1	Original Research Article
2	Prevalence and Intensity of Gastrointestinal Parasites of Goats in Belo Sub
3	Division, Boyo Division, North West Region of Cameroon
4	
5	
6	ABSTRACT
7	Background: Goats are one of the oldest domesticated animals by man, about10000 years
8	ago, increases economic wellbeing of the rural populations Worldwide. Goats provide milk,
9	meat, fiber and are also companion pets. They are easy to rear though production is affected by
10	environmental conditions and parasitism.
11	Objectives. : A study was conducted in Belo Sub Division from July 2016 to October 2016
12	with the main objective to investigate the prevalence and intensity of gastrointestinal (GI)
13	parasitic infections in goats. The specific objectives., to identify gastrointestinal parasites of
14	goats and evaluate their intensity., to determine if prevalence and intensity is influenced by
15	either age or sex., to evaluate the effects of the type of management systems on the prevalence
16	and intensity of gastrointestinal parasites and to determine if prevalence and intensity is
17	influenced by locality.
18	Materials and Methods : A total of 499 fecal samples were ramdomly collected directly
19	from the recta of 499 goats in six villages and analyzed for the detection of any parasitic ova
20	or oocysts using standard saturated sodium chloride flotation technique, while fecal
21	egg/oocyst count were estimated using the modified McMaster technique.
22	Results : The study found that all 499 goats with a mean EPG value of $494,3 \pm 374,8$) were
23	found to harbor at least two gastrointestinal parasites. The prevalence and intensity of various
24	parasites encountered respectively were: Eimeria spp (86%), (455,2 ± 400,8), Haemonchus
25	spp (74,5%),(1282.9 ± 1244,4),Toxocara spp (72,5%) (953,3 ± 814,3),Charbertia spp
26	$(55,9\%), (448,2 \pm 416,0), Fasciola spp (45,4\%), (475,0 \pm 338,1), Moniezia spp (42,2\%),$
27	$(828,6 \pm 793,9)$., Oesophagostomum spp $(33,1\%)$, $(638,3 \pm 463,5)$., Strongyloides spp $(32,5)$,
28	$(200,0 \pm 00)$, Trichostrongylus spp (28,3%) (200,0± 00), Trichuris spp (23,7%) (200,0±
29	00), Teladorsagia spp (14,6), (200,0 \pm 00) and Nematodorius spp (8,1%), (50,0 \pm 0,0). There
30	was no significant difference in prevalence (100%) in the different age groups, type of
31	husbandry management system and locality (P>0.05) except for gender where there was
32	significant difference. Female goats had mean EPG value of 526,5± 388,3 compared to males
33	$462,0 \pm 283,6$. Tethered goats had mean EPG value of $513,5\pm 412,4$ compared to free range
34	goats 446,2 \pm 333,1. Belo had mean EPG value of 1233,6 \pm 1145,3) compared to Njinikejem
35	562,0 ± 172,7, Anjin 466,9 ± 197,8 ,Kitchu 373,3 ± 199.6, Baingo 220,8 ± 139,3, Mbessa

36 197,2 ±308,6. Young goats had mean EPG value of 558,1± 331,2) compared to the adult

goats 529,3 \pm 349,5 and the old goats 463,0 \pm 330,7. There was no significant difference in all cases (P>0.05).

39 Conclusion: It can be concluded that gastrointestinal parasitic infections in goats from Belo 40 Sub Division are common, with a very high prevalence. This high prevalence of 41 gastrointestinal parasitism among the goats possibly reflected grazing, low immunity due to 42 malnutrition and lack of anthelminthic treatment programs.

43 Key Words: Prevalence, Intensity, Gastrointestinal Parasites, Goats, Belo Sub Division.

44

45 **INTRODUCTION**

Livestock increases economic status of the rural population and plays a crucial role in the 46 economic well-being of populations Worldwide. Goats are the oldest domesticated animals by 47 48 man¹. Evolutionary biology indicates that goats were domesticated about 10,000 years ago at the dawn of the Neolithic age^2 . The West African dwarf goats are popular as hobby goats due 49 to their easy maintenance resilience and small stature. In rearing them, they do not require as 50 51 much space as the larger dairy goat counterparts. Their gentle and friendly natures make them good companion pets³. Goats are important to man in different spheres and aspects of life. 52 They provide milk which is more easily digestible than cow milk⁴. Their milk is also used 53 in industries in the production of cheese. The rearing of goats provides employment 54 and income to rural populations. In order to rear goats, a minimum investment of money is 55 required, even without specific arrangement for housing and homemade supplied feed. 56 Grazing is mostly done on road-side grass lands and fields⁵ 57

According to Gadahi et *al.*⁶, improper care, unhygienic environment, extreme climate and close contact with infected animals, goats get infected with a variety of parasites Parasitism in goat is a substantial problem plaguing farmers across the nation and it has a highly detrimental effect on the goat industry⁷. Production potential of livestock development programs is plagued in tropical and subtropical areas by prevalence of helminthiasis which causes high mortality and great economic losses⁸.

Goat production and rearing in Belo is challenged with gastrointestinal parasitism being one of the main obstacles. The prevalence of gastrointestinal parasites is related to agroclimatic conditions like quantity and quality of pasture, temperature, humidity and grazing behavior of the host⁹. Infection with gastrointestinal parasites of goats depends on the quantity and species of goats present, general health, age, nutritional and immunological status of the animal. These infections occur mostly as mixed infections of different GIT parasites. Emaciation, persistent diarrhea and weight loss are usually the main symptoms¹⁰.Villous atrophy causes impaired digestion and malabsorption of nutrients, leading to decrease in liveweight gain, fiber and milk production as well as reproductive performance of goats and therefore has a serious impact on animal health and productivity. Hence, GIT parasitism of goats represents the greatest economic constraint and the most important limiting factor of small ruminant production^{11,12,13}.

76 The prevalence of GI parasitic infections may vary and clinical disease may or may 77 not occur due to various environmental factors in different areas. The rich potential from the 78 small ruminant sector is not efficiently exploited due to several constraints already highlighted¹⁴. Amongst the gastrointestinal parasitic diseases of greatest importance in goats 79 are: Nematodes (roundworms), Cestodes (tapeworms), Trematodes (liverflukes) and 80 81 Coccidia¹⁵. Compared to sheep which develop a strong natural immunity around 12 82 months of age, goats have a lower level of immunity to gastrointestinal parasites. This 83 results in goats having greater populations of adult parasites with high egg output. Studies on 84 the epidemiology and other aspects of parasitic infection in goats have been carried out 85 extensively in different parts of the world. On the contrary not much attention has been given to this important area of husbandry in Cameroon in general and in Belo Sub Division 86 87 in particular. Therefore this study was designed to determine the prevalence and intensity of gastrointestinal parasites of goats in Belo Sub division, specifically to identify gastrointestinal 88 parasites and evaluate their intensity, determine if prevalence and intensity is influenced by 89 90 either age or sex, management systems and locality.

91

92 MATERIALS AND METHOD

93 Study area Description

This study was carried out in Belo Sub Division, Boyo Division, North West Region, 94 Cameroon from July 2016 to October 2016. Belo Sub Division is located about 50 km from 95 Bamenda. It is found between latitude $6^{0}4^{I}$ and $6^{0}20^{I}$ North, between longitude $10^{0}11^{1}$ and 96 $10^{0}30^{1}$ East. The entire Sub Division covers a surface area of about 46.068 square kilometers 97 and situated within part of the most mountainous sections of the Western highlands of 98 Cameroon. In terms of relief, its morphology is characterized by two broad valleys separated 99 100 from each other by mountain ridges which all join in the east to form part of the Ijim-Kilum 101 Mountain of more than 3000m above sea level. Four types of vegetation can be distinguished: 102 the montane forest, domesticated sub montane landscape, shrub savanna and semi deciduous 103 forest. The main food crops cultivated in Belo Sub Division include: maize, beans and Irish 104 potatoes for consumption, though some small quantities are sold within and without the Sub 105 Division. The main cash crop is coffee.

106 Two seasons distinguish themselves here. The rainy season is longer starting from 107 March to late October with often torrential rains (about 2900mm on average) accompanied by 108 thunderstorms. The dry season is shorter and starts from November to March, during which 109 temperatures exceed 23⁰C during the Day and drop to about 18⁰C during the night.

110 The total population of Belo Sub Division is about 80775.4735. Only Belo town can111 be classified as being a semi urban town. The rest of the villages are rural.

Belo Sub Division is bounded to the East by Oku Sub Division, to the North by
Njinnikom Sub Division, to the South by Tubah Sub Division and to the West by Bafut Sub
Division.

115 Characteristics of sampled animals

The goats are grazed in open spaces, along the road, yard, and garbage sites and around houses in the municipalities. The age of the goats considered for the study ranged between 0–5 years, characterized as young goats (Less than 6 months old), adult goats (6 to 24 months old inclusive), and old goats (more than 24 months, but Less than 5 years old). Goats of both sexes were involved in the study.

121 **2.3.** Parasitological Techniques

122 **2.3.1.** Collection of samples

During sample collection, information relating to their, age, sex, village and type of 123 husbandry management system were recorded and labels placed on plastic containers. A total 124 of four hundred and ninety nine (499) fecal samples from goats in six villages of Belo sub 125 division were collected and submitted to the Laboratory of Biology and Applied Ecology 126 127 (LABEA) at the University of Dschang, between July and October 2016 to be analyzed for 128 the confirmation of gastrointestinal parasitic infections. About 15 warm, moist, soft fecal pellets were taken directly from the rectum with the finger and placed in sealable plastic 129 containers. Few minutes after collection, the samples were stored in a refrigerator at 10^oC in 130 Belo, from where they were transported within 5 days to the refrigerator in the laboratory in 131 Dschang for analysis. All along, samples were kept cool until analysis, in order to prevent 132 133 trichostrongylid larvae from hatching.

134 Examination of Sample

135 Fecal egg count

The McMaster's fecal egg count is an easy, rapid, and accurate method to determine the quantity of parasite eggs passed in the feces, which helps to determine the effectiveness of various dewormers for a given worm population¹⁶

139 The number of eggs per gram was determined as follows: The egg counts (X_1, X_2) 140 from both chambers of the McMaster's slide were summed (for each type of egg counted) and 141 the total (N) multiplied by a factor of 200.

142	Egg counts in the first Chamber = X_1
143	Egg counts in the second Chamber = X ₂
144	Total egg counts (N) = X_1+X_2
145	Total number of eggs in 2g of fecal sample = 60/0.15X N
146	Therefore number of eggs per gram of feces = 60/0.15 X N/2
147	= 200 X N
148	Parameters studied

149 *** Prevalence** (**P**)

The prevalence of infecton is defined as the percentage of infected individuals on the total number of individuals' examined¹⁸ and was calculated generally and for each parasite using the formula:

153 **Prevalence of infection (P) Number of infected Goats** X 100 154 = Total number of Goats examined 155 156 Intensity (I) 157 The intensity of infection is the mean number of eggs / oocysts of each parasite 158 species per gram of feces of each goat expressed as follows. 159 160 161 162 Intensity of infection (I) = Number of eggs/oocysts per gram of feces (EPG/OPG) obtained by multiplying 200 by N (200XN) 163 164 Classification of GI parasitic infections by virtue of mean EPG. 165 166 The animals were categorized as lightly, moderately and severely (heavily) infected

according to their egg per gram of feces (EPG) counts. Egg counts from 50-799, 800-1200 and over 1200 eggs per gram of feces were considered as light, moderate and heavy infection, respectively ¹⁹.

170 Statistical Analysis

Data was stored in a Microsoft Excel spread, cleaned by checking for errors or missing 171 variables and then exported to SPSS (Statistical Package for Social Science, Version 20) 172 Software for analysis. Summary statistics were generated using the same software. For the 173 174 purpose of modeling these data, explanatory variables were first explored for associations 175 between parasites using χ^2 test. The prevalence of helminth parasites was compared between 176 demographic parameters using the chi square test. Non parametric test of Krustal Wallis was 177 used to compare mean intensity between age group and locality while Mann Whitney was 178 used to compare intensity with animal gender, breeding system and state of health. Before 179 comparison of intensity of infection (EPG), non infected host were discarded. EPG of each 180 parasite was used as variable and breeding system, gender, state of health of animal, age 181 group and locality as factor. They were all tested at 5% significance level.

182

183 **RESULTS.**

184 Overall Prevalence and Intensity of gastrointestinal parasites.

185 The analysis of fecal samples (Table1) revealed that all 499 samples examined, were

186 positive with mixed gastrointestinal parasite infections. There was an overall prevalence of

187 100 percent and a mean EPG value of $(494,3 \pm 374,8)$.

Para	sites	Number	Number of	Prevalence %	Intensity
		examined	infested animals	of infestation	(mEPG/OPD ±SD)*
Nematodes	Nematodirius spp		40	8.1	50.0 ± 0.0
	Haemonchus spp		372	74.5	1282.9 ± 1244.4
	Oesophagostomum spp		163	33.1	638.3 ± 463.5
	Chabertia spp		279	55.9	448 ± 416.0
	Trichuris spp	499	117	23.7	200.0 ± 00
	Strongyloides spp		162	32.5	200.0 ± 00
	Teladorsagia spp		73	14.6	200.0 ± 00
	Toxocara spp		362	72.5	953.3 ± 814.3
	Trichostrongylus spp		141	28.3	200.0 ± 00
Trematodes	Fasciola spp	499	224	45.4	475.0 ± 338.1
Cestodes	Monieza spp	499	208	42.2	828.6 ± 793.9
Protozoa	Eimeria spp	499	429	86	455.2 ± 400.8

188 Table1: Prevalence and Intensity of gastrointestinal parasites

189

190 Influence of gender on Prevalence and Intensity infections

Globally, out of the 499 goats examined, 236 were males, while 263 were females
Both sexes each had 100% prevalence of GI parasitic infections with statistical significance
difference (P<0.05). Multiple infections were more prevalent in female goats than male goats.

- 194 (Table 2). Female goats had the highest mean EPG value of $526,5 \pm 388,3$ compared to 462,0
- $\pm 283,6$ in male goats with no significant difference (P>0.05) (Table 3)..

Parasite			Gender				Total	P-value
		Males			Females		N(%)	
	N0. examined	N0. infected	Prevalence (%)	N0. examined	N0. infected	Prevalence (%)		
Nematodirius spp	chummeu	12	2.4	chuinneu	28	7.7	40 (8.1)	0.028
Haemonchus spp		206	41.3		166	33.3	372 (74.6)	0.001
Oesophagostomum spp		31	6.3		132	26.8	163 (33.1)	0.000
Chabertia spp		86	17.2		193	38.7	279 (55.9)	0.000
Trichuris spp		38	7.7		79	16.0	117 (23.7)	0.000
Eimeria spp		194	38.9		235	47.1	429 (86.0)	0.022
Fasciola spp	236	60	12.2	263	164	33.3	224 (45.4)	0.000
Monieza spp		68	13.8		140	28.4	208 (42.2)	0.000
Strongyloides spp		49	9.8		113 👞	22.6	162 (32.5)	0.000
Teladorsagia spp		19	3.8		54	10.8	73 (14.6)	0.000
Toxocara spp		170	34,1		192	38,5	362(72,5)	0.80
Trichostrongylus		43	8.6		98	19.6	141 (28.3)	0.000

196 Table 2: Prevalence of infections by gender

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198 Table 3: Gender related intensity (mEPG/OPG)

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Parasite		Gender			Total	Р-
		Males	Fema	ales	(mEPG/OPG±SD)	value
	NO	Intensity	NO	Intensity		
	examined	(mEPG/OPG±SD)*	examined	(mEPG/OPG±SD)		
Nematodirius spp		50.0 ± 0.0		50.0 ± 0.0	50.0 ± 0.0	1
Haemonchus spp		798.4± 680.1		1767.3±1808.4	1282.9 ± 1244.4	0.00
Oesophagostomum spp		600.0 ± 0.0	\checkmark	676.6 ± 463.5	638.3 ± 463.5	0.47
Chabertia spp		400.0 ± 0.0		496.3 ± 416.0	448 ± 416.0	0.00
Trichuris spp		200.0 ± 0.0		200.0 ± 00	200.0 ± 0.0	1
Eimeria spp	236	463.9 ± 374.8	263	446.4 ± 426.7	455.2 ± 400.8	0.08
Fasciola spp		460.0 ± 393.7		489.9 ± 282.4	475.0 ± 338.1	0.04
Monieza spp		823.5 ± 810.0		833.6 ± 777.8	828.6 ± 793.9	0.57
Strongyloides spp		200.0 ± 0.0		200.0 ± 00	200.0 ± 0.0	1
Teladorsagia spp		200.0 ± 0.0		200.0 ± 00	200.0 ± 0.0	1
Toxocara spp		1148.2 ± 1144.4		758.3 ± 484.2	953.3 ± 814.3	0.02
Trichostrongylus		200.0 ± 0.0		200.0 ± 00	200.0 ± 0.0	1

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202 Influence of age on Prevalence and Intensity of infection

Table 4 shows the prevalence of infection by age group of the goats examined.70 were young goats, 303 were adults goats, while 126 were old goats. A prevalence of 100% was recorded in each of the 3 age groups with no significance difference (P>0.05). Multiple infections were more prevalent in adult goats than young and old goats Table 5 shows the intensity of GI parasites by age group of the study. The highest mean EPG was recorded by the young goats (558,1 ± 331,2), followed by the adult goats (529,3 ± 349,5) and old goats (463,0 ± 330,7) with no significant difference (P>0.05).

210 Table 4: Age related Prevalence of infections.

Parasite			Age				Total (N)	Р-
	Young		Adults				Prevalence	value
	NO	Prevalence	N0	Prevalence	N0.	Prevalence	(%)	
	infected	(%)	infected	(%)	infected	(%)	_	
Nematodirius spp	13	2.6	7	1.4	20	4.1	40 (8.1)	0.00
Haemonchus spp	40	8.0	240	48.1	92	18.4	372 (74.5)	0.001
Oesophagostomum spp	33	6.7	86	17.4	44	8.9	163 (33.1)	0.001
Chabertia spp	34	6.8	168	33.7	77	15.4	279 (55.9)	0.23
Trichuris spp	13	2.6	85	17.2	19	3.9	117 (23.7)	0.01
Eimeria spp	44	8.8	291	58.3	94	18.8	429 (86.0)	0.00
Fasciola spp	27	5.5	121	24.5	76	15.4	224 (45.4)	0.00
Monieza spp	20	4.1	130	26.4	58	11.8	208 (42.2)	0.14
Strongyloides spp	21	4.2	98	19.6	43	8.6	162 (32.5)	0.84
Teladorsagia spp	0	0	47	9.4	26	5.2	73 (14.6)	0.00
Toxocara spp	58	11.6	210	4.1	94	18.8	362 (72.5)	0.06
Trichostrongylus	12	2.4	85	17.0	44	8.8	141 (28.3)	0.03

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213 Table 5: Age related intensity of infection

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Parasite		Age			7		P value
	Young		Adult		Old		
	N0	Intensity	N0	Intensity	N0	Intensity	_
	examined	(mEPG/OPG)	examined	(mEPG/OPG)	examined	(mEPG/OPG)	
Nematodirius spp		50.0 ± 0.0		50.0±0.0		50.0 ± 0.0	0.06
Haemonchus spp		1300.7 ±1220.1		1473.9±1434.2		1072.9 ±1079.6	0.001
			\sim				
Oesophagostomum spp		566.7 ± 196.6		1052.2±614.7		296.0 ± 102.0	0.001
Chabertia spp		450.0 ± 227.7		750.0±385.9		509.4 ± 437.8	0.23
Trichuris spp		200 ± 00	A	200.0±0.0		200.0 ± 0.0	0.07
Eimeria spp	70	404.5 ± 281.2	303	563.2±422.2	126	397.9 ± 499.0	0.00
Fasciola spp		850.2 ± 498.8		418.2±272.0		475.1±243.5	0.00
Monieza spp		840.0 ±409.3		578.0±594.0		1067.8 ± 972.0	0.14
Strongyloides spp		200.0 ±0.0		200.0±0.0		200.0 ± 0.0	0.84
Teladorsagia spp		200.0 ± 0.0		200.0±0.0		200.0 ± 0.0	0.70
Toxocara spp		1435.6 ± 1140.7		710.6±470.8		714.3 ± 531.4	0.06
Trichostrongylus		200.0 ± 0.0		200.0±0.0		200.0 ± 0.0	0.07

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217 Influence of husbandry systems on infection

The prevalence of GI parasites by type of husbandry management system of the goats is shown in (Table 6). 210 goats were on free range, while 289 goats were tethered. Both types of husbandry management systems recorded each 100% prevalence of GI parasitic infections with no significant difference (P>0.05). Multiple infections were more prevalent in tethered goats than free range goats. The highest mean EPG value (513,5± 412,4) was recorded by tethered goats compared to (446,2 ± 333,1) on free range system with no significant difference (P>0.05).(Table 7)

225 Table 6: Influence of husbandry systems on infection

rasite		Husbandry sy			Total	P-
	Т	ethered	Free rai	nge	N(%)	value
	N0. infected	Prevalence (%)	N0. infected	Prevalence (%)		
Nematodirius spp					40	
	20	4.1	20	4.1	(8.1) 372	0.323
Haemonchus spp	209	41.9	163	32.7	(74.5) 163	0.180
Oesophagostomum spp	104	21.1	59	12.0	(33.1) 73	0.101
Charbertia spp	32	6.4	41	8.2	(14.6) 117	0.008
Trichuris spp	78	15.8	39	7.9	(23.7) 429	0.043
Eimeria spp	239	47.9	190	38.1	(86.0) 224	0.014
Fasciola spp	133	27.0	91	18.5	(45.4) 208	0.756
Monieza spp	109	22.1	99	20.1	(42.2) 162	0.170
Strongyloides spp	71	14.2	91	18.2	(32.5) 73	0.000
Teladorsagia spp	32	6.4	41	8.2	(14.6) 362	0.008
Toxocara spp	196	39.3	166	33.3	(72.5) 141	0.006
Trichostrongylus	95	19.0	46	9.2	(28.2)	0.007

227 Table 7: Influence of husbandry system on intensity of infections

Parasite Husbandry	system	N# N		Total Intensity	P-	
	Tethered	Free range	•	(mEPG/OPG) ±SD	value	
N0 examined	Intensity N0 (mEPG/OPG)±SD exan	nined (mEP	Intensity G/OPG)±SD			
Nematodirius spp	50.00±0.0	V	50.0±0.00	50.0 ± 0	0.0	1
Haemonchus spp	1283.9±1253.3		1331.9±1237.5	$5 1282.9 \pm 1$	244.4 0).774
Oesophagostomum spp Charbertia spp Trichuris spp	642.5±485.1 505.7±445.5 200.0±0.0		674.1±441.9 390.7±386.5 200.0±0.0	638.3 ± 4 448 ± 41 $200.0 \pm$	6.0 0).512).001 1
Eimeria spp 23	36 591.6±525.1	263	318.8±278.7	455.2 ± 4	00.8 0	0.022
Fasciola spp	515.1±345.2		435.1±331.5	475.0 ± 3	38.1 0	0.001
Monieza spp	614.7±848.2		1042.5±739.6	828.6 ± 7	93.9 0	0.000
Strongyloides spp	200.0±0.0		200.0±0.0	$200.0 \pm$	0.0	1
Teladorsagia spp	200.0±0.0		200.0±0.0	$200.0 \pm$	0.0	1
Toxocara spp	1158.5±1046.5		748.1±582.1	953.3 ± 8	14.3	1
Trichostrongylus	200.0±0.0		200.0±0.0	$200.0 \pm$	0.0 0	0.908

228

229 Influence of locality on Prevalence and Intensity of GI parasite infections

The spectrum of gastrointestinal parasites presented in figure 1 shows the prevalence of GI parasites by locality of sampled goats.62 goats were examined from Anjin, 219 from Belo, 60 from Baingo, 47 from Kitchu, 90 from Mbessa, and 21 from Njinikejem. All 6 Villages recorded 100% prevalence each with no significant difference (P>0.05) of GIparasitic infections. Multiple infections were also more prevalent in Belo goats than goats in the other village. The highest mean EPG value was recorded in Belo with no significance difference (p>0.05).

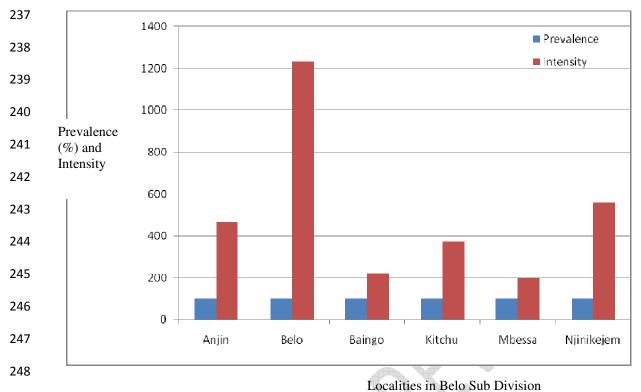


Figure 1: Prevalence and intensity of GI parasitic infection in some Localities

249 **DISCUSSION**

250 **Prevalence and Intensity of GI parasite infections**

251 Goats harbor a variety of gastrointestinal (GI) parasites. Data from this study indicated 252 that gastrointestinal parasitic infections in goats from Belo Sub Division were common, with 253 an overall prevalence of 100%. All the 499 goats examined were infected with at least two 254 gastrointestinal parasites amongst which were Haemonchus spp, Nematodirius spp, Oesophagostomum spp, Chabertia spp, Strongyloides spp, Teladorsagia spp, Toxocara spp, 255 256 Trichostrongylus spp, Trichuris spp, Moniezia spp. Fasciola spp. and coccidian (Eimeria spp) giving a total of twelve parasites (9 Nematodes, 1 cestode, 1 trematode and 1 protozoan). 257 Sathaporn et al.²⁰, Nuraddis et al.¹⁹ and Dogo et al.²¹ also reported slightly similar types of GI 258 parasites. The gastrointestinal parasitic infection rate of 100% recorded in goats during this 259 study agrees with the 100% prevalence reported by Dogo et al.²¹ in Vom and 90.4 reported by 260 Ntonifor et $al.^{22}$ in Jakiri. This is higher than the (87.2%) prevalence reported by Nuraddis et 261 al¹⁹ and (72%) reported by Paul et al²³ in Maiduguri. This is guite high and shows that the 262 agro-ecological and geo-climatic conditions of the study area favor the growth and 263 264 multiplication of these parasites. Climatic conditions, particularly rainfall, are frequently associated with differences in the prevalence of GI parasitic infections, because free-living 265 infective stages (eggs, larvae, cysts, and oocysts) survive longer in moist conditions¹⁹.Belo 266 267 Sub Division expereinces about eight months of rainy season from mid March to mid November and about four months of dry season from mid November to mid March. Since the 268 269 study was conducted from July to October towards the end of the rainy season, higher parasitic infections might be related to the availability of browse and a longer browsing time 270 271 in the warm-rainy season by the host, sufficient moisture and optimum temperature. These create favorable conditions allowing for the larval development, oocyst sporulation and 272 survival of the infective larvae stage²⁴. The high prevalence in this study could also be 273 attributed to illiteracy on the side of the goat keepers and their ignorance or avoidance 274 tendency of preventive measures²⁵. For example, effective pasture management, applied 275 knowledge about host-parasite interactions and interrelations building the base for low 276 277 pasture infection rates for grazing animals, stocking rate reduction and regular intensive monitoring of animal condition that can help optimize animal health status and anthelmintic 278 treatments²⁶. The overall higher prevalence of GI parasitic infections in this study area could 279 also be attributed to lower immunity of hosts as a result of malnutrition^{25, 23}. Among other 280 281 factors that may have further contributed to these discrepancies observed are host breeds

and different husbandry practices. The physiological status of the animals like parturition,
lactation stage and pasture contamination can also influence the prevalence of GI parasites
in different areas²⁴.

Most important to the findings of Nuraddis et al.¹⁹ compared to the present study, 285 Monezia spp. and Emeria spp. were the only cestode and protozoa types found 286 respectively, a finding similar to Kanyari et al.²⁷ Encountered in this study were 287 Nematodorius spp and Toxocara spp, that Nuraddis et al.¹⁹ did not encounter in Jimma, 288 Ethiopia. This difference may be due to variation in climate, parasite evolution or mixed 289 rearing that affect parasitic infection. The most prevalent and commonly observed parasite 290 291 was *Eimeria spp*, with a significant infection rate of (86%), which is higher compared to the low prevalence (48%) reported by Kanyari et al^{27} in Kenya and (20.6%) reported by 292 Nuraddis et al.¹⁹ in Jimma, Ethiopia. Similarly, low prevalence of (18.6) was reported by 293 Dogo et $al.^{21}$,(2017) and Gebeyehu et $al.^{24}$ for *Eimeria spp* in Daegu, Korea. This high 294 prevalence of Eimeria spp in Belo Sub Division may be associated to the fact that Eimeria 295 296 oocysts are much resistant to disinfectants, and can remain in the environment (particularly 297 moist, shady areas) for long periods of time and maintain their infectivity. Stress factors such 298 as tethering, post weaning, dietary changes and other problems can precipitate an outbreak of 299 coccidiosis. In this study, the severity of GI parasitic infection depended on the number of eggs per gram of feces. The intensity of infection measured by fecal egg or oocyst count 300 varied from light to heavy infection. In a high percentage of animals, light parasitic 301 infections were found, while heavy infections were less common. Among these 302 gastrointestinal parasites observed, Haemonchus spp had the highest overall mean EPG value 303 of 1445,2 \pm 1594,4 which is higher than that reported by Ntonifor et al.²² 304

Female goats had higher multiple infections and mean EPG value than male goats from our study and this agrees with the findings of Paul et $al.^{23}$ In a study by Sathaporn et $al.^{20}$, male goats actually had a higher prevalence than female goats which disagrees with our findings. This could be because most of the goats that are tethered in Belo Sub Division are females.

In age related infections, multiple infections and mean EPG value was higher in adults goats than the old and the young goats similar to the report of Gebeyehu et $al.^{24}$ However, this result did not agree with the reports of Kanyari et $al.^{27}$, Gwaze et $al.^{28}$ and Sathaporn et $al.^{20}$ who showed that young goats had higher prevalence of GI parasites than

adult goats. This middle age group had a significant higher prevalence of Eimeria spp 314 315 infections (58,3%) and higher oocyst numbers compared to other age groups in the present study. This did not also agree with Sathaporn et al.²⁰ in Satun, Thailand who reported that 316 317 young goats had a higher prevalence (94.9%) of coccidial infections and higher oocyst 318 numbers in young goats (< 1 year) than older goats (> 2 years). This higher prevalence of GI parasites and of coccidial infections in this age group might be due to the fact that a higher 319 incidence occurs during post weaning stress (since, coccidia is most frequently observed in 320 kids 2 to 4 weeks post weaning), tethering stress and stress related to dietary changes^{29,30} in 321 322 addition to the fact that immunity is low. The low prevalence of coccidial infections in the young goats is probably due to the absence of this stress factors and in old goats probably 323 because of acquired immunity. Although natural immunity develops with repeated exposure³¹ 324 325 younger goats remain highly susceptible. The Institute for International Cooperation in Animal Biologics³² reported that most ruminants stop shedding *Toxocara spp* eggs by the time they 326 327 are 2 to 4 months old and that T.vitulorum infections can be controlled by eliminating 328 patent infections, which occur only in 3 to 10 week old ruminants. Young goats (<6 months 329 old) had the highest mean EPG value of (630.8 ± 268.3) than other age groups, with the 330 highest parasitic intensity $(2137,9 \pm 4493,0)$ shown by *Toxocara spp* in this age group. This high Toxocara spp intensity might probably be due to Transcolostral transmission in the life 331 cycle and sanitation standards related to Toxocara spp.32 This finding even though was not 332 consistent with the reports of Nuraddis et al.¹⁹, was not surprising because naive young and 333 old carriers frequently graze the same areas, coupled with the fact that young goats have low 334 immunity. The intensity of infection is also reportedly related to hygiene level³³ 335

Goats examined in this study were either on free range or tethered systems all under 336 extensive management (grazing). Tethered goats actually had a higher multiple infections and 337 mean EPG than the free range goats. The highest infection rate of (47,9%) was recorded by 338 *Eimeria spp.* in tethered goats This high infection rate and intensity in tethered goats could be 339 explained by the fact that tethering is a stress factor^{29,30} Again most people in Belo Sub 340 341 Division tether goats in the same area throughout the tethering period with little rotation. Consequently, the grazing environment becomes contaminated with various GI parasites eggs 342 and oocysts which infect the goats²⁵ 343

All Villages recorded 100% prevalence each of GI parasitic infection with no statistical significance. These results differed from those of Sathaporn et *al.*²⁰ who reported in Satun, that the prevalence of GI parasites of goats in seven Districts statistically varied from 347 60% to 86.4% (P< 0.05). Belo had a higher multiple infections and mean EPG of $1233.6\pm$ 348 1145,3 compared to other five villages. Geographical consistence of prevalence in Belo Sub 349 Division might be due to the climatic conditions that are consistent in this area. *Eimeria spp* 350 recorded the highest prevalence of 38,2% and Haemonchus spp had the highest mean EPG of 351 4467.3±4396.2 in Belo. Only Belo town can be classified as being a semi urban town. The 352 rest of the villages are rural. These geographical differences in the prevalence of coccidial 353 infections and other infections and high mean EPG value in Belo might be due to the high 354 population density and unhygienic conditions of the area compared to other Villages, which 355 leads to the high infection rates. Inadequate nutrition, however, which is common in this area, may exacerbate the course of GI parasitic infections. The animals are generally malnourished 356 and suffer from other diseases, and are thus not resistant to nematode infection³⁴ 357

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359 CONCLUSION

Goats in Belo Sub Division are infested by gastrointestinal parasites. The adult goats 360 361 recorded higher multiple gastrointestinal parasites and mean EPG value than the young goats 362 and the old goats. Female goats recorded higher multiple gastrointestinal parasites and mean 363 EPG value than male goats. Tethered goats recorded higher multiple gastrointestinal parasites and mean EPG value than free range. Belo recorded higher multiple gastrointestinal parasites 364 365 and mean EPG value than Njinikejem, Anjin, Kitchu, Baingo, Mbessa. Prevailing agro-366 ecological and geo-climatic conditions, illiteracy on the side of goat keepers, avoidance 367 tendency of preventive measures and lack of anthelmintic treatments provide an ideal condition for the transmission of the GI parasitic infections. 368

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370 **REFERENCES**

- 1. Hirsto K.K. (2008). The History of the domestication of goat. 84.
- 2. Zeder M.A. & Hesse B. (2000). The initial domestication of goats (*Capra hircus*) in the
 Zagros Mountains 10,000 years ago. Science 28: 2254 2257.
- 374 3. Mcloed L.A. (1998) Goats as pets, 25.
- 4. Edwards B.A.& Charlotte J.C. (2005). Fresh goat milk for infants: Myths and Realities. *London Daily Telegraph*. 5-7.

- 5. Devendra C. (1992). Studies in the nutrition of the indigenous goat at Malaya and
 requirement of liveweight gain. *Malay. Agri. J.*, 46: 98-118.
- 6. Gadahi J. A., Arshed M. J., Ali Q., Javaid S. B. & Shah S. I. (2009). Prevalence of
 gastrointestinal parasites of sheep and goat in and around Rawalpindi and
 Islamabad, Pakistan. *Vet. World.*,2:51–53.
- 382 7. Jones R. (2001): Sheep Parasites and Diseases .http:// www. kt. iger. bbsrc. ac. uk/ FACT
 383 %20 sheet % 20 PDF % 20 files / kt 36 . pd f . p.2 .
- Al-Quaisy, H.H.K., Al-Zubaidy A.J., Altaf K.I. & Makkawi T.A., (1987):The
 pathogenecity of haemonchosis in sheep and goat in Iraq. Clinical, parasitological and
 haematological findings. *Vet. Parasitol.* 24:221-228.
- 9. Pal R.A. & Qayyum M. (1993): Prevalence of gastrointestinal nematodes of sheep and
 goats in upper Punjab, Pakistan. Pak. Vet. J. 13 (3): 138-141.

389 Parasitology, 46: 33-53

- 390 10. Scheuerle M.(2009). Anthelmintic resistance of *Haemonchus contortus* and the Famacha
 -Method as a tool to delay the development of Anthelmintic resistance.*Inaugural-* 392 *Dissertation*.1st edition
- 393 11. Perry B.D., Randolph T.F. (1999). Improving the assessment of the economic
 394 impact of parasitic diseases and of their control in Production animals. *Vet.* 395 *Parasitol.* 84, 145-168.
- Harper C. K.& Penzhorn B. L.(1999).Occurrence and diversity of coccidia in indigenous,
 Saanen and crossbred goats in South Africa.*Veterinary Parasitology.*, 82: 1-9
- 13. Kagira J. M.&Kanyari P. W. N.(2001). The role of parasitic diseases in causing
 mortalities in small ruminants in a highly productive area of Central Province,
 Kenya. Journal of South Africa *Veterinary Association*, 72: 147-149
- 401 14. Nuraddis I., Mulugeta T., Mihreteab B.& Sisay A.(2014).Prevalence of Gastrointestinal
 402 Parasites of Small Ruminants in and Around Jimma Town, Western Ethiopia.*Acta*403 *Parasitologica Globalis.*,5 (1): 26-32
- 404 15. Bagley C.V. (1997). Internal Parasites. Utah State University Extension, Logan UT.
 405 84322-5600. Available from: http://www.tvsp.org/pdf/sheep/internal-parasites.pdf.

406	16. Cooperative Extension Program (CEP) (2013). Integrated Management of Internal
407	Parasites in Goats Workshop Proceedings.Livestock Production.College of
408	Agriculture, Environment, and Nutrition Sciences.Tuskegee University.Publication
409	Number TUAG 0513-01
410	17. Storey B. (2013).Integrated Management of Internal Parasites in Goats Workshop
411	Proceedings.Livestock production. Publication Number TUAG 0513-01 Editor: Uma
412	Karki
413	18. Thienpont, D., Rochette, F.R., and Vanperijs, O.F.J., (1979). Diagnosis of verminosis by
414	coprological examinations. Beerse, Belgium, Janssen Research Foundation. P. 48-
415	67.
416	19. Nuraddis I., Mulugeta T., Mihreteab B.& Sisay A.(2014).Prevalence of Gastrointestinal
417	Parasites of Small Ruminants in and Around Jimma Town, Western Ethiopia.Acta
418	Parasitologica Globalis .,5 (1): 26-32
419	20. Sathaporn J. Sinsamut S. Nongnuch P. Wissanuwat C. Witaya K.&Roger W. (2012).
420	Gastrointestinal Helminthic and Protozoal Infections of Goats in Satun,
421	Thailand. Trop Med Parasitol., 35:48-54.
422	21. Dogo G. I., Karaye P., Patrobas M.G., Galadima M.& Gosomji I.J.,(2017). Prevalence of
423	Gastrointestinal Parasites and their impact in Domestic animals in Vom, Nigeria.
424	Saudi Journal of Medical and Pharmaceutical Sciences. Vol-3, Iss-3B (Mar, 2017):
425	211- 216
426	22. Ntonifor H. N., Shei S. J., Ndaleh N. W.& Mbunkur G.N., (2013). Epidemiological
427	studies of gastrointestinal parasitic infections in ruminants in Jakiri, Bui Division,
428	North West Region of Cameroon. Journal of Veterinary Medicine and Animal. Vol.
429	5 (12), pp. 344-352,
430	23. Paul B.T., Biu A.A., Gadzam M.A., Ali M., Mana H.P., Jairus Y., (2016).Point
431	prevalence and intensity of gastrointestinal parasite ova/oocysts and its association
432	with body condition score(BCS) of Sheep and goats in Maiduguri, Nigeria. The
433	journal of Advances in Parasitology. Vol 3.pp 88.
434	24. Gebeyehu E. B., Seo M. G., Jung B. Y., Byun J. WOem., J. K., Kim H. Y. & Kwak D.
435	(2013). Prevalence of Gastrointestinal Parasites in Korean Native Goats. The Journal
436	of Animal & Plant Sciences. 23(4): Page: 986-989. ISSN: 1018-7081

- 437 25. William J.A.P. (2001). An Introduction to Animal Husbandry in the Tropics. Bailliere
 438 Tindall, London., 1:92–97.
- 439 26. Gerold R. & Hannah S. (2007). Alternative management strategies to prevent and control
 440 endoparasite diseases in sheep and goat farming systems a review of the recent
 441 scientific knowledge.*Landbauforschung Völkenrode.*, 2 (57):193-206
- 27. Kanyari P.W.N., KagiraJ.M. & MhomaR.J. 2009. Prevalence and intensity of
 Endoparasites in small ruminants kept by farmers in Kisumu Municipality,
 Department of Veterinary Pathology, Microbiology and Parasitology, Faculty of
 Veterinary Medicine, University of Nairobi, Kenya.
- 28. Gwaze F.R., Chimonoyo & DzamaK. (2009). Prevalence and loads of gastrointestinal
 parasites of goats in the communal areas of the Eastern Cape Province of South
 Africa. Small Rumin. Res.84:132–134.
- 29. John M. (2007) .Ucd Vet Views .*California Cattlemen's Magazine* School of Veterinary
 Medicine, University of California-Davis fluke vaccines. *Parasitology Today.*,14,
 224-228.
- 452 30. Leite-Browning M. (2009).Coccidiosis of Goats and Prevention. Alabama Cooperative
 453 Extension System Alabama ,A&M University.
- 31. Newton S.E.& Munn E.A.(1999). The development of vaccines against gastrointestinal
 nematode parasites particularly *Haemonchus contortus*. *Parasitol Today* ., 15: 16222.
- 457 32. Institute For International Cooperation in Animal Biologics (IICAB) (2005).Toxocariasis.
 458 Toxocarosis, Visceral Larva Migrans, Ocular Larva Migrans, Larval Granulomatosis,
 459 Toxocaral Retinitis. Lowa State University. College Of Veterinary Medicine
 460 www.cfsph.iastste.edu/IICAB
- 33. Foreyt W.J. (1990). Coccidiosis and cryptosporidiosis in sheep and goats. Vet Clin North
 Am Food *Anim Pract.*, 6:655-70.
- 463 34. Maichomo M.W., Kagira J.M & Walker T (2004). point prevalence of gastrointestinal parasites in
 464 calves, sheep and goats in Magadi Division, South-Western Kenya. Onderstepoort. *Journal of Veterinary*465 *Research.*,71:257–261