Transfer and Accumulation of some heavy metals in native vegetation plants

4

1

5 **ABSTRACT**

6 Phytoremediation is the use of selected plants in order to eliminate some heavy 7 metals from soil, or wastewater in a cost-effective method. This study aimed to investigate the concentrations of heavy metals such Cd, Pb, Cu, Zn and Cr in soils 8 9 and vegetation plants grown in Wadi Hanifa, Riyadh city, Kingdom of Saudi Arabia. Five sites have been chosen for collected plant samples (shoot and root) 10 for one year, and five plant species have chosen which distributed in the study 11 area including: Ziziphus spina-christi, Prosopis juliflora, Rhazya stricta, 12 Ochradenus baccatus and Conocarpus erectus. Determination of Cd, Pb, Cu, Zn 13 and Cr has been done with ICP. Accumulation coefficient (AC), and translocation 14 15 factor (TF) have been calculated to evaluate the ability of selected plants to extract the heavy metals from soil. The results indicated that Ziziphus spina-christi and 16 17 *Conocarpus erectus* shows the high ability for accumulate the Pb and Zn in its 18 root and shoot compare with other plants. The trend of heavy metal transfer factors for different plants was in the order of Cd > Cr > Pb > Cu >Zn. The 19 20 accumulation coefficient (AC) of the Cd, Pb, Zn, Cu and Cr in the roots/soil of Ziziphus spina-christi, Prosopis juliflora, Rhazya stricta, Ochradenus baccatus 21 22 and Conocarpus erectus were varied from 0.80 to 3.60. The order of AC in the 23 shoot as follows: Pb>Cu>Zn>Cr>Cd, while in roots of as follows: Cd>Cr>Pb>Cr> 24 Zn.

Key words: Heavy metals, transfer, accumulation, Ziziphus spina-christi, Prosopis juliflora,
 Rhazya stricta, Ochradenus baccatus and Conocarpus erectus.

27

28 **1. INTRODUCTION**

Heavy metals are becoming increasingly prevalent in soil environments as a result of wastewater irrigation, sludge application, solid waste disposal, automobiles exhaust and industrial wastewater discharge [1, 2]. The release of heavy metals into the environment

32 by industrial activities presents a serious environmental threat. Heavy metal 33 contamination is considered as a dominant source of pollution and a potentially growing 34 environmental and human health concern worldwide [3, 4]. Copper, Pb and Cd can become a sanitary and ecological threat to drinking water resources, even at very low 35 36 concentrations [5]. In addition, Cd and Zn are common industrial pollutants [6, 7]. Both 37 Cd and Zn are harmful to plant at relatively low concentrations [8]. Plant uptake of some heavy metals from soil occurs either passively with the mass flow of water into the roots, 38 or through active transport crosses the plasma membrane of root epidermal cells [8]. 39

Some kind of plants can potentially accumulate certain heavy metal ions an order 40 41 of magnitude greater than the surrounding medium [9]. Therefore, clean alternatives 42 must be developed in order to remove heavy metals from effluents. Heavy metals can be removed from industrial wastewater and contained soil by a range of physico-chemical 43 precipitation, ion exchange, 44 remediation technologies such as adsorption. electrochemical processes and membrane processes [10-12]. However, these 45 technologies are expensive and energy-intensive, driving towards a search of cheaper 46 47 alternatives [13].

Phytoremediation is a method of environmental treatment that makes use of the 48 ability of some plant species to accumulate certain heavy metals in amounts exceeding 49 the nutrients requirements of plants. Phytoextraction is one of the elements of 50 phytoremediation in which heavy metals from the contaminated site are taken up by 51 plants and then transported from roots to shoots and removed with crops from a 52 53 specified area of nature [14]. Phytoremediation is phytostablization where plants are 54 used to minimize some of heavy metals mobility in contaminated soils. Nowadays, than 500 plants are known as hyper accumulation of heavy metals into their aboveground 55 biomass including weeds, trees, grasses and vegetable crops [15, 16]. 56

57 The objectives of this research were to determine the concentration of some heavy 58 metals in some native plant species growing on a contaminated soil which located in the 59 Wadi Hanifa, Riyadh city, Kingdom of Saudi Arabia and to assessment and evaluation of 60 heavy metals pollution.

61

62 2. Materials and Methods

63 **2.1 Experimental sit description and soil chemical analysis**

Wadi Hanifa (24°14'27.0"N 47°00'00.0"E) is a valley in the Najd region, Riyadh province, 64 in central Saudi Arabia. The valley runs for a length of 120 km from northwest to 65 southeast, cutting through the city of Riyadh, the capital of Saudi Arabia. Temperatures 66 in summer reach an average of 43.9 °C and precipitation averages only 62 mm per year 67 68 in the driest places. Rain falls with great intensity for short periods, causing flash floods. 69 The nature of the dry, warm climate leads to a high percentage of the scarce rainfall 70 being instantly evaporated [17, 18]. Represented soil samples were collected from the surface layer (0-20 cm depth) by 20x20 m. The soil samples were air-dried at room 71 72 temperature for two weeks and then sieved by 2-mm stainless steel sieve. The pH and EC of samples were measured (using 1:5 ratio of w/v with demonized distilled water) by 73 74 pH-meter and the electrical conductivity (EC) meter respectively. Complex metric EDTA titration was employed for determining Ca⁺⁺ and Mg⁺⁺ simultaneously and individually 75 [19]. Sodium and potassium was determined using flame photometer (Corning 400). 76 Carbonate and bicarbonate were determined by titration with H₂SO₄ while silver nitrate 77 78 was used to determine chloride [19]. Sulphate was determined by turbidity method as 79 described by [20]. Particle size distribution was analyzed according to [21]. Calcium Carbonate content was determined using the Calcimeter [22]. Some selected soil 80 physical and chemical properties are presented in Table (1). The total content of heavy 81 metals (Cd, Pb, Zn, Cu, Cr and Zn) in the soil samples were determined after digestion 82 with HNO₃-HCIO₄-HF as described by Hossner [23], then total heavy metals content 83 were determined using ICP (Perkin Elmer, Model 4300DV). 84

85

86 **2.2 Plant sample collection and plant analysis**

Five plant native plant species (Ziziphus spina-christi, Prosopis juliflora, Rhazya stricta, 87 88 Ochradenus baccatus and Conocarpus erectus) based on their coverage at the Wady 89 Henifia, Riyadh city, Saudi Arabia were collected in acid-washed polyethylene bags 90 according to the sampling procedures of Australian National Botanic Garden [23, 24]. Table 2 shows the botanical and vernacular names of plants species collected from the 91 study area. The collected plant samples separated into shoot and root, washed gently 92 93 with demonized water for approximately 5 minutes to remove soil particles adhered to 94 the plant roots, then, air-dried at 60 and finally ground to powder using a Wiley mill. The plant samples were acid digested with HNO₃-HClO₄ mixture according to Chapman and
Pratt, (1996). After digestion, the cooled samples was diluted to 50 mL with distilled
water and filtered into plastic bottles pre washed with acid. The concentrations of heavy
metals mainly (Cd, Pb, Zn, Cu, Cr and Fe) were measured using ICP (Perkin Elmer,
Model 4300DV). Reagent blanks and standards were used where appropriate to ensure
accuracy and precision in heavy metals analysis.

101

102 2.3 Estimation heavy metals between soil and plant

103 2.3.1 Accumulation coefficient

Accumulation coefficient was used for evaluation of heavy metals accumulationin the plant according to [25] as flows:

$$Ac = \frac{C}{C}$$

107 Ac= concentration of heavy metal in plant shoot $(mg kg^{-1})/$ concentration of heavy metal

108 in background soil (mg kg⁻¹).

109 2.3.2 Translocation factor

Transfer factor (TF) describes the amount of heavy metal transferred from the soil to the 110 111 plant under equilibrium conditions [26]. Heavy metals from the soil are consumed by 112 plant roots and then distributed in various plant tissues. Transfer of this heavy metal from soil to plant tissues is measured using the TF indicator, which measures the ratio of 113 the concentration of a specific metal in plant tissue to the concentration of the same 114 metal in soil. If the TF values are \geq 1.0 it shows a higher uptake of metal from soil by the 115 116 plant, while lower values mean less absorption of the metal from the soil, and the plant can be used for consumption [27]. 117

118 Translocation Factor (TF) was calculated as follows:

$$TF = \frac{Metal}{Metal}_{roots}$$

119

120

121 3. RESULTS AND DISCUSSION

122 **3.1 Soil characteristics**

Selected physical and chemical properties of soil collected from the Wadi Hanifa are 123 listed in Table 1. The soil texture was sandy; the soil pH was 7.80 with EC 4.50 dS m 124 1in. In addition, results indicated that the percentage of CaCO₃ was 19.2%. Also A wide 125 range of soil heavy metals concentration was observed of soil at start of the collecting 126 the plant samples. The total content of Cu 20±0.10 mg kg⁻¹; Zn 35±1.20 mg kg⁻¹; Pb 127 15 ± 0.70 mg kg⁻¹; Cd 0.20±0.01 mg kg⁻¹ and Cr 30±1.15 mg kg⁻¹. According to Lindsay, 128 [28] who reported that, 35 mg kg⁻¹ Zn 40 mg kg⁻¹ Pb, 10 mg kg⁻¹ Cd, 0.05 mg kg⁻¹ and Cr 129 95 mg kg⁻¹ the average of heavy metals concentrations in common soils. The results 130 indicated that, the total concentration of Cr, Pb, Zn and Cu within the normal range of 131 132 common soils expect the Cd. Wastewater samples were collected from different 133 locations in the Wadi Hanifa. The results indicated that the water quality is characterized 134 by high pH and contains high concentrations of Fe, Mn, Zn, Cd, Ni, Pb, and Mo compared with Kingdom of Saudi Arabia [6]. 135

136

137 **3.2 Concentration of heavy metal in plant species**

Table 3 show the heavy metals content in shoot and roots of selected plant species. In 138 general, the concentration of Cd, Pb, Zn, Cu and Cr in roots of Ziziphus spina-christi, 139 prosopis juliflora, Rhazya stricta, Ochradenus baccatus and Conocarpus erectus plant 140 species was higher than that in shoots. Concentration of heavy metals by shoots and 141 roots varied with heavy metals type and plant species. Plant Ziziphus spina-christi and 142 143 Conocarpus erectus shows the high ability for accumulate the Pb and Zn in its root and 144 shoot compare with other plants. All the studied plant species showed ability to accumulate the Pb, Zn, Cu, Cr expect Cd. The results in agreement with reported by [29, 145 146 30]. Some of heavy metal from soil occurs either passively with the mass flow of water

147 and accumulated plant roots [9, 31]. Some of plants can accumulate certain metal ions 148 in higher concentration than the surrounding soil [32]. On the study of heavy metals 149 accumulation on second industrial wastewater of Riyadh city, [7] founded that concentrations of Cr, Ni, Cu, Zn, and Pb in Fagonia indica and Cenchrus ciliaris plants 150 151 were markedly higher than in *Rhazya stricta* plant. According to State Environmental Protection Administration and China, the average concentration of Cr, Ni, Cu and Cd 152 elements were 0.50, 9.0, 20 and 0.20 mg kg⁻¹ on a dry weight basis, respectively. In the 153 current study, the concentration of Cr, Ni and Zn concentrations in roots and shoot of 154 155 Ziziphus spina-christi, Prosopis juliflora, Rhazya stricta, Ochradenus baccatus and Conocarpus erectus were exceeded the concentration limits. The heavy metals enter the 156 environment from both natural processes which included weathering, parent material 157 rock erosions and atmospheric deposition and from anthropogenic activities such as 158 159 using chemicals, sewage sludge disposal, mining [33-35]. The heavy metal accumulated 160 would be contaminated the soil and vegetables at high concentrations.

161 **3.2 Accumulation coefficient (AC)**

The accumulation coefficient (AC) of the Cd, Pb, Zn, Cu and Cr in the roots/soil of 162 Ziziphus spina-christi, prosopis juliflora, Rhazya stricta, Ochradenus baccatus and 163 Conocarpus erectus were varied from 0.80 to 3.60 (Table 4). The results indicated that 164 the AC values depend on the heavy metal type and plant species. Regardless the AC 165 values were 0.02-4.0, 0.02-3.40, 0.70-3.60, 0.71-3.90 and 0.46-5.60 for Ziziphus spina-166 christi, Prosopis juliflora, Rhazya stricta, Ochradenus baccatus and Conocarpus erectus, 167 respectively. The order of AC in the shoot as follows: Pb>Cu>Zn>Cr>Cd, while in roots 168 of as follows: Cd>Cr>Pb>Cr> Zn. The calculated AC values of the heavy metals, in the 169 roots of Fagonia indica and Cenchrus ciliaris were 0.31-2.30 [2, 36]. The higher AC 170 values of studied plant in this study indicated that, these plant species could be 171 172 accumulated heavy metals and also are suitable for heavy metal phytoextraction from 173 contaminated soil.

174 **3.2 Transfer Factor (TF)**

The calculated (TF) values of transfer the heavy metals from soil to different plant spices are presented in (Table 5). The TF values varied among the plant species and highest TF for Cd in *Prosopis juliflora*, *Ochradenus baccatus* flowed by Cr in *Rhazya stricta*, *Ochradenus baccatus* and and *prosopis juliflora* plant, while the lowest TF for Zn and Cu 179 was recorded in all studied plants. The high TF for Cd, Cr and Pb heavy metals from soil 180 to plant indicated that a strong accumulation of those metals. The calculated TF values 181 for heavy metals were found in the order: Cd>Cr>Pb>Zn>Cu. The results indicated that the plant species have a different ability to accumulate the heavy metals. The results 182 183 shows clearly that the plant species differ to use its ability to accumulate the heavy from the soil (Figure 1). For example, the ability of the Ochradenus baccatus and prosopis 184 185 juliflora plant for accumulation of Cd was higher than other plant species, while Rhazya stricta have ability to accumulate Cr. The higher the value of TF means that, the more 186 187 heavy metal can be accumulated by plants parts. Cadmium, Cr and Pb is the highest TF values, which agrees with Yang et al., [37]. By comparing TF values, for Ziziphus spina-188 christi, Prosopis juliflora, Rhazya stricta, Ochradenus baccatus and Conocarpus erectus 189 190 it could be compare the ability of these plant species for accumulation of heavy metals from the contaminated soil and translocations them in the plant canopy. Plants showing 191 192 TF values less than one are unsuitable for photo extraction of some heavy metals [38] and but can be used as an indicator for soil contamination with some heavy metals. 193

194

195 **4. CONCLUSION**

This study was conducted to screen Ziziphus spina-christi, Prosopis juliflora, Rhazya 196 197 stricta, Ochradenus baccatus and Conocarpus erectus plant species growing on Wadi 198 Hanifa, Riyadh city, Kingdom of Saudi Arabia for their potential for removal of some heavy metal. The study confirmed that there were differences in the heavy element 199 200 contents in plant species. The results showed that the existence Cd, Pb and Cr in the shoot and roots of Prosopis juliflora, Ochradenus baccatus and Prosopis juliflora plant 201 species. The current study shows clearly that the plant species differ to use its ability to 202 accumulate the heavy from the soil and wastewater and the recommendation that those 203 204 species, with high concentration in heavy elements.

205

206 COMPETING INTERESTS

208 Authors have declared that no competing interests exist.

209

207

210 **REFERENCES**

Sternberg SP, Dom RW. Cadmium removal using Cladophora in batch, semi-batch
 and flow reactors. Bioresour. Technol., 2002; 81: 249-255.

Shi Z., Tao, S., Pan, B, Fan, W, He, XC., Zuo, Q, Wu, SP., Li, BG., Cao, J, Liu, FL.,
 Xu WX., Wang XJ, Shen WR, Wong PK. Contamination of rivers in Tianjin, China, by
 polycyclic aromatic hydrocarbons. Environ., Pollut., 2005; 134: 97-111.

- 3. Jarup L. Hazards of heavy metal contamination. Br Med Bull., 2003; 68: 167-182.
- 4. Atta S, Moore F, Modabberi S. (2009). Heavy metal contamination and distribution in
 the Shiraz Industrial Complex zone soil, South Shiraz, Iran. World Appl. Sci., J., 2009; 6:
 413-425.

5. Lasat MM. Phytoextraction of toxic metals: A review of biological mechanisms.
Journal of Environmental Quality, 2002; 31:109-129.

Al-Farraj A, Al-Sewailem M, Aly A, Al-Wabel M, El-Maghraby S. Assessment and
heavy metal behaviours of industrial waste water: A case study of Riyadh city, Saudi
Arabia. Proceedings of the International Academy of Ecology and Environmental
Sciences, 2013; 3: 266-277.

7. Ghoneim AM, Al-Zahrani S, El-Maghraby S, Al-Farraj AS. Heavy metals
accumulation in *Rhazya stricta* L. plant growing on industrial wastewater of Riyadh city,
Saudi Arabia. Journal of Applied Sciences, 2014; 14: 2007-2010.

Chapman HD, Pratt PF. Methods of Analysis for Soils, Plants and Water. California
 Univ., Berkeley, CA, USA, 1996.

Schen Y, Wang C, Wang Z. Residues and source identification of persistent organic
 pollutants in farmland soils irrigated by effluents from biological treatment plants.
 Environ. Int., 2005; 31: 778-783.

10. Kim IS, Kang HK, Johnson-Green P, Lee EJ. Investigation of heavy metal
accumulation in Polygonum thunbergii for phytoextraction. Environ Pollut., 2003; 126:
235-43.

11. Kurniawan TA, Chan GY, Lo WH, Babel S. Physico-chemical treatment techniques
for wastewater laden with heavy metals. Chemical Engineering Journal, 2006; 118: 8398.

12. Mulligan CN, Yong RN Gibbs BF. Remediation technology for meat-contaminated
soils and groundwater, an evaluation. Eng. Geol., 2001; 60: 193-207.

13. Alhawas M, Alwabel M, Ghoneim A. Alfarraj, A, Sallam A. Removal of nickel from
aqueous solution by low-cost clay adsorbents. Proceedings of the International Academy
of Ecology and Environmental Sciences, 2013; 3(2): 160-169.

- 14. Esawy KM, Ghoneim AM. Effect of polluted water on soil and plant contamination by
 heavy metals in El-Mahla El-Kobra, Egypt. Solid Earth; 2016; 7: 703-716.
- Liu HA, Probst B, Liab B. Metal contamination of soils and crops affected by the
 Chenzhou Lead/Zinc mine spill (Hunan, China). Science of the Total Environment, 2005;
 399:153-166.

16. Afzal M, Awais A, Alderfasi, A, Ghoneim, A, Saqib M. Physiological tolerance and
cation accumulation of different genotypes of Capsicum annum under varying salinity
stress. Proceedings of the International Academy of Ecology and Environmental
Sciences, 2014; 4(1): 39-49.

17. Vitz WJ, Wenzel WW. Arsenic transformation in the soil-rhizosphere-plant system,
fundamentals and potential application of phytoremediation. J. Biotechnol., 2002; 99,
259-278.

18. Nagham AO., (2010). The use of local sawdust as an adsorbent for the removal of

- copper lon from wastewater using fixed bed adsorption, J., 28 (2), 859–860.
- 19. Sparks DL. Methods of Soil Analysis. Part 3. Chemical Methods. SSSA Book Ser. 5.
 ASA and SSSA, Madison, WI, USA. 1996.
- 261 20. Lindsay W. Chemical Equilibrai in Soils. 1st edition. A Wiley-Interscience Publication.
 262 Jones Wiley and Sons. New York. 1979.
- 263 21. Gee GW, Bauder JW. Particle Size Analysis, ³th Ed. In: Methods of soil Analysis.
- Part 1: Physical and Mineralogical Methods, S.S.S.A. and American Society ofAgronomy, Madison, WI, pp. 377-382, 1996.
- 266 22. Loeppert, R. H., Suarez, D. L. (1996). Carbonate and Gypsum: Manometer Method.
- In: Methods of Soil Analysis. Part 3: Chemical Methods ^{3th} Ed. Soil Sci. Soc. Am.,
- 268 Madison, WI, pp. 437-474.

269 23. Hossner LR. Dissolution for Total Elemental Analysis. *In.* Methods of soil analysis.
270 Part 3. Chemical Methods. Edited by Sparks et al., SSSA and ASA. Madison. WI. pp:
271 46-64, 1996.

272 24. Australian National Botanic Gardens (2016). Plant collection procedures and273 specimen preservation. Australian Government, Canberra.

274 25. Tony B. Collecting and preserving plant specimens, a manual. Department of
275 Science, Information Technology and Innovation, 2016; 2 (1).

276 26. EPA, (2000). Introduction to Phytoremediation. 600/R-9910/. February. U.S
277 Environmental Protection Agency. Cincinnati. Ohio.

278 27. Alina L, Isidora R, Adina B, Laţo K F.The transfer factor of metals in soil-plant
279 system. Research Journal of Agricultural Science, 2012; 44 (3).

280 28. Rangnekar SS, Sahu SK, Pandit GG, Gaikwad VB. Accumulation and translocation
of nickel and cobalt in nutritionally important Indian vegetables grown in artificially
282 contaminated soil of Mumbai, India. Re-search Journal of Agricultural and Forest
283 Sciences, 2013b, 1, 15.

284 29. Al-Wahaibi MH. Accumulation Phenomenon of Heavy Metals in Plants. Saudi
285 Journal of Biological Sciences, 2007; 14(2) : 73-96.

30. Al-Otabi TG, Al-Farraj AS. (2009). Heavy Metals Accumulation by Ochradenus
baccatus Plant Grown on Mining Area at Mahad AD'Dahab, Saudi Arabian. Jouranl of
the Saudi Society of Agricultural Sciences, 2009; 7: 459-468.

31. Melendo M, Benítez E, Nogales R. Assessment of the feasibility of endogenous
Mediterranean species for phytoremediation of lead contaminated areas. Fresenius
Environ Bull., 2002; 11: 1105-1109.

32. Héctor MC, Ángel F, Raquel A. Heavy metal accumulation and tolerance in plants
from mine tailings of the semiarid Cartagena-La Unión mining district (SE Spain).
Science of the Total Environment, 2006; 366:1-11.

33. Singh RB. Heavy metals in soils: Sources, chemical reactions and forms. *In* Smith
D, Fityus S, Allman M (eds.) Proceedings of the 2nd Australia and New Zealand
Conference on Environmental Geotechnics. Australian Geochemical Society, Newcastle,
2001, pp. 77-93.

- 34. Waqas M, Li, G, Khan, S, Shamshad I, Reid BJ, Qamar Z, Chao, C. Application of
 sewage sludge and sewage sludge biochar to reduce polycyclic aromatic hydrocarbons
 (PAH) and potentially toxic elements (PTE) accumulation in tomato. Environ. Sci. Pollut.
 Res., 2015; 22: 12114-12123.
- 303 35. Oti WJO. Pollution indices and bioaccumulation factors of heavy metals in selected 304 fruits and vegetables from a Derelict mine and their associated health implications. Int. J. 305 Environ. Sci. Toxic. Res., 2015; 3: 9-15.
- 306 36. Yang q, Zeng X, Zeng J, Kuang W, Liu D. Heavy metal enrichment and 307 bioaccumulation of equisetum Ramosissimum in Pb-Zn Tailings. Academia Journal of 308 Scientific Research,2016; 4 (10), 368.
- 309 37. Khan S, Aijun L, Zhang S, Hu Q, Zhu Y. Accumulation of polycyclic aromatic
 310 hydrocarbons and heavy metals in lettuce grown in the soils contaminated with long311 term wastewater irrigation. J. Hazard Mater., 2007; 152 (2): 506–515.
- 312 38. Zapata F. Isotope techniques in soil fertility and plant nutrition studies. *In* Use of
 313 Nuclear Techniques in Studies of Soil-Plant Relationships. G. Hardarson (Ed).Training
 314 Course Series No. 2. International Atomic Energy Agency, Vienna; 1990; pp 61-128.
- 315
- 316

317

- 319
- 320
- 321

322	Table 1. Soil physical a	nd chemical properties o	f Wadi Hanifa, Riyadh city	, Kingdom of Saudi Arabia

		Cations (meq L ⁻¹) Anions (meq L ⁻¹)						Particles size distribution						
									*EC		CaCO₃%			
Ŗ	ъH	K⁺	Na⁺	Mg ²⁺	Ca ²⁺	Cl	HCO ₃ -	SO4=	CO ₃ =	dS m⁻¹	Clay	Silt	Sand	
7	.80	5.00	0.90	0.50	0.90	4.50	0.80	0.08	0.00	4.30	10.0	8.00	82.0	19.2

*EC: Electrical conductivity. Results of soil properties expressed as average of three replicates.

Name of plants	Common names	Latin name	Binomial name	Species	Genus	Family name	Order name
Ziziphus spina-christi	Ziziphus, crown of thorns, sidr	Ziziphus spina- christi(L.) Desf.	Ziziphus jujuba	Z. jujuba	Ziziphus	Rhamnaceae	Rosales
Prosopis juliflora	Prosopis	<i>Prosopis juliflora</i> (Sw.) DC	Prosopis juliflora	P. juliflora	Prosopis	Mimosaceae	Fabaceae
Rhazya stricta	Harmal	<i>Rhazya</i> <i>stricta</i> Decne	Rhazya stricta	R. stricta	Rhazya	Zygophyllaceae	Apocynaceae
Ochradenus baccatus	Pearl plant, qardi, Qurdi, Taily Weed	<i>Ochradenus</i> <i>baccatus</i> Delile	Ochradenus baccatus	O. baccatus	Ochradenus	Resedaceae	Resedaceae
Conocarpus erectus	Buttonwood, button mangrove	<i>Conocarpus</i> <i>acutifolius</i> <i>Willd.</i> ex Schult.	Conocarpus erectus	C. erectus	Conocarpus	Combretaceae	Myrtales

Table 2. Description (botanical and vernacular names) of plants species grown in the study area

337	Table 3. Heavy metal concentration (mg kg ⁻¹ dry weight) in the shoot and root
338	tissues of some native plants

		<u> </u>	DI	-	a	G				
Plant type	Plant	Cd	Pb	Zn	Cu	Cr				
Flaint type	part		Concentration (mg kg ⁻¹)							
Ochradenus	Shoot	0.20±0.01	0.50±0.03	20.1±1.10	6.20±0.10	20.1±1.50				
baccatus	Root	0.10±0.01	30.5±2.50	25.1±2.00	18.1±1.02	10.5±0.80				
Rhazya stricta	Shoot	0.01±0.01	0.50±0.03	4.10±0.04	0.50±0.01	15.0±1.20				
	Root	0.30±0.01	18.2±1.10	29.1±1.20	15.1±0.30	7.50±0.80				
Conocarpus	Shoot	0.25±0.10	34.0±2.50	25.0±1.30	8.50±0.06	13.0±0.90				
erectus	Root	0.20±0.01	42.1±2.70	44.8±2.30	16.0±1.20	16.1±1.30				
Ziziphus spina-	Shoot	0.23±0.01	45.0±2.90	46.0±2.40	14.0±1.30	12.0±0.90				
christi	Root	0.19±0.01	45.1±2.90	55.2±2.90	22.1±1.5	29.1±2.20				
Prosopis	Shoot	0.55±0.10	47.0±2.60	30.1±2.66	15.0±0.80	19.0±1.50				
juliflora	Root	0.17±0.02	45.1±2.55	55.1±2.75	25.1±0.75	15.1±1.20				

Results of heavy metals are expressed as average ± standard deviation of the three
replicates.

Table 4. Heavy metals accumulation factors (AC) on dry weight basis for some native plants grown in Wadi Hanifa

Heavy	Accumulation	Ochradenus	Rhazya	Conocarpus	Ziziphus	Prosopis
Metal	Factor (AF)	baccatus	stricta	erectus	spina- christi	juliflora
Cd	Shoot/Soil	1.00	0.00	1.25	1.15	2.75
Cu	Roots/Soil	0.50	1.50	1.00	1.00	0.90
Pb	Shoot/Soil	0.02	0.02	1.40	1.86	1.94
PD	Roots/Soil	1.30	0.80	1.70	1.90	1.90
7.5	Shoot/Soil	0.31	0.06	0.38	0.71	0.46
Zn	Roots/Soil	0.40	0.40	0.70	0.80	0.80
<u></u>	Shoot/Soil	1.38	0.11	1.89	3.11	3.33
Cu	Roots/Soil	4.00	3.40	3.60	3.90	3.60
0.1	Shoot/Soil	0.87	0.65	0.56	0.52	0.82
Cr	Roots/Soil	0.50	0.30	0.70	1.30	0.70

Table 5. Heavy metals translocation factors (TF) on dry weight basis for some native plants grown in Wadi Hanifa

	Diant turna	Translocation Factor (TF)							
	Plant type	Cd	Pb	Zn	Cu	Cr			
	Ochradenus baccatus	2.00	0.02	0.80	0.34	1.90			
•	Rhazya stricta	0.01	0.03	0.14	0.03	2.00			
	Conocarpus erectus	1.25	0.81	0.56	0.53	0.81			
	Ziziphus spina- christi	1.21	1.00	0.83	0.63	0.41			
	Prosopis juliflora	3.23	1.04	0.55	0.59	1.26			

