# Original Research Article THE EFFECT OF INDUCERS AND INHIBITORS OF MONOOXYGENASE ON THE ACTIVITY NITRERGIC SYSTEM IN THE MICROSOMES IN THE ISCHEMIC LIVER

## 7 Abstract

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8 In 62 experiments on white out bred rats male weighing 180-220g. Found that inducers and 9 inhibitors of monooxygenase opposite effects on the activity of NOS in the ischemic liver 10 microsomes. Benzonal and cimetidine after 1 night of their introduction had no significant 11 effect on all studied parameters. After 3 and 10 of the daily introduction of the inducer of 12 drug metabolism - benzonal slow speed nitrate reductase system, stimulates nitroxylenes 13 system (eNOS), and cimetidine, on the contrary – even more nitrate reductase activates the 14 speed system, inhibits eNOS nitroxylenes.

Keywords: ischemia, liver, monooxygenases, NO-system, benzonal,
cimetidine.

## 17 INTRODUCTION

An important aspect of modern drug therapy is a personalized medicine 18 based on research and implementations in practical health care of medicines 19 influencing on the system of biotransformation of xenobiotics in the liver 20 21 (Sivkov et al., 2010; Archakov et al. 2008). The inducers and inhibitors of drug 22 metabolism – regulating activity of the monooxygenase system (MOS) of the liver in this problem plays the key role (Kukes et al. 2007; Villeneuve et al. 23 2004). In the last decade, thanks to basic research in molecular biology and 24 medicine found that in vascular endothelium the synthesis of nitric oxide (NO<sup>o</sup>) 25 is the family of cytochrome P-450-like hemoproteins - NO-synthase in 5-26 electron oxidation of L-arginine with the formation of L-citrulline and NO<sup>o</sup> 27 (Minamiyama et al., 2001; Manuhina et al., 2000). A family of isoenzymes of 28 29 NO-synthase (NOS) synthesize NO from L-arginine by three major isoforms –

two constitutive (neuronal (nNOS) and endothelial (eNOS) and one inducible 30 (iNOS) (Ivashkin et al., 2000). For the production of NO with the participation 31 of NOS along with a variety of cofactors, substrates is an important arginine, 32 oxygen and oxidized nicotinamide dinuceotid phosphate (NADPH) (Markov, 33 2005; Vinogradov et al., 2005). In pathological processes accompanied by 34 hypoxia or ischemia, the role of NO- sinus mechanism is reduced and induced 35 activity nitrate reductase systems (Reutov, 2000). It is now established that 36 NOS and inactive nitrate reductase system (LDCs) is found in hepatocytes, 37 endothelium of sinusoids, the Kupffer cells/macrophages (Habib and Ali, 2011), 38 as well as in the endothelium of the portal vein and hepatic artery (Hirst and 39 Robson, 2011; Jaeschke et al., 2001). The presence of NOS in hepatocytes 40 suggests a correlation with the enzymes of the MOS. However, in the literature 41 there is practically no data on the effect of inducers and inhibitors of drug 42 metabolism on the activity of NOS in microsomes isolated from hepatocytes in 43 the development of liver pathological process. 44

In connection with the above, the aim of the study was to study the activity of NOS in the liver microsomes after administration to animals in the dynamics of postischemic period of benzonal and cimetidine.

# 48 MATERIAL AND METHODS

The study was carried out on 62 male rats of mixed population weighing 180-220g., which were divided into 3 groups. First group animals after 1, 2 and 3 days ischemia/hypoxia of the liver caused by occlusion of it during the 180 min of the vascular pedicle of the left lateral and middle lobes.

The study drugs were administered after restoration of blood flow to the liver. An inducer of drug metabolism benzonal was administered intragastrically in the form of a 1% solution in 0.5% starch gel single dose of 56 50mg/kg for 1, 3 and 10 days in a row (2ml). Inhibitor of drug metabolism

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cimetidine also was injected intraperitoneally in a 0.1% aqueous solution daily,
once daily for 1, 3 and 10 days in a row (3<sup>rd</sup> group). Control for all research
groups served as data of intact animals. Each group consisted of 6-8 animals.

The animals were sacrificed by instant decapitation method under light 60 Rausch-anesthesia. The extracted liver was perfused through the inferior vena 61 cava by chilled (0±4°C) 50 mM Tris <u>HCl</u> buffer, pH 7.4, containing 0.05 M KCl 62 and 0.25 M saccharose. After washing the liver from the blood it was ground 63 and homogenized in the same solution (1:3). From that fraction, which was 64 obtained by centrifugation at VAC-602 (Germany) after 20 minutes of 65 unscrewing, with 12 thousand g, had been beset microsomes thousand at 105 g 66 for 60 min. All procedures were performed in the refrigerating chamber KHS-67 12(Russia) at 0±4°C. In microsomes, resuspended in 100 mM Tris - HCl buffer; 68 pH 7.4 was evaluated activity of monooxigenase system that content of 69 cytochromes P-450, P-420, and b5 by classic method of T. Omura, R. Sato 70 71 (1964), the activity of NADPH-reductase (NADPH-op.-ed.) by C. H. Williams, H. Kamin (1961), benzo( $\alpha$ )pyrene hydroxylase (B(a)PG) by C. H. Yang, 72 L.P.Kicha (1978). Aniline hydroxylase (AG) by A. I. Archakov et al. (1975), N-73 demethylase amidopyrine (N-AP) by A. Bast, J. Nordhosck (1981), glucose-6-74 phosphatase (G-6-Phase) by N. S. Gnosh, N. C. Kar (1983) were assessed. 75

76 Nitrooxygenase activity was determined by the content of stable metabolites nitrite and nitrate NO -, NO<sub>2</sub> and NO<sub>3</sub> - by the method of P. P. 77 Golikov et al.(2000), activity of endothelial NOS (eNOS) by Sumbaev V.V., 78 Yasinska, I.M. (2000), inducible NOS (iNOS) and the concentration of 79 peroxynitrite (ONO<sub>2</sub>) in Ravaeva M. Yu, E. N. Chuyan (2011). Content, 80 activity of monooxygenase and oxidoreductase of nitrooxygenase systems were 81 recorded on computerized dual beam spectrophotometer UV-2100 (Ltd, China). 82 The content and activity of oxidoreductase was calculated in microsomes per 83

milligram of protein in 1 ml (mg/ml), which was determined by the method of
O. N. Lowry et al. (1951).

The obtained results were subjected to statistical analysis using the 86 software package Excel, Statistic for Windows V.6.0. Normality of distribution 87 of quantitative parameters was checked using the criteria Kolmogorov-Smirnov 88 and Shapiro-Wilk test. Calculated arithmetic mean (M), standard deviation ( $\sigma$ ), 89 error arithmetic average (m), sample standard deviation (S). The distribution of 90 the samples was carried out on the basis of student's criterion (t) with the 91 computation of error probability (P). The correlations for the indicators was 92 carried out using correlation analysis Pearson (r). For comparison, samples 93 were used Student's t-test. Data were considered significant at p < 0.05. 94

#### 95 **RESULTS AND DISCUSSION**

Benzonal and cimetidine after 1 night of their introduction had no 96 significant effect on all studied parameters characterizing the activity of NOS in 97 the liver microsomes postchemotherapy, compared to groups, which drugs are 98 99 not injected, the corresponding term monitoring (1day.) (Table.1). In 100 subsequent periods after 3 and 10 days benzonal significantly reduced the expression of NO, iNOS and ONO<sub>2</sub> on the background of the dynamic of the 101 102 studied follow-up period of increasing eNOS activity and content of microsomal protein. At the same time after 3 and 10 of the daily administration of 103 cimetidine in a selected microsomal fractions of the liver shows a dynamic 104 period of observation the decrease in the activity of eNOS and increased 105 expression of NO, iNOS and ONO<sub>2</sub><sup>-</sup>, marked inhibition of microsomal protein 106 concentration. Therefore, the introduction of animals with ischemic liver 107 108 benzonal optimizes the processes of NOS in microsomal system in the body, and cimetidine on the contrary an even greater extent, potentiates the effects of 109 damage to this system. When analyzing the performance of NOS is therefore 110 with the activity of eNOS associated changes in the level of iNOS reaction rate, 111

the content of microsomal NO and ONO2<sup>-</sup> in all studied groups of animals. In 112 this regard, it is quite possible to believe that the increased NO and  $ONO_2^{-1}$  is 113 due to inhibition of eNOS and overexpression of iNOS. Benzonal positively 114 influenced changes in the level of NO in microsomes, reducing the activity of 115 iNOS and content of cytotoxic ONO<sub>2</sub>. You can put that with the decreased 116 activity of iNOS and the level of ONO2 was associated, although not 117 significantly increasing the activity of eNOS and restore to control values the 118 concentrations in the ischemic liver microsomes NO administered to animals of 119 benzonal. 120

As follows from literature data, iNOS and ONO<sub>2</sub>. and NO are 121 components of the expression system of nitrate reductase. Its gain during 122 ischemia/hypoxia involves an increase in the cytotoxic compounds, including 123 NO and ONO<sub>2</sub><sup>-</sup> which block the active centers of cytochrome P450 in 124 microsomes ischemic liver [11]. Cimetidine as follows from the data obtained, 125 reinforces these processes in microsomes of animals with ischemic liver and 126 suppresses NOS way. However, as shown by a number of researchers during 127 ischemia/hypoxia blockade of the active site of the isoforms of cytochrome P-128 450 activated oxygen metabolites, including NO and  $ONO_2^{-1}$  have a fragile 129 relationship [12]. In this regard, we can assume that the inducer of drug 130 metabolism benzonal, promotes the release of the connection of active center of 131 cytochrome P-450 with NO and ONO<sub>2</sub> ischemic liver. As a result of increased 132 accessibility to the substrates of oxidation in particular L-arginine, which plays 133 a major role in the regulation of functional metabolic and regenerative functions 134 of liver [13, 14]. This is evidenced by the increase of eNOS activity in 135 microsomes when administered to animals with ischemic liver of benzonal. 136 Therefore, benzonal as an inducer of drug metabolism when administered to 137 animals with ischemic liver microsomes increases in NOS activity, through 138

mechanisms of oppression nitrate reductase components, thus reducing the level in hepatocytes toxic compounds, the overexpression of NO and  $ONO_2^-$ .

Thus, inducers and inhibitors have opposite effects on the activity of NOS 141 in the ischemic liver microsomes. Benzonal - slow speed nitrate reductase 142 system, stimulates nitroxygenase system (eNOS), and cimetidine, on the 143 contrary that even more nitrate reductase system activates the speed system, 144 145 inhibits eNOS nitroxylenes. The difference in activity of benzonal and cimetidine explain through what mechanisms can regulate the enzyme 146 monooxygenase, thereby positively impact on pathological processes in the 147 liver that is critical to its hypoxic conditions. At the present time in connection 148 with the growth of liver disease and aggressive exposure to xenobiotic with 149 induction and inhibitory action on the person, this problem acquires a special 150 urgency and, of course, requires further study. 151

152 Table 1. Dynamics of indicators of activity of NO – system in the liver microsomes after

- 153 playing it acute ischemia/ hypoxia and different periods (day) of benzonal and cimetidine,
- 154 M±M.

Group	NO	aNOS	INOS	ONO2	Drotain No
Gloup	NO,	enos,	INOS,	UNO2-,	гюс,
	mkM/mg	mkM/min/mg	mkM/min/mg	mkM/mg	mg/ml
Control	5,5±0,16	17,4±0,62	0,10±0,002	0,080±0,016	36,8±1,22
Ischemia:					
1 day	8,6±0,33*	7,9±0,29*	0,35±0,017*	0,23±0,010*	29,5±1,13*
3 day	8,1±0,27*	8,5±0,35*	0,23±0,009*	0,19±0,009*	30,8±1,09*
10 day	7,6±0,28*	9,7±0,42*	0,17±0,006*	0,14±0,007*	31,2±1,18
Ischemia+B					
1 day	8,7±0,29*	8,3±0,21*	0,32±0,019*	0,22±0,011*	29,1±1,26*
3 day	$6,3\pm0,26^{*\Delta}$	12,5±0,43* <sup>Δ</sup>	$0,17\pm0,005^{*\Delta}$	$0,16\pm0,006^{*\Delta}$	31,7±1,31
10 day	$5,8\pm0,22^{\Delta}$	$18,4\pm0,59^{\Delta}$	$0,11\pm0,004^{*\Delta}$	$0,07{\pm}0,005^{*\Delta}$	37,5±1,42
Ischemia+C:					
1 day	8,9±0,39*	8,1±0,28*	0,36±0,019*	0,25±0,013*	28,7±1,26*
3 day	$10,6\pm0,37^{*\Delta}$	8,4±0,15*	0,33±0,012* <sup>Δ</sup>	0,21±0,011*	28,3±1,33*
10 day	$13.5\pm0.52^{*\Delta}$	$7.2\pm0.18^{*\Delta}$	$0.46\pm0.021^{*\Delta}$	$0.35\pm0.014^{*\Delta}$	$32.6\pm1.40$

15 \* - P<0.05 compared with control,  $\delta$  - P<0.05 compared to hypoxia of the corresponding

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<sup>156</sup> period

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