# EFFECT OF SAGO EFFLUENT ON THE LEVELS OF THE GROWTH HORMONE IN THE BLOOD SAMPLE OF THE FRESH WATER FISH *CLARIAS BATRACHUS*

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5 Abstract: The aim of the study was to determine the effect of Sago effluent on the levels of 6 growth hormone in the blood samples of the fresh water fish *Clarias batrachus*. The fish were 7 exposed to control and different concentrations of treated sago effluents. The concentrations 8 chosen were 25%, 50% and 75% of treated sago effluent. The levels of the growth hormone was 9 significantly in the blood sample of the experimental fish *Clarias batrachus*, when compared 10 with that of controls.

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12 Keywords: Growth hormone, Sago effluent, *Clarias batrachus*.

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### 14 Introduction

The aquatic environment is the ultimate sink for all the environment pollutants any chemical 15 pollutant either natural or synthetic is most likely to reach the aquatic environment sooner or 16 later. The toxicity may be either acute or chronic to all forms of biota in aquatic system and also 17 varies to different aquatic organisms. The toxic effects may include both lethal and sublethal 18 concentrations, which may change the growth rate, development, reproduction, histopathology, 19 biochemistry, physiology and behavior [1]. Alterations in the physiological and biochemical 20 parameters of toxicant treated fish have recently emerged as an important tool for the water 21 quality assessment and to know the pathological status of fish in the field of environmental 22 toxicology [2, 3]. The alteration in various physiological and biochemical parameters of an 23 24 aquatic animal due to exposure of different toxicant has been shown to be directly or indirectly related to the behaviour, immune system, neurotransmission, energy metabolism and 25 reproduction [4, 5]. Accumulation of the environmental pollutants and toxicants has been shown 26 to cause alteration in the activity of many enzymes concerning to cellular energy metabolism [6, 27 28 7, 8, 9]. Alteration in enzyme activities of the fish is one of the major biomarker indicating the 29 level of changes consequent of pollutants in the tissues, organs and body fluid of the fish that can be recognized and associated with established health impairment process [10]. Moreover, 30 31 Gabriel and Akinrotimi [11] noted that enzymes can be used to confirm and asses fish exposure to toxicants, providing a link between external and internal structure and degree of responses to 32 toxicant exposure observed between different individuals. However, the applications of enzyme 33 determinations in fish, as an indicator of chemical intoxication seem to be promising. It is most 34 relevant and appropriate in sublethal exposure which spans over many days [12]. Toxicants also 35 can inhibit the activity or synthesis of enzymes[13], resulting in decreased activities in the 36 organs. 37

Growth hormone is a major participant in control of several complex physiologic processes, including growth and metabolism. Growth hormone is also of considerable interest as a drug used in both human and animals. Growth is a very complex process and requires the coordinated

- 41 action of several hormones. The major role of growth hormones in stimulating body growth is 42 to stimulate the liver and other tissues to secrete IGF -1. IGF -1 stimulates proliferation of 43 chondrocytes (cartilage cells), resulting in bone growth. Growth hormone has important effects 44 on protein, lipid and carbohydrate metabolism. Growth hormone is the primary hormone 45 responsible for stimulating tissue repair, cell replacement, brain function and enzyme production 46 [14]
- 46 [14].

Fish are sensitive indicators of pollutants present in water. These pollutants cause various physiological and physical alterations in fishes. In the present work an attempt has been taken to study the alterations in the levels of Growth hormone has been evaluated in the liver tissue of the

50 fresh water fish *Clarias batrachus*.

# 51 Materials and Methods

The Sago industry effluents were collected from a private Sago industry, situated at Ponnachi near Ammapet of Erode District, Tamil Nadu, India. The effluent from the industry was collected and transported to the laboratory and used for further experiments. Fingerlings of healthy *Clarias batrachus* were brought to the laboratory and acclimatized for 15 days. The fish were well fed during the acclimatized period. Then fish were exposed to control and 25%, 50%, 75% concentrations of treated sago effluents for period of 28 days. Feeding was stopped one day

58 before commencement of the experiment.

After the experimental period the fish exposed to sago effluent were sacrificed. Blood samples were collected from the caudal vein by using the hypodermic micro syringes pre-rinsed with heparin. Blood was centrifuged at 3200 rpm for 15 min and plasma was stored at -26°C until it was used for the estimation of plasma cortisol and growth hormone. The growth hormone level was estimated by ELISA method.

## 64 **Results**

The growth hormone level in the muscle of *Clarias batrachus* was increased with increase in the concentrations of treated sago effluent. The control fish were able to record 0.20ng/ml and the fish treated with the effluents recorded 0.32ng/ml for 25%, 0.35ng/ml for 50% and 0.46ng/ml for 75% respectively.

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- 76 Table.1. Levels of Growth Hormone activity in the blood sample of *Clarias batrachus*
- 77 exposed to control and different concentrations of sago effluent.
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Effluent	Growth	Hormone	level
Concentration	ng/ml		
Control	0.20 ng/ml		
25%	0.32 ng/ml		
50%	0.35 ng/ml		
75%	0.46 ng/ml		

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# Fig.1. Growth Hormone levels in the blood sample of *Clarias* batrachus on exposure to control and different concentrations of treated sago effluent.



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### 82 **Discussion**

Growth hormone affects almost all body tissues. Growth Hormone is considered as a master
hormone which controls many organ and body function. It also regulates itself. The rejuvenating
effects of Growth Hormone are all encompassing, acting on both the mind and body.

Growth hormone is the primary hormone responsible for stimulating tissue repair, cell replacement, and brain function and enzyme production. Growth hormone is the ultimate antiaging therapy and affects almost every cell in the body, rejuvenating the skin and bones, regenerating the heart, liver, lungs and kidneys, bringing back organ and tissue function to more youthful level.

- 91 Growth hormone (GH) has multiple targets and diverse effects in vertebrates. It is a principal
- promoter of growth, and also influences the metabolism. During the past years, it has become
- 93 clear that GH alters the behaviour of fish as it increases appetite, swimming activity, aggression,
- 94 and reduces anti-predator behavior [15].
- Lescroart [16] have reported that the several neurotransmitters and intraperitoneal injections
  induce the secretion of growth hormone and increase in plasma Growth hormone levels in the
  African Cat fish (*Clarias gariepinus*) by sensitive radio immuno assay.
- Peterson et al. [17] have studied the effect of recombinant bovine growth hormone (rbGH) on
  growth rate, feed efficiency, body composition and insulin-like growth factor binding proteins
  (IGFBPs) in Norris.
- 101 The scientists have discovered few synthetic growth hormones like methyl testosterone and ethyl 102 oestranol, which evidensus that the synthetic growth hormone promotes weight in several fishes.
- 103 The fish *Betta splendeus* were given  $17\alpha$  methyl testosterone at different dietary levels under 104 laboratory conditions for 15 days. The maximum growth was found in methyl testosterone 105 treated fish than the control fish [18]. Higher dose of methyl testosterone induced growth in 106 different fish species was reported by various studies [19, 20, 21].
- Sumera et al. [22] have studied the changes in growth hormone and cortisol profile due to lead induced toxicity in *Labeo rohita* and according to their study; Pb acts as endocrine disruptor and has profound influence on the hormonal profiles and specific growth rate of carp. El-Shebly [23] reported that exposing fish to Pb significantly interferes with the activity of serum GH.
- 111 Moreover, exposure to toxicants disrupts hormone signaling cellular pathways favoring the 112 findings of present study [24].

## 113 Conclusion

In the present study the level of GH has shown steady increase, which is more in effluent exposed fish than in control. According to some recent studies, growth hormone levels are increased by sleep, stress, exercise and low glucose levels in the blood. In the current study

- 117 increased plasma GH level in the treated group confirms that sago effluent acts as a stressor.
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### 119 **References:**

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- Rand GM, Petrocelli SR, In: 'Fundamentals of Aquatic Toxicology Methods and Applications'. (Eds.) Hemisphere Publishing Corporation, Washington, U.S.A. (1985): 1-28.
- Racicot YC, Gander M, Leay C, Blood and liver enzymes in rainbow trout (*Salmogairdneri* Rich.) with emphasis in their diagnostic study of CCl4 toxicity and a case of aeromonas infection J Fish Biol 7 (1975): 825.
- Wieser W, Hinterleintner S, Serum enzymes in rainbow trout as tools in the diagnosis of water quality. Bull Environ contam Toxicol 25 (1980): 188-193.
- 4. Ekweozor IKE, Bobmanuel NOK, Gabriel UU, Sublethal effect of ammonial fertilizer
  effluents on the three commercial species from Niger Delta area. J app Sci Environ
  Mange 5 (2001): 63-68.
- Adeyemo OK, Hematological and histopathological effects of cassava mill effluent in *Clarias gariepinus*. Afr J Biomed Res 8 (2005): 179-183.
- 138
  6. Niwelinski J, Zamorska L, Kaczarski F, Pawlicki R, Enzyme histochemistry and microstructure of the human placenta as indicators of environmental pollution Archiwumochrony Srodowiska 3 (1990): 53-59.
- 142 7. Claireaux G, Dutil JD, Physiological responses of Atlantic cod (*Gadusmorrhna*) to
   143 hypoxia at various environmental salinities J *Ep Biol 163 (1992)*: 97-118.
  - 8. Sebert P, Simon B, Barthelemy, Hydrostatic pressure induces a state resembling histotoxic hypoxia in *Anguilla Anguilla* Comp Biochem Physiol105 B (1993) : 255-258.
- Almeida–Val VME, Farias IP, Silva MNP, Duncan WP, Val AL, Biochemical adjustments to hypoxia by Amazon cichids *Braz* J Med Biol Res 28 (1995) : 1257-1263.
  - 10. Akinrotimi OA, Abu OMG, Ansa EJ, Edun OM, George OS, Haematological responses of *Tilapia guineensis* to acute stress. J Nat Appi Sci 5 (2009) : 338 343.
- 154 11. Gabriel UU, Akinrotimi OA, Management of stress in fish for aquaculture development,
   155 Researcher 3(4) (2011): 28- 38.
- 12. Cengiz EJ, Vnlu E, Sublethal effects of commercial deltamethrin on the structure of the
   gill, liver and gut tissues of mosquito fish, *Gambusia affiis* microscopic study Environ
   Toxicol Pharmacol 21 (2006) : 246-253.
- 13. Jung SH, Sim DS, Kim Y, Effects of formalin on haematological and blood chemistry in
   olive flounder *Paralichthysolivaceus* Aquat Res 34 (2000) : 1269-1275.
- 163

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- 14. Wilmore D W. Growth hormone and growth factors in catabolic illness. *Endocrinal metab*, 2 (Supp B) (1995) : 77-84.
- 15. Elisabeth J, Björn TB. Physiological functions of growth hormone in fish with special
   reference to its influence on behavior. Fisheries science 68 Issue sup1 (2002): 742-748
- 16. Lescroart O, Roelats I, Mikolajczyk T, Bosma PT, Schulz RW, Kuhn R and Ollevier
  F. A Radio immuno assay for African cat fish growth hormone: Validation and effects of
  substance modulating the release of Growth Hormone. General and comparative
  Endocrinology 104 (1996): 147-155.
- 175 17. Peterson BC, Small BC, Bosworth BG. Effects of Bovine Growth hormone (Polisac) on
  176 growth performance, body composition and IGFBPs in two strains of channel cat fist.
  177 Aquaculture 232 (2004): 651-663.
- 18. Adsul AD, Singh H. fect of 17α- methyl testosterone on growth and maturation of the
   fish *Betta splendeus*. J Ecobiol, 15(1) (2003):23-27
- 19. Guerrero RD. Use of androgens for the production of male *Tilapia aurea* (Staindanchner)
   Trans. Am. Fish Soc, 4(2) (1975):342-348.
- 20. Nirmala ARC, Pandian TJ. The effect of steroid injection on the food utilization in
   *Channa striatus*. Proc.Indian Acad.Sci, 92 (3) (1983): 221-229.
- 188 21. Sindhu S, Pandian TJ. Effect of administration of different doses of 17α methyl
   189 testosterone in Heteropneustes fossilis (Bloch). Proc. Indian Acad.Sci, 93(6) (1984): 511 190 516.
- 192 22. Sumera S, Husna M, Laiba S, Aqsa C. Changes in Growth Hormone and Cortisol Profile
  193 due to Lead Induced Toxicity in *Labeo rohita*. Turkish Journal of Fisheries and Aquatic
  194 Sciences 18 (2018): 921-926
- 196 23. El-Shebly AA. Protection of Nile Tilapia (*Oreochromis niloticus*) from lead pollution
   197 and enhancement of its growth by α- tocopherol vitamin E. *Research Journal of Fisheries* 198 and Hydrobiology, 4(1) (2009): 17-21.
- 24. Gagnon A, Jumarie C, Hontela A. Effects of Cu on plasma cortisol and cortisol secretion
   by adrenocortical cells of rainbow trout (*Oncorhynchus mykiss*). Aquatic Toxicology,
   78(1) (2006): 59–65. https://doi.org/10.1016/j.aquatox.2006.02.004

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