

Original Research Paper

Comparative Effect of Aspirin, Meloxicam and *Terminalia catappa* leaf extract on serum levels of some Inflammatory Markers in Alloxan induced Diabetic rats

ABSTRACT

Background: The association between diabetes mellitus and inflammation is established.

Aim: The study was aimed at comparing the levels of some inflammatory biomarkers in diabetic rats treated with aqueous leaf extract of *Terminalia catappa*, non steroidal anti-inflammatory drugs (NSAIDs) and exogenous insulin.

Materials and Methods: Thirty six (36) Wistar rats were assigned to 6 groups of 6 animals each. Group 1 and 2 served as normal and diabetic controls and received orally 5ml/kg body weight of distilled water. Group 3 was diabetic treated orally with 130mg/kg body weight of aqueous leaf extract of *Terminalia catappa*. And groups 4, 5 and 6 were administered orally with aspirin (30mg/kg), meloxicam (2mg/kg) and 0.75U/kg body weight of insulin subcutaneously. Diabetes was induced with intraperitoneal injection of 150mg/kg body weight of alloxan solution and diabetes confirmed after 72 hours with blood glucose levels ≥ 200 mg/dl. The experiment lasted for 14 days.

Results: The results showed significant ($P < 0.05$) increase in serum levels of C-reactive protein, Interleukin-6 and blood fibrinogen in diabetic group compared to control. These inflammatory biomarkers was significantly ($P < 0.05$) reduced by the extract, aspirin, meloxicam and insulin.

Conclusion: In conclusion, levels of reduction of these inflammatory biomarkers by leaf extract of *Terminalia catappa* was much compared to aspirin and meloxicam. This may present the extract as a more potent anti-inflammatory agent than NSAIDs and could complement the function of insulin in diabetes treatment.

Key words: C-reactive protein, Interleukin-6, fibrinogen, inflammation, diabetes mellitus, *Terminalia catappa*, Insulin, aspirin, meloxicam, NSAIDs

1. INTRODUCTION

Several studies have reported the existing association between hyperglycaemia, inflammation and diabetes complications [3]. Investigations on various markers of inflammation in different population groups have

confirmed this association [8, 9]. Biomarkers of inflammation such as interleukin-6, C-reactive protein, tissue necrosis factor, fibrinogen and even total white blood cell count [15] are increased in cases of injury, infection [13] or obesity [36]. It is now well appreciated that low grade chronic inflammation is central to the pathology of the pancreatic islet in type 1 diabetes and also plays an important role in the pathogenesis of type 2 diabetes, including obesity-related insulin resistance, impaired insulin secretion, and diabetes-related vascular complications [42]. The development of overt diabetes of any category results in hyperglycemia. Other findings have shown reverse causality in which hyperglycemia is itself pro-inflammatory and this through the generation of oxidative stress [10]. Thus there is a vicious cycle of inflammation – hyperglycemia and hyperglycemia – inflammation association. Following the first line of causality and effect, since inflammation leads to poor glycaemic control, then treatment of inflammation with non-steroidal anti-inflammatory drugs (NSAIDs) may help improve glycaemic control. On the other hand, hypoglycemic and anti-diabetic drugs can reduce inflammation by reducing the hyperglycemia. But accumulated evidence on association between inflammation and complications of diabetes [2, 1] has attracted the consideration of targeting inflammation to ameliorate diabetes, prevent its progression and diminish vascular complications [34]. However, the effects of immunomodulatory treatments are not limited to tissues involved in disease pathophysiology and thus might have unwarranted side effects. It is reported that some anti-diabetic agents may alleviate systemic and tissue-specific inflammation [5, 6, 7].

The anti-inflammatory effects might be mediated via their metabolic effects on hyperglycemia and hyperlipidemia or by directly modulating the immune system [3]. For example, glitazones that reduce insulin resistance, metformin which improves insulin sensitivity partly through activation of Adenosine Monophosphate-activated Kinase (AMPK), a key regulator of cellular energy homeostasis to exert both anti-inflammatory and antioxidant effects [4] and exogenous insulin which increases glucose uptake and disposal by stimulation of insulin dependent glucose transporter are all found to reduce inflammation [16]. Moreover, vasoactive drugs such as statins and Adenosine Converting Enzyme (ACE) inhibitors/angiotensin receptor antagonists often prescribed to people with diabetes also counteract inflammation and reduce the risk of diabetes complications in type 2 diabetes [15, 36]. The complications of diabetes are divided into macrovascular (myocardial infarction and stroke) [6] and microvascular (nephropathy and retinopathy) complications. Several researches has supported that reduction in hyperglycemia may ameliorate the microvascular complications but such certainty is not attained in macrovascular complication [14]. This is because many cytokines have been released thus activating some inflammatory pathways which may not only be inhibited by glycaemic control. Therefore the use of drugs targeting some more specific biomarker and inflammatory signaling pathways may provide the needed reduction in morbidity and mortality resulting from diabetic complications. This research aims at comparing the effect of *Terminalia catappa* leaf extract, non steroidal anti-inflammatory drugs on some inflammatory biomarkers in diabetic rats.

2. MATERIALS AND METHODS

2.1 Preparation of Plant Extract

Fresh leaves of *Terminalia catappa* were collected at the premises of the University of Uyo and the area was free of pesticides and other contaminants. The leaves were authenticated by a botanist at the Department of Botany and Ecological studies, University of Uyo. The leaves were then washed with clean water to remove debris. The water was blotted out and kept overnight at room temperature to dry up. The clean leaves were pulverized and 5000g of the pulverized leaves were soaked in 5 litres of deionised water for 18 hours. The mixture was filtered using muslin cloth and evaporated to dryness using thermostatic water bath at 45°C until a semi solid paste of 204.18g of the extract was obtained after evaporation representing a percentage yield of 4.08%. The extract was stored in refrigerator for later use.

2.2 Preparation of Experimental Animal

Healthy adult male albino Wister rats weighting between 150-200g were used for the study. The animals were procured from the animal house, Department of Physiology, Faculty of Basic Medical Sciences, University of Uyo. The animals were housed in a well ventilated cage in the animal house and they were allowed to acclimatize for two weeks and maintained in a 24 hours dark and light cycle. The animals were fed with standard pellets (from Guinea Feeds, Plc Nigeria) and have access to water *ad libitum*.

2.3 Induction of diabetes

Diabetes was induced by intraperitoneal injection of alloxan monohydrate at a dose of 150mg/Kg body weight [19, 18, 20]. The animals were assessed for development of diabetes after 72 hours [17] by obtaining blood sample from the tip of the tail. The blood sample was dropped into glucose strip to measure the glucose level using a glucometer (One Touch Ultra, Life Scan Inc, U.S.A). Blood glucose of $\geq 200\text{mg/dl}$ was considered diabetic (normal range of blood glucose in rat is 80 – 120mg/dl) and were used for the experiments [18, 17].

2.4 Experimental Design

The experimental animals were randomly distributed into Six (6) groups of six (n=6) rats per group as follows:

Group 1: Control group administered with only distilled water orally at a dose of 5ml/kg body weight.

Group 2: Diabetic group administered with only distilled water orally at a dose of 5ml/Kg body weight.

Group 3: Diabetic group treated with aqueous leaf extract of *Terminalia catappa* at a dose of 130mg/Kg body weight by oral administration.

Group 4: Diabetic group treated orally with 30mg/kg body weight of aspirin

Group 5: Diabetic group treated orally with 2mg/kg body weight of meloxicam

Group 6: Diabetic group treated with exogenous Insulin at a dose of 0.75U/Kg body weight by subcutaneous administration.

2.5 Rat C-reactive protein assay

Serum C-reactive protein level was analysed by ELISA method. Commercial rat C-reactive protein analysis kit using Sandwich-ELISA (Enzyme Linked Immunosorbent Assay) method. Standards or samples were added to the appropriate microelisa strip plate wells and combined to the specific antibody. The absorbance or optical density (OD) is measured spectrophotometrically at a wavelength of 450nm and readings are obtained using Microtiter Plate Reader and the Optical Density value is proportional to the concentration of C-reactive protein.

2.6 Rat Interleukin-6 assay

Serum from the blood of the experimental animals was used to analyse the Interleukin-6. Interleukin-6 assay was carried out with commercially prepared rat Interleukin-6 analysis kit using Sandwich-ELISA (Enzyme Linked Immunosorbent Assay) method. Standards or samples are added to the appropriate microelisa strip plate wells and combined to the specific antibody. The absorbance or optical density (OD) was measured spectrophotometrically at a wavelength of 450nm and readings are obtained using Microtiter Plate Reader and the Optical Density value is proportional to the concentration of Interleukin-6.

2.7 Rat Fibrinogen Assay

Whole blood was collected into plain sample bottle and used for determination of fibrinogen. The fibrinogen level was determined by Enzyme Linked Immunosorbent Assay (ELISA) method. Standards or samples were added to the appropriate microelisa strip plate wells and combined to the fibrinogen antibody. The absorbance or optical density (OD) was measured by spectrophotometry at wavelength of 450nm and the result read on a microplate immediately. The Optical Density value is proportional to the concentration of fibrinogen. The assay was performed at room temperature (18-25°C).

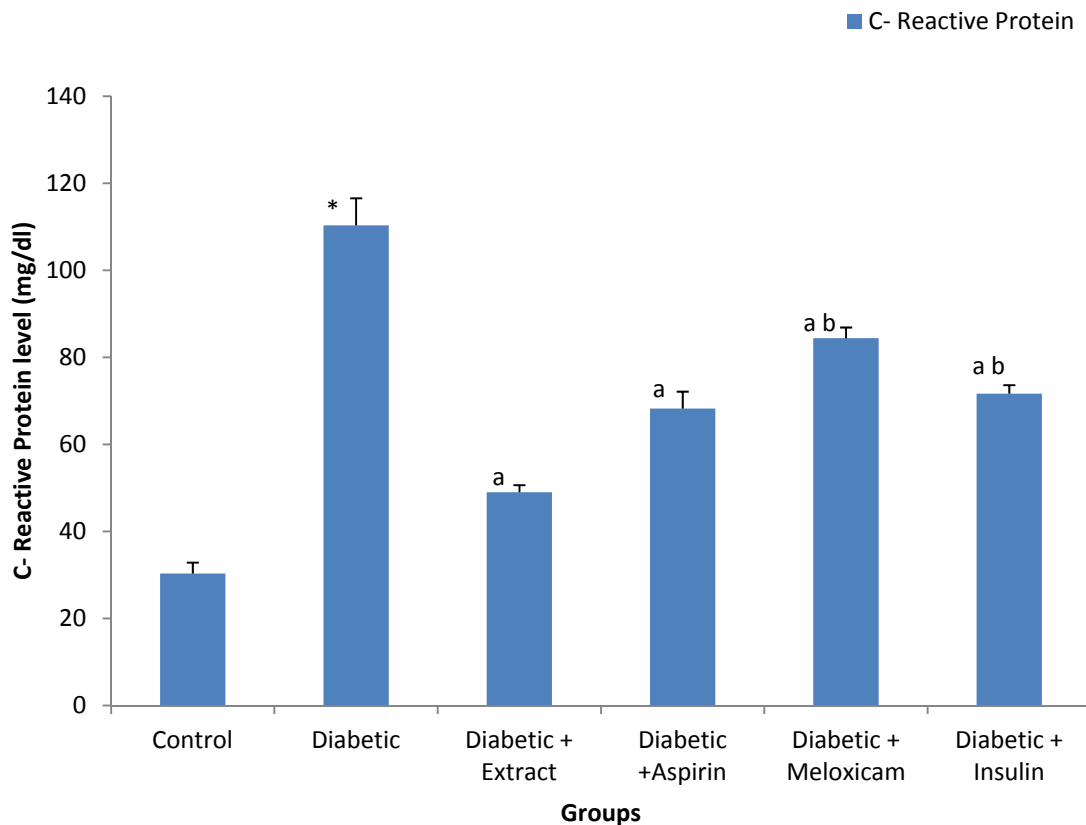
2.8 Statistical Analysis

The data obtained from the result was subjected to statistical testing using one way ANOVA followed by Tukey test using Graph Pad Prism software 6.0. Data was expressed as mean \pm standard error of mean (SEM). Results with values of $P < 0.05$ were considered significant.

3. RESULTS

3.1 C- reactive protein

The C-reactive protein analysis is shown in (figure 1). The serum level of C-reactive protein in control group was 30.33 ± 2.51 mg/dl while in the diabetic none treated group, the serum level significantly ($P < 0.05$) rise to 110.33 ± 6.24 mg/dl. This was reduced significantly to 49.0 ± 1.63 mg/dl, 68.25 ± 3.89 mg/dl and 84.4 ± 2.48 g/dl in the diabetic extract, aspirin and meloxicam treated groups respectively. Insulin treated group also show significant ($P < 0.05$) reduction to a mean value of 71.67 ± 1.95 mg/dl.

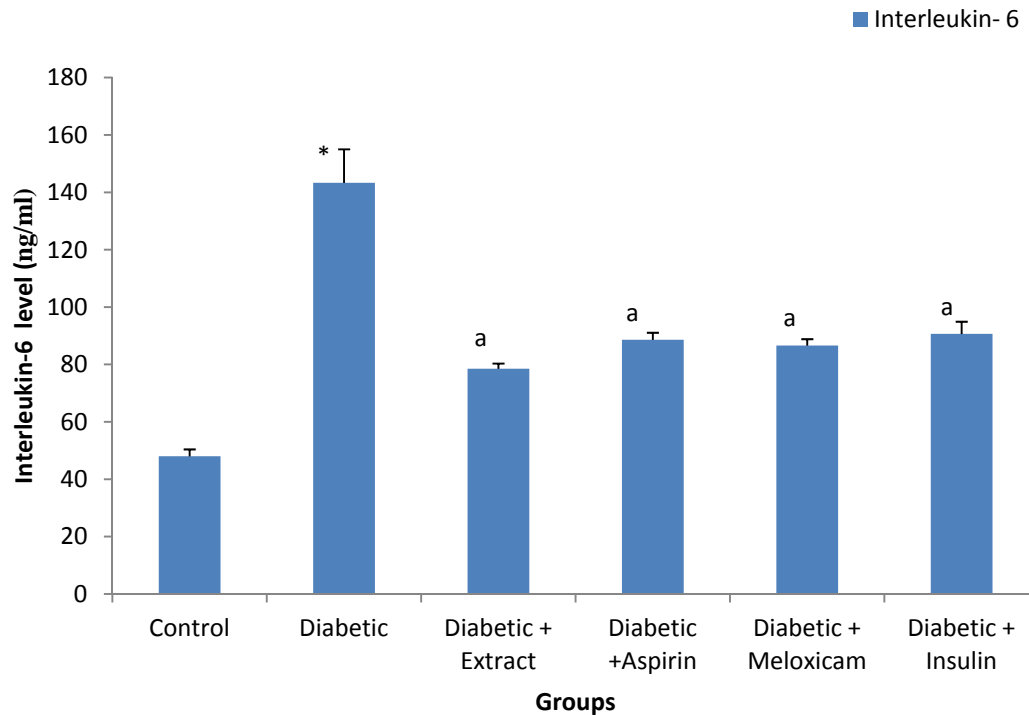


Values are mean \pm SEM. * =Significant change compared with control group ($P < 0.05$)
a, b= Significant change compared with diabetic control group ($P < 0.05$)

Fig. 1 Serum C - reactive protein in experimental group compared with control and diabetic control groups.

3.2 Interleukin - 6

The result of the inflammatory marker, interleukin-6 is represented in (figure 2). Serum level of Interleukin-6 in the control group was $48.0 \pm 2.44 \text{ ng/ml}$ and $143.33 \pm 11.69 \text{ ng/ml}$ diabetic group. Diabetic extract treated group has a serum level of $78.5 \pm 1.84 \text{ ng/ml}$ and this was significantly ($P < 0.05$) lower than the diabetic group. The diabetic aspirin and meloxicam treated groups have respectively the mean values of $88.6 \pm 2.47 \text{ ng/ml}$ and $86.6 \pm 2.24 \text{ ng/ml}$ which was significant when compared to diabetic group. The mean value in the insulin treated diabetic group also showed significant ($P < 0.05$) reduction to a mean value of $90.67 \pm 4.29 \text{ ng/ml}$.

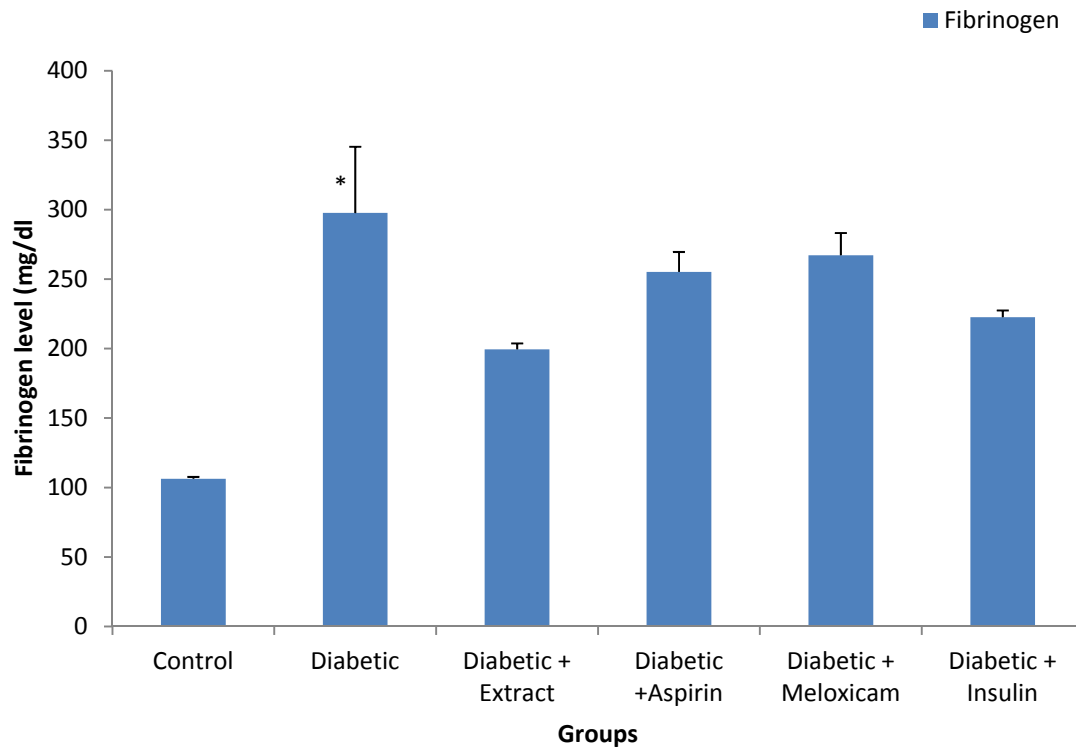


Values are mean \pm SEM. * =Significant change compared with control group ($P < 0.05$)
a= Significant change compared with diabetic control group ($P < 0.05$)

Fig. 2 Serum level of Interleukin-6 in experimental group compared with control and diabetic control groups.

3.3 Fibrinogen

The results of serum level of fibrinogen are as shown in (figure 3). The result showed that the serum level of fibrinogen increased from mean value of 106.33 ± 1.33 mg/dl in the control to 148.33 ± 9.51 mg/dl in diabetic group. The mean value reduced significantly ($P < 0.05$) to 297.67 ± 47.64 mg/dl in the diabetic extract treated group. Also there was a significant reduction to a mean value of 255.2 ± 14.46 mg/dl, 267.2 ± 16.02 mg/dl and 222.67 ± 4.81 mg/dl in diabetic groups which received aspirin, meloxicam and insulin treatments respectively.



Values are mean \pm SEM. * =Significant change compared with control group ($P < 0.05$)

Fig. 3 Serum level of Fibrinogen in experimental group compared with control and diabetic control groups

4. DISCUSSION

The levels of inflammatory biomarkers in diabetic rats treated with aqueous leave extract of *Terminalia catappa*, non-steroidal anti-inflammatory drugs (NSAIDs) and exogenous insulin were analysed. The result showed that C-reactive protein level was high in the diabetic group compared to control. The result of this study is consistent with our previous result and with other research findings which has established elevated C-reactive protein levels in people with impaired glucose tolerance and frank diabetes [32, 31]. However, the observed increase was reduced by the aqueous extract of *Terminalia catappa*, aspirin, meloxicam and insulin in their respective diabetic treated groups. These results reveal that the reduction in C-reactive protein and Interleukin-6 by the extract was stronger than that of aspirin and meloxicam. Moreover, fibrinogen level follows a similar pattern of being increased significantly in the diabetic group and was lowered significantly by the extract, aspirin, meloxicam and insulin, with insulin presenting a stronger effect compared to the other agents. Both aspirin and meloxicam are non steroidal anti-inflammatory drugs used in the treatment of inflammation besides other functions as analgesia and anti-pyretic. These anti-inflammatory drugs inactivate cyclooxygenase (COX) enzymes with meloxicam being more COX-2 specific blocker than COX-1 [26]. Blocking COX enzymes leads to suppressed production of prostaglandins and thromboxanes [41] as the mechanism of reducing inflammation.

The level of reduction of these biomarkers varies between the two anti-inflammatory drugs. It was observed that C-reactive protein was more reduced by aspirin than meloxicam while the level of reduction of Interleukin-6 and fibrinogen were similar between the two drugs. The irreversible effect of aspirin on COX-1 enzyme [25] makes the drug different from other anti-inflammatory drugs in addition to regular pathways such as the formation of nitric oxide free radicals in the body as independent mechanism in reducing inflammation [23], uncoupling of oxidative phosphorylation in mitochondria [24] and signal modulation through NF-kB [22]. The present study cannot explain if the specificity in inactivation of the COX isoforms; COX-1 and COX-2 may play a role in the effectiveness of aspirin in the reduction of the acute phase inflammatory marker, C-reactive protein compared to meloxicam despite the similarity in the reduction of pro-inflammatory cytokine; Interleukin-6.

It was observed that insulin reduced C-reactive protein and Interleukin-6 but have comparatively higher effect on fibrinogen than the other agents. Following the established association between hyperglycaemia and inflammation, the anti-inflammatory effect of insulin will first involve lowering of blood glucose and this might show existing positive link between hyperglycaemia, fibrinogen level and risk of cardiovascular disease in diabetes. It is reported that insulin suppresses important inflammatory mediators namely: Intercellular adhesion molecule-1(I-CAM-1), Monocyte Chemoattractant Protein-1 (MCP-1) expression, necrosis factor NF-kB binding [11, 12] in human aortic endothelial cells *in vitro* via nitric oxide signalling pathway [27,28, 29]. It is suggested that Toll like receptors (TLRs) which plays an important role in tissue inflammation and damage such as cardiac ischemia and atherosclerosis [30] is also suppressed by insulin. Although the reduction of C-reactive protein and Interleukin-6 by insulin in this study is not as much as the

non steroidal anti-inflammatory drugs, the more pronounced effect on fibrinogen may suggest the usefulness of insulin in thrombosis more than the non steroidal anti-inflammatory drugs.

In view of these findings, NSAIDs such as aspirin or meloxicam adjunct to insulin therapy in diabetes may be advocated. But reports on various side effects of these drugs are documented. Gastrointestinal toxicity and increased risk of cardiovascular events such as heart attack and stroke are associated with many non steroidal anti-inflammatory drugs [21]. Meloxicam is specifically contraindicated in persons with hypertension and diabetes despite its modification for gastrointestinal tolerability thereby ruling out its use in diabetes condition. Moreover, recent clinical findings have revealed the development of NSAIDs induced enteropathy [39, 40]. It is reported that the permeability of the small intestinal mucosa in a single dose of NSAID is within 12 hours and this is by uncoupling of the mitochondrial phosphorylation which breaks the integrity of the mucosa junction normally protected by COX1 and COX 2 [37, 38] however this enteropathy is asymptomatic and physicians continue to treat with this drug because the side effect is vague [35].

5. CONCLUSION

In conclusion, more pronounced reduction was observed in serum levels of C-reactive protein and Interleukin-6 but not fibrinogen by the aqueous leaf extract of *Terminalia catappa* compared to aspirin and meloxicam. And aspirin effect was more marked on C-reactive protein than meloxicam while the two affected Interleukin-6 and fibrinogen similarly. This suggests that the extract may activate the pathways used by these agents and other pathways not known to express its anti-inflammatory actions. However this assertion requires a more empirical investigation to ascertain the signalling pathway(s) involved. The result is in agreement with current position that natural treatments can help to reduce inflammation in the blood [33]. Therefore aqueous leaf extract of *Terminalia catappa* may present as a more potent anti-inflammatory agent than non steroidal anti-inflammatory drugs and could complement the function of insulin in diabetes treatment.

CONSENT

It is not applicable.

ETHICAL APPROVAL

As per International standard or University standard ethical approval has been collected and preserved by the authors.

REFERENCES

1. Alam, U., Asghar, O., Azmi, S., and Malik, R. General aspects of diabetes mellitus. *Handb. Clin. Neurol.* 2014;126: 211–222.
2. Bergman, M. Pathophysiology of prediabetes and treatment implications for the prevention of type 2 diabetes mellitus. *Endocrine.* 2013; 43: 504–513.
3. Pollack, R., Donath, M., LeRoith, D. and Leibowitz, G. Anti-inflammatory Agents in the Treatment of Diabetes and Its Vascular Complications. *Diabetes Care* 2016;39(Suppl. 2):244–252.
4. Foretz, M., Guigas, B., Bertrand, L., Pollak, M. and Viollet, B. Metformin: from mechanisms of action to therapies. *Cell Metab* 2014;20:953–966.
5. Zhou, R., Tardivel, A., Thorens, B., Choi, I. and Tschopp, J. Thioredoxin interacting protein links oxidative stress to inflammasome activation. *Nat Immunol* 2010;11:136–140
6. Isoda, K., Young, J., Zirlik, A., MacFarlane, L., Tsuboi, N., Gerdes, N.. Metformin inhibits proinflammatory responses and nuclear factor-kappaB in human vascular wall cells. *Arterioscler Thromb Vasc Biol.* 2006;26: 611–617
7. Lee, H., Kim, J., Kim, H., Shong, M., Ku, B., and Jo, E. Upregulated NLRP3 inflammasome activation in patients with type 2 diabetes. *Diabetes* 2013; 62:194–204
8. Pradhan, A., Manson, J., Rifai, N., Buring, J. and Ridker P. C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA.* 2001; 286:327–334.
9. Han, T., Sattar, N., Williams, K., Gonzalez-Villalpando C., Lean, M., Haffner, S. Prospective study of C-reactive protein in relation to the development of diabetes and metabolic syndrome in the Mexico City Diabetes Study. *Diabetes Care.* 2002; 25:2016–2021.
10. Esposito, K., Nappo, F., Marfella, R., Giugliano, G., Giugliano, F., Ciotola, M., Quagliaro, L., Ceriello, A. and Giugliano, D. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation.* 2002;106: 2067–2072.
11. Kopp, E., and Ghosh, S. Inhibition of NF- κ B by sodium salicylate and aspirin. *Science.* 1994; 265: 956–959.
12. Grilli, M., Pizzi, M., Memo, M. and Spano, P. Neuroprotection by aspirin and sodium salicylate through blockade of NF- κ B activation. *Science.* 1996; 274: 1383–1385.
13. Gulhar, R. and Jialal, I. *Physiology, Acute phase reactant.* Bethesda: StatPearls Publishing LLC; 2018
14. Fowler, M. Microvascular and Macrovascular Complications of Diabetes. *Clinical Diabetes.* 2011; 29(3): 116-122
15. Sjöholm, A. and Nystrom, T. Inflammation and the etiology of type 2 diabetes. *Diabetes Metab Res Rev.* 2006; 22: 4–10
16. Hansen, T., Thiel, S., Wouters, P., Christiansen, J. and Van den Berghe, G. Intensive insulin therapy exerts antiinflammatory effects in critically ill patients and counteracts the adverse effect of low mannose-binding lectin levels. *J Clin Endocrinol Metab.* 2003;88:1082–1088
17. Borgohain, R., Lahon, K., Das, S. and Gohain, K. Evaluation of mechanism of anti-diabetic activity of Terminalia chebula on alloxan and Adrenaline induced Diabetic albino rats. *International Journal of Pharmaceutical and Biological Sciences.* 2012; 3(3): 256-266.
18. Kulkarni, S. Commonly used drugs, their doses and nature of action in laboratory animals. 3rd ed. Vallabh Prakashan Delhi: Hand book of Experimental Pharmacology, pp 190-195: 2005

19. Katsumata, K., Katsumata, Y., Ozawa, T., Katsumata, J. Potentiating effect of combined usage of three sulfonylurea drugs on the occurrence of alloxan diabetes in rats. *Hormone and Metabolic Research*. 1993;25: 125-126
20. Etuk, E. Animals models for studying diabetes mellitus. *International Journal of Agriculture and Biology*. 2010; 1:130-4
21. Fanelli, A., Ghisi, D., Aprile, P. and Lapi, F. Cardiovascular and cerebrovascular risk with nonsteroidal anti-inflammatory drugs and cyclooxygenase 2 inhibitors: latest evidence and clinical implications. *Ther Adv Drug Saf*. 2017; 8(6): 173–182
22. McCarty, M. and Block, K. Pre-administration of high-dose salicylates, suppressors of NF-kappaB activation, may increase the chemosensitivity of many cancers: and example of proapoptotic signal modulation therapy. *Integr. Cancer Ther*. 2006; 5(3): 252-268
23. Paul-Cark, M., Van Cao, T., Moradi-Bidhendi, N., Cooper, D. and Gilrow D. 15-epilipoxin A4-mediated induction of nitric oxide explains how aspirin inhibits acute inflammation. *The Journal of Experimental Medicine*. 2004; 200(1): 69-78.
24. Somasundaram, S. Sigthorsson, G., Simpson, R., Watts, J. Jacob, M., Tavares, I., Rafi, S. and Roseth, A. Uncoupling of Intestinal mitochondrial oxidative phosphorylation and inhibition of cyclooxygenase are required for the development of NSAID-enteropathy in the rat. *Ailment Pharmacol. Ther*. 2000; 14 (5):639-650.
25. Toth, L., Muszbek, L. and Komoromi, I. Mechanism of the irreversible inhibition of human cyclooxygenase-1 by aspirin as predicted by QM/MM calculations. *Journal of Molecular Graphics and modelling*. 2013; 40:90-109
26. Noble, S. and Balfour, J. Meloxicam. *Drugs*. 1996; 51(3): 424-30
27. Aljada, A., Ghanim, H., Saadeh, R. and Dandona, P. Insulin inhibits NFkappaB and MCP-1 expression in human aortic endothelial cells. *J Clin Endocrinol Metab*. 2001; 86:450-453.
28. Aljada, A., Saadeh, R., Assian, E., Ghanim, H. and Dandona, P. Insulin inhibits the expression of intercellular adhesion molecule-1 by human aortic endothelial cell stimulation of nitric oxide. *J. Clin Endocrinol. Metab*. 2000; 85:2572-2575.
29. Bazzoni, F. and Beutler, B. The tumor necrosis factor ligand and receptor families. *N Engl J Med*. 1996; 334: 1717-1725.
30. Chao, W. Toll-like receptor signalling: a critical modulator of cell survival and ischemic injury in the heart. *Am J Physiol Heart Circ Physiol*. 2009; 296: H1-12.
31. Wu, T., Dorn, J., Donahue, R., Sempos, C. and Trevisan M. Associations of serum C-reactive protein with fasting insulin, glucose, and glycosylated hemoglobin. *Am J Epidemiol*. 2002; 155:65–71
32. Ford, E. Body mass index, diabetes, and C-reactive protein among U.S. Adults. *Diabetes Care*. 1999; 22:1971–1977
33. Rao, M. and Gopal, S. C-Reactive Protein - A critical Review. *Int. J. Curr.Microbiol. App.Sci*. 2015; 4(12): 55-61.
34. Teodoro, J., Nunes, S., Rolo, A., Reis, F. and Palmeira, C. Therapeutic Options Targeting Oxidative Stress, Mitochondrial Dysfunction and Inflammation to Hinder the Progression of Vascular Complications of Diabetes *Front. Physiol*. 2019;9:1-18
35. Chaitanya, A., Mohit, T. Aucar, John, A and Greenberg, E. Meloxicam-induced enteropathy of the small bowel. *CMAJ*. 2011; 183(5): 577 - 580
36. Hotamisligil, G., Shargill, N., Spiegelman, B. Adipose expression of tumor necrosis factor alpha: direct role in obesity-linked insulin resistance. *Science*. 1993; 259:87–91.
37. Sigthorsson, G., Crane, R., Simon, T., Hover, M., Quan, H., Bolognese, J. and Bjarnason, I. COX-2 inhibition with rofecoxib does not increase intestinal permeability in healthy subjects: a double blind crossover study comparing rofecoxib with placebo and indomethacin. *Gut*. 2000;47:527-32.
38. Maiden, L., Thjodleifsson, B., Seigal, A., Bjarnason, I., Scott, D., Birgisson, S. and Bjarnason, I. Long-term effects of Non-steroidal anti-inflammatory drugs and cyclooxygenase-2 selective agents on the small bowel: a cross-sectional capsule enteroscopy study. *Clin Gastroenterol Hepatol*. 2007; 5:1040-5.
39. Higuchi, K., Umegaki, E., Watanabe, T. Yoda, Y., Morita, E., Murano, E. and Tokioka, S. Present status and strategy of NSAIDs-induced small bowel injury. *J Gastroenterol*. 2009;44:879-88.
40. Endo, H., Hosono, K., Inamori, M., Nozaki, Y., Yoneda, K., Fujita, K. and Takahashi, H.. Characteristics of small bowel injury in symptomatic chronic low-dose aspirin users: the experience of two medical centers in capsule endoscopy. *J Gastroenterol*. 2009;44:544–549.

41. Patrono, C., Ciabattoni, G., Pinca, E., Pugliese, F., Castrucci, G., De Salvo, A., Satta, M. and Peskar, B. 1980. Low dose aspirin and inhibition of thromboxane B₂ production in healthy subjects. *Thromb. Res.* 1980 17: 317–327
42. Duncan, B., Schmidt, M., Pankow, J., Ballantyne, C., Couper, D., Vigo, A., Hoogeveen, R., Folsom, A. and Heiss, G. Low-Grade Systemic Inflammation and the Development of Type 2 Diabetes. *Diabetes.* 2003; 52; 1799-1804.

UNDER PEER REVIEW