

External Quality Assessment: Microscopy Diagnosis Of *Plasmodium Falciparum* For A Better Management Of Malaria In The Regional Health Center In Côte D'ivoire

ABSTRACT

Context: In Côte d'Ivoire, malaria is transmitted throughout the year with an increased rate during the rainy season. This pathology is endemic on the whole territory with seasonal variations. The major vector is *Anopheles gambiae*. The external microbiology quality assessment programs organized by both Institut Pasteur of Côte d'Ivoire (IPCI) and PEPFAR, malaria microscopy was randomly carried out in 1/3 of the country regional health center laboratories. Laboratory technicians play a key role in malaria control programs because care services such as the disease monitoring depend on their diagnosis and technical skills.

Aim: The aim of this evaluation was to control the quality of the microscopic diagnosis and the performance of on-duty technicians for the management of feverish patients and efforts to strengthen laboratory services.

Méthodology: Six (6) RHC (Regional Health Center) laboratories were involved in the evaluation. Anonymity code was assigned to each of the participating laboratories.

There were many discrepancies in External Quality Assessment (EQA) results on the field not with standing the parasitemia, low or high.

Results: Only 30% of correct answers were recorded for *P. falciparum* identification. For *P. ovale*, we found a failure rate of 100% for laboratories.

Conclusion: Parasitemia was approximate and many confusions were observed regarding the different stages of parasites.

Keywords: EQA, *Plasmodium*, parasitemia, microscopy, thick blood. *Plasmodium falciparum* Microscopy diagnosis, Côte d'Ivoire

1. INTRODUCTION

According to WHO last estimations published in December (2016), there were 212 million malaria cases and 429,000 deaths (2015). In Côte d'Ivoire the vulnerable population was estimated at 8 million (2015). 4 million cases of malaria were confirmed with 14 thousand 14,000 deaths recorded. The number of cases of malaria confirmed through microscopy was estimated to 500 thousand. All cases of diagnosed malaria were caused by *Plasmodium falciparum*, the most dangerous and dreadful plasmodial species and a good reduction in mortality depends inevitably on good management of malaria cases (Abrogoua *et al.*, 2006)

For an efficient treatment, WHO recommends that malaria is confirmed in all suspected cases by a diagnosis based on parasite research (by microscopy or rapid diagnostic test) prior to treatment. According to the National Malaria Control Program (NMCP) report published (2004) Not listed in reference, malaria represents 80% of medical consultations and hospitalisations in Côte d'Ivoire and accounts for 33% of mortality causes. In Côte d'Ivoire, malaria is transmitted throughout the year with an increased rate during the rainy season. This pathology is having a field in the form of stable malaria, endemic on the whole territory with seasonal variations. The major vector is *Anopheles gambiae* (Konan *et al.*, 2008). Treatment only based on symptoms should not be considered if the parasitological diagnosis is not possible. Early diagnosis and treatment of malaria reduce disease intensity and prevent death. They also help reducing malaria transmission.

In a series of external microbiology quality assessment programs organized by both Institut Pasteur of Côte d'Ivoire (IPCI) and PEPFAR, malaria microscopy was randomly carried out in 1/3 of the country regional health center laboratories. Laboratory technicians play a key role in malaria control programs because care services such as the disease monitoring depend on their diagnosis and technical skills.

The objective of this evaluation was to control the quality of the microscopic diagnosis and the performance of on-duty technicians for the management of feverish patients and efforts aiming at strengthening laboratory services. Several short courses of malaria through microscopy courses will significantly increase the knowledge and the level of microscopy skills of the trainees and will bridge up the significant difference in baseline microscopy skills of the different categories of trainees those

59 who participated in the courses.
60 For this purpose, six 6 laboratories of the Regional Health Centers (RHC) of the health districts of
61 Côte d'Ivoire participated in the study of EQA in connection with the WHO guideline external quality
62 assessment (WHO, 2014).
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65 2. METHODOLOGY

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67 Six (6) RHC (Regional Health Center) laboratories were involved in the evaluation. Anonymity code
68 was assigned to each of the participating laboratories. It was about a Giemsa staining thick blood
69 smears and thin blood film. The panel was made of twenty-four (24) thin blood film slides with the
70 same staining. Thin blood films were made and stained 21 days before the expected date and results
71 were validated by the National reference center for malaria chemo-resistance hosted by IPCI through
72 the unit of malariology. Slide transportation was done by road and samples were given to the head of
73 the medical analyzes laboratory.

74 A questionnaire was submitted to participants including slide code, clinical information about the
75 patient, the result of thick blood smear with parasite density, the result of thin blood film with
76 identification of species and sexual and non-sexual forms and results interpretation interpretation of
77 results

78 Only the mean of parasitemia estimated by participants was calculated and reported. The results
79 were saved as an Excel file. They were classified as: correct (parasitic density inferior or equal to
80 10%), minor errors (non-significant difference or parasitic density inferior or equal to 20%) or major
81 errors (incorrect diagnosis on species or sexual or non-sexual forms or Incorrect interpretation or
82 parasitic density exceeding 20%).
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86 3. RESULTS AND DISCUSSION

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88 Many discrepancies were noticed in the parasitic microscopy results for both the density and
89 determination of species. Some results were over estimated (table 1) and others were underestimated
90 by participants (Table 1). Only 30% of correct answers were recorded for *P. falciparum unbold*
91 identification. For *P. ovale*, we found a failure rate of 100% for laboratories (Table 2). Parasitemia was
92 approximate and many confusions were much confusion was observed regarding the different stages
93 of parasites (table 1). However, there is a laboratory (lab. 007) which from the point of view of
94 parasitological diagnosis and diagnosis of species has nothing found. Some technicians could use
95 today the so-called "plus system" which is an old, simple, but much less precise method for
96 establishing parasitic density in thick blood smears. Because of its unreliability, it was replaced by the
97 method of determination of parasitic density by calculation, a simple mathematical formula, which
98 multiplies the number of parasites by 8000 (standard number of leukocytes/ μ l) dividing by the number
99 of leukocytes (200 or 500). The result is the number of parasites/ μ l of blood. Studies showed that
100 many technicians forgot the details of the plus system and were mistaken about the code (the number
101 of signs +) and numeration (the number of parasites per field or for 100 fields), which leads to
102 unreliable information about the parasitic density. The number of red blood cells infected with *P.*
103 *falciparum unbold* parasites is essential and the percentage of parasitemia should always be
104 reported as this has effects on the prognosis and the mode of treatment used. This qualitative
105 assessment may be considered insufficient in malaria-endemic areas (Benasseni *et al.*, 1987; Baudon
106 *et al.*, 1988). But some biologist technicians who do not want to get rid of this method would have
107 difficulty adapting to the counting methods by in force.

108 Overestimation of parasitemia observed in some participants could be due to counting errors. A red
109 blood cell infected with multiple parasites counts as a parasitic red blood cell. Another reason could
110 include gametocytes when calculating parasitemia. When calculating the *Plasmodium falciparum*
111 parasitemia, only the trophozoite stages are were counted. Gametocytes and other species of
112 malaria parasites are excluded from the result, but it was important to know them and to differentiate
113 all forms of the parasite biomass (Table I and II). Participants who underestimated parasitemia might
114 not have counted a sufficient number of fields (Table I). It is recommended to count 40 fields of a thin
115 blood smear (Table II), and Thick films should be examined by two observers, each viewing 200 high
116 power fields or counted 200 leukocytes. Especially when parasitaemia is low due to possible unequal
distribution of parasites or to count in the case of a thick blood smear, it is recommended to Count

117 until 500 leukocytes (WHO, 2009). Considering the diagnostic results per laboratory, no laboratory
 118 scored less than 80%, level of technical skill and accuracy expected for the examination of a series of
 119 slides for accreditation is inevitable, whereas the scores of all assessed laboratories range from 0% to
 120 50% (fig1). Technicians, once the certificate is obtained once certified, it is important to ensure that
 121 the level reached during the training is maintained. To achieve this, it is agreed that their work is
 122 regularly monitored by a supervisor at all times to help them improve their techniques and skills. This
 123 is called quality control which is part of the general activities of quality assurance applied in all
 124 microscopic malaria diagnostic services (WHO, 2010).

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 126 **Table 1: Analysis of laboratory performance according to parasitemia results**
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Order number of assessed Lab	Plasmodium strain Slide 1		Plasmodium strain Slide 2		Assessed Lab results categorization	
	NRC Lab	Assessed Lab	NRC Lab	Assessed Lab	Slide 1	Slide 2
001	440 gam/µl	2400 tr/µl	4440 sch/µl	4800 tr/µl	Major error	Major error
002	7080 tr/µl	3600 tr/µl	1600 sch/µl	Nc	Major error	Major error
003	840 gam/µl	Nc	20120 tr/µl	8500 sch/µl	Major error	Major error
004	1638 tr/µl	1700 tr/µl	4120 sch/µl	Nc	Correct	Major error
006	360 gam/µl	600 tr/µl	2400 sch/µl	Nc	Major error	Major error
007	850 gam/µl	Nc	3890 sch/µl	Nc	Major error	Major error

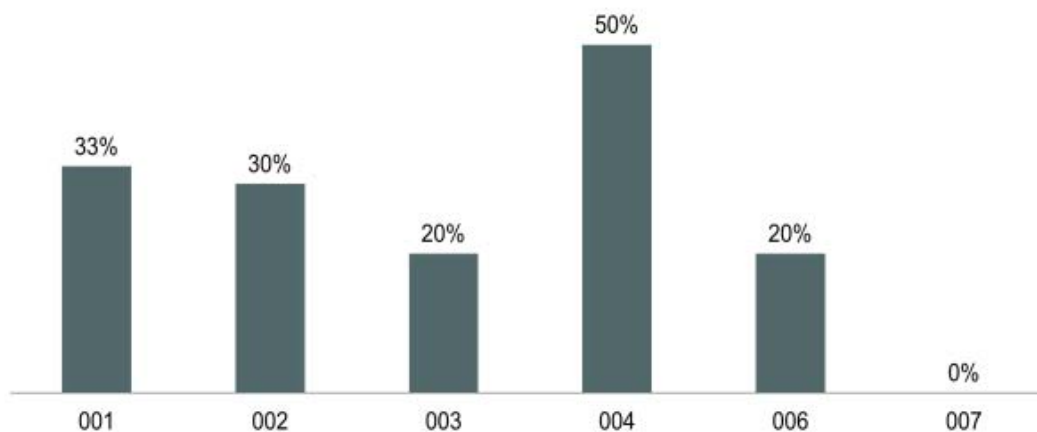
128
 129 gam: gametocyte; tr: trophozoite; sch: schizonte
 130 Nc: Not counted
 131 NRC: National Reference Center
 132 Lab: laboratory
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134 **Table 2 Laboratory performance analysis according to the results of the identification of plasmodial strain**
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Order number of assessed Lab	Plasmodium strain Slide 3		Plasmodium strain Slide 4		Assessed Lab results categorization	
	NRC Lab	Assessed Lab	NRC Lab	Assessed Lab	Lame 3	Lame 4
001	<i>Pf</i>	<i>Pf</i>	<i>Po</i>	<i>Pm</i>	Correct	Major error
002	<i>Pf</i>	<i>Po</i>	<i>Po</i>	Ni	Major error	Major error
003	<i>Pf</i>	Ni	<i>Po</i>	<i>Pf</i>	Major error	Major error
004	<i>Pf</i>	<i>Pf</i>	<i>Po</i>	Ni	Correct	Major error
006	<i>Pf</i>	<i>Po</i>	<i>Po</i>	Ni	Major error	Major error
007	<i>Pf</i>	Ni	<i>Po</i>	Ni	Major error	Major error

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 139 *Pf* : *Plasmodium falciparum* ; *Po* : *Plasmodium ovale* ; *Pm* : *Plasmodium malariae*
 140 Ni : Nothing identified
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142 Grid lines should be removed in all the tables
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Figure 1 Mean obtained by participating laboratory

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4. CONCLUSION

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Microscopy identification of parasite though being WHO standard method opens up a current reflection for a better management of malaria. Rapid diagnostic tests (RDTs) for a malarial antigen cannot replace microscopy but are indicated as a supplementary test when malaria diagnosis is performed by relatively inexperienced staff. Microscopy requires a high qualified staff which is not always available in areas where malaria is endemic. It is therefore important to maintain the level reached by technicians during their training. This requires supervision and a regular supervision of their work. This will help to continuously improve their skills. Quality control should be part of the general activities applied in all microscopic malaria diagnostic services.

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COMPLIANCE WITH ETHICAL

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This study was carried out according to the guidelines of the Ivorian National reference center for malaria chemo-resistance created by the interministerial decree number 393/08/ 2006, and conduct research according to the Ivorian National Ethical Committee and Research **with due approval.** **Therefore, this study was performed after receiving approval from the Ivorian National Ethical Committee and Research.**

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- 196 NOTE TO AUTHOR
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