

1 **IN VITRO-MODULATION OF HbS ERYTHROCYTE PARAMETERS BY**
2 **PREDNISOLONE TESTING FOR Fe²⁺/Fe³⁺ RATIO, HBS GELATION AND**
3 **OSMOTIC FRAGILITY**
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6
7 **ABSTRACT**
8

9 **Aim:** It has been discovered that some drugs or medications taken for certain ailments are
10 either pro-sickling or anti-sickling in nature. In this study, a corticosteroid by the name of
11 prednisolone was investigated to determine its possible effects on human haemoglobin-S
12 gelation, erythrocyte fragility and Fe²⁺ and Fe³⁺ concentrations.

13 **Materials and method:** Various concentrations of the drug (0.05, 0.1, 0.3, 0.5 and 1mg/ml)
14 were used to determine the effects on human haemoglobin-s, gelation rate, erythrocyte
15 fragility, Fe²⁺ & Fe³⁺ concentrations. Absorbance reading was taken at 540nm using a
16 spectrophotometer.

17 **Results:** The results showed that Prednisolone increased haemoglobin S gelation at all
18 concentrations (p < 0.05) when compared to the control. The Fe²⁺/Fe³⁺ ratio showed a
19 reduction in haemoglobin values at 0.3, 0.5 and 1.0mg/ml concentrations when compared to
20 the control and a slight increase at 0.05 and 0.1mg/ml. For Erythrocyte Fragility, there was
21 destabilization of red cell in all concentrations.

22 **Conclusion:** This study suggests that this drug could have some undesirable effects on sickle
23 cell subjects.
24

25 **KEY WORDS:** Erythrocyte, Fragility, Gelation, Haemoglobin, Prednisolone, Sickle cell.
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28 **1. INTRODUCTION**

29 Sickle-cell disease (SCD) is a group of genetically passed down blood disorders. The most
30 common type is known as sickle-cell anaemia. It results in an abnormality in the oxygen-
31 carrying protein haemoglobin found in red blood cells. This leads to a rigid, sickle-like shape
32 under certain circumstances. Problems in sickle cell disease typically begin around 5 to 6
33 months of age. A number of health problems may develop, such as attacks of pain ("sickle-

34 cell crisis"), anaemia, bacterial infections, and stroke. Long term pain may develop as people
35 get older. The average life expectancy in the developed world is 40 to 60 years [1, 2, 3].

36 Sickle-cell disease occurs when a person inherits two abnormal copies of the haemoglobin
37 gene, one from each parent. Several subtypes exist, depending on the exact mutation in each
38 haemoglobin gene. An attack can be set off by temperature changes, stress, dehydration, and
39 high altitude. A person with a single abnormal copy does not usually have symptoms and is
40 said to have sickle-cell trait. Such people are also referred to as carriers. Diagnosis is by a
41 blood test and some countries test all babies at birth for the disease. Diagnosis is also possible
42 during pregnancy. The care of people with sickle-cell disease may include blood transfusion.
43 A small proportion of people can be cured by a transplant of bone marrow cells [4, 5].

44 As of 2013 about 3.2 million people have sickle-cell disease while an additional 43 million
45 have sickle-cell trait. About 80% of sickle-cell disease cases are believed to occur in sub-
46 Saharan Africa. It also occurs relatively frequently in parts of India, the Arabian peninsula,
47 and among people of African origin living in other parts of the world. In 2013, it resulted in
48 176,000 deaths, up from 113,000 deaths in 1990. The condition was first described in the
49 medical literature by the American physician James B. Herrick in 1910. In 1949 the genetic
50 transmission was determined and in 1954 the protective effect against malaria of sickle-cell
51 trait was described [6, 7]. In Nigeria about 10,000 sickle cell anemia are born yearly. The
52 carrier frequency ranges between 10% and 40% across Africa [7, 8].

53 Sickle cell disease is also a single-gene aspect that results in sickle shaped red blood cells.
54 Although the manifestations of sickle cell disease have been described, variations in the
55 severity and a number of manifestation as well as interactions with other health events leads
56 to significant gap in the understanding of the natural history of the disorder for as
57 impairments to renal, cardiac and pulmonary organ function are known to occur in sickle cell

58 patients. However the descriptions of these outcomes is generally retrospective in nature and
59 occurs when organ damage is severe [9, 10, 11].

60 In sickle hemoglobin (Hbs), a valine is substituted for glutamic acid on the surface of the
61 Hbs molecule in the sixth codon of the betaglobin (HBB_{glu}val). This change endows
62 Haemoglobin s when deoxygenated with a new property, the capacity to polymerize
63 conspires against an indispensable feature of the red cell [12, 13].

64 The polymerization of deoxy Hbs is the primary and indispensable event in the molecular
65 pathogenesis of sickle cell disease. The polymer has the form of an elongated rope- like fibre
66 which usually aligns with other fibres forming a fascicle and distorting the red cell into the
67 classic crescent or sickle shape, among other many abnormal shapes, and resulting in a
68 marked degree in cell deformability [14].

69 Haemoglobin-S can polymerize when oxygenated and depolymerize when reoxygenated
70 infinitely, however, the sickle erythrocyte membrane can withstand only a finite number of
71 these cycles before it is irreversibly deformed and recognized via the many abnormal “sickle”
72 shapes that circulate in patients [3].

73 Polymerization alone does not account for the physiology of sickle cell disease. Changes in
74 red cell membrane structure and function, disordered red cell volume control, also contribute
75 to the physiology of the disease. Furthermore, increased red cell adherence to vascular
76 endothelium, misregulation of vaso-activity also contribute to sickle vaso-occlusion and
77 haemolysis. In genetics, these are called pleiotropic effects, because they go beyond the
78 immediate consequences of the abnormal gene [15].

79 It is known that most sickle cell patients usually experience severe painful crisis and over the
80 years there has been the use of synthetic drugs to manage or alleviate this pain. This research
81 work is concerned with the ultimate goal of determining the modulatory effects of the drug

82 commonly known as prednisolone and how it affects the human erythrocyte through its effect
83 on $\text{Fe}^{2+}/\text{Fe}^{3+}$ ratio, gelation rate and osmotic fragility in ranging concentrations

84 The objectives of this work are to determine the effect of prednisolone on human Hbs
85 gelation rate, secondly to determine the effect of prednisolone on haemoglobin-S
86 erythrocyte osmotic fragility and finally, to determine the effect of prednisolone on human
87 Hbs erythrocyte haemoglobin (Fe^{2+}) and methaemoglobin (Fe^{3+}) ratios.

88 Prednisolone is a steroid under the class of corticosteroids, gluco corticoids and anti-
89 inflammatory agent. The brand name is prelone. It prevents the release of substances in the
90 body that causes inflammation. Prednisolone is used to treat many different conditions such
91 as allergic disorders, skin conditions, ulcerative colitis, arthritis, lupus, psoriasis or breathing
92 disorders. It is the active metabolite of the drugs prednisone and is used especially in patients
93 with liver failure as these individuals are unable to metabolize prednisone into active
94 prednisolone. Adverse effects are not generally seen with short term therapy, but weight gain,
95 impaired immune response and disturbances in behaviour commonly occur with longer
96 durations of treatment [16].

97

98 **2. MATERIALS AND METHOD**

99 The blood sample of 5ml was collected from adult male and female donors by vein puncture
100 using a 5ml syringe and needle. The blood samples were confirmed as HbSS using standard
101 haemoglobin electrophoresis. All experiments were carried out with fresh heparinized blood.
102 Prednisolone tablets were bought from a Pharmacy in Port Harcourt Rivers State.

103 **2.1. Preparation of five serial dilutions of prednisolone and tablet solution**

104 Prednisolone of 50mg tablet was crushed using mortar and pestle then added to a
105 50ml of distilled water to form the solution. The solution was then stirred properly. After this,
106 different concentrations of prednisolone were made using different reagent bottles.

Bottles	Conc/mg/ml	Vol. of Prednisolone (ml)	Volume of distilled water (ml)
1	0.05	1	19
2	0.10	2	18
3	0.30	6	14
4	0.50	10	10
5	1.00	20	0

107

108 The reagent bottles containing the solution were then stored in the refrigerator to preserve the
109 freshness of the solution.

110 Control procedure for prednisolone:

111 Exactly 5ml of distilled water was put in a test tube using a micro pipette, 0.1 ml of blood
112 sample was then added to the 5ml of water in the test tube using a micropipette. It was shaken
113 and the mixture was transferred into a cuvette. The absorbance reading was taken at different
114 wavelength of 540nm and 630nm using a spectrophotometer.

115

116 Test experiment for prednisolone:

117 Five test tubes were labeled at different concentrations in mg/ml and 5ml of distilled water
118 was added to each of the test tubes.

119 Then 0.1 ml of the blood sample was added to the 5ml of water in the test tubes.

120 The test tubes were shaken by inversion until equal mixture was obtained. Exactly
121 0.1 ml of the test compound (prednisolone solution) was added into the 5ml of water and
122 then 0.1 ml of blood was added into each of the test tubes with the different concentrations in
123 (mg/ml).

124 The mixtures were transferred into a cuvette and then the absorbance reading were taken at
125 540nm using a spectrophotometer.

126

127 Preparation of control sample for osmotic fragility:

128 Exactly 5ml of normal saline was measured into a test tube containing 0.05ml of blood this
129 was thoroughly mixed by inverting the tube several times. The suspension was allowed to
130 stand for 30minutes after which the content was centrifuged for 5 minutes at 1200rpm.

131 **2.2. Osmotic fragility procedure**

132 5ml of normal saline, was measured into 5 test tubes. Then 0.1ml of the test compound was
133 also added into each of the test tubes. To each of the test tubes 0.05ml of blood sample was
134 added and mixed thoroughly by inverting the tubes several times. The suspensions were
135 allowed to stand at room temperature after which the content was centrifuged for 5 minutes at
136 1200rpm. The relative amount of haemoglobin released into the supernatant was determined
137 using a Spectrophotometer at the maximum wave length of 540nm. The physiological saline
138 solution and distilled water served as 100% lysis point and blank respectively [17, 18].

139

140 **2.3. Haemoglobin-S gelation (polymerization) experiment.**

141 Principle: Haemoglobin-S undergoes gelation when deprived of oxygen.

142 Sodium metabisulphite was used as a reductant. The haemoglobin polymerization (gelation)
143 experiment was based on the method described by Noguchi and Schechter [19].

144 Control experiment: Sodium metabisulphite (4.4ml), 0.1ml of HbSS haemosylate and 0.1ml
145 of normal saline were quickly mixed in a cuvette and the absorbance read at 540nm, and at 1
146 minute intervals for 5 minutes.

147 Test experiment: Sodium metabisulphite (4.4ml), 0.1ml of HbSS haemosylate and 0.1ml of
148 test compound were quickly mixed in a cuvette and the absorbance read as was the control.

149

150 Preparation of control sample for Fe^{2+} and Fe^{3+} ratio:

151 Exactly 5ml of distilled water was measured in a test tube containing 0.1ml of blood. They
152 were mixed and allowed to stand for 5 minutes. Fe^{2+} and Fe^{3+} concentration were measured
153 using spectrophotometer at 540nm and 630nm respectively.

154

155 **2.4. Procedure for Fe^{2+} and Fe^{3+} ratio of prednisolone**

156 Five test tubes were labeled 0.05, 0.1, 0.3, 0.5, 1.0mg/ml respectively representing the
157 different concentrations of the drug prednisolone. Then 5ml of distilled water was added into
158 the five labeled test tubes.

159 Exactly 0.2 ml of normal saline was added into each of the test tubes containing the distilled
160 water using a micropipette. Then 0.1ml of the erythrocyte haemosylate was added into each
161 of the test tubes. Subsequently, 0.1ml of the test solution (the drug) was added into each of
162 the test tubes with the respective concentration except the controls. The solution was properly
163 mixed. The absorbance readings were taken respectively [17, 18, 19].

164

165 **2.5. Statistical analysis**

166 All data were subjected to statistical analysis. Values are reported as mean \pm standard error of
167 mean (SEM) while one way ANOVA was used to test for differences between treatment
168 groups. The results were considered significant at p-values of less than 0.05, that is, at 95%
169 confidence level ($p < 0.05$).

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172 **3. RESULTS**

173 The results of the *in vitro* modulation of some Hbs erythrocytes parameters by prednisolone
174 are illustrated in the Tables 1 to 3.

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180 **Table 1: Effect of Predinosolone on Hbs gelation rate**

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182

Predinosolone	Erythrocyte fragility and Hb gelation (optical density) standard = 0.03. (Five minutes duration at one minute interval)
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(mg/ml)	0	1	2	3	4	5
Control	0.00 ±	0.015 ±	0.015 ±	0.015 ±	0.015 ±	0.015 ±
0.00	0.00	0.00	0.00	0.00	0.00	0.00
0.05	0.00 ±	0.160 ±	0.161 ±	0.162 ±	0.163 ±	0.164 ±
	0.00	0.00	0.00	0.00	0.00	0.01*
0.10	0.00 ±	0.180±	0.181 ±	0.182 ±	0.183 ±	0.184 ±
	0.00	0.00	0.00	0.00	0.00	0.01*
0.30	0.00 ±	0.184 ±	0.185 ±	0.186 ±	0.187 ±	0.188 ±
	0.00	0.00	0.00	0.00	0.00	0.00*
0.50	0.00 ±	0.186 ±	0.187 ±	0.188 ±	0.189 ±	0.190 ±
	0.00	0.00	0.00	0.00	0.00	0.00*
1.00	0.00 ±	0.188 ±	0.189 ±	0.190 ±	0.191 ±	0.192 ±
	0.00	0.00	0.00	0.00	0.00	0.00*

183 Results are means of three determinations ± standard deviation.

184 *Statistically significant at 95% confidence level, (P < 0.05).

185

186 This result shows that Prednisolone increased Hbs- Gelation in a concentration and time
 187 dependent manner.

188 **Table 2: Effect of Predinosolone on Hbs Fe²⁺/Fe³⁺**

Prednisolone mg/ml	Fe ²⁺ /OD 540 nm	Fe ³⁺ /OD 630 nm

Control	0.142 ± 0.00	0.142 ± 0.00
0.05	0.171 ± 0.00	1.184 ± 0.00
0.10	0.174 ± 0.01*	0.187 ± 0.00
0.30	0.178 ± 0.00	1.194 ± 0.01*
0.50	0.179 ± 0.01*	1.200 ± 0.01*
1.00	0.183 ± 0.01*	1.215 ± 0.00

189 Results are means of three determinations ± standard deviation.

190 *Statistically significant at 95% confidence level, (P < 0.05).

191

192 **Table 3: Effect of prednisolone on Hbs osmotic fragility**

Prednisolone mg/ml	OD	% Hemolysis
Control	0.400	0
0.01	0.352 ± 0.02	4.60 ± 0.02*
0.05	0.462 ± 0.02	6.00 ± 0.02*
0.3	0.581 ± 0.02	18.00 ± 0.02*
0.5	0.623 ± 0.02	22.00 ± 0.02*
0.1	0.672 ± 0.03	27.00 ± 0.03*

193 Results are means of three determinations ± standard deviation.

194 *Statistically significant at 95% confidence level, (P < 0.05).

195

196 **4. DISCUSSION**

197 Sickle cell disease is genetic disorder which has over the years proved difficult to manage
198 which results from the abnormal haemoglobin S.

199 Sickle cell diseases has caused lots of problem to human life, that is why efforts are
200 constantly being made to find a permanent cure to the disease.

201 For Fe^{2+} and Fe^{3+} the result indicates a progressive effect in the values obtained with
202 increasing concentration of the drug from 0.01mg/ml to 0.1 mg/ml. The results also show that
203 Prednisolone inhibited Hbs- Gelation at increasing concentration and so a reduction in the
204 polymerization rate.

205 In Erythrocyte fragility, at 1mg/ml concentration, Prednisolone stabilized the red cell, but
206 destabilized the cells at lower concentrations which include 0.5mg/ml, 0.3 mg/ml, 0.05mg/ml
207 and 0.1mg/ml. this indicates that Prednisolone inhibits Erythrocyte fragility in Haemoglobin-
208 S patient but can only be administered at 1mg/ml concentration if so desired.

209 Several works have been carried out in the past to determine the effect of various drugs taken
210 for other ailments on sickle cell haemoglobin gelation [20, 21, 22, 23, 24, 25]. The
211 biochemistry of haemoglobin S and the hypothesized pathophysiologic mechanisms of
212 complications provide principles for treatment of complications in sickle cell syndromes.
213 These principles must be applied in preventing and treating almost every complication
214 observed during the clinical course of patients. The pathophysiology of pain and principles of
215 pain management are shown to provide a rational basis for the use of analgesics to treat pain
216 associated with complications [26, 27].

217 Also good medical as well as nutritional management has helped immensely in reducing the
218 incidence of death among sicklers. Most of these treatments are aimed at alleviating one or

219 more of the complications that accompanies crisis [28, 29]. At present time, the important
220 aspect of treatment is supportive care with fluids and analgesics and the judicious use of
221 transfusion. Most currently some chemicals have been found to effective in preventing or
222 managing sickle cell disease in various way. These includes nutritional supplement
223 “sicklevit” an antisickling formula have been formulated. It exerts its effects in reducing the
224 frequency of crisis occurring and act as a maintenance formula [7, 27, 30].

225 **5. CONCLUSION**

226 In conclusion, the present study has shown that Prednisolone has a sickling effect through its
227 inhibition of Haemoglobin-S Gelation rate. Furthermore, the drug had negative effect on Hbs
228 erythrocyte fragility where there was a gross destabilization of the erythrocyte at varying
229 concentrations except at 1mg/ml and Fe^{2+} and Fe^{3+} concentration which was also seen to
230 decrease when compared to the control. Therefore Prednisolone shouldn't be administered to
231 HBS patients.

232 **Competing Interests**

233 Authors have declared that no competing interests exist.

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236 **6. ETHICAL APPROVAL:**

237 This research work was carried out with the approval of the University of Port Harcourt
238 research ethics committee.

239

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UNDER PEER REVIEW