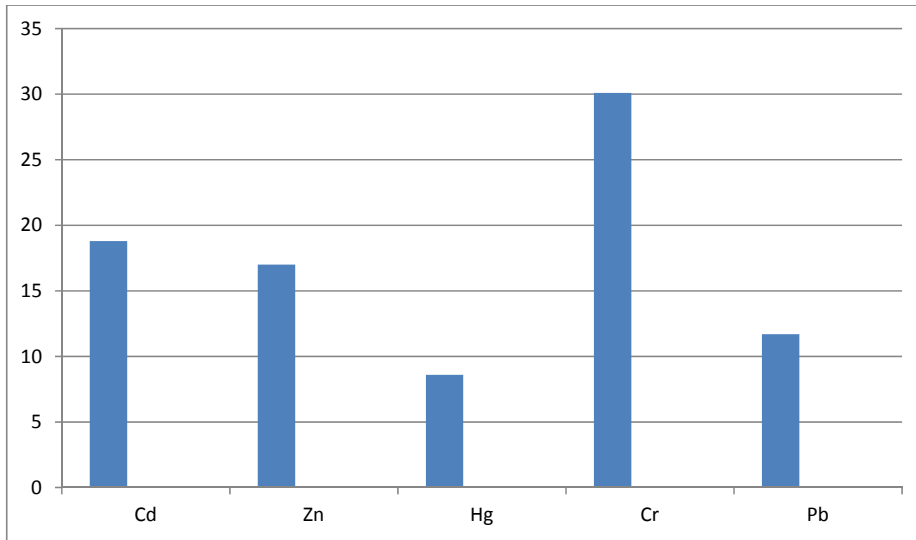
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**Graph 3.11: Heavy metal concentration in Stems of *B.scoparia***

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308 **Graph 3.12: Heavy metal concentration in Stems of *P. grandiflora***

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311 **Table 8: Presence of heavy metals in the Leaves (mg/kg<sup>-1</sup>) of plants**

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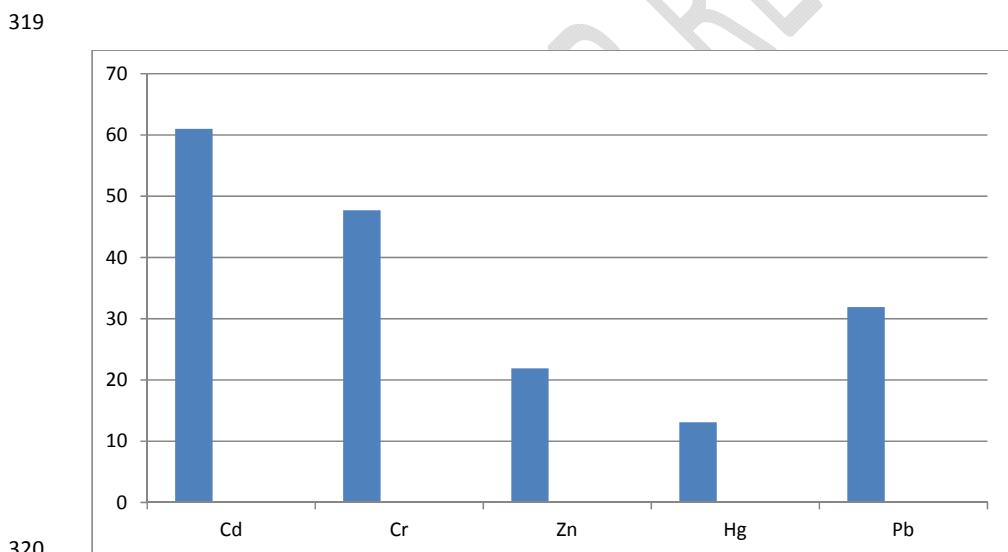
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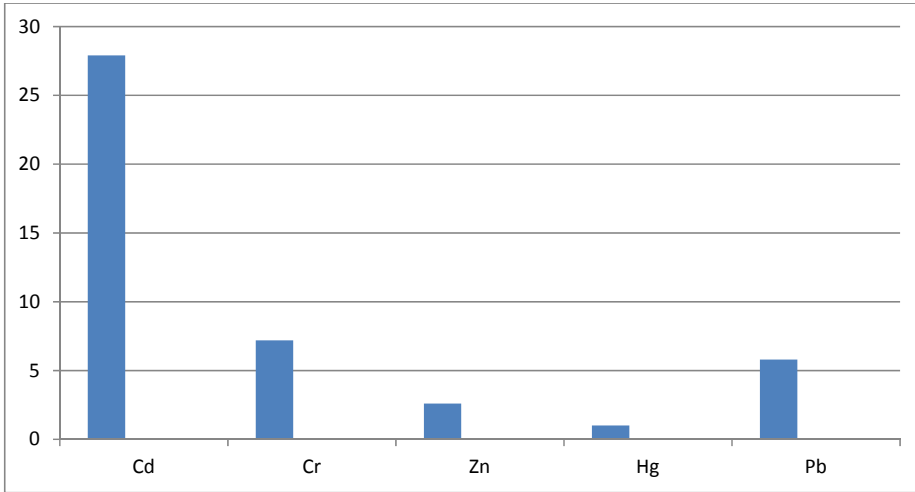


Metals	<i>Tagetes patula</i>	<i>Portulaca grandiflora</i>	<i>Bassia scoparia</i>
<b>Cd</b>	61	36.1	27.9
<b>Cr</b>	47.7	20.8	7.2
<b>Zn</b>	21.9	2.3	2.6
<b>Hg</b>	13.11	4.6	1
<b>Pb</b>	31.9	4.6	5.8

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 316 According to the above table, *T. patula* accumulated highest amount of heavy metals in its leaves  
 317 and *P. grandiflora* and *B. scoparia* accumulated a great amount of cadmium in their leaves. *P.*  
 318 *grandiflora* has also accumulated a significant level of chromium in its leaves.



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 322 **Graph 3.13: Heavy metal concentration in Leaves of *T. patula***  
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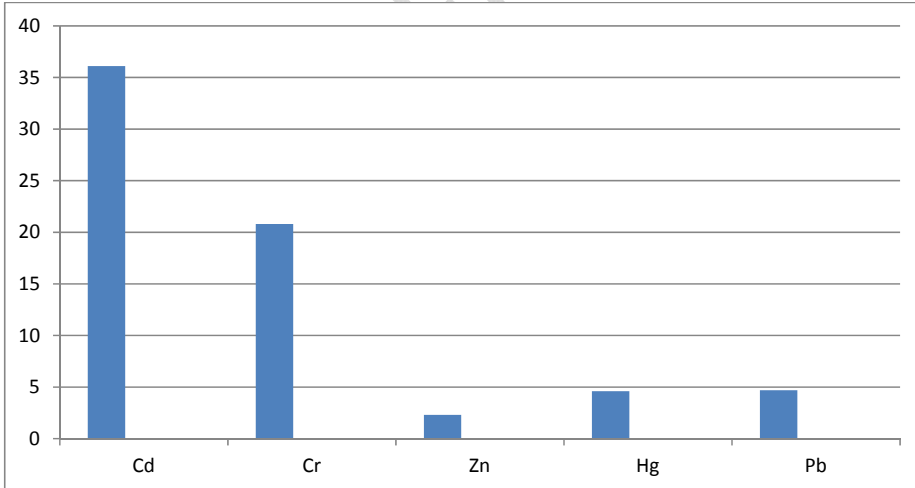
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**Graph 3.14: Heavy metal concentration in Leaves of *B.scoparia***

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330 **Graph 3.15: Heavy metal concentration in Leaves of *P.grandiflora***

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332 **Conclusion:** Phytoremediation is an effective, cheap or low maintenance technique for removal  
333 of heavy metals from environment. Out of all the three plants, *T.patula* shows a better growth in  
334 size and also shows the highest bio accumulating capacity for heavy metals. It can be concluded  
335 from the above study that the water quality of Yamuna river is good before entering national  
336 capital Delhi. The main disastrous impact is from Najafgarh drains. From the above experiment,  
337 it can be said that phytoremediation, phytoextraction technique can be used for making Yamuna  
338 river pollution free, but we have to stop mixing untreated sewage water in Yamuna river. This  
339 project is a little attempt towards the big problem of Yamuna river pollution. This study showed  
340 the phytoremediation capacity of all of three selected plants: *Tagetes patula*, *Bassica scoparia*,  
341 and *Portulaca grandiflora*. Bioaccumulation of heavy metals in various parts of plants has also  
342 been analysed. The study concludes that the non-edible plants can be used for treatment of  
343 wastewater and contaminated soil in in-situ techniques. Further, Phytomining can be done to  
344 recover and reuse the heavy metals from plant tissues after phytoremediation.

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