

# **Plant protease inhibitors as a potential source for novel therapeutic agents: a review**

---

## **ABSTRACT**

Most of the currently available therapeutic agents, particularly for cardiovascular disorders and cancers are very expensive and induce some serious side effects. Some of these drugs have also become less effective due to the emergence of antibiotic resistance. There is a necessity and great demand for the development of novel efficacious plant-based agents that are of pharmacologically effective. In this connection, this review focuses on therapeutic potential of plant protease inhibitors. Protease inhibitors are of a particular concern at present due to their potent ability to inhibit protease enzymes that are involved in pathogenesis of various human diseases. In addition to their function as protein-degrading enzymes, protease inhibitors are now well-known for their capability to involve in many biological activities as signaling molecules. Plant protease inhibitors are also engaged in several physiological and pathological processes, such as blood clotting, inflammation, immune regulation, apoptosis and carcinogenesis. Therefore, isolation of plant protease inhibitors and evaluation of their therapeutic capacity against chronic human diseases have become a major research interest. Nevertheless, protease inhibitor content and protease specificity vary significantly even in the same plant species depending on the geographical location and environmental factors. Consequently, it is important to identify potent therapeutic potential of each plant protease inhibitor on human health individually.

*Keywords: Plant-based drugs, Proteases inhibitors, Therapeutic capacity, Chronic human diseases*

## **1. INTRODUCTION**

Plants are naturally well-off with diverse health promoting properties including excellent nutritional and medicinal potentials and play a crucial role in human life since their existence [1]. A considerable percentage of world populations still rely entirely on plants for their nutritional and medicinal needs. Plant parts have been served as the primary raw material for synthesis of many drugs in pharmaceutical industry thus far [2]. From prehistoric times, plants and plant extracts have been used by traditional medical practitioners due to their promising curative capacities [3]. Treatment methods use in Ayurveda, Unani, Homeopathy and Allopathic systems were primarily built on plant-based medication [1].

Epidemiological studies and clinical trials have revealed that plant-based diet is also as a key factor for disease prevention. It has been identified that consumption of diets that are rich in fruits, vegetables and legumes decrease the occurrence of chronic diseases including cancer [4]. For example, incidence of cancer, particularly colon, prostate and breast cancers are significantly less among people who consume large amounts of cereals and pulses including legumes [5]. Likewise, Block [6] reported that probability of having cancers among

people who consume plant-based diets is approximately one-half when compared to the people who consume less amounts of fruits and vegetables. Tusso and Ismail [7] stated that plant-based diet also decreases the chances of having diabetes and heart diseases as well as reduces the possibilities of developing unhealthy conditions like obesity and high blood pressure. It also improves the health status of people who are suffering from diabetes, heart diseases, obesity and high blood pressure. In this context, World Health Organization has recognized medicinal plants as a reliable natural source to discover new drugs and highly recommended to explore new therapeutically effective agents from herbs to treat chronic diseases [8].

Researchers have identified that the therapeutic potential exhibited by plants are mainly due to large array of natural compounds present in them [9]. Among those natural compounds, most of the secondary metabolites are well-studied at biochemical and molecular levels and their therapeutic capabilities have well-recognized. In addition to phytoconstituents, broad spectrum of other compounds that accounts for the medicinal property of plants is identified. Among them, low molecular weight proteins including protease inhibitors (PIs) are one of the most important groups of such compounds [1].

Protease inhibitors are a group of proteins that hinder the function of proteases, the enzymes that are capable of catalyzing the breakdown of proteins via a process known as proteolysis [10]. Presence of PIs in plants is well identified since 1938 and several plants have been screened for presence of PIs [11]. As PIs occur naturally in angiosperms as well as in gymnosperms, they are one of the most abundant types of proteins found in plant kingdom [12]. PIs are small proteins with majority of them having the molecular mass of 8-20 kDa [13]. They are found plentifully in storage organs like tubers and seeds, comprising about 10 % of the overall protein content of plants [14]. However, they also occur in other parts of the plants including leaves [15]. Many types of naturally occurring plant PIs have been isolated and purified from various plants across the globe to evaluate their nutritional and medicinal properties [16].

Many researchers have recently focused on therapeutic capability of PIs present in medicinal plants. It has been identified that plant PIs are effective for inactivating target proteases that are involved in pathogenesis of various human diseases including cardiovascular disorders, cancers, HIV-AIDs, neurodegenerative diseases, inflammatory conditions, ulcerative colitis, pancreatitis and osteoporosis [1,17, 18,19]. Therefore, in future, PIs isolated from plants may add to the existing drug lists or even may replace some of the expensive compounds utilize in drug preparation in pharmaceutical industry at present [1]. As a result, screening of plants for PIs and evaluation of their therapeutic potential have become an important field of study. This review emphasizes on the therapeutic potential of plant PIs for the treatment of chronic human diseases and some of their applications in clinical trials as novel drugs.

## **2. FAMILIES OF PLANT PROTEASE INHIBITORS**

Protease inhibitors are categorized into several groups either by the type of protease they inhibit or by their mechanism of action [20]. Major families of plant PIs include Bowman-Birk Inhibitors (BBI), Serine Protease Inhibitors, Soybean Trypsin Inhibitors (Kunitz), Cereal Trypsin/ $\alpha$ -Amylase Inhibitors, Cysteine Protease Inhibitors, Potato Type I and Type II Inhibitors, Metalloprotease Inhibitors, Serpin Inhibitors, Mustard Trypsin Inhibitors and Squash Inhibitors [15]. Among them, BBI and kunitz are widely distributed in legume seeds and have been isolated and extensively studied on their biophysical, physicochemical and therapeutic properties [14]. Depending on the type of proteases they inhibit, PIs are broadly grouped into four classes known as Cysteine, Serine, Metalloprotease and Aspartic.

Among them, PIs that inhibit cysteine, serine and metallocarboxy proteases are found ubiquitously in plants, whereas PIs active against aspartic proteases have not been found in seeds [21]. All those different types of plant PIs play significant role in plants.

### **3. EFFECT OF PLANT PROTEASE INHIBITORS ON DISEASE CONTROL AND HUMAN HEALTH PROMOTION**

Proteases are proteolytic enzymes that involved in many biological processes of living organisms including regulation of activities of other proteins, modulation of protein-protein interactions, generation of new bioactive molecules, contribution to the processing of cellular information and magnification of molecular signals, etc [22]. Hence, they engage in important biological activities of living organisms, such as cell proliferation and differentiation, tissue remodeling, DNA replication and transcription, wound healing, immunity, inflammation, blood coagulation, necrosis, apoptosis, etc [26]. Nevertheless, imbalance of proteolytic activities results in severe adverse pathological conditions including cancer, neurodegenerative disorders, cardiovascular diseases and inflammatory diseases [20]. Thus, proteases play a key role in pathogenesis of many chronic human diseases. The activity of proteases can be regulated by inhibitors and the process of proteolytic degradation that exhibit variable degrees of affinity with those proteases [24]. Therefore, PIs have been recognized as useful agents to control the progression of such human diseases by inactivating target proteases that are involved in pathogenic processes [1].

In this connection, many plant PIs are explored extensively in pharmaceutical industry for their ability to inhibit or regulate proteases that are engaged in disease progression. Despite their role as protein-degrading enzymes, currently PIs have also been recognized as essential mediators in several physiological and pathological processes, such as inflammation, blood clotting, apoptosis and endocrine activities [1]. Further, diverse plant PIs are known to be efficient against cardiovascular disorders, inflammatory conditions, osteoporosis, neurodegenerative diseases and many viral and parasitic infections [25, 1]. It is well-known that the preventive role of plant PIs against the progression of human diseases is largely depend on their bioactivities.

Previous studies have revealed that plant PIs exhibit various biological activities, such as anti-oxidant, anti-cancer, anti-inflammatory, anti-microbial, etc, suggesting the therapeutic importance of plant PIs for the treatment of chronic diseases. Trypsin PI has been recognized for its high anti-oxidant and free radical scavenging activities that could possibly be useful in preventing oxidative stress mediated diseases including cancers [26]. BBI demonstrates its anti-inflammatory activity through suppression of functions of proteases, such as trypsin, leukocyte elastase and human cathepsin G, which are released from human inflammatory cells [27, 28, 29].

Due to PIs capacity to inhibit bacterial proteases that are involved in vital physiological activities, they have also been recognized as potent anti-bacterial agents [30]. Fistulin, a PI present in the leaves of *Cassia fistula* demonstrated anti-bacterial activity against wide range of pathogenic microorganisms, such as *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Bacillus subtilis* and *Klebsiella pneumonia* [31]. Besides, trypsin inhibitor detected from *Moringa oleifera* flowers (MoFTI) demonstrated potent anti-bacterial activity against broad spectrum of both Gram-negative and Gram-positive bacteria [32]. Despite the anti-bacterial activity, Kunitz PI isolated from *Nicotiana tabacum* (NtKT11) expressed potent anti-fungal activity against *Rhizoctonia solani* and moderate activity towards *Rhizopus nigricans* and *Phytophthora parasitica* var. *nicotianae* [33]. Further, trypsin inhibitor present in *Pseudostellaria heterophylla* also showed anti-fungal potential against *Fusarium oxysporum* [34].

Protease inhibitors have also identified as useful agents for immunosuppressive activities [35] and interestingly, they have also been evaluated comprehensively for their capacity to serve as anti-tumor agents against several human cancers. For instance, plant BBI is believed as a therapeutically important agent for cancer prevention due to its potential to inhibit cancer initiation irrevocably especially at the initial stages [36, 37]. Further, Yavelow *et al.* [38] and Fields *et al.* [37] reported that BBIs also have an ability to stop malignant transformation *in vitro*.

Importantly, several plant PIs are currently under the assessment of human clinical studies, such as blood coagulation, immune regulation, platelet aggregation and anti-carcinogenesis [39, 40]. The above evidence suggests that plant PIs possibly exhibit a significant therapeutic potential against multiple chronic diseases. Thus, PIs are a promising source for the development of new therapeutic agents. Yet, the most extensively studied plant PIs origin from Poaceae, Fabaceae and Solanaceae plant families [41]. Among the plants, potato, pineapple, squash, wheat, buckwheat, chickpea, pigeonpea, bean, barley, millet, groundnut and corn are recognized as good sources of PIs [1]. Besides, it has been identified that these naturally occurring PIs differ in protease specificity and several types of PIs might present in a single plant [1, 41, 42]. Additionally, PIs isolated from various plants showed different physicochemical properties and bioactivities. As a result, it is important to identify potent therapeutic potential of each plant PI on human health individually.

#### **4. THERAPEUTIC IMPORTANCE OF DIFFERENT GROUPS OF PLANT PROTEASE INHIBITORS**

Evaluation of plant PIs for their ability to combat against various clinical diseases has begun in 1950's [43]. Since then, researchers have isolated and purified PIs from diverse plant families and studied them for their medicinal potentials using both *in vitro* and *in vivo* techniques. Among them, BBI, serine, kunitz, trypsin and cysteine PIs have been examined extensively for their applications in biotechnology and biomedicine as novel therapeutic agents for human illnesses [42, 44]. Besides, mainly Fabaceae, Poaceae, Solanaceae, Malvaceae, Rutaceae and Moringaceae plant families have been studied comprehensively for PIs to date [41, 45]. Evidence from previous research has revealed that various types of plant PIs play a role in disease prevention and management due their potent biological activities.

##### **4.1. BBI and BBI-type PIs**

Da Costa Souza *et al.* [46] have purified a BBI-type PI from seeds of *Vigna unguiculata* and it is termed as the black-eyed pea trypsin/chymotrypsin inhibitor (BTCI). It has been identified that BTCI has an effect on the catalytic activities of MCF-7 breast cancer cells and purified 20S proteasomes in which BTCI mainly crosses the breast cancer cell membranes and restrain the proteasomes present in the cytoplasm and nucleus. As a result of the above activities, BTCI has been recognized as a PI with a potential to inhibit trypsin, chymotrypsin and caspase activities of 20S proteasomes [46].

BBI and BBI-type PIs have been isolated from some other leguminous seeds as well, particularly from *Glycine max* L. and *Cicer arietinum* L. BBIs present in both plants have exhibited high level of inhibitory activity against trypsin and chymotrypsin [19, 47]. An elevated level of trypsin is found in ovarian and colorectal tumors, and therefore inhibitory activity against trypsin makes those plant PIs to act as possible candidates for anti-cancer agents. Imbalance levels of trypsin and chymotrypsin are also responsible for pancreatitis and inhibitory activity against those proteolytic enzymes possibly can reduce the development of pancreatitis. Additionally, BBI-type PI isolated from *Cicer arietinum* L. has an

ability to considerably inhibit the progression of breast cancer (MDAMB-231) and prostate cancer (LNCaP) cells [19].

In addition to the inhibition of trypsin and chymotrypsin, BBIs present in seeds of *Glycine max* L. showed an ability to decrease proteolytic activities of other proteases, such as cathepsin G, elastase, chymase, urokinase protein activator, serine protease-dependent matrix metalloproteinases, PI3 kinase and mitogen activated protein kinase. It also mediated the up regulation of expression of connexin 43 [48]. An *in vivo* study conducted in rodents showed that BBI therapy of *Glycine max* L. has a significant suppressive activity on inflammation in anal gland and colon after subjected to tumorigenic agents [49]. BBIs are also effective against development of colorectal carcinomas with no any adverse outcomes or side effects given the fact that they can diminish the activation and rapidity of such malignancies even at very low concentrations of BBIs in the diet of dimethylhydrazine (DMH) using rat models [39]. Moreover, BBIs of *Glycine max* L. are capable of suppressing the progression of prostate cancer cells [50]. Recent studies have also revealed the potential of BBI to prevent the development of prostate tumors via its anti-proliferative activity by stimulating connexin 43 present in the gap junctions of transgenic rats [51, 52]. Further, *Glycine max* L. BBI exerts its anti-cancer effect against breast cancer due to its capacity to involve in proteasome inhibition [53].

Despite the application of BBI in animal models, it has also been used in several clinical trials to understand its therapeutic importance in human diseases. For instance, clinical trials were conducted using BBI as a treatment in patients with prostate cancer, lung cancer, ulcerative colitis, oral leukoplakia, encephalomyelitis, gingivitis and multiple sclerosis. In those human trials, BBI was given in the form of Bowman-Birk inhibitors concentrate (BBIC), an investigational novel drug prepared from a concentrated soybean protein extract with high content of BBI. Each patient was administered with varying concentrations of BBIC doses per day with the highest dosage up to 1066 chymotrypsin inhibitory units (C.I.U) [54]. None of the completed clinical trials of phase I and phase II-a reported toxic effect, antibody neutralization effect or any other adverse outcome of BBIC [55, 56]. Interestingly, BBIC has exhibited a dose-related response against oral leukoplakia and prostate cancer [56]. Similarly, Malkowicz *et al.* [57] conducted a randomized and double-blinded phase I human trial among 19 men who were with an early stage of prostate cancer. This study showed that six months treatment of BBIC decreases the amounts of prostate-specific antigen (PSA), which is clinically important indicator for the diagnosis of prostate cancer. BBI has also exhibited a potent anti-inflammatory activity without any toxicity in patients who have ulcerative colitis [18]. Even with the limited number of clinical trials completed up to date, BBI has been suggested as an excellent therapeutic agent against common chronic diseases. At present, various other PIs are also subjected to assessment in clinical trials to evaluate their therapeutic activity against chronic human diseases [1].

#### **4.2. Serine PIs**

Serine PIs are also proven as one of the prime candidates for novel therapeutics in drug discovery due to their ability to control proteolysis. In fact, serine plant PIs have been found to be effective in treating inflammatory diseases, respiratory disorders, cardiovascular diseases, HIV-AIDS, immune diseases, neurodegenerative disorders (i.e., Alzheimer disease) and cancers [58, 59]. At present, eight classes of serine plant PIs have been identified on the basis of the sequence of their amino acid chains [60] and among them, soybean-derived serine PI, PI 1 and PI 2 have been well-examined [61, 62]. Almost all of the formerly examined serine PIs were isolated from Cucurbitaceae, Euphorbiaceae, Fabaceae, Poaceae and Solanaceae plant families [59]. At present, other plant families are also being explored for serine PIs.

Bacha *et al.* [59] have isolated serine PI from the leaves of *Rhamnus frangula* (RfIP1) that belongs to the family Rhamnaceae. It has been identified that serine PI from *Rhamnus frangula* exhibited high level of inhibitory activity against some pharmacologically important proteases, such as chymotrypsin, thrombin, trypsin, cathepsin B and collagenase. The ability of RfIP1 to inhibit thrombin allows it to act as a potent anti-coagulant agent. Cathepsin B is associated with tumor invasion, and thereby the capability of RfIP1 in restraining cathepsin B exhibited its ability to act as an anti-tumor agent. Besides, cathepsin B is known as the most harmful and toxic protease in many protozoan infections including Leishmaniasis [63]. Thus, RfIP1 is also capable of playing protective role against parasitic growth and recognized as a novel therapeutic agent for Leishmaniasis [64]. RfIP1 also possessed an *in vitro* anti-bacterial activity against broad spectrum of Gram-positive and Gram-negative bacteria. Accordingly, RfIP1 can be used as a potent drug in pharmaceutical industry due to its therapeutic importance as an anti-coagulant and anti-cancer.

#### **4.3. Kunitz and Kunitz-type inhibitors**

Kunitz and kunitz-type inhibitors (KTIs) occur naturally in wide range of plants. KTIs are about 8-22 kDa proteins that contain reactive site for trypsin and two disulphide bonds [65]. Neuhoof *et al.* [66] stated that KTIs detected from seeds of *Bauhinia bauhinioides* and *Bauhinia rufa* have an ability to decrease the formation of edema in perfused rabbit lungs. Wang and Ng [34] have isolated and purified a Kunitz-type trypsin inhibitor with a molecular mass of 20.5 kDa from roots of *Pseudostellaria heterophylla* and it has expressed anti-fungal and trypsin inhibitory activities comparable to soybean trypsin inhibitor. Additionally, KTIs are also capable of preventing cell invasiveness via reducing extracellular signal transduction in human ovarian cancer cells [67, 68].

Nakahata *et al.* [69] have screened *Enterolobium contortisiliquum*, which belongs to the family Fabaceae for PIs. From *Enterolobium contortisiliquum* seeds, they have purified a trypsin inhibitor (EcTI) of molecular mass of 20 kDa and it has been recognized as a KTI. EcTI has demonstrated an inhibitory activity for clinically important proteases, such as trypsin, chymotrypsin, plasmin and plasma kallikrein. It has also demonstrated an inhibitory activity against gastric cancer cells via controlling integrin-dependent signal transduction pathways [70]. Importantly, it is capable of inhibiting human cancer cell lines namely breast (SkBr-3 and MCF-7), leukemia (K562 and THP-1) and colorectal (HCT116 and HT29), in addition to its effect on human mesenchymal stem cells (hMSCs) and primary fibroblasts. Furthermore, EcTI is identified as a cytotoxic agent against cancer cells, which is not harmful for normal body tissue [69].

#### **4.4. Cysteine PIs**

Like most of the other PIs, cysteine PIs are also widespread in plant kingdom. They occur in seeds of several monocotyledonous including rice (*oryzacystatin*) and maize as well as in dicotyledonous plants, such as apple fruit [71, 72, 73, 74, 75, 76]. Researchers have identified cysteine PIs as an important tool for the utilization in pharmaceutical industry due to their potent health benefits. Obayomi *et al.* [77] purified cysteine PI from *Tetracarpidium conophorum* which is competitively inhibit papain, a proteolytic enzyme that aids in digestion. The biological activities and physicochemical properties illustrated by cysteine PI suggest it as an effective therapeutic agent for the treatment of clinical conditions, such as cancer, neurodegenerative diseases and atherosclerosis.

#### 4.5. Novel plant PIs

In addition to known plant PIs, researches have also focused on purifying novel PIs from plants. For example, CGPIs is one of such PIs extracted from *Coccinia grandis* leaves with a molecular mass of 14.3 kDa [78]. CGPIs have exhibited a significant inhibitory effect for chymotrypsin and bovine pancreatic trypsin. The CGPIs demonstrated a dose-dependent inhibitory activity against the progression of colon cancer. Further, CGPIs also capable of hindering sporulation and mycelial growth in infective microorganisms including both bacteria and fungus, such as *Staphylococcus aureus*, *Escherichia coli*, *Klebsiella pneumonia*, *Bacillus subtilis*, *Proteus vulgaris*, *Aspergillus flavus*, *Mucor indicus*, *Cryptococcus neoformans*, *Candida albicans* and *Penicillium notatum* [78].

Four different types of new PIs were purified from the seeds of *Lavatera cashmeriana* and they were termed as LC-pi I, II, III and IV. According to their molecular size, these four types of PIs have been identified as KTIs [79]. These PIs were found to be involving in inhibiting trypsin, chymotrypsin and elastase. All of them have exhibited *in vitro* anti-cancer effect on colon (HCT-116), leukemia (THP-1) and lung (NCIH322) cancer cell lines [79, 80, 81]. Besides, LC-pi I has also showed potent inhibitory activity against prostate and breast cancer cell lines due to its capacity to hinder elastase, trypsin and chymotrypsin [81]. Further, LC-pi I has demonstrated a significant anti-bacterial activity against *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* [80].

Protease inhibitors isolated from mature leaves and seeds of *Moringa oleifera* showed inhibitory activity against proteases that are of pathological importance, such as trypsin, chymotrypsin, elastase, thrombin, cathepsin B, papain and cysteine. They have also demonstrated complete inhibitory activity against proteases isolated from *Bacillus licheniformis* and *Aspergillus oryzae* [45]. Thereby, *Moringa oleifera* PI controls a wide range of protease mediated activities including fibrinolysis, tissue remodeling, aging and development of cancer [24, 58, 82]. The affinity of *Moringa oleifera* PI towards thrombin expressed its potential to act as an anti-coagulant. Further, trypsin inhibitor detected from *Moringa oleifera* flowers (MoFTI) demonstrated potent anti-bacterial activity against broad spectrum of Gram-negative bacteria including, *Escherichia coli*, *Proteus mirabilis* and *Salmonella enteritidis*, and Gram-positive bacteria, such as *Bacillus subtilis*, *Enterococcus faecalis* and *Staphylococcus aureus* [32]. Bijina *et al.* [45] also reported the possible use of PIs isolated from *Moringa oleifera* in sea food preservation. As a result, *Moringa oleifera* is a promising source for the development of useful drugs in pharmaceutical industry.

Some researchers have studied on anti-HIV effect of PIs from plant sources using pepsin as a substitute for HIV protease. In this context, a potent anti HIV-1 protease activity of *Adhatodavasic* was detected using pepsin [83]. Besides, water extracts of *Sesbania grandiflora* leaves and flowers have also demonstrated mild anti-HIV activity [84]. Another study also stated the inhibitory potential of PIs isolated from *Terminalia arjuna* (fruit and bark), *Terminalia chebula* (aerial parts and fruit) and *Terminalia horrida* (resin) of Combretaceae family against HIV-1 protease [85].

A study conducted in Sri Lanka on five medicinal plants, such as *Mangifera zeylanica*, *Sesbania grandiflora*, *Terminalia bellerica*, *Terminalia catappa* and *Terminalia chebula* revealed the presence of therapeutically important PIs. In this study, pepsin and trypsin were used as model enzymes to examine aspartic and serine protease inhibitory activities, respectively. Among the tested extracts, the highest inhibitory activity against pepsin and trypsin was detected from the water extracts of the bark of *Terminalia catappa* and *Sesbania grandiflora*. Additionally, inhibitory activities exhibited by those PIs remained stable with varying temperature [86].

## 5. CONCLUSION AND FUTURE RECOMMENDATIONS

Plant PIs are recognized as very effective curative agents for various human diseases because of their potent ability to inhibit proteases that are involved in the process of pathogenesis. In addition, therapeutically important plant PIs can also be provided naturally through routine diet containing legumes, soybean, rice, potato, etc. Depending on the geographical location and environmental factors, PI content and activity may differ even in the same plant species. Therefore, there is a necessity to conduct further studies to identify possible plant sources for therapeutically important PIs in various geographical locations and also important to characterize isolated PIs and assess their therapeutic applications individually. Investigation of PIs from diverse plant families may lead to discover novel PIs with different pharmacological and therapeutic importance. Identification and scientifically validation of the therapeutic effect of plant PIs could also be useful for people to identify plants that are rich in nutritional and medicinal properties.

## CONSENT

Not applicable

## ETHICAL APPROVAL

Not applicable

## REFERENCES

- [1] Srikanth S, Chen Z. Plant protease inhibitors in therapeutics-focus on cancer therapy. *Front Pharmacol.* 2016; 7(470):1–19.
- [2] De Feo V. Medicinal and magical plants in the northern Peruvian Andes. *Fitoterapia.* 1992; 63(5):417–40.
- [3] Chaudhury RR, Rafei UM. Traditional medicine in Asia. World Health Organization Regional Office for South-East Asia (New Delhi): SEARO Regional Publications; 2001. Accessed 30 October 2017  
Available: [http://apps.searo.who.int/pds\\_docs/b0104.pdf](http://apps.searo.who.int/pds_docs/b0104.pdf).
- [4] Hasler CM. Functional foods: their role in disease prevention and health promotion. *J Food Technol.* 1998; 52(2):57–62.
- [5] Correa P. Epidemiological correlations between diet and cancer frequency. *Cancer Res.* 1981; 41(9 Part 2):3685–90.
- [6] Block E. The organosulfur chemistry of the genus *Allium*-implications for the organic chemistry of sulfur. *Angew Chem Int Ed.* 1992; 31(9):1135–78.
- [7] Tusso PJ, Ismail MH, Ha BP, Bartolotto C. Nutritional update for physicians: plant-based diets. *Perm J.* 2013; 17(2):61–6.
- [8] Nascimento GG, Locatelli J, Freitas PC, Silva GL. Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. *Braz J Microbiol.* 2000; 31(4):247–56.

- [9] Ghosh S, Parihar VS, More P, Dhavale DD, Chopade BA. Phytochemistry and therapeutic potential of medicinal plant: *Dioscorea bulbifera*. J Med Chem. 2015; 5(4):154–59.
- [10] Farady CJ, Craik CS. Mechanisms of macromolecular protease inhibitors. Chembiochem. 2010; 11(17):2341–46.
- [11] Ryan CA. Proteolytic enzymes and their inhibitors in plants. Annu Rev Plant Physiol. 1973; 24(1):173–96.
- [12] Mutlu A, Gal S. Plant aspartic proteinases: enzymes on the way to a function. Physiol Plant. 1999; 105(3):569–76.
- [13] Chye ML, Sin SF, Xu ZF, Yeung EC. Serine proteinase inhibitor proteins: exogenous and endogenous functions. In Vitro Cell Dev Boil Plant. 2006; 42(2):100–8.
- [14] Ryan CA. Protease inhibitors in plants: genes for improving defenses against insects and pathogens. Annu Rev Phytopathol. 1990; 28(1):425–49.
- [15] Leo FD, Volpicella M, Licciulli F, Liuni S, Gallerani R, Ceci LR. Plant-PIs: a database for plant protease inhibitors and their genes. Nucleic Acids Res. 2002; 30(1):347–48.
- [16] Majumdar DD. Recent updates on pharmaceutical potential of plant protease inhibitors. Int J Med Pharm Sci. 2013; 3(4):101–20.
- [17] Hussein G, Miyashiro H, Nakamura N, Hattori M, Kawahata T, Otake T, Kakiuchi N, Shimotohno K. Inhibitory effects of Sudanese plant extracts on HIV-1 replication and HIV-1 protease. Phytother Res. 1999; 13(1):31–6.
- [18] Lichtenstein GR, Deren JJ, Katz S, Lewis JD, Kennedy AR, Ware JH. Bowman-birk inhibitor concentrate: a novel therapeutic agent for patients with active ulcerative colitis. Dig Dis Sci. 2008; 53(1):175–80.
- [19] Magee PJ, Owusu-Apenten R, McCann MJ, Gill CI, Rowland IR. Chickpea (*Cicer arietinum*) and other plant-derived protease inhibitor concentrates inhibit breast and prostate cancer cell proliferation in vitro. Nutr Cancer. 2012; 64(5):741–8.
- [20] Habib H, Fazili KM. Plant protease inhibitors: a defense strategy in plants. Biotechnol Mol Biol Rev. 2007; 2(3):68–85.
- [21] Valueva TA, Mosolov VV. Protein inhibitors of proteinases in seeds: 1. classification, distribution, structure and properties. Russ J Plant Physiol. 1999; 46(3):307–21.
- [22] López-Otín C, Bond JS. Proteases: multifunctional enzymes in life and disease. J Biol Chem. 2008; 283(45):30433–37.
- [23] Turk B. Targeting proteases: successes, failures and future prospects. Nat Rev Drug Discov. 2006; 5(9):785–99.
- [24] Laskowski M Jr, Kato I. 1980. Protein inhibitors of proteinases. Annu Rev Biochem. 49(1):593–626.
- [25] Jayawardana BDS, Perera HKI, Rajapakse S. Protease inhibitory activity of some medicinal plants in Sri Lanka [Abstract]. Book of Abstracts of the Peradeniya University Research Sessions. 2012; 17(1):126.
- [26] Shamsi TN, Parveen R, Fatima S. Trypsin inhibitors demonstrate antioxidant activities, inhibit A549 cell proliferation and increase activities of reactive oxygen species scavenging enzymes. Indian J Pharmacol. 2017; 49(2):155–60.
- [27] Larionova NI, Gladysheva IP, Tikhonova TV, Kazanskaia NF. Inhibition of cathepsin G and elastase from human granulocytes by multiple forms of the bowman-birk type of soy inhibitor. Biokhimiia. 1993; 58(9):1437–44.

- [28] Gladysheva IP, Larionova NI, Gladyshev DP, Tikhonova TV, Kazanskaia NF. The classical bowman-birk soy inhibitor is an effective inhibitor of human granulocyte alpha-chymotrypsin and cathepsin G. *Biokhimiia*. 1994; 59(4):513–18.
- [29] Tikhonova TV, Gladysheva IP, Kazanskaia NF, Larionova NI. Inhibition of elastin hydrolysis, catalyzed by human leukocyte elastase and cathepsin G, by the bowman-birk type soy inhibitor. *Biokhimiia*. 1994; 59(11):1739–45.
- [30] Supuran CT, Scozzafava A, Clare BW. Bacterial protease inhibitors. *Med Res Rev*. 2002; 22(4):329–72.
- [31] Arulpandi I, Sangeetha R. Antibacterial activity of fistulin: a protease inhibitor purified from the leaves of *Cassia fistula*. *ISRN Pharm*. 2012; (1):1–4.
- [32] Pontual EV, Napoleão TH, Assis CRD, Bezerra RS, Xavier HS, Navarro DMAF, Coelho LCBB, Paiva PMG. Effect of *Moringa oleifera* flower extract on larval trypsin and acetylcholinesterase activities in *Aedes aegypti*. *Arch Insect Biochem Physiol*. 2012; 79(3):135–52.
- [33] Huang H, Qi SD, Qi F, Wu CA, Yang GD, Zheng CC. NtKT11, a kunitz trypsin inhibitor with antifungal activity from *Nicotiana tabacum*, plays an important role in tobacco's defense response. *FEBS J*. 2010; 277(19):4076–88.
- [34] Wang HX, Ng TB. Concurrent isolation of a kunitz-type trypsin inhibitor with antifungal activity and a novel lectin from *Pseudostellaria heterophylla* roots. *Biochem Biophys Res Commun*. 2006; 342(1):349–53.
- [35] Mahajan SG, Mehta AA. Immunosuppressive activity of ethanolic extract of seeds of *Moringa oleifera* Lam. in experimental immune inflammation. *J Ethnopharmacol*. 2010; 130(1):183–6.
- [36] Wattenberg LW. Inhibition of carcinogenesis by minor dietary constituents. *Cancer Res*. 1992; 52(7):2085–91.
- [37] Fields C, Mallee P, Muzard J, Lee GU. Isolation of bowman-birk-inhibitor from soy bean extracts using novel peptide probes and high gradient magnetic separation. *J Food Chem*. 2012; 134(4):1831–38.
- [38] Yavelow J, Collins M, Birk Y, Troll W, Kennedy AR. Nanomolar concentrations of bowman-birk soy bean protease inhibitor suppress x-ray induced transformation in vitro. *Proc Natl Acad Sci*. 1985; 82(16):5395–99.
- [39] Kennedy AR. The bowman-birk inhibitor from soybeans as an anticarcinogenic agent. *Am J Clin Nutr*. 1998; 68(6):1406–12.
- [40] Chaudhary NS, Shee C, Islam A, Ahmad F, Yernool D, Kumar P, Sharma AK. Purification and characterization of a trypsin inhibitor from *Putranjiva roxburghii* Seeds. *Phytochemistry*. 2008; 69(11):2120–26.
- [41] Richardson MJ. Seed storage proteins: the enzyme inhibitors. 1991. New York (NY): Academic Press.
- [42] Majumdar DD. Recent updates on pharmaceutical potential of plant protease inhibitors. *Int J Med Pharm Sci*. 2013; 3(4):101–20.
- [43] Vogel R, Trautschold I, Werle E. Natural Proteinase Inhibitors. 1968. New York (NY): Academic Press.
- [44] Tamir S, Bell J, Finlay TH, Sakal E, Smirnoff P, Gaur S, Birk Y. Isolation, characterization and properties of a trypsin-chymotrypsin inhibitor from Amaranth seeds. *J Protein Chem*. 1996; 15(2):219–29.
- [45] Bijina B, Chellappan S, Krishna JG, Basheer SM, Elyas KK, Bahkali AH, Chandrasekaran M. Protease inhibitor from *Moringa oleifera* with potential for use as therapeutic drug and as seafood preservative. *Saudi J Biol Sci*. 2011; 18(3):273–81.

- [46] Da Costa Souza L, Camargo R, Demasi M, Santana JM, De Sa CM, De Freitas SM. Effects of an anticarcinogenic bowman-birk protease inhibitor on purified 20S proteasome and MCF-7 breast cancer cells. *PLoS One*. 2014; 9(1):e86600–e86610.
- [47] Hatano KI, Kojima M, Tanokura M, Takahashi K. Solution structure of bromelain inhibitor VI from pineapple stem: structural similarity with bowman-birk trypsin/chymotrypsin inhibitor from soybean. *Biochemistry*. 1996; 35(17):5379–84.
- [48] Losso JN. The biochemical and functional food properties of the bowman-birk inhibitor. *Crit Rev Food Sci Nutr*. 2008; 48(1):94–118.
- [49] Billings PC, Newberne PM, Kennedy AR. Protease inhibitor suppression of colon and anal gland carcinogenesis induced by dimethylhydrazine. *Carcinogenesis*. 1990; 11(7):1083–86.
- [50] Sun XY, Donald SP, Phang JM. Testosterone and prostate specific antigen stimulate generation of reactive oxygen species in prostate cancer cells. *Carcinogenesis*. 2001; 22(11):1775–80.
- [51] McCormick DL, Johnson WD, Bosland MC, Lubet RA, Steele VE. Chemoprevention of rat prostate carcinogenesis by soy isoflavones and by bowman-birk inhibitor. *Nutr Cancer*. 2007; 57(2):184–93.
- [52] Tang M, Asamoto M, Ogawa K, Naiki-Ito A, Sato S, Takahashi S, Shirai T. Induction of apoptosis in the LNCap human prostate carcinoma cell line and prostate adenocarcinomas of SV40T antigen transgenic rats by the bowman-birk inhibitor. *Pathol Int*. 2009; 59(11):790–6.
- [53] Chen YW, Huang SC, Lin-Shiau SY, Lin JK. Bowman-birk inhibitor abates proteasome function and suppresses the proliferation of MCF-7 breast cancer cells through accumulation of MAP kinase phosphatase-1. *Carcinogenesis*. 2005; 26(7):1296–306.
- [54] Sugano M. *Soy in Health and Disease Prevention*. 2005. NewYork (NY): CRC Press.
- [55] Armstrong WB, Kennedy AR, Wan XS, Atiba J, McLaren CE, Meyskens FL. Single-dose administration of bowman-birk inhibitor concentrate in patients with oral leukoplakia. *Cancer Epidemiol Biomarkers Prev*. 2000; 9(1):43–7.
- [56] Armstrong WB, Wan XS, Kennedy AR, Taylor TH, Meyskens FL. Development of the bowman-birk inhibitor for oral cancer chemoprevention and analysis of neu immunohistochemical staining intensity with bowman-birk inhibitor concentrate treatment. *Laryngoscope*. 2003; 113(10):1687–702.
- [57] Malkowicz SB, McKenna WG, Vaughn DJ, Wan XS, Probert KJ, Rockwell K, Marks SH, Wein AJ, Kennedy AR. Effects of bowman-birk inhibitor concentrate (BBIC) in patients with benign prostatic hyperplasia. *The Prostate*. 2001; 48(1):16–28.
- [58] Koivunen E, Ristimäki A, Itkonen O, Osman S, Vuento M, Stenman UH. Tumor-associated trypsin participates in cancer cell-mediated degradation of extracellular matrix. *Cancer Res*. 1991; 51(8):2107–12.
- [59] Bacha AB, Jemel I, Moubayed NM, Abdelmalek IB. Purification and characterization of a newly serine protease inhibitor from *Rhamnus frangula* with potential for use as therapeutic drug. *3 Biotech*. 2017; 7(148):1–13.
- [60] Mosolov VV, Valueva TA. Proteinase inhibitors and their function in plants: a review. *Appl Biochem Microbiol*. 2005; 41(3):227–46.
- [61] Kim JY, Park SC, Hwang I, Cheong H, Nah JW, Hahm KS, Park Y. Protease inhibitors from plants with antimicrobial activity. *Int J Mol Sci*. 2009; 10(6):2860–72.
- [62] Meulenbroek EM, Thomassen EA, Pouvreau L, Abrahams JP, Gruppen H, Pannu NS. Structure of a post-translationally processed heterodimeric double-headed

- kunitz-type serine protease inhibitor from potato. *Acta Crystallogr D Struct Biol*. 2012; 68(7):794–99.
- [63] Keppler D, Pagano M, Dalet-Fumeron V, Engler R. Regulation of neoplasm-specific cathepsin B by cysteine-protease inhibitors present in cancerous exudates. *Acc Acad Sci Ser III Life Sci*. 1985; 300(13):471–4.
- [64] Mottram JC, Brooks DR, Coombs GH. Roles of cysteine proteinases of trypanosomes and leishmania in host-parasite interactions. *Curr Opin Microbiol*. 1998; 1(4):455–60.
- [65] Laskowski M, Qasim MA. What can the structures of enzyme-inhibitor complexes tell us about the structures of enzyme substrate complexes?. *Biochim Biophys Acta Protein Struct Molec Enzym*. 2000; 1477(1):324–37.
- [66] Neuhofer C, Oliva MLV, Maybauer D, Maybauer M, Oliveira CD, Sampaio MU, Sampaio CA, Neuhofer H. Effect of plant kunitz inhibitors from *Bauhinia bauhinioides* and *Bauhinia rufa* on pulmonary edema caused by activated neutrophils. *J Biol Chem*. 2003; 384(6):939–44.
- [67] Kobayashi H, Suzuki M, Kanayama N, Terao T. A soybean kunitz trypsin inhibitor suppresses ovarian cancer cell invasion by blocking urokinase upregulation. *Clin Exp Metastasis*. 2004; 21(2):159–66.
- [68] Suzuki K, Yano T, Sadzuka Y, Sugiyama T, Seki T, Asano R. Restoration of connexin 43 by Bowman-Birk protease inhibitor in M5076 bearing mice. *Oncol Rep*. 2005; 13(6):1247–50.
- [69] Nakahata AM, Mayer B, Ries C, De Paula CAA, Karow M, Neth P, Sampaio MU, Jochum M, Oliva MLV. The effects of a plant proteinase inhibitor from *Enterolobium contortisiliquum* on human tumor cell lines. *J Biol Chem*. 2011; 392(4):327–36.
- [70] De Paula CAA, Coulson-Thomas VJ, Ferreira JG, Maza PK, Suzuki E, Nakahata AM, Nader HB, Sampaio MU, Oliva MLV. *Enterolobium contortisiliquum* trypsin inhibitor (EcTI), a plant proteinase inhibitor, decreases in vitro cell adhesion and invasion by inhibition of Src protein-focal adhesion kinase (FAK) signaling pathways. *J Biol Chem*. 2012; 287(1):170–82.
- [71] Abe K, Emori Y, Kondo H, Suzuki K, Arai S. Molecular cloning of a cysteine proteinase inhibitor of rice (Oryzacystatin), homology with animal cystatins and transient expression in the ripening process of rice seeds. *J Biol Chem*. 1987; 262(35):16793–97.
- [72] Abe K, Kondo H, Arai S. Purification and characterization of a rice cysteine proteinase inhibitor. *Agric Biol Chem*. 1987; 51(10):2763–8.
- [73] Abe M, Abe K, Kuroda M, Arai S. Corn kernel cysteine proteinase inhibitor as a novel cystatin superfamily member of plant origin. *Eur J Biochem*. 1992; 209(3):933–7.
- [74] Pernas M, Sánchez-Monge R, Gómez L, Salcedo G. A chestnut seed cystatin differentially effective against cysteine proteinases from closely related pests. *Plant Mol Biol*. 1998; 38(6):1235–42.
- [75] Ryan SN, Laing WA, McManus MT. A cysteine proteinase inhibitor purified from apple fruit. *Phytochemistry*. 1998; 49(4):957–63.
- [76] Sakuta C, Oda A, Konishi M, Yamakawa S, Kamada H, Satoh S. Cysteine proteinase gene expression in the endosperm of germinating carrot seeds. *Biosci Biotechnol Biochem*. 2001; 65(10):2243–8.
- [77] Obayomi A, Adeola SA, Bankole HA, Raimi OG. Characterization of partially purified cysteine protease inhibitor from *Tetracarpidium conophorum* (African walnut). *Afr J Biochem Res*. 2015; 9(2):26–34.

- [78] Satheesh LS, Murugan K. Antimicrobial activity of protease inhibitor from leaves of *Coccinia grandis* (L.) Voigt. Indian J Exp Biol. 2011; 49(5):366–74.
- [79] Rakashanda S, Ishaq M, Masood A, Amin S. Antibacterial activity of a trypsin-chymotrypsin-elastase inhibitor isolated from *Lavatera cashmeriana* Camb. seeds. J Anim Plant Sci. 2012; 22(4):983–6.
- [80] Rakashanda S, Mubashir S, Qurishi Y, Hamid A, Masood A, Amin S. Trypsin inhibitors from *Lavatera cashmeriana* Camb. seeds: isolation, characterization and in vitro cytotoxicity activity. Int J Pharm Sci Invent. 2013; 2(5):55–65.
- [81] Rakashanda S, Qazi AK, Majeed R, Rafiq S, Dar IM, Masood A, Hamid A, Amin S. Antiproliferative activity of *Lavatera cashmeriana*-protease inhibitors towards human cancer cells. Asian Pac J Cancer Prev. 2013; 14(6):3975–8.
- [82] Higgins GA, Oyler GA, Neve RL, Chen KS, Gage FH. Altered levels of amyloid protein precursor transcripts in the basal forebrain of behaviorally impaired aged rats. Proc Natl Acad Sci. 1990; 87(8):3032–6.
- [83] Kennedy AR, Troll W. Protease Inhibitors as Cancer Chemopreventive Agents. 2012. New York (NY): Springer Science and Business Media.
- [84] Tewtrakul S, Subhadhirasakul S, Rattanasuwan P. HIV-1 protease inhibitory effects of some selected plants in Caesalpiniaceae and Papilionaceae families. J Sci Technol. 25(4):509–514.
- [85] Filho JR, De Sousa Falcão H, Batista LM, Filho JM, Piuvezam MR. 2010. Effects of Plant Extracts on HIV-1 Protease. Curr HIV Res. 2003; 8(7):531–44.
- [86] Perera HKI, Jayawardana BDS, Rajapakse S. Heat stable protease inhibitors from *Sesbania grandiflora* and *Terminalia catappa*. Br J Pharm Res. 2016; 11(4):1–9.