

# 1 Isolation and characterization of bergapten from the root bark of 2 *Ficus exasperata* (Vahl)

## 3 4 ABSTRACT

5 Despite the wide ethnomedicinal applications of *Ficus exasperata*, little is known about the  
6 active principles responsible for the observed biological effects, thus limiting opportunities for  
7 further therapeutic applications. In this study we isolated a furocoumarin (**D-1**) shown to be  
8 partly responsible for the acclaimed anti-diabetic effect of the plant

## 9 10 1. Introduction

11 *Ficus exasperata* (Moraceae) also known as the sandpaper tree is found in different parts of  
12 tropical Africa and Asia, and has been widely used for the treatment of various ailments in these  
13 regions. In Nigeria, decoctions of its roots and leaves are traditionally used as remedies for  
14 hypertension, cough, ulcer and microbial infections<sup>1,2,3</sup>. Attempts to provide scientific  
15 rationalization for these uses have unraveled several pharmacological activities of the plant  
16 including antiulcer, anti-inflammatory, antidiabetic, antihypertensive, antioxidant and  
17 hypolipidemic properties<sup>3-7</sup>.

18 Phytochemical investigation of extracts from the genus *Ficus* revealed the presence of several  
19 bioactive secondary metabolites including flavonoids, alkaloids, phenolic acids, steroids,  
20 saponins, tannins, terpenoids and coumarins<sup>8-13</sup>. In 2016, Nnamonu *et al*<sup>14</sup> reported the isolation  
21 of  $\alpha$ -amyrin from the ethyl acetate fraction of the stem bark extract of *Ficus exasperata*  
22 harvested in North Central Nigeria.

23 Whilst the ethnomedicinal value of *Ficus exasperata* continues to increase, relatively little  
24 achievement has been recorded in isolating and identifying its active principles. Against this  
25 backdrop, we aimed to investigate the plant for bioactive contents. We herein report the isolation  
26 and characterization of bergapten, a furocoumarin, as a major constituent in the root bark of  
27 *Ficus exasperata* as well as its hypoglycemic activity.

## 28 29 2. Materials and Methods

### 30 2.1 Chemicals

31 Analytical grade solvents and chemicals, thin layer chromatography (TLC) silica gel 60 F254  
32 plates, and silica gel 60 (70-230 mesh) used for column chromatography were purchased from  
33 Merck (Germany), Sigma Aldrich (USA), and AK Scientific (USA).

### 34 2.2 Plant material

35 The roots of *Ficus exasperata* were collected in a plantation in Ondo, Ondo State, Nigeria. The  
36 samples were identified at the Department of Crop, Soil and Pest Management, Federal  
37 University of Technology, Akure, Nigeria where voucher specimens have been deposited in the  
38 herbarium (CSPH2614).

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### 40 2.3 Extraction and Isolation

41 The root bark of *Ficus exasperata* was sun dried for 48 hours followed by oven drying at 40 °C  
42 for further 48 hours. The dried material (0.5 Kg) was coarsely milled and subjected to soxhlet  
43 extraction using chloroform for 72 hr. Obtained extract (0.0025 Kg, 0.5% yield) was  
44 concentrated under reduced pressure by means of a rotary evaporator (R-114, Buchi,  
45 Switzerland), and fractionated using 100% pet ether, 80% pet ether in EtOAc, 60% pet ether in  
46 EtOAc, 100% EtOAc, and 80% EtOAc in MeOH.

47 The 100% EtOAc fraction termed CLE4 was dissolved in chloroform and loaded onto a pipette  
48 previously packed with silica gel (70-230 mesh ASTM). It was then successively eluted with a  
49 mixture of EtOAc and MeOH in increasing concentration. 12 fractions of 10 ml each were  
50 collected; fractions containing the same compounds as determined by their TLC profiles were  
51 combined and concentrated to dryness under reduced pressure. Five fractions were obtained  
52 (CLE4A-E). Fraction CLE4B (0.7 g) was reconstituted in chloroform and chromatographed  
53 using Silica gel as described above. Three bulked fractions were obtained, termed 4B1 – 4B3.

54 Fraction 4B2 was further column chromatographed over silica gel (70-230 mesh) using a column  
55 of diameter 2.5 cm and length 40 cm and isocratically eluting with pet-ether: ethyl acetate 85:15.  
56 40 fractions of 2 ml each were collected. Isolate D-1 (250 mg, >99% by HPLC) was obtained  
57 from the fractions 1-10 as off-white needle-like crystals with a characteristic odour.

### 58 2.4 Structural Elucidation

59 Proton (1H, 500 MHz), carbon-13 (13C, 101 MHz), and two-dimensional NMR were acquired  
60 on Varian 400 spectrometer (Varian, California, United States of America). **D-1** was measured in  
61 deuterated chloroform (CDCl<sub>3</sub>), with 1H and 13C chemical shifts of 7.26 and 77.16 ppm  
62 respectively. High resolution mass spectra of the isolate were obtained using a Bruker  
63 microTOF-Q spectrometer (Bruker, Bremen, Germany) with an electrospray ionization (ESI)  
64 source.

### 65 2.5 Oral Glucose Tolerance Test

66 A total number of 20 Wistar male rats weighing between 120-180 g were purchased from the  
67 Animal House of the Federal University of Technology, Akure, Nigeria. Animal grooming and  
68 collection and testing of blood samples were conducted as described by Akhtar *et al.*,2008<sup>15</sup>. In  
69 summary, the animals were divided into four groups of five and kept in standard rat cages where  
70 they were adequately fed. All five groups were fasted for 16 hours, after which groups II-IV  
71 were induced with diabetes intragastric administration of glucose (3g/Kg body weight) The  
72 baseline glucose level was then measured by glucometer (Accu-Check glucose test meter).  
73 Group I represented the negative control group. Group II was treated with metformin (100 mg/kg

74 body weight). While groups III and IV received 10 and 30 mg/Kg body weight of **D-1**  
75 respectively, administered by intraperitoneal injection. Serum glucose was determined for all  
76 groups at 0, 30, 60, 90 and 150 min. Data obtained was statistically compared using two-way  
77 ANOVA (multiple comparison), observed difference was termed significant at  $P < 0.05$ .  
78 Statistical analysis was conducted using GraphPad Prism 7.03.

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### 80 3. Results and Discussion

#### 81 Structural determination of **D-1**

82 The structure of **D-1** was elucidated using ESI-MS and NMR, and was found to be  
83 spectroscopically similar to previously reported bergapten structure by Chunyan *et al.*(ref). As  
84 shown in table 1 and figure 1, the compound contains a total of twelve carbon atoms, eleven of  
85 which are  $sp^2$  hybridized resonating at  $\delta$  93.8 –161.3 and the remaining one being a methoxy  
86 carbon resonated at  $\delta$  60.1. The  $^1H$  NMR spectrum revealed the presence of five  $sp^2$  protons ( $\delta$   
87 6.27 – 8.16), and three highly deshielded methyl protons ( $\delta$  4.27). The full assignment of the  
88 proton and carbon signals was conducted with the aid of 2D NMR experiments. Heteronuclear  
89 single quantum coherence (HSQC) was used to assign individual protons to their corresponding  
90 carbon atoms as given in table 1, and also to confirm the absence of methylene protons.  
91 Homonuclear correlation spectroscopy ( $^1H - ^1H$  COSY) revealed protons that are coupled in an  
92 AB system, specifically  $\delta$  6.27 (1H, *d*,  $J = 9.8$  Hz, H-3) and 8.16 (1H, *d*,  $J = 9.8$  Hz, H-4); 7.59  
93 (1H, *d*,  $J = 2.4$  Hz, H-2') and 7.02 (1H, *d*,  $J = 2.4$  Hz, H-3'). ESI-MS showed the sodiated mass  
94 ion  $[M + Na]^+$  peak at 239.028, consistent with the 216.042 molecular weight of **D1**. Thus  
95 allowing the structural confirmation of **D1** as bergapten.

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98 **Table 1.**  $^{13}C$  and  $^1H$ -NMR chemical shifts (ppm) of **D-1**.

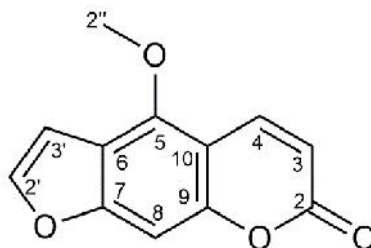
Position	$^1H$ , $\delta$ (multi, Hz)	$^{13}C$ , $\delta$
C-2	-	161.3
C-3	6.27 (1H, <i>d</i> , $J = 9.8$ )	112.5
C-4	8.16 (1H, <i>d</i> , $J = 9.8$ )	139.2
C-5	-	149.3
C-6	-	112.3
C-7	-	158.3
C-8	7.14 (1H, <i>s</i> )	93.8
C-9	-	152.6
C-4a	-	106.4
C-2'	7.59 (1H, <i>d</i> , $J = 2.4$ )	144.8

C-3'	7.02 (1H, <i>d</i> , <i>J</i> = 2.4)	105.0
C-2''	4.27 (3H, <i>s</i> )	60.1

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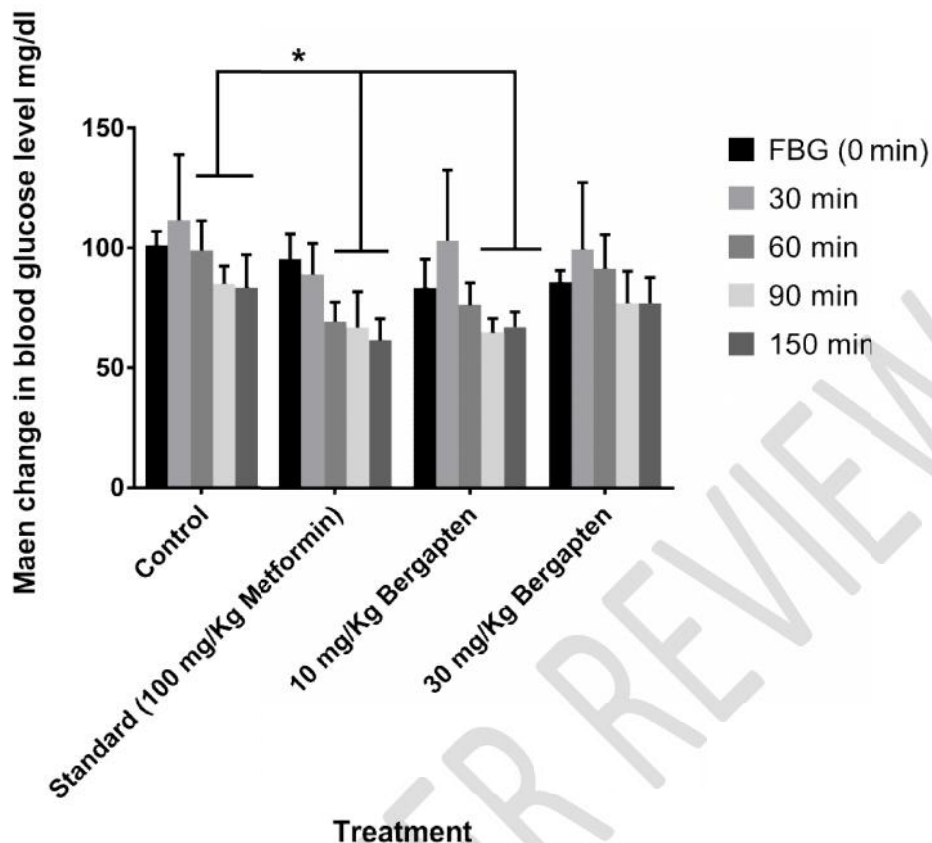
102 **Figure 1.** Structure of Compound **D-1** (Bergapten).

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#### 104 **Hypoglycemic activity of D-1**

105 As shown in figure 2, hyperglycemia was observed approximately 30 minutes after intragastric  
106 administration of glucose. Upon administration of the standard drug metformin, and **D-1**, the  
107 most significant blood glucose reduction was observed for metformin and 10mg/Kg **D-1** at 60  
108 min and 90 min relative to the control. Metformin produced a reduction of 29% at 60 min and  
109 22% at 90 min, while 10mg/Kg **D-1** gave a reduction of 23% and 24% at 60 and 90 min  
110 respectively. However, 30 mg/Kg **D-1** showed a significantly lower decrease in blood glucose  
111 both at 60 min (7%) and at 90 min (9%) compared to 10mg/Kg **D-1**; also at 150 min, a slight  
112 increase in blood sugar compared to what was observed at 90 min. The observed non-dose  
113 dependent hypoglycemic activity of **D-1** is most likely due to lack of target specificity.

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115  
 116 **Figure 2.** Effect of **D-1** on blood glucose levels of glucose-fed rats. Error bars indicate SEM  
 117 obtained from experimental replicates (minimum of 5). \*P < 0.05, significant difference in mean  
 118 glucose levels at similar time intervals in comparison to the control. Intraperitoneal metformin as  
 119 standard drug.

120  
 121 **Conclusion**

122 Overall, this study shows that bergapten has considerable hypoglycemic effect, which in part  
 123 justifies the reported anti-diabetic activity of *Ficus exasperata*. The comparable hypoglycemic  
 124 effect displayed by bergapten at 10 mg/Kg body weight, and the clinically used metformin,  
 125 positions bergapten as a viable lead molecule for the development of novel anti-diabetic agents.  
 126 It is thus research-worthy to unravel the precise mechanism of action of the molecule as a  
 127 hypoglycemic agent, followed by rational structural modification to optimize it for potency and  
 128 safety.

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133 **References**

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