

46 sanitation is an important prerequisite to hygienic safety, prosperity and political stability
47 [4]. The conventional method of water purification using aluminum sulphate (alum) and
48 calcium hypochlorite exerts pressure on nations' over-burdened financial resources since
49 they are imported thereby making treated water very expensive in most developing
50 countries and beyond the reach of most rural folks. The use of alternative, non-
51 conventional, relatively cheap, sustainable and readily available water purification
52 methods could be the most suitable intervention for developing countries.

53

54 **The application of *Moringa oleifera* seeds in water treatment**

55 River water drawn for human consumption and general household use can be highly
56 turbid particularly during the rainy season. River silt is churned into suspension and run
57 off from fields and other surfaces carries solid material, bacteria and other
58 microorganisms into the river. It is of paramount importance to remove as much of this
59 suspended matter as possible prior to a disinfection stage and subsequent consumption.
60 This, generally, can be achieved by the addition of coagulants to the raw water, within a
61 controlled treatment sequence [7].

62 In many developing countries, chemical coagulants, such as aluminium sulphate and
63 synthetic poly-electrolytes are usually unavailable [7]. Moringa tree seeds, when crushed
64 into powder, can be used as a water-soluble extract resulting in an effective natural
65 clarification agent for highly turbid and untreated pathogenic surface water [8]. Besides
66 improving water drinkability, this technique reduces water turbidity (cloudiness) resulting
67 in water being both aesthetically as well as microbiologically more acceptable for human
68 consumption [9]. The application of this low cost *Moringa oleifera* seeds is
69 recommended for eco-friendly, nontoxic, simplified water treatment for rural and peri-
70 urban people living in extreme poverty.

71 Using natural materials to clarify water is a technique that has been practiced for
72 centuries and of all the materials that have been used, seeds of Moringa plant have been
73 found to be one of the most effective [10]. Studies have been conducted since the early
74 1970's to test the effectiveness of *Moringa* seeds for treating water [11]. These studies
75 have confirmed that the seeds are highly effective in removing suspended particles from
76 water with medium to high levels of turbidity.

77 *Moringa oleifera* seeds can be applied to treat water on two levels, acting both as a
78 coagulant and an antimicrobial agent [12]. It is generally accepted that *Moringa* plant
79 works as a coagulant that leads to the formation of "flocs" that settle at the bottom of
80 water [12]. The antimicrobial aspects of *Moringa* plant continue to be investigated [11].
81 While there are on-going research work being conducted on the nature and characteristics
82 of these components, it is accepted that treatments with *Moringa* solutions remove 90-
83 99.9% of the impurities in water [13]. A viable alternative to the chemical coagulants is
84 natural coagulant [14]. *Moringa* seed pods are allowed to dry naturally on the tree prior to
85 harvesting. The mature seeds are readily removed from the pods, easily shelled and then
86 may be crushed and sieved using traditional techniques such as those employed for the

87 production of maize flour [15]. The crushed seeds' powder, when mixed with water,
88 yields a solution [16]. To treat surface water, the equivalent weight of seed powder
89 required to make up a crude extract solution is dependent upon the turbidity [9].

90 *Moringa oleifera* derived coagulants offers several advantages over conventional
91 coagulants such as aluminium sulphate [16]. This includes its activity being maintained
92 over a wide range of influent pH values i.e. no pH correction is required. Natural
93 alkalinity of the raw water also remains unchanged following coagulation i.e. no addition
94 of alkalinity is required. Sludge production is also greatly reduced and is essentially
95 organic in nature with no aluminium residuals sludge volumes are reduced by a factor of
96 up to 5 [17].

97

98 **Charcoal filter system and its application in household water purification**

99 Charcoal filters have been used for several hundred years and are considered one of the
100 oldest means of water purification [18]. Historians have shown evidence that carbon
101 filtration may have been used in ancient Egyptian cultures for medical purposes and as a
102 purifying agent [19]. The first recorded use of a charcoal filter to purify potable water on
103 a large scale occurred in 19th century England [19]. Currently, carbon filters are used in
104 individual homes as point-of-use water filters, groundwater remediation, landfill leachate,
105 industrial wastewater and, occasionally, in municipal water treatment facilities.

106 Charcoal filters consist of either compressed charcoal/carbon block which is the best type
107 of charcoal filter since it can remove chemicals and lead. It is however easily clogged,
108 hence should be used with sediment pre-filter. Charcoal filter can also comprise of
109 granular charcoal. This is cheaper, but water can flow around the granules without being
110 treated. Powdered charcoal is a very fine dust useful for spot cleaning larger bodies of
111 water, but is messy and can pass through some filters and be consumed. Activated carbon
112 filters for home-based water treatment typically contain either granular activated carbon
113 or powdered carbon block. Both forms of filters are effective; however, carbon block
114 filters generally have a higher contaminant removal ratio.

115 **Mechanism of action of Charcoal filters**

116 The charcoal filter functions primarily by the process of adsorption. Adsorption, which
117 signifies a surface interaction between dissolved species and the charcoal, is distinct from
118 absorption, which essentially means "to soak up" or "to take into." In water treatment,
119 contaminants diffuse into char pores (absorption) where they bind to charcoal surfaces
120 (adsorption).

121 The porosity and large surface area of charcoal provides a multitude of reactive sites for
122 the attachment of dissolved compounds. These reactive sites can bind non-problematic
123 dissolved organic compounds as well as targeted hazardous contaminants. Background
124 dissolved organic matter, present in all natural waters, can occupy sites on charcoal
125 surfaces and thereby exclude contaminants of concern. This is called "fouling." Fouling
126 in charcoal filters is mitigated by upstream unit processes – in our case, the *Moringa* seed

127 treatment – that act to remove a substantial portion of background dissolved organic
128 matter from the source water before it encounters the charcoal. The principle is to achieve
129 a high level of treatment prior to the charcoal filter, in order to “save the carbon” for
130 removal of targeted problematic dissolved compounds that make it through the previous
131 treatment steps.

132 The charcoal filter in this case functions as a post-coagulation adsorber. The charcoal
133 filter is placed after the *Moringa* seed treatment in order to target specific components of
134 background organic matter (for example, compounds that cause undesirable tastes, odors,
135 or appearance) or synthetic organic compounds (SOCs) such as pesticides,
136 pharmaceuticals, fuel compounds, etc., that are not well removed by the preceding unit
137 processes.

138 The two most important factors affecting the efficiency of charcoal filtration are the
139 amount of charcoal in the unit and the amount of time the contaminant spends in contact
140 with it. The more the charcoal used the better. Similarly, the lower the flow rate of the
141 water, the more time that the contaminants will be in contact with the charcoal, and the
142 more absorption that will take place. Particle size also affects removal rates. The effective
143 lifetime of the charcoal filter media depends upon the quality of the charcoal, as well as
144 the characteristics of the source water and efficacy of upstream treatment steps.

145
146 **Broad objective**

147 The main objective of the study was to evaluate the effectiveness of using *Moringa*
148 *oleifera* seed powder as a coagulant and wattle stem charcoal as filter material in
149 purification of stream water from unprotected sources in Kapseret.

150
151 **Specific objective**

152 To compare the antimicrobial activity of *Moringa oleifera* seed extract integrated with
153 wattle stem charcoal filtration alongside the independent performance of *Moringa*
154 *oleifera* and charcoal filter system against microbial populations in raw water sample.

155
156 **MATERIALS AND METHODS**

157 **Study Area**

158 The study was conducted in Kapseret division, Uasin Gishu County, Kenya. The region
159 covers an area of 148.30 sq. Km. It comprises of Simat, Chepkatet and Lemook locations.
160 It receives an average rainfall ranging between 900-1200mm and this occurs between
161 March and September with two distinct peaks in May and August. The dry spells begin in
162 November and end in February while temperatures range between 8.4 and 26°C but these
163 features are changing probably due to climate change [20]. According to the 2009
164 Population and Housing Census, the total population of area stood at 31,030.

165 The area is a peri-urban setup with an increasing population owing to outward expansion
166 of Eldoret town and rural-urban migration. Major water sources in the area include

167 streams, shallow wells and springs. These sources are usually unprotected and therefore
168 exposed to pollution.

169 **Sampling and Sample Preparation**

170 Sampling procedures described by American Public Health Association [21] were
171 followed. Glass sample bottles (2000 ml) were sterilized in an autoclave at 121 °C for 15
172 minutes at 121 kPa. Two litres of sample was fetched from each of the five streams i.e
173 Leberio, Malanymaina, Lemook, Nganiat and Kapbodigita in sample bottles and the
174 bottles stoppered. Samples were collected from these streams in the study area since they
175 were found to be the commonly used water sources by locals. The sampling sites were
176 identified to represent even distribution of unprotected streams across the study area.
177 Random sampling was used in the study. Samples collected were labeled and placed in a
178 cooler box containing ice blocks and then transported within six hours to Eldoret Water
179 and Sanitation (ELDOWAS) laboratories for analysis.

180 **Preparation of *Moringa oleifera* seed extract**

181 Fully matured *Moringa oleifera* seeds were collected from Marigat forest. The seeds
182 were air-dried in direct sun for a week. The shells surrounding the seed kernels were
183 removed using a knife and the kernels were pounded using laboratory mortar and pestle
184 into fine powder. The powder was sieved using a strainer with a pore size of 2.0mm to
185 separate the coarse powder and obtain only the fine powder to achieve solubilization of
186 active ingredients in the seed. This powder was used to prepare *M. oleifera* stock solution
187 for water purification. The stock solution was prepared by mixing 10, 20, 30, 40, 50 and
188 60g of fine seed powder in 1000ml of distilled water and solution later filtered. The
189 suspension was vigorously shaken for 30 min using a stirrer to promote water extraction
190 of the coagulant proteins and this was then passed through filter paper (Whatman No. 1).
191 The filtrate was used within an hour.

192 **Designing an Improved Charcoal Water Filter**

193 Fresh charcoal that had cooled completely was collected. Wattle tree charcoal was used
194 as it was readily available and has no known side effects. The charcoal was crushed into
195 small bits up to the size of aquarium gravel. The particle sizes of the charcoal were
196 graded from 0.5 mm to 5mm using standard sieves at the Ministry of Public Works
197 laboratory in Eldoret.

198 The graded charcoal sample was sterilized by boiling in water for 15 minutes before use
199 in the filter. A 2-litre cylindrical plastic container with the lower part cut open was
200 obtained. The smaller opening was covered with a piece of fabric that acted to prevent the
201 charcoal from falling out or running through with the water. Approximately 500g of
202 crushed charcoal of varying sizes was packed into the container tightly. This was meant
203 to create as fine a matrix as possible for the water to drip through slowly, thus trapping
204 more sediment. The crushed charcoal was filled up to about halfway the cylinder.
205 Another piece of cloth was placed on top of the charcoal to prevent it from becoming
206 displaced when water was added. The filter was placed atop a sterile container to collect

207 the filtered water. Sterile boiled water was poured through the filter to clean up the
208 charcoal dust and any other soluble particles in the filter.

209 **Sample Filtration**

210 A 500 ml sample of raw water was slowly poured into the filter and allowed to slowly
211 percolate through. The filtrate was collected in a sterilized beaker. The raw and the
212 filtered samples were later analysed for total coliforms, fecal coliforms and biological
213 oxygen demand. To determine the effectiveness of combined activity of *M. oleifera* and
214 charcoal filter, stream water was initially treated with optimum stock solution of *M.*
215 *oleifera*. The treated sample was then passed through the charcoal filter in a similar
216 procedure of filtration undertaken above.

217

218 **Determination of the antimicrobial activity of Wattle tree charcoal-*Moringa oleifera*** 219 **seed filter in water purification.**

220 **Estimation of Total and fecal coliforms**

221 Analysis of collected raw water samples and treated water samples) to estimate the
222 populations of total coliforms and fecal coliforms was done using the Colilert-18 test
223 procedure. This analysis represented one aspect of water quality whose findings were
224 used to draw inferences about the suitability of the water for use based on average
225 microbial populations as per WHO recommendations.

226 One pack of Colilert reagent was added to a 100 ml room temperature water sample in a
227 sterile water container. The container was capped and shaken until its contents dissolved.
228 The sample/ reagent mixture was poured into a quanti tray and sealed in a quanti tray
229 sealer. The quanti-tray 2000 of 97 wells was used. The sealed tray was incubated at 37°C
230 for 18 hours. The results were read according to an interpretation table as described by
231 [22].

232 Fluorescence to detect the presence of *Escherichia coli* was checked using a 6-Watt, 365-
233 nm Ultra violet light lamp within 5 inches of the sample in a dark environment. This
234 procedure ensured that the UV light was directed away from the experimenter's eyes and
235 towards the sample. Colilert results were read after 18 hours, however if the results were
236 ambiguous based on the initial reading, incubating up to additional four hours to allow
237 the color and/or fluorescence to intensify was done. Only sterile, none buffered, oxidant
238 free water for dilutions was used. Aseptic techniques were followed during analysis and
239 good laboratory practice GLP for disposal. Sample tests were stored at 25°C away from
240 light.

241 **Measurements of Biochemical Oxygen Demand (BOD)**

242 Biochemical Oxygen Demand (BOD) is a measure of the oxygen in the water that is
243 required by the aerobic organisms. The biodegradation of organic materials exerts oxygen
244 tension in the water and increases the biochemical oxygen demand (22).

245 In the current study initial DO values were recorded in the field and the same samples
246 incubated at 20 °C for 5 days in dark bottles. This was done in order to avoid some
247 processes like photosynthesis and respiration that could have released or consumed
248 oxygen hence affecting its concentration. Final DO was recorded at the end of 5 days.
249 Biological Oxygen Demand after the 5th day was determined in the formula given below:

250 $BOD_5 = \text{Final DO} - \text{Initial (20)}$. Similar procedure was done for *Moringa oleifera* treated
251 samples and charcoal filtered samples in the laboratory.

252 **Data processing and analysis**

253 Analysis of variance (ANOVA) was conducted to assess whether significant ($p < 0.05$)
254 variations existed among the treatments given to assess their effectiveness as water
255 coagulants. Analysis of data was computed using GenStat Discovery Edition III, 2008.

256

257 **RESULTS**

258 Based on the objectives of the study, the findings of the study were as follows;

259 **Effects of *Moringa oleifera* and charcoal filter on assessment of microbiological** 260 **parameters**

261 In this study, the microbiological parameters under investigation were total coliforms,
262 fecal coliforms and biological oxygen demand. Summaries of the findings for these
263 parameters are shown in tables below.

264 The interactions between *Moringa oleifera* and charcoal filter had significant ($p \leq 0.05$)
265 effects on all microbiological parameters.

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285 **Table 4.1 Analysis of variance (ANOVA) summary on the effect of treatment**
 286 **on percentage reduction of biological parameters (TC, FC and BOD) in**
 287 **sampled water**

Source of variation	Total Coliforms		Fecal Coliforms		BOD	
	F-Value	P-Value	F-Value	P-Value	F-Value	P-Value
Treatment	23.38	0.000*	60.996	0.000*	29.402	0.000*

288 ** Denotes significance at p<0.05

289
290
291
292
293
294
295
296
297
298
299
300
301

302 **Total coliforms**

303 The results of total coliforms in the sample water were as shown in Table 4.2. There
 304 were significant differences in total coliforms count (p<0.05) among the different
 305 treatments of the sample water in the area of study. *Moringa oleifera* reduced the total
 306 coliforms by approximately 33%. Filtration over charcoal reduced the population
 307 significantly by a further 33%. A combination of *M. oleifera* and charcoal filtration
 308 reduced the total coliform population by 92%. This reduction was significantly different
 309 from either using charcoal or *M. oleifera* singly (Table 4.2).

310 **Fecal coliforms**

311 The results of fecal coliforms (Table 4.2), shows that there were significant differences in
 312 fecal coliforms count (p<0.05) among the different treatments. *Moringa oleifera* reduced
 313 the population by 21%. Charcoal filtration further reduced the population by a significant
 314 82%. A combination of the two treatments reduced the population by approximately
 315 99%. This reduction was significantly different from either using charcoal or *M. oleifera*
 316 singly (Table 4.2).

317 **Biological Oxygen Demand (BOD)**

318 The BOD levels were found to be significantly different (p< 0.05) among the different
 319 treatments (Table 4.2). BOD reduction by *Moringa oleifera* was 20%. Filtration over
 320 charcoal reduced the BOD concentration by a further 12%. A combination of the two
 321 treatments reduced the BOD concentration by 51%.

322

323 **Table 4.2 Mean (\pm) percent reductions of microbiological parameters using *Moringa*
 324 *oleifera*, charcoal filter and *Moringa oleifera* and charcoal filter combined in water
 325 treatment.**

Treat ment	Total Colifor ms	Fecal Colifor ms	BOD
B(<i>M</i> <i>oleifera</i> a)	32.56 \pm 1 5.88a	21.37 \pm 1 6.94a	19.95 \pm 9.36 a 31.51
C(Charcoal)	66.05 \pm 1 0.68b	82.44 \pm 1 1.19b	\pm 4.76 b
D(Combine d)	92.36 \pm 1 4.48c	99.23 \pm 0. 84c	50.66 \pm 3.52 c

326 Means followed by different letters within a column are significantly different at
 327 p<0.05
 328

329 DISCUSSION

330 Effect of *Moringa oleifera* and charcoal filter on microbiological parameters

331 *Moringa oleifera* reduced the total coliforms by approximately 33%. Filtration over
 332 charcoal reduced the population significantly by a further 33%. A combination of *M.*
 333 *oleifera* and charcoal filtration reduced the total coliform population by 92%. This
 334 reduction was significantly different from either using charcoal or *M. oleifera* singly.

335 Processing the water by coagulation using *M. oleifera* as natural coagulant showed that
 336 the treatment with *M. oleifera* provided additional advantage of reduced total coliforms
 337 With proper mixing, the moving particles enlarged and formed flocs that fall to the
 338 bottom of the vessel due to gravity. This confirms the effectiveness of *Moringa oleifera*
 339 as coagulant for the purification of dirty water. Furthermore, the decrease in total
 340 coliform number was also affected by alkaline condition generated by *Moringa oleifera*.
 341 Most microorganisms grow well at pH 6.0-8.0, but some of them can grow well at pH 3
 342 (acidophiles) and at pH 10.5 (alkaliphiles). Coliform bacteria are facultative anaerobic
 343 microorganisms that can grow in aerobic environments and in fermentation condition that
 344 produces lactic acid. Therefore these bacteria can still grow at low pH environment,
 345 coliform bacteria can still grow, but they cannot survive alkaline pH 14. Additions of
 346 *Moringa* as coagulant affect the increase in pH which in turn stops bacteria from
 347 growing.

348 Bacterial species *S. faecalis* and *P. aeruginosa* which were cultured in water, stop
 349 growing back after *M. oleifera* seeds were added [23]. When the seeds of *M. oleifera* are
 350 crushed and dissolved into the water, protein produces a positive charge that acts like a
 351 magnet and attracts dominant negatively charged particles such as clay, silk, and other
 352 toxic particles. This is in accordance with the invention that the flocculation process
 353 removes about 90-99% of bacteria that are usually attached to solid particles, so the
 354 bacteria will be aggregated together to form flocs and can be removed from the water
 355 [23]. The control treatment had the highest counts of coliform. This affirms earlier stated
 356 recommendation above that raw water without treatment is not safe for drinking.

357 It was observed that the BOD of raw water was very high. This was due to the presence
 358 of high amount of decomposable organic matter in the water samples. Generally, use of
 359 *M. oleifera* jointly with charcoal filters had significant effects on nutrients and BOD.
 360 BOD reduction by *Moringa oleifera* was 20%. Filtration over charcoal reduced the BOD
 361 concentration by a further 12%. A combination of the two treatments reduced the BOD
 362 concentration by 51%. This was probably due to the fact that the phosphates and nitrates
 363 were filtered out mechanically by adsorption.

364
 365

366 REFERENCE

- 367 1). Sobsey, M. D. and Bartram J. (2003). Water Quality and Health in the New
 368 Millennium: The Role of World
- 369 2). Aulia, H. (1994). *Personal and domestic hygiene and its relationship to the*
 370 *incidence of diarrhea in South Sumatra*. Journal of Diarrhoea diseases
 371 Research, 12(1):428-432.
- 372 3). Sterritt, R. M. and Lester, J. N. (1988). Microbiology for Environmental and
 373 public Health Engineers, E. and F. N. Sponpress, London. p. 247-250.
- 374 4). Bartram, J. and Richard B. (1996). *Water quality monitoring: a practical*
 375 *guide to the design and implementation of freshwater quality studies and*
 376 *monitoring programmes*. 2nd Edition. 2-6 Boundar Row, London SE 18HN
 377 UK. 3:245-274.
- 378 5). Rheingans, R. Dreibelbis, R. and Freeman, M. C. (2006). Beyond the
 379 Millennium Development
- 380 6). Goals; Public Health Challenges in Water and Sanitation: *Global Public*
 381 *Health 1(1): 31-48*.
- 382 7). WHO, Guidelines for Drinking Water Quality, First Addendum to Third
 383 Edition, Recommendations, Vol. 1, 2006, available
 384 online at: [http://www.who.int/water_sanitation_health/dwq/gdwq05](http://www.who.int/water_sanitation_health/dwq/gdwq0506.pdf)
 385 06.pdf. (accessed Aug. 5, 2010)
- 386 8). Sutherland, J. P. (2000). *The application of Moringa oleifera as a coagulant*
 387 *for water treatment in developing countries* (Doctoral dissertation,
 388 Engineering).
- 389 9). Mangale, S. M., Chonde, S. G., Jadhav, A. S., & Raut, P. D. (2012). Study of
 390 *Moringa oleifera* (Drumstick) seed as natural Absorbent and Antimicrobial
 391 agent for River water treatment. *J Nat Prod Plant Resour*, 2(1), 89-100.
- 392 10). Lea, M. (2010). Bioremediation of turbid surface water using seed extract
 393 from *Moringa oleifera* Lam.(drumstick) tree. *Current protocols in*
 394 *microbiology*, 16(1G).
- 395
- 396 11). Sutherland, J. P., Folkard, G. K., & Grant, W. D. (1990). Natural coagulants
 397 for appropriate Water Treatment: a novel approach. *Waterlines*, 8(4), 30-
 398 32.
- 399
- 400 12). Doerr, B., & Staff, E. C. H. O. (2005). *Moringa water treatment*. *ECHO*
 401 *Technical Note*. Educational Concerns for Hunger Organization, Florida,
 402 4.

- 403 13). Ferreira, R. S., Napoleão, T. H., Santos, A. F., Sá, R. A., Carneiro da Cunha,
404 M. G., Morais, M. M. C., & Paiva, P. M. (2011). Coagulant and
405 antibacterial activities of the water soluble seed lectin from *Moringa*
406 *oleifera*. *Letters in applied microbiology*, 53(2), 186-192.
- 407 14). Gambhir, R. S., Sohi, R. K., Bansal, V., Nirola, A., Randhawa, A. K., Sogi,
408 G. M., & Veerasha, K. L. (2013). Newer Water Purification Techniques-A
409 Review. *Indian Journal of Public Health Research & Development*, 4(1),
410 249-252.
- 411 15). Anwar, F., Latif, S., Ashraf, M., & Gilani, A. H. (2007). *Moringa oleifera: a*
412 *food plant with multiple medicinal uses*. *Phytotherapy research*, 21(1), 17-
413 25.
- 414 16). Joshua, R., & Vasu, V. (2013). Characteristics of stored rain water and its
415 treatment technology using moringa seeds. *Int. J. Life Sc. Bt & Pharm.*
416 *Res*, 2(1), 155-174.
- 417 17). Folkard, G., Sutherland, J., & Al Khalili, R. (1995, September). Natural
418 coagulants-a sustainable approach. In *WEDC CONFERENCE* (Vol. 21,
419 pp. 263-265). WATER, ENGINEERING AND DEVELOPMENT
420 CENTRE.
- 421 18). Ndabigengesere, A. & Narasiah, K.S. 1998, Quality of water treated by
422 coagulation using *Moringaoleiferaseed*, *Water Research*, 32 , 3, pp
423 781-791.
- 424 19). Cheremisinoff, Paul N.; Angelo C. Morresi (1980). *Carbon Adsorption*
425 *Handbook*. Ann Arbor, Michigan: Ann Arbor Science Publishers, Inc.
426 pp. 1-54.
- 427 20). Luke, K. K., Job, K. L., & Bernard, K. N. (2011). The Role of Non-farm
428 Investments in Agricultural Risk Management in Kenya. *Current*
429 *Research Journal of Economic Theory*, 3(2), 62-68.
- 430 21). APHA (American Public Health Association), *Standard Methods for the*
431 *Examination of Water and Wastewater*, 19th ed., Repress Springfield,
432 1983, pp. 22-54.
- 433 22). Abida, B. & Harikrishna, (2008), *Study on the Quality of Water in Some*
434 *Streams of Cauvery River*, *Journal of Chemistry*, 5(2), pp 377-384.
- 435 23). Chao, W. L. (2006). Evaluation of Colilert 18 for the detection of coliforms
436 and *Escherichia coli* in tropical fresh water. *Letters in applied*
437 *microbiology*, 42(2), 115-120.
- 438
439