Microorganisms Isolated from Hospital Environmental Surfaces in Akure Metropolis, Ondo State, Nigeria.

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5 ABSTRACT

Microorganisms Isolated from Hospital Environmental Surfaces in Akure Metropolis, Ondo 6 7 State, Nigeria were investigated. The study revealed that bacteria were the most predominant 8 microorganisms found in the hospital environmental surfaces than fungi. Staphylococcus 9 aureus, Streptococcus Escherichia coli. Pseudomonas pyogenes, aeruginosa, Klebsiellapneumoniae and Bacillus cereus were the bacterial isolates while fungi include 10 11 Aspergillus fumigatus, Aspergillus niger and Candida albicans. Staphylococcus aureus was 12 found to be predominant bacteria but Aspergillus funmigatus was the predominant fungi. The 13 result showed that the microbial loads of the public hospitals were higher than that of the 14 private hospitals. The bacteria load of the male ward was found to be higher than that of the 15 female ward while the fungal loads of each of the hospital environmental surfaces of female 16 were higher than that of the male. The study revealed that bacteria were the most 17 predominant microorganisms found in the hospital environment than fungi. Staphylococcus 18 aureus, Streptococcus pyogenes, Escherichia coli, Pseudomonas aeruginosa, Klebsiella 19 pneumoniae and Bacillus cereus were the bacterial isolates while fungi include Aspergillu 20 sfumigatus, Aspergillu sniger and Candida albicans. Staphylococcus aureus was found to be 21 predominant bacteria. All the hospital environmental surfaces were contaminated with one or 22 more microorganisms in the course of the research.

23 Keywords: Hospital; ward; environmental; bacteria; fungi.

24 Introduction

Nosocomial infection is an infection occurring in a patient in a hospital or other health care facility in whom the infection was not present or incubating at the time of admission. It is estimated that in developing countries, nosocomial infections concern above 25% of hospitalized patients, and in the developed countries from 5 to 10% (Wenzel, 1999). This includes infections acquired in the hospital but appearing after discharge, and also occupational infections among staff of the facility (Benenson, 1995). The sources of infections can be: patients, medical personnel, visitors or parts of the environment: equipmentand hospital items, also arthropods inhabiting hospitals.

33 Patient care is provided in facilities which range from highly equipped clinics and 34 technologically advanced university hospitals to front-line units with only basic facilities 35 (World Health Oraganization, 2002). Despite progress in public health and hospital care, infections continue to develop in hospitalized patients, and may also affect hospital staff. 36 37 Many factors promote infection among hospitalized patients: decreased immunity among patients; the increasing variety of medical procedures and invasive techniques creating 38 39 potential routes of infection; and the transmission of drug-resistant bacteria among crowded 40 hospital populations, where poor infection control practices may facilitate transmission 41 (World Health Oraganization, 2002).

42 Hospital acquired infection is an additional affliction to the patient admitted to the hospital 43 for some serious illness and is caused by pathogens which are prevalent in hospital 44 environment (Davaneet al., 2014). In the hospital, microbes are ubiquitous; and can reach 45 the sick patient through various sources, such as air, water, food, contaminated equipments, 46 linen, catheters, scopes, ventilators, contaminated disinfectants and other preparations used 47 for treatment, visitors, infected patients, etc (Davaneet al., 2014). Recently, the probable 48 involvement of surfaces and equipment from the hospital environment as a disseminating 49 source of pathogens, including resistant bacteria, has been highlighted (Schulster *et al.*, 2003). 50 There are no meaningful standards for permissible levels of microbial contamination of 51 inanimate surfaces in hospital environment, but an increased microbial load on surfaces may 52 imply the possibility of finding a pathogen (Dancer, 2004). Microorganisms that are often 53 associated with hospital acquired infections are Staphylococcus aureus, Micrococcus sp., 54 Pseudomonas sp., Proteus sp., Escherichia coli, Enterobacter, Bacillus cereus, Cladosporium 55 sp., Aspergillus sp., and viruses (Ekhaiseet al., 2008). Pseudomonas aeruginosa has been particularly incriminated in nosocomial infection because of its intrinsic resistance to most 56 57 antibiotics and its ability to survive and multiply at low temperatures and in disinfectant 58 solutions (Ohsakiet al., 2007).

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62 MATERIALS AND METHODS

63 **Description of study location**

This research work was carried out from September 2016 to April, 2017 in Akure metropolis, 64 Ondo state, Nigeria. Akure covers an area of 14,798.8,993.7 square kilometers and lies at 65 latitude 7°15'0"N, 7⁰ 11' N 5°11'42"E and longitude 5°11'42"E, 5⁰35'E. Akure is one of the 66 18 local government areas of Ondo State with a population of 484,798 based on the 2006 67 68 population census. It is situated in the peripheral zone of the rainforest of Ondo state. Akure is the administrative capital of Ondo state. Akure lies about 70°15' north of the equator and 69 70 50°15' east of the Meridian. It is about 700 km Southwest of Abuja and 311 km north of Lagos State. The town is situated in the tropic rainforest zone in Nigeria. 71

72 Collection of samples

73 Samples were collected by swab sticks from Male Accident and Emergency Bed, Female 74 Accident and Emergency Bed, Male Toilet, Female Toilet, Male Surgical Ward Chair, 75 Female Surgical Ward Chair, Male Medical Ward Floor, Female Medical Ward Floor, Male 76 Ward Air flora, Female Ward Air flora, Theatre Couch, Injection Room Tables, Neonatal 77 Ward Couch and Maternity Ward Couch from Health Centre FUTA, Don Bosco Catholic 78 Hospital and State Specialist Hospital Akure. The date, time, conditions and sites of sampling 79 were noted. Basically, swabs were used, at least, for each sampling site. For sampling, swabs were moistened in 2 ml sterile saline solution and rolled several times over a surface area of 80 around 25 cm^2 , and the swab sticks were transported to the laboratory. Sampling was always 81 done between 8-10am 82

83 Isolation of microorganisms from hospital environment

Isolation of microorganisms from hospital environment were carried out as described 84 85 by Bakkaliet al. (2005) with slight modification. Basically, swabs were used, at least, for each sampling site. For sampling, swabs were moistened in 2 ml sterile saline solution and 86 rolled several times over a surface area of around 25 cm², and the swab sticks were 87 transported to the laboratory. Sampling was always done between 8-10am. A five-fold serial 88 dilution was made and 0.1 ml of the 10^{-3} and 10^{-5} dilutions were uniformly pour-plated onto 89 14 cm diameter wide agar plates and of nutrient agar, Potato dextrose agar, MacConkey agar 90 91 and EMB agar.

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93 Characterization of bacterial isolates

The pure culture of each isolate was examined. Microscopic examination, staining techniques and biochemical tests were carried out on the isolates according to the methods described by Olutiola*et al.* (2000) and Cheesbrough (2010).

97 Identification of fungal isolates

Fungal isolates were characterized and identified based on macroscopic and
microscopic details with reference to Barnett and Hunter (1998).

100 Statistical analysis of data

All experiments were carried out in triplicate, and data obtained were subjected to one way
analysis of variance, while the means were compared by Duncan's New Multiple Range Test
at 95 % confidence interval using Statistical Package for Social Sciences version 16.0.
Differences were considered significant at p≤0.05.

105 **RESULTS**

Study Area(Source)	FUTA Health Centre (Cfu/ml)	State Specialist Hospital (Cfu/ml)	DonBosco Hospital (Cfu/ml)
MAEB	4.1×10^4	4.6×10 ⁴	2.0×10^{3}
FAEB	2.9×10^4	3.1×10^4	1.0×10^{3}
MT	TNC	TNC	TNC
FT	TNC	TNC	TNC
MSWC	ND	4.5×10 ⁴	3.0×10^{3}
FSWC	ND	2.0×10^{3}	1.0×10^{3}
MMWF	3.3×10^4	4.1×10^4	3.0×10^{3}
FMWF	3.0×10^{3}	3.4×10^4	2.0×10^{3}
MWA	3.9×10 ⁴	4.5×10^{4}	3.0×10^{3}
FWA	3.0×10 ³	5.0×10 ³	2.0×10^{3}
TC	ND	2.0×10^{3}	1.0×10^{3}

106 Table 1: Bacterial load of hospital environmental surfaces.

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IRT	3.0×10^{3}	4.0×10^{3}	1.0×10^{3}
NWC	3.0×10^{3}	4.0×10^{3}	2.0×10 ³
MWC	4.0×10^{3}	5.0×10 ³	2.0×10^{3}

LEGEND: Not Determine (ND), No Growth (NG), Male Accident and Emergency Bed
(MAEB), Female Accident and Emergency Bed (FAEB), Male Toilet (MT), Female Toilet
(FT), Male Surgical Ward Chair (MSWC), Female Surgical Ward Chair (FSWC), Male
Medical Ward Floor (MMWF), Female Medical Ward Floor (FMWF), Male Ward Air flora
(MWA), Female Ward Air flora (FWA), Theatre Couch (TC), Injection Room Tables (IRT),
Neonatal Ward Couch (NWC) and Maternity Ward Couch (MWC).

Table 1: The bacterial load of each of the items isolated from different hospital environmental surfaces is shown in Table 1, it was observed that bacterial load of the toilet were higher than other surfaces, while the bacterial load from each of the male hospital environmental

surfaces was higher than that of the female hospital environmental surfaces. It was also

observed that the bacterial loads isolated from government own hospital was higher than

those microorganisms isolated from private hospital

Study Area(Source)	FUTA Health Centre (Sfu/ml)	State Specialist Hospital (Sfu/ml)	Don Bosco Hospital ((Sfu/ml))
MAEB	2.0×10^{3}	1.5×10^4	1.0×10^{3}
FAEB	3.0×10^{3}	3.0×10 ³	2.0×10^{3}
MT	4.0×10^{3}	4.4×10 ⁴	3.0×10^{3}
FT	6.0×10^3	7.1×10^4	4.0×10^{3}
MSWC	ND	NG	NG
FSWC	ND	NG	NG
MMWF	1.0×10^{3}	2.0×10 ³	NG
FMWF	2.0×10^{3}	4.0×10 ³	1.0×10^{3}
MWA	2.0×10^{3}	3.0×10 ³	NG
FWA	3.0×10^3	4.0×10 ³	1.0×10^{3}
TC	ND	NG	NG
IRT	NG	NG	NG

119 Table 2: Fungal load of hospital environmental surfaces.

NWC	2.0×10^{3}	3.0×10^{3}	1.0×10^{3}	
MWC	NG	NG	NG	

LEGEND: Not Determine (ND), No Growth (NG), Male Accident and Emergency Bed
(MAEB), Female Accident and Emergency Bed (FAEB), Male Toilet (MT), Female Toilet
(FT), Male Surgical Ward Chair (MSWC), Female Surgical Ward Chair (FSWC), Male
Medical Ward Floor (MMWF), Female Medical Ward Floor (FMWF), Male Ward Air flora
(MWA), Female Ward Air flora (FWA), Theatre Couch (TC), Injection Room Tables (IRT),
Neonatal Ward Couch (NWC) and Maternity Ward Couch (MWC).

Table 2: The fungal load of each of the fungi isolated from different hospital environmental surfaces are shown in Table 2, it was observed that fungal load of the toilet was found to be higher than other surfaces, while the fungal load isolated from each of the female hospital environmental surfaces was higher than that of the male hospital environmental surfaces. It was also observed that the fungal loads isolated from government own hospital was higher than those microorganisms isolated from private hospital

132Table 3:Rate of occurrence of different bacteria isolated from FUTA Health133Centre, State Specialist hospital Akure and Don Bosco HospitalAkure

Bacteria	Number of surfaces Tested Positive	Percentage positivity (%)
Staphylococcus aureus	39	22.81
Streptococcus pyogenes	24	14.04
Escherichia coli	21	12.28
Pseudomonas aeruginosa	27	15.79
Klebsiella pneumonia	33	19.30
Bacillus cereus	27	15.79
Total	171	100.01

Table 3: The rate of occurrence of different bacteria isolated from different hospital environmentalsurfaces is presented in Table 3. It was observed that *Staphylococcus aureus*had the highest rate of occurrence, while *Esherichia coli* had the lowest rate of occurrence out of the bacteria isolated for different hospital environment surfaces

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Fungi	Number of surfaces Tested Positive	Percentage positivity (%)
Aspergillusfumigatus	21	36.84
Aspergillusflavus	18	31.58
Candida albicans	18	31.58
Total	57	100

140Table 4:Rate of occurrence of different fungi isolated from FUTA Health Centre,141State Specialist hospital Akure and Don Bosco Hospital Akure

Table 4: The rate of occurrence of different fungi isolated from different hospital environmentalsurfaces is presented in Table 4. It was observed that *Aspergillusfumigatus had* the highest rate of occurrence followed by *Candida albicans* and *Aspergillusniger* which share the same number percentage positivity.

146 DISCUSSION

147 Hospital associated infections have been linked with many factors among which is the 148 microbial quality of the indoor air of different wards and units of each hospital (Ekhaise *et al.*, 149 2010). This type of infection occurs in 5% of all acute care hospitalization in the United State 150 and has been reported to be responsible for the death of one out of every five thousand 151 patients attending an American hospital (Putsept, 1981). In Nigeria, the rate of nosocomial 152 infection ranges between 2.7%-3.8% (Onipedeet al., 2004). This calls for looking at every possible measure to control the rise including (among other investigations) examining the 153 154 quality of indoor air of the hospital wards and units. Each of the hospital environmental 155 surfaces was contaminated with microorganisms.

156 Bacteria were found to be more predominant than fungi, the bacteria isolated from the 157 hospital surfaces were Bacillus cereus, Escherichia coli, Klebsiella pneumonia, Pseudomonas 158 aeruginosa, Staphylococcus aureus, and Streptococcus pyogeneswhile fungi include 159 Aspergillus fumigatus, Aspergillus niger and Candida albicans. Staphylococcus aureus was 160 found to be predominant bacteria with the occurrence of 22.81%, this correlate with the report of Awosika et al. (2012) who reported Staphylococcus aureusas the most frequently 161 162 isolated bacterium from hospital surface. Staphylococcus aureus as the most frequently 163 isolated bacterium from hospital surface has been incriminated in various diseases such as 164 post-operative infections, urinary tract infections, skin infections, respiratory infections and 165 food poisoning (Murray *et al.*, 1995). Proper control measures, such as increase in hygiene, 166 are required to combat infections by Staphylococcus aureus in these hospital wards and units 167 (Awosika et al., 2012). The occurrence of bacteria in hospitals has been commonly 168 related to some possible sources of dissemination: bottle soap (Buffet-Bataillonet al., 2009), hands of healthcare professionals (Tan et al., 2013), gloves and gowns (Rock et 169 170 al., 2014), mobile phones (Ustun and Cihangiroglu, 2012) paper money and coins 171 (Angelakiset al., 2014). Aspergillus funmigatus was found to be predominant fungi with 172 frequency occurrence of 36.84%, this correlate with the report of Cagginaoet al. (2014)who 173 reported that Aspergillus fumigates was the most commonly isolated (68.5%).

The bacterial load of the male ward was found to be higher than that of the female, this could be due personal hygiene of the female, this in line with the report of Ekhaise*et al.* (2008) who reported that quantitative study of different hospital units showed that the children ward and female ward had the highest total bacterial count followed by the bacteriology laboratory.

179 The fungal loads of each of the hospital environment surfaces of female were higher than that of the male. In hospital environments, airborne molds are a potential risk for 180 181 patients because of possible inhalation of conidia (Augustowska and Dutkiewicz, 2006). 182 Because surgical procedures expose patients to infective complications, the operating theater 183 is considered a complex habitat in which all sources of pollution have to be kept under 184 control (Partridge-Hinckley et al., 2009; Grossiet al., 2011). In particular, the widespread 185 presence of *Aspergillus* spp. is the major extrinsic risk factor for invasive aspergillosis, 186 caused by A. fumigates and other species of Aspergillus, such as A. flavus, A. niger, and A. 187 *terreus*, depending on the local epidemiology (Singh and Paterson, 2005) and according to 188 the season (Panagopoulouet al., 2007).

The microbial load of the public hospital were higher than that of the private, this 189 190 tallied with the report Ekhaise et al. (2008) who reported high microbial counts recorded for 191 the public hospital (Central Hospital) as compared to private hospital (Faith Medical 192 Center), could be due to the subsidizes rate of the public hospital so as accommodate 193 more people, compared to the private hospital, where high fees are charged and are not 194 within the reach of the poor people in the society. These findings could be explained by many 195 factors including the number of visitors visiting the children and female wards, which 196 exceeded visitors in other hospital units. It was noted that the amount of materials brought

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197 from outside such as personal belongings, food and fruits were more common in children and 198 female wards. These are recognized as sources of hospital contamination. Hospital surfaces 199 are contaminated by factors inherent to the presence of patients, such as biological 200 fluids, sometimes associated to assistance techniques and hygiene. Another 201 contamination factor would be the circulation of vectors as carrier agents for fungi and bacteria resistant to antimicrobials (Prado et al., 2006; Rodovalhoet al., 2007) 202

The microbial load of the theater couch and surgical ward were found to be the lowest, this could probably due to the fact that there is high sanitary standards in this area as compared to other hospital areas and also theater is a restricted area, this tallied with submission of Ekhaise*et al.* (2008) who reported that the number of microorganisms in the theater was extremely low.

Although, surfaces are not directly connected to transmission in most hospital infections, the impact of hygiene and cleaning procedures in microbial control is evident. It is suggested that microorganisms associated to hospital infections are able to survive during large periods of time, thus being a continuous source of contamination in cases where population control is not efficiently conducted (Kramer *et al.*, 2006; Rossi *et al.*, 2008).

Regular surveillance, cleaning and restriction of patients relative might be among the strict measures necessary to reduce or totally eliminate the microbial load of indoor air of this hospital wards and units (Awosika*et al.*, 2012)

217 CONCLUSION

This investigation has been able to identify and prove the sensitivity patterns of microorganism isolated from hospital environment surfaces. The study has shown that all the hospital environmental surfaces examine in the course of the study in Akure, Nigeria were contaminated with one or more microorganisms.

222 **REFERENCES**

Angelakis, E., Azhar, E. I., Bibi, F., Yasir, M., Al-Ghamdi, A. K, Ashshi, A. M., Elshemi,
A. G. and Raoult, D (2014). Paper money and coins as potential vectors of
transmissible disease. *Future Microbiology*, 9: 249-261.

- Augustowska, M. and Dutkiewicz, J. (2006). Variability of airborne mikroflora in a hospital
 ward within a period of one year. *Annals of Agricultural and Environmental Medicine*,
 13: 99–106.
- Awosika, S. A., Olajubu, F. A., and Amusa, N. A. (2012).Microbiological assessment of
 indoor air of a teaching hospital in Nigeria.*Asian Pacific Journal of Tropical Biomedicine*, 2(6): 465-458
- Bakkali, M. E., Hmid, K., Kari, K. E., Zouhdi, M., Mzibri, M. E. and Lalaoui, A.
 (2015).Characterization of bacterial strains and their resistance status in hospital
 environment.*Journal of Tropical Disease*, 4: 180.
- Barnett, H. L. and Hunter, B. B. (1998).Illustrated genera of imperfect fungi.4th edition. St.
 Paul, Minn: APS Press. pp. 2, 70, 92-94.
- Benenson, A. S. (1995). *Control of communicable diseases manual*, 16th edition.
 Washington, American Public Health Association.
- Buffet-Bataillon, S., Rabier, V., Bétrémieux, P., Beuchée, A., Bauer, M., Pladys, P., Le,
 G. E., Cormier, M. and Jolivet-Gougeon, A. (2009). Outbreak of *Serratiamarcescens*in a neonatal intensive care unit: contaminated unmedicated liquid soap and risk
 factors. *Journal of Hospital Infection*, 72: 17-22.
- Caggiano, G., Napoli, C., Coretti, C., Lovero, G., Scarafile, G., Gigilio, O. D. and Montagna,
 T. (2014). Mold contamination in the controlled hospital environment: A 3 years
 surveillance in southern Italy. *BMC Infectious Diseases*, 14: 595
- Cheesbrough, M. (2010).*District Laboratory Practice in Tropical Countries*, 2ndedition,
 Cambridge University press, Cambridge University Press, New York, pp. 70 -71.
- Dancer, S. J. (2004) How Do We Assess Hospital Cleaning? A Proposal for Microbiological
 Standards for Surfaces Hygiene in Hospital. *Journal of Hospital Infection*, 56: 10-15.
- Davane, M., Suryawanshi, P.A and Nagoba, B. (2014) *Pseudomonas aeruginosa* from
 hospital environment. *Journal of Microbiology and Infectious Diseases*, 4(1):42-43.
- Ekhaise, F. O., Ighosewe O. U., and Ajakpori, O. D. (2008). Hospital indoor air microflora in
 private and government owned hospital in Benin city, Nigeria. *World Journal of Medical Sciences*, 3(1): 19-23
- Ekhaise, F. O., Isitor, E. E., Idehen, O. and Emogbene, O. A. (2010). Airborne microflora in
 the atmosphere of an hospital environment of University of Benin Teaching Hospital
 (UBTH), Benin City, Nigeria. *World Journal of Agricultural Science*, 6(2): 166-170.

Grossi, P. A., Gasperina, D. D., Barchiesi, F., Biancofiore, G., Carafiello, G., De, G. A., Sganga, G., Menichetti, F., Montagna, M. T., Pea, F., Venditti, M., Viale, P., Viscoli, C. and Nanni, C. A. (2011). Italian guidelines for diagnosis, prevention, and

261 262	treatment of invasive fungal infections in solid organ transplant recipients. <i>Transplantation Proceedings</i> , 43 :2463–2471.
263 264	Kramer, A., Schwebke, I. and Kampf, G. (2006). How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. <i>BMC Infectious Diseases</i> , 6 : 130.
265 266 267	Murray, P. R., Baron, E. J., Pfaller, M. A., Tenover, F. C. and Yolken, R. H. (1995). Manual of clinical microbiology. 6th ed. Washington, DC: American Society of Microbiology, p. 282-293.
268 269 270	Ohsaki, Y., Koyano, S., Tachibana, M., Shibukawa, K., Kuroki, M. and Yoshida, I. (2007). Undetected Bacillus pseudo-outbreak after renovation work in a teaching hospital. <i>Journal of Infection</i> , 54 : 617-622.
271 272 273	Olutiola, P.O., Famurewa, O. and Senntag, H.G. (2000). <i>An introduction to General microbiology</i> , Hygiene institute Der UniversitalHeideberg. Federal Republic of Germany. Pp 267.
274 275 276 277	Onipede, A. O., Oluyede, C. O., Aboderin, A. O., Zailami, S. B., Adedosu, A. M., Oyelese AO, et al. (2004). A survey of hospital acquired infection in ObafemiAwolowo University Teaching Hospital, Ile-Ife. <i>African Journal of Clinical and Experimental</i> <i>Microbiology</i> , 5: 108-118.
278 279	Panagopoulou, P., Filioti, J., Farmaki, E., Maloukou, A., Roilides, E. (2007). Filamentous drugs. <i>Infection Control and Hospital Epidemiology</i> , 28: 60–67.
280 281 282	Partridge-Hinckley, K., Liddell, G. M, Almyroudis, N. G. and Segal, B. H. (2009). Infection control measures to prevent invasive mould diseases in hematopoietic stem cell transplant recipients. <i>Mycopathologia</i> , 168 : 329–337.
283	Prado, M. A., Gir, E., Pereira, M. S., Reis, C. and Pimenta, F. C. (2006). Profile of
284	antimicrobial resistance of bacteria isolated from cockroaches
285	(Periplanetaamericana) in a Brazilian health care institution. Brazilian Journal of
286	Infectious Diseases, 10: 26-32.
287	Putsept, E. (1981). Modern hospital. New York: Apen System Cooperation, p. 426.
288 289 290 291	 Rock, C., Thom, K. A., Masnick, M., Johnson J. K., Harris, A. D. and Morgan, D. J. (2014). Frequency of <i>Klebsiellapneumoniae</i>carbapenemase (KPC)-producing and non-KPC-producing <i>Klebsiella</i> species contamination of healthcare workers and the environment. <i>Infection Control and Hospital Epidemiology</i>, 35: 426-429.
292	Rodovalho, C. M., Santos, A. L., Marcolino, M. T., Bonetti, A. M. and Brandeburgo, M. A.
293	(2007). Urban ants and transportation of nosocomial bacteria. Neotropical Entomology,
294	36 : 454-458.

- Rossi, D., Devienne, K. F. and Raddi, M. S. G. (2008). Influence of biological fluids on
 survival of *Staphylococcus aureus* on various dried surfaces. *Revista de CiênciasFarmacêuticasBásica e Aplicada*, 29: 211-214.
- 298 Sehulster, L. M., Chinn, R. Y. W., Arduino, M. J., Carpenter, J., Donlan, R. and Ashford, D. Environmental Infection Control 299 (2003).Guidelines for in Health-Care 300 Facilities.Recommendations from Centers for Disease Control and Prevention (CDC) 301 and the Healthcare Infection Control Practices Advisory Committee (HICPAC). Chicago: American Society for Healthcare Engineering/American Hospital 302 Association. http://www.cdc.gov/hicpac/pdf/guidelines/eic in HCF 03.pdf. Accessed 303 304 13 January 2012.
- Singh, N. and Paterson, D. L. (2005). *Aspergillus* infections in transplant recipients. *Clinical Microbiology Review*, 18: 44–69.
- Tan, T. Y., Tan, J. S., Tay. H., Chua, G. H., Ng, L. S. and Syahidah, N. (2013). Multidrug resistant organisms in a routine ward environment: differential propensity for
 environmental dissemination and implications for infection control. *Journal of Medical Microbiology*, 62: 766-772.
- Ustun, C. and Cihangiroglu.M. (2012). Health care workers' mobile phones: a potential
 cause of microbial cross-contamination between hospitals and community. *Journal* of OccupationalEvironmental Hygiene, 9: 538-42.
- Wenzel, R., Edmond, M., Pittet, D., Devaster, J-M., Brewek, T., Geddes, A., Butzler, J-P.
 (1999). *KontrolazakażeńszpitalnychVademecum.*,Bielsko Biała, α- Medica Press.
- World Health Organization (2002). Prevention of hospital-acquired infections.
 WHO/CDS/EPH/2002.12