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3 **Mycorrhizal Colonization in *Atriplex nummularia* Lind.**
4 **Subjected to Desalinizador Reject**
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8 **ABSTRACT**
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10 This work has the objective of evaluating the mycorrhizal colonization of AMF
11 *Claroideoglobus etunicatum* in *Atriplex nummularia* Lind. subjected to desalinador
12 reject. The experiment was conducted in a greenhouse at the headquarters of Agronomic
13 Institute of Pernambuco - IPA, Recife, Pernambuco, Brazil. The experimental design
14 was randomized blocks with the treatments constituted in a factorial scheme of five
15 levels of salinity in AC= 2.86 mS/cm; T1= 11.54 mS/cm; T2= 12.04 mS/cm; T3= 13.13
16 mS/cm and T4= 14.16 mS/cm, associated with the presence and absence of fungus,
17 presence and absence of nutrient solution, and autoclaved and non-autoclaved soil. 8.0g
18 of Hoagland & Arnon complete nutrient solution was added every fortnight. After five
19 months, the roots of the treatments were collected and the root colonization was
20 evaluated. It was found that in all treatments the association between *Claroideoglobus*
21 *etunicatum* and *Atriplex nummularia* was beneficial. The correlation was positive for
22 the treatment T4 (Reject + 14gNaCl) + AMF. Thus, it was observed that salinity had no
23 negative effect on the association as well as on the growth of the vegetable.
24

25 **Keywords:** Mutualism, plant growth, halophyte plants, AMF, saline reject.
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29 **1. INTRODUCTION**
30

31 Salinity is an abiotic stress limiting plant growth and productivity worldwide, which can
32 induce physiological changes and compromise plant growth and development [1].

33 Amidst this, some species of the *Chenopodiaceae* family are very tolerant to high
34 salinity and many of them are of great economic and environmental importance.

35 *Atriplex nummularia* is one of the species tolerant to salinity, which has been used as a
36 model plant in biosaline conditions due to its halophytic nature and also to extract salts
37 from the soil and accumulate in the aerial part [2, 3].

38 Mycorrhiza is a mutualist association between certain soil fungi and plant roots. Its
39 classification is divided into three groups according to its morphological and anatomical
40 aspect of root colonization: ectomycorrhiza, endomycorrhiza and ectoendomycorrhiza.
41 AMF are endomycorrhizal due to penetration of the internal mycelium into the
42 intercellular and intercellular root cortex. One of the main characteristics of this
43 association is to increase the surface area of the root and to allow greater capacity of
44 absorption of water and nutrients of the soil, providing a higher rate of growth and
45 survival [4, 5, 6].

46 Therefore, the objective of this work is to analyze the mycorrhizal colonization of
47 *Claroideoglossum etunicatum* in *Atriplex nummularia* Lind. irrigation with the
48 desalinator reject.

49

50 **2. MATERIALS AND METHODS**

51 The experiment was conducted in the period from November / 2018 to March / 2019 in
52 a greenhouse at the headquarters of Agronomic Institute of Pernambuco - IPA, Recife,
53 Pernambuco, Brazil.

54 The soil was obtained from the Experimental Station of the IPA of São Bento do Una,
55 air dried, dewormed, homogenized and sieved in 2 mm mesh. Part of the soil was
56 autoclaved at 120 °C for one hour. Then the sterile and the natural soils were transferred
57 to the 80 polyethylene vessels.

58 In the Laboratory of Soil Fertility of the IPA a chemical analysis was carried out,
59 determining $P = 350 \text{ mg/dm}^3$, $\text{pH (H}_2\text{O)} = 7.8$, $\text{Ca} = 16 \text{ cmolc/dm}^3$, $\text{Mg} = 3.9$
60 cmolc/dm^3 , $\text{Na} = 3 \text{ cmolc/dm}^3$, $\text{K} = 0.7 \text{ cmolc/dm}^3$ and $\text{Al} = 0 \text{ cmolc/dm}^3$. The physical
61 characteristics of the soil were $\text{Dap} = 1.29 \text{ g/cm}^3$, $\text{Dr} = 2.62 \text{ g/cm}^3$, Coarse sand = 7%,
62 Sand = 21%, Silte = 56%, Flocculation = 100%, Clay = 19%, Texture = Franco-silty,
63 Residual humidity = 1.7%.

64 The reject for irrigation was obtained from the desalinator located in the municipality of
65 Riacho das Almas, Pernambuco, Brazil. The physicochemical analysis was performed at
66 the IPA Plant, Ration and Water Analysis Laboratory - LAPRA with the following
67 characteristics: electrical conductivity = 11.541 $\mu\text{S/cm}$ at 25 °C, $\text{Ca}^{+2} = 403 \text{ mg/L}$, Mg^{+2}
68 = 393.09 mg/L, $\text{Na}^+ = 200 \text{ mg/L}$ and $\text{K}^+ = 40 \text{ mg/L}$, $\text{RAS} = 23.67$, $\text{pH} = 7.9$,
69 classification for irrigation = C4S4 (very high salinity water and high sodium
70 concentration).

71 The seedlings of *Atriplex* were obtained with 120 days of age, multiplied by the cutting
72 technique, and then planted in commercial substratum. After 30 days, the best seedlings
73 were chosen to be transplanted into the vessels.

74 The AMF used was purchased from the AMF Inoculum Bank of the Laboratory of Soil
75 Microbiology of IPA. During the transplantation into the vessels, the soil was
76 inoculated - inoculum containing 50 g of *Claroideoglomus etunicatum*.

77 In all treatments, 8mL of [7] was applied fortnightly. In addition, the temperature and
78 humidity of the greenhouse were monitored daily.

79 The experimental design was randomized blocks consisting of a factorial scheme with 5
80 levels - AC: water – EC = 2.86 mS/cm; T1: reject – EC = 11.54 mS/cm; T2: reject plus
81 7g NaCl – EC = 12.4 mS/cm; T3: reject plus 14g NaCl – EC = 13.13 and T4: reject plus
82 21g NaCl – EC = 14.16 mS/cm, associated with two levels of the fungus (presence and
83 absence), two levels of the nutrient solution (presence and absence) and two levels of
84 the soil (autoclaved and non-autoclaved). A 5x2x2x2 factorial is then used, with two
85 replicates, totaling 80 experimental units.

86 After five months of experiment, height was measured. Then the roots were collected,
87 washed and sent to the Laboratory of Soil Microbiology - IPA, to perform the root
88 coloration.

89 Root staining was performed using the methodology described by [8], where five grams
90 of secondary roots were removed from the plants, washed and placed in 10% KOH
91 solution and heated in a 90 °C water bath, for ten minutes. The roots were then washed
92 in running water to remove excess KOH, placed in 10% H₂O₂ solution for two minutes,
93 washed in running water and placed in 1% HCl solution for five minutes. The HCl was
94 then discarded and 0.05% trypan blue solution was added, heated at 90 °C for ten
95 minutes, excess dye removed and the roots were placed in lactoglycerol.

96 The evaluation of the mycorrhizal colonization was through the technique of [9], by
97 observing the fungal structures (hyphas, arbuscules, vesicles and glomerospores) in the
98 interior of the roots, in the cortex region. Root segments of approximately one cm of
99 stained sample were randomly selected and assembled in parallel in slices, in groups of
100 10 (10 slides with 10 root segments each). One hundred root segments were sufficient
101 for evaluation by the method chosen. The root segments were fixed with lactoglycerol,
102 covering the entire surface of the slides, which were then covered with cover slip,
103 without forming air bubbles.

104 The method consists in evaluating the presence or absence of colonization in each
 105 segment and the result expressed in percentage of colonized roots.
 106 For statistical analysis, the means between the treatments were compared through the
 107 analysis of variance Anova and the Tukey test at 5% probability. In addition, Pearson's
 108 (r) simple correlation analysis was performed between plant height and percentage of
 109 mycorrhizal colonization. These analyzes were performed using the software Statistica
 110 version 10.

111

112 3. RESULTS AND DISCUSSION

113

114 During the conduction of the experiment, the average temperature and atmospheric
 115 humidity of the greenhouse was 32°C and 61%, respectively.

116 All treatments presented a significant difference between means at the significance level
 117 of $p < 0.001$. The difference between the means of the treatments for the mycorrhizal
 118 colonization of *Atriplex* roots and for the height are presented in Table 1.

119 The best result obtained for the colonization of *Claroideoglomus etunicatum* in the
 120 *Atriplex* was the treatment T3 (reject plus 14g NaCl - EC = 13.13 mS / cm) in the
 121 presence of AMF in autoclaved soil (9.6%).

122 The root colonization observed was of the *Paris* type, characterized by the intracellular
 123 growth of hyphas, in a linear and longitudinal way along the cortical space. The fungal
 124 structures found in all treatments are shown in Figure 1.

125

126 **Table 1. Percentage of mycorrhizal colonization and height of *Atriplex nummularia***
 127 ***Lind.* cultivated under different conditions**

128

| Treatments | Colonization (%) | Height (cm) |
|-----------------------------------|------------------|---------------|
| AC= Water + AMF | 8.7 ± 0,3 abcd | 72 ± 5,0 bcd |
| AC= Water + AMF + SHA | 8.2 ± 0,2 cd | 80 ± 1,0 acdb |
| T1= Reject + AMF | 8.4 ± 0,2 bcd | 106 ± 22 ab |
| T1= Reject + AMF + SHA | 9.0 ± 0,1 abcd | 73 ± 3,0 bcd |
| T2= Reject + 7g NaCl + AMF | 7.3 ± 0,4 f | 88 ± 2,0 abc |
| T2= Reject + 7g NaCl + AMF + SHA | 8.1 ± 0,1 de | 73 ± 5,0 bcd |
| T3= Reject + 14g NaCl + AMF | 9.4 ± 0,1 ab | 87 ± 1,0 abc |
| T3= Reject + 14g NaCl + AMF + SHA | 9.3 ± 0,1 abc | 70 ± 6,0 cd |
| T4= Reject + 21g NaCl + AMF | 9.3 ± 0,2 abc | 83 ± 2,0 abcd |
| T4= Reject + 21g NaCl + AMF + SHA | 9.5 ± 0,1 ab | 110 ± 2,0 a |
| * AC= Water + AMF | 8.2 ± 0,1 de | 102 ± 1,0 ab |
| * AC= Water + AMF + SHA | 9.4 ± 0,1 abc | 92 ± 1,0 abc |

| | | |
|------------------------------------|----------------|---------------|
| *T1= Reject + AMF | 8.5 ± 0,1 abcd | 70 ± 4,0 bcd |
| *T1= Reject + AMF + SHA | 9.1 ± 0,2 abcd | 82 ± 1,0 abcd |
| *T2= Reject + 7g NaCl + AMF | 9.1 ± 0,2 abc | 75 ± 3,3 abcd |
| *T2= Reject + 7g NaCl + AMF + SHA | 8.8 ± 0,2 abcd | 91 ± 3,0 abc |
| *T3= Reject + 14g NaCl + AMF | 9.6 ± 0,2 a | 87 ± 3,0 abc |
| *T3= Reject + 14g NaCl + AMF + SHA | 8.1 ± 0,4 de | 54 ± 6,0 d |
| *T4= Reject + 21g NaCl + AMF | 9.3 ± 0,2 abc | 72 ± 6,0 bcd |
| *T4= Reject + 21g NaCl + AMF + SHA | 8.1 ± 0,2 de | 75 ± 1,0 abcd |

129 AMF = Arbuscular mycorrhizal fungus (*Claroideoglossum etunicatum*); SHA = Solution of Hoagland and Arnon (1950).
130 Different letters indicate a significant difference at the 5% probability level.
131 Values represent mean ± standard error.
132 * Autoclaved soil.
133

134 Furthermore, arbuscules were not observed; however, there were several septate hyphas
135 in non-autoclaved soil of T3 and T4 treatments (reject plus 14g NaCl – EC = 14.16
136 mS/cm), which is not characterized by fungal hyphas (14 g NaCl - EC = 14.16 mS /
137 cm).

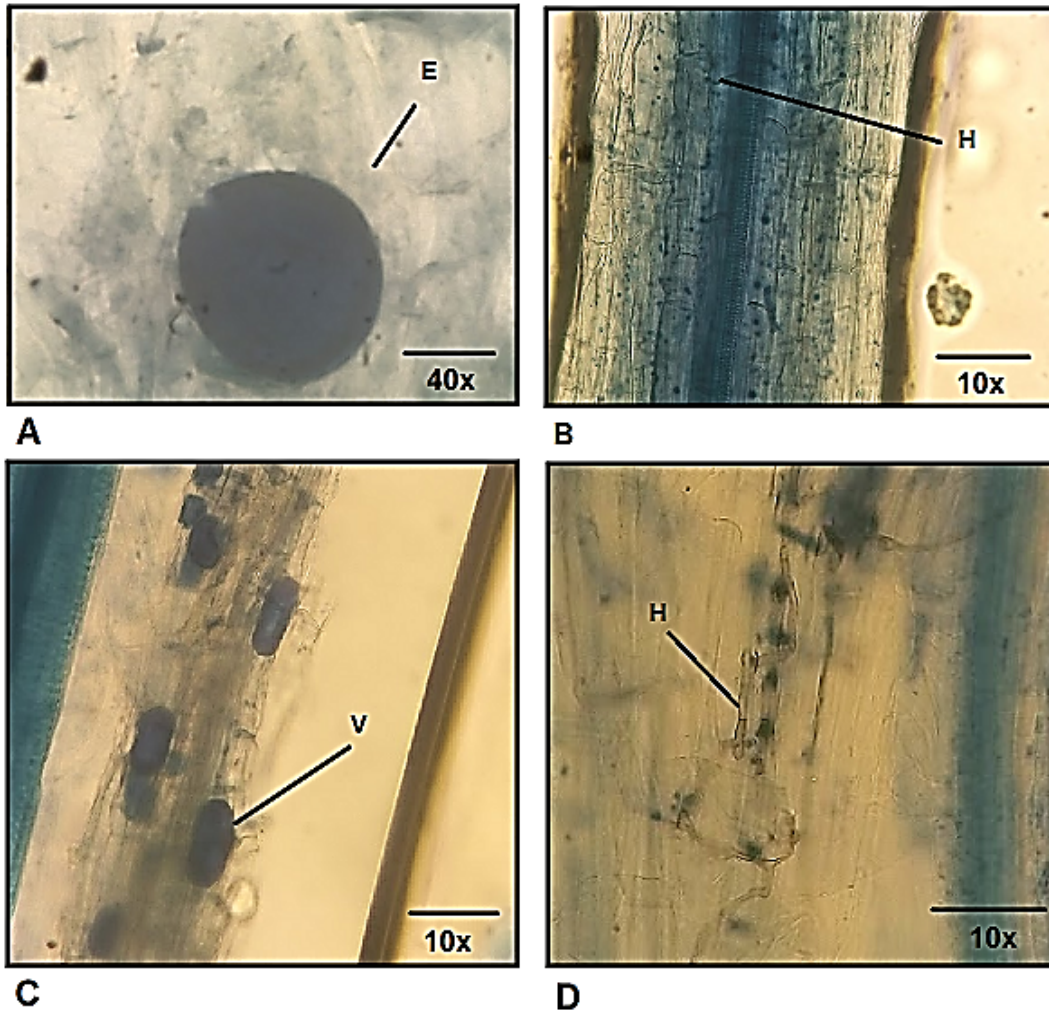
138 It was observed by [10] that the percentage of mycorrhizal colonization in *Atriplex*
139 *cordobensis* inoculated with *Funneliformis geosporum* in saline soils was 90%. In the
140 work of [11], the colonization of *A. nummularia* roots by *G. intraradices* was well
141 developed (77%), and only hyphas and internal vesicles were observed, the arbuscules
142 were not found.

143

144

145

146 **Figure 1. Structures of the *Claroideoglossum etunicatum* observed in the roots of**
147 ***Atriplex nummularia* in non-autoclaved soil**



148
149 (A) spore, (B) intracellular hyphas, (C) vesicles, (D) hyphas septated from another fungus.

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152 In the treatment with autoclaved and non-autoclaved soil, it was observed that, in the
153 presence of AMF and in the combination AMF x SHA (Nutrition Solution of Hoagland
154 & Arnon), mycorrhizal colonization increases according to the electrical conductivity,
155 except for the AMF x SHA treatment in autoclaved soil. Thus, in accordance with Table
156 1, in the presence of the fungus, in autoclaved soil the following the following order is
157 observed: AC (8.2%)>T1(8.5%)>T2(9.1%)>T3(9.3%), except for T4 (8.1%) and, in
158 non-autoclaved soil: AC>(8.7)>T1(8.4)>T3(9.4) and T4(9.3), except for T2 (7.3%).

159 In the combination, for the non-autoclaved soil: AC (8.2%)>T1 (8.8%)>T2 (28.1%)>T3
160 (9.3%)>T4 (9.5%). And, finally, the autoclaved soil: AC (9.4%), T1 (9.1%), T2 (9.0%),
161 T3 (8.1%) and T4 (8.1%).

162 In the work of [12] the colonization with *Glomus etunicatum* (*Claroideoglomus*
163 *etunicatum*), in sterile soil, was low, not exceeding 7% and in the non-disinfested soil
164 reached 55% of colonization. These values were not observed in this study, because the
165 percentage of colonization was higher in the treatments in non-autoclaved soil.

166 [13], observed that the percentage of colonization in *Aeluropus littoralis* with
167 *Claroideoglomus etunicatum* in the treatments with NaCl addition was reduced, showing
168 saline irrigation water in three treatments: the control and addition of 50mM and
169 200mM of NaCl, which resulted in colonization percentage of 33%, 16% and 10%
170 respectively.

171 According to [14], for the species *Tamarox ramosissima*, tree species highly resistant to
172 salt and drought, the colonization of AMF increased with EC of soil at low and medium
173 salinity levels (0.4 to 4.3 dS m⁻¹) but decreased at high salinity levels (> 7 dS m⁻¹).

174 Soil pH ranged from 7.6 to 7.9. In relation to the pH presented by the autoclaved and
175 non-autoclaved soil, [15] state that the pH near neutrality favors the *Glomus* species.

176 The correlation between height and colonization of *Atriplex* was significant at p <0.05
177 level for the treatments with *Claroideoglomus etunicatum*. In non-autoclaved soil, with
178 T2 + Fungus (R = -0.91) and T4 + Fungus (R = 0.90). For the autoclaved soil the
179 correlation was as follows: in the treatment AC + Fungus (R = 1,00) and in T1 + Fungus
180 (R = -0.97).

181 [16] observed that *A. nummularia* at low salinity level 2.2 dS.m⁻¹ had a colonization
182 percentage of 2%; even though, despite the low colonization, the inoculation promoted
183 the growth of the plants and affected the nutrient absorption positively.

184

185 **4. CONCLUSION**

186

187 In view of the results obtained it is concluded that: the mycorrhizal association was
188 beneficial for all treatments; the best treatment involving the percentage of mycorrhizal
189 colonization consisted of T3 (reject plus 14g NaCl - EC = 13.13 mS / cm) in the
190 presence of AMF *Claroideoglomus etunicatum* in autoclaved soil; for the height of the
191 *Atriplex nummularia*, it was observed that the treatment T2 (reject plus 7g NaCl - EC =
192 12.4 mS / cm), in the presence of fungus, in the non-autoclaved soil, presented better
193 result; when positively correlated, the height with the percentage of colonization, the
194 most significant treatments were T4 (reject plus 21g NaCl - EC = 14.16 mS / cm) in the
195 presence of *Claroideoglomus etunicatum* in non-autoclaved soil and AC (water - EC =

196 2.86 mS / cm) in the presence of fungus in autoclaved soil; in non-autoclaved soil the
197 presence of microorganisms may have promoted a combination of beneficial
198 associations that favored the growth of *Atriplex*; in autoclaved soil, there is a possibility
199 of having suffered with the abiotic stress.

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202 **COMPETING INTERESTS**

203 Authors have declared that no competing interests exist.

204

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