Original Research Article

2 ASSESSMENT OF THE PREVALENCE OF

3 Chlamydia trachomatis INFECTION IN

4 DONATED BLOOD AT REGIONAL BLOOD

5 TRANSFUSION CENTRE, ELDORET,

6 KENYA.

7 ABSTRACT

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Aim: To detect *Chlamydia trachomatis* in donated blood at Regional Blood Transfusion center,
 Eldoret.

10 Methodology: A random selection blood sample was done for this study for a period of one month.

11 Blood bags were numbered 1-87 and picked amongst unnumbered blood bags by each donor in order

12 of donation. The samples obtained were subjected to centrifugation and serum used in the

13 immunoassay test by using the *Chlamydia trachomatis* IgG/IgM Elisa test kit.

14 **Result:** This study showed that *C. trachomatis* was present in 8% (7 samples) of the blood samples 15 analyzed and 92% were found to be negative.

16 **Conclusion:** With 8% of the samples testing positive for *Chlamydia trachomatis* this indicated that 17 some of the blood donated to the Regional Transfusion Center is contaminated.

18 Keywords: Blood, Chlamydia trachomatis, Detection, Kenya, Transfusion

19 1. INTRODUCTION

Chlamydia is a genus of pathogenic bacteria that are obligate intracellular parasites. Chlamydia infections are the most common bacterial sexually transmitted diseases in humans and are the leading cause of infectious blindness worldwide [1]. Chlamydia trachomatis, commonly known as chlamydia, is one of four bacterial species in the genus Chlamydia.

C. trachomatis is globally a major cause for sexually transmitted diseases (STDs) including, urogenital and Chlamydia. Chlamydia is one of the most common sexually transmitted infections worldwide affecting about 4.2% of women and 2.7% of men [2]. *C. trachomatis* cause various infections that include trachoma (conjunctivitis which is the most reported reason for blindness globally), pneumonic infections, and genital infections in people of both sexes and conjunctivitis in newborns. It is estimated that there are over 50 million new cases of *C. trachomatis* infection annually [3]. However another study done in 2015 showed that about 61 million new cases occurred globally [4].

C trachomatis causes various contaminations among among which is donated blood. A study estimated that about 92 million blood donations are collected per year in different parts of the world [5]. In addition, WHO published that 39 countries in the world still do not routinely screen for transfusion-transmissible infections (TTIs) including HIV, Hepatitis B, Hepatitis C and syphilis [6]. About 47% of blood donations in low-income countries are screened in laboratories which do not have

36 capacity for *C. trachomatis* contamination [7]. Blood transfusion centers are expected to provide blood

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that is free of contaminants. However, without this there is likeliness of transmitting fatal agents like *C. trachomatis* that may result in deadly diseases that may be found in blood [8].

39 Weström [9] asserted that one of the factors that may contribute to unnoticed C. trachomatis

40 contamination of donated blood is that its infections are to a bigger percentage asymptomatic. This

41 has commonly been reported in both males and females with about 50% males and 75% females of

42 those affected having no symptoms.

43 2. MATERIALS AND METHODS

44 2.1. Sample Collection

Blood samples used in this study were collected aseptically using venipuncture techniques by qualified personnel of the center following the Pendergraph procedure of phlebotomy [10].

Use of aseptic techniques were observed to preserve the integrity of the specimen. Blood samples obtained were labelled with the donors code for *C. trachomatis* test. The sample blood obtained was subjected to centrifugation to separate the serum from the other blood components. The serum samples were then refrigerated at 2°-8°C in RBTC Hematology Laboratory to be used in the immunoassay test by using the *Chlamydia trachomatis* IgG/IgM Elisa test kit manufactured by Vircell Company Spain to find out the presence of *C. trachomatis* in donated blood using the ELISA

53 procedure as shown below.

54 2.2. Sample Processing

55 10ml of blood sample from each donor was centrifuged at 1600rpm for 15 minutes. This was done 56 when clotting had occurred and within 1 hour after blood sample was drawn. Serum was then used for

- 57 the indirect immunoassay test.
- 58 2.2.1. Indirect Immunoassay

59 Preparations and Assay Procedure

- 60 The washing solution was prepared in advance. This was done by using distilled water to fill 50 ml of
- 61 20x washing solution up to 1 liter after which the solution was warmed up to 37°C in the washing
- 62 concentrate during storage before diluting. Once diluted, it was stored at 4°C.

63 The Indirect Immunoassay Procedure

The incubator was set at 37°C and all the reagents brought to room temperature before use (approximately 1hour), without removing the plate from the bag. The components were then shaken well.

- 67 The plate was removed from the package and the numbers of wells to be employed determined that
- is: Four wells for the controls; two for the cut off serum and one each for the negative and positivesera. Wells not required for the test were returned to the pouch and sealed.
- For IgG test, 100 µl of serum diluent **2** was added to all wells followed by **5** µl of each sample, **5** µl of
- 71 positive control 3G, 5 µl of cut off control 4G (in duplicate) and 5 µl of negative control 5G into the
- 72 corresponding wells. The plates were then shaken in a plate shaker for 2 min in order to achieve a 73 homogenous mixture of the reagents. A sealing sheet was then used to cover the wells and incubated
- 74 at 37°C for 45 min.
- For IgM test, 25 µl of VIRCELL IgG sorbent was added to each of the required wells, except for the wells where controls will be dispensed. 5 µl of sample was added and then 75 µl of the serum diluent
- 2 to each well. Control wells were prepared by adding first 100 µl of the serum diluent 2 to each well
- and then 5 µl of the positive control **3M**, 5 µl of the cut off control **4M** (in duplicate) and 5 µl of the
- 79 negative control **5M** to the corresponding wells. This was followed by plate shaking in a plate shaker
- 80 for 2 min in order to achieve a homogenous mixture of the reagents. A sealing sheet was then used to
- 81 cover the wells and incubated at 37°C for 45 min.

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82 After incubation, the seal was removed, liquid aspirated from all wells and washed five times with 0.3

ml of washing solution **9** per well and any remaining liquid was drained off.100 µl of IgG conjugate

solution **6G** or IgM conjugate solution **6M** was immediately added into each well and a sealing sheet

85 used to cover followed by incubation at 37°C for 30 minutes.

86 After 30mins the seal was removed, liquid aspirated from all wells and washed five times with 0.3 ml

of washing solution 9 per well. Any remaining liquid was drained off and 100 µl of substrate solution 7
immediately added into each well followed by incubation at room temperature for 20 minutes
protected from light. After incubation 50 µl of stopping solution 8 was immediately added into all wells
and finally readings were carried out within 1 hour of stopping.

and finally readings were carried out wit91

92 3. RESULTS

In this study, a greater percentage of the samples were from the donors between 20-29 years of age (60.9%), while the least were below 20 years old group. There was a higher proportion of females (55.2%) than males (44.8%). Samples from single donors were 64.4% while divorced/separated and widowed were both 3.4%. 75% of the samples were from donors who were currently at tertiary education level while none was from those who had never attended attained any form of education. The highest proportion of the samples was from unemployed donors (73.6%), 11.5% from self-employed and 14.5% from employed persons (Table 1).

100 The largest percentage of samples were from donors who felt they were currently in good health 101 (97.7%), had no sexual encounters with persons whom they did not know their background (96.6%),

- 101 (97.7%), had no sexual encounters102 had irregular sexual partners (85.1).
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122 Table 1. Overall characteristics of blood donors whose samples were analyzed at RBTC,

123 Eldoret.

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Characteristics	Categories	Total (N=87) No (%)
ge groups	Below 20 years	4 (4.6)
	20-29 years	53(60.9)
	30-39 years	25(28.7)
	40 and above	5(5.7)
ender	Male	39(44.8)
	Female	48(55.2)
larital status	Single	56(64.4)
	Married	25(28.7)
	Divorced/Separated	3(3.4)
	Widowed	3(3.4)
ducation level	None	0(0)
	Primary	7(8.0)
	Secondary	14(16.1)
	Tertiary	66(75.9)
mployment status	Unemployed	64(73.6)
	Self-employed	10(11.5)
	Employed	13(14.9)
Good health	Yes	85(97.7)
	No	2(2.3)
exual activity	Yes	3(3.4)
110.	No	84(96.6)
regular sexual	No	13(14.9)
artners	Yes	74(85.1)

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125 Detection of Chlamydia trachomatis in donated blood using IgG/IgM Elisa kit

126 Samples with indexes below 9 were considered as not having IgG or IgM specific antibodies against

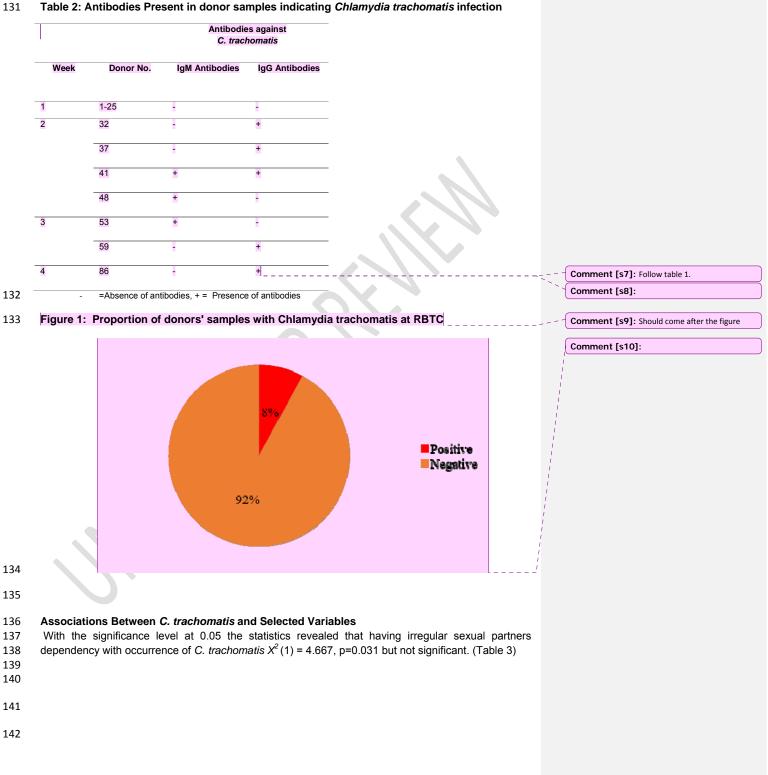
127 C. trachomatis while samples with indexes above 11 were considered as having IgG or IgM specific

128 antibodies against C. trachomatis. Therefore this study showed that C. trachomatis in blood donors'

129 samples at RBTC was found to be present in 8% (7 samples) of the blood samples analyzed at RBTC

130 and 92% were found to be negative. (Table 2).





143 **Table 3:** Detection of C. trachomatis infection in donated blood with respect to risk factors at RBTC,

144 Kenya.

Characteristics		C. trachomatis status	Chi Square tests			
		Positive N=7 Out of 87 samples	Pearson Chi-Square	Valid cases	Asymp. Sig. (2-sided)	
Age groups	Below 30 years	4	0.236	87	0.627	-
	30 years and above	3				
Gender	Male	2	0.706	87	0.401	
	Female	5				
Marital status	Single	4	0.173	87	0.677	
	Non-singles	3				
Education level	Primary and below	1	0.401	87	0.527	
	Secondary and above	6				
Employment status	Unemployed	4	1.055	87	0.304	
	Self-employed and/ Employed	3	\sim			
Irregular sexual	No	3	4.667	87	0.031*	
partners	Yes	4				

- 146 * Significant at p<0.05.
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150 4. DISCUSSION

151 The presence of C. trachomatis in blood samples was determined based upon the reaction of 152 antibodies in the donor sample tested with the antigen adsorbed on the polystyrene surface. Unbound 153 immunoglobulins were washed off, this was then followed by an enzyme anti-human globulin binding 154 the antigen-antibody complex in a second step. After a new washing step, bound conjugate was 155 developed with the aid of a substrate solution (TMB) to render a blue colored soluble product which 156 turned into yellow after adding the acid stopping solution [11]. IgG/IgM antibody detection are markers 157 for a Chlamydia-positive immune response, either for current, chronic or past infections. The assay 158 used in this study was the COMP (Complexes of Outer Membrane Proteins) of C. trachomatis, free 159 from LPS which is responsible for most cross-reaction with other Chlamydia species. ELISA yields very good agreement with the target results (IgM100%, IgG 98%) in quality assessment schemes 160 161 [12]. It provides good agreement with characterized samples from quality assessment schemes of 162 IgM 100%, IgG 98% [12]. Therefore ELISA technique is paramount for detection of C. trachomatis 163 antibodies.

164 Occurrence of IgM antibodies without IgG being present is an indication of primary infection while 165 presence of IgG antibodies without IgM antibodies in the blood sample is an indication of re-infection 166 or presence of previous infections since IgG antibodies persists in the body for a long period of time. 167 Presence of both IgG and IgM antibodies in the sample shows an ongoing infection.

168 In a study from Makkah Saudi Arabia, 8.7% of the women were positive for IgG antibodies to C. 169 trachomatis with antibody indexes of 1.4–2.0 [13]. Low prevalence in Saudi patient population may be due to the adherence of strict moral principles and code of ethics in Saudi Arabia. In comparison to

171 this study, 71.4% of women were positive for both IgG/IgM antibodies which is much higher.

Similarly, it has been shown in the USA and Europe that demographic factors which increase the risk
of chlamydial infection include youth, single marital status and multiple sexual partners [14].
Therefore, a closer attempt should be made to correlate risk factors and disease entity when
screening for *C. trachomatis*.

According to a study done by CDC,[15] on the prevalence of *C. trachomatis* among the adults, the highest rates is between 20 to 24 years old, however the age-specific rates for women was as low as to 16 to 26 years old. This current study however slightly differs from the statistics by CDC. The age between 20 to 29 years of age showed the highest prevalence.

Multiple partnerships may increase the likelihood of encountering a sexually transmitted pathogen through the increased probability of choosing a partner with infection, while having new or casual sexual contacts may be related to increased risk because of a reduced familiarity between partners [16]. Statistics from this study revealed that having irregular sexual partners might have had dependency with occurrence of *C. trachomatis* X^2 (1) = 4.667, p=0.031.Therefore, study concurs with

- 185 the later however it is not conclusive as per the statistics on the *chi* square table.
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193 **5. CONCLUSION**

The findings of this study therefore indicates that some blood donated to the Regional Transfusion Center was contaminated with *Chlamydia trachomatis* with 8% of the total samples indicating positive for *C. trachomatis* infection while 92 % were negative. Moreover the age group of between 20 to 29

197 vears old had the greatest percentage of those with *C. trachomatis*. Having multiple irregular number

- of sexual partners increases the chances of having *Chlamydia trachomatis* infection. However this is
- 199 not be conclusive according to the statistics of this study.
- 200 The sex partners of donors with Chlamydial infection should therefore be referred for medical care
- 201 through the efforts of individual donors. A further study is also recommended on the exact level of C.
- 202 trachomatis by use of absorbance ranges.

203 6. COMPETING INTERESTS

- 204 "The authors declares that they have no competing interests"
- 205 7. ACKNOWLEGDMENTS
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- 207 Role for her advice, encouragement and her role in interpretation of data.
- 208 8. AUTHORS' CONTRIBUTIONS
- 209 The authors' responsibilities were as follows;
- 210 **REFERENCES**

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Comment [s13]: Many grammatical errors still there. It still needs editing.

Comment [s14]: Discussion part still not satisfied.

Comment [s15]: Table???????

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