

ISOLATION AND IDENTIFICATION OF MICROORGANISMS ASSOCIATED WITH BIOREMEDIATION OF OIL SPILLED SITE IN BODO RIVERS STATE, NIGERIA.

ABSTRACT

The samples of spilled water collected from Bodo West in Gokana Local Government of Rivers State were isolated to identify microorganisms associated with bioremediation and the isolate was used for biodegradation of hydrocarbons. The microorganisms isolated may have the potential of biodegradation of hydrocarbon which may show evidences of bioremediation. Carbon source from petrol, diesel and kerosene were used. The evidence of degradation was based on the increase in the number of cells from day 1 to day 4. The colony forming unit was used to count the increase in the number of cells. The results of the analysis show that notable number of microorganism of which seven bacteria and seven fungi were isolated and identified. The bacteria are *Micrococcus Luteus*, *Streptococcus Lactic*, *Streptococcus Epidemidis*, *Streptococcus Faecalis*, *Clostridium Sprogenes*, *Aerococcus Viridems*, and *Bacillus Anthracis*. The fungi are *Articulosspara inflata*, *Dendospora Erecta*, *Aspergillus Niger*, *Liodioderium Species*, *Geotichrum Albdum*, *Aspergillus Funigatus* and *Sreptothric Atrax*. On the strength of the result, it is inferred that microorganisms are associated with bioremediation and can be used for environmental and petroleum cleanup exercise in an oil spilled site.

Keywords: microorganisms, biodegradation, bioremediation, hydrocarbons, oilspilled, isolation, identification.

1. INTRODUCTION

Petroleum exploration is a lucrative business especially in Nigeria [7,12,14,15,34]. Nigeria since the discovery of oil has survived on the proceeds from oil production, as capital projects and paying of workers' salaries is dependent on income generated from the oil business [7,12,14,15,36]. Though, there had been calls from different quarters for diversification of the economy from the solo means of petroleum exploration into other sectors like agriculture, commerce and manufacturing [14]; however the current gains for petroleum resources has overshadowed government interest in other areas of the economy [13,32].

Petroleum exploration involves a complex process; from drilling, refining to the distribution of the products to the different marketers and end users [13,36]. The processes has its own associated environmental problems as oil spills on a large scale on the land, sea or river and massive air pollution has been reported [1,8,9,11,12,16,17,18,21,29,30]. Government had in the past carried out environmental programmes to educate the people on the consequences of pollution [22,32,35]; but the people has always these rejected government programmes due to their non-participation in the decision making process [31]. Such agitations by the people in the local communities have always resulted in to violent conflicts [10,19,23,24,25].

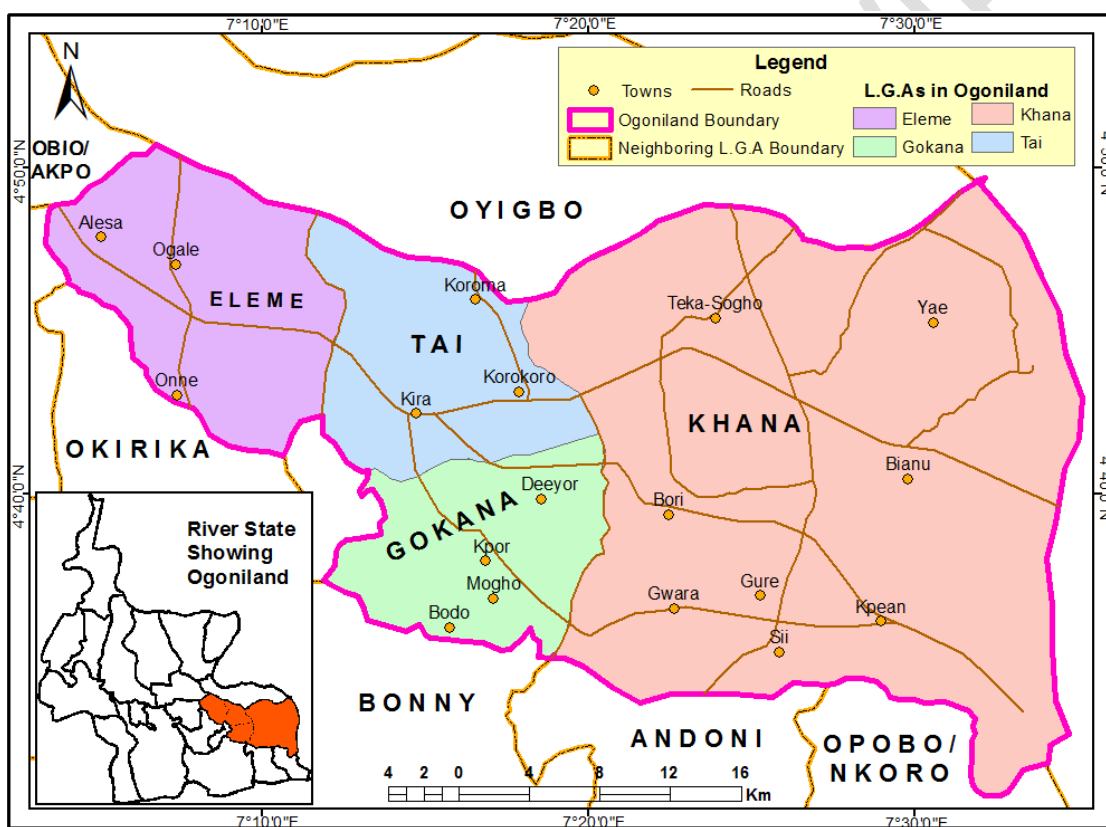
Hydrocarbon contamination of the environment has not only destroyed the ecosystem, but has also resulted in several health challenges and deaths [33]. Thus, there had been calls for remediation of polluted land in the Niger Delta [31]. Mechanical and chemical methods are generally used to remove hydrocarbons from contaminated sites [26-28]. These methods have limited effectiveness and can be expensive; so bioremediation is a promising technology for the treatment of these contaminated sites,

44 since it is cost effective and will lead to complete mineralisation [26-28]. The process of
45 bioremediation is simply the used of microorganisms to remove pollutants from the polluted
46 environment through the establishment and maintenance of a condition that favours oil biodegradation
47 rates in the contaminated environment [26-28,2-5].

48 Bioremediation become a process of interest in the petroleum industry due to the success in the
49 cleanup of the oil tanker Exxon Valdez of oil spill of 1989 [6,26-28). Bioremediation is an attractive
50 technology that has gained popularity in the global conservation and sustainability strategies [26-28].

51 2. THE STUDY AREA

52 Bodo West is a village settlement in Gokana Local Government Area in Ogoni. Ogoni which is a
53 superset of Bodo West lies between latitude $4^{\circ}05^1$ and $4^{\circ}20^1$ North and longitude between $7^{\circ}10^1$ and
54 $7^{\circ}30^1$ East [36] as shown on figure 1. The site is in Rivers Basin. It is accessible by roads and footpath
55 and some parts that are covered by thick vegetation were inaccessible.



56
57 Map of Ogoni indicating the position of Bodo West [36]

59 3. MATERIALS AND METHODS

60 Source of Samples

61 Samples were collected from contaminated oil spilled sites at Bodo West, Gokana Local Government
62 Area of Rivers State and analysed in Jossy laboratory, Port Harcourt.

63 Sterilization Techniques

64 Throughout the case of this project, a sterile condition was maintained. The material used was
65 sterilized in different form according to the nature of the materials.

66 **Method of Sample Collection**

67 Samples were collected using aluminium foil or glass wares. PVC materials, plastics or polythene
68 materials were avoided because they contain hydrocarbon. This was done to ensure quality control.

69 **Culture Media**

70 The sterilization of the media was carried out by heating under pressure autoclave. This was carried
71 out at 121°C for 15 minutes.

72 **Inoculating Chambers**

73 Since air always contained dust particles that generally have a population of micro organism, it was
74 carried out in inoculating chamber. The chamber was sterilizer with ethanol by swabbing.

75 **Inoculatin Loops**

76 Aseptic transfer of culture from one tube of another was accomplished with an inoculating loop. This
77 was sterilized by incinerating the loop in a flame while forceps was sterilized by dipping it in alcohol
78 and flaming.

79 **Glass Ware**

80 This glass ware was washed, rinsed with tap water and finally with distilled water. The glass ware was
81 sterilized first in autoclave at 121°C for 15 minutes to get rid of all the microorganisms before normal
82 washing. After washing, they were heated at a mode rate temperature with the use of hot air oven. The
83 sterilization period was 160°C for 1 hour. In the process precaution was taken to pack them so tightly
84 to impede the circulation of trapped air.

85 **Bench Tops**

86 Bench tops and cabinet was sterilized using 40% formalin absolute alcohol dilated to a concentration
87 of 70%.

88 **Incubator**

89 The incubator was used to create an enabling environment for microorganism to grow on any material.
90 The pleasant and conditioning of an already inoculated culture medium is the specific environment
91 that favours the growth of microorganisms in the laboratory. This was carried out by using incubator
92 with the door and the entire air closed for 48 hours.

93 **Isolation of Organism**

94 Millilitre of sample of water liquor was withdrawn and transferred into test tubes containing sterile
95 distilled water in preparation for sterile distilled water in preparation for serial dilution to 10^{-8} dilution
96 which 10^{-1} , 10^{-4} , 10^{-8} and stock was used. It was carried out with the aid of sterile syringes. Nutrient
97 Agar (2.8g was dispersed into conical flask containing 100m/s distilled water and autoclaved at 121°C
98 at 15 minutes.

99 About 0.5mls of selected four dilution factors source was dispensed into sterile Petri dishes. Potato
100 dextrose Agar (3.6g) into 100mls distilled water in a conical flask was used for fungi by pour plate
101 method, the Petric dishes already inoculated with diluted isolate and incubated at 37°C for 48 hours for

102 bacteria and normal room temperature for fungi for 48-72 hours. After incubation, growth was
103 observed and discrete colonies developed on the surface of the plate. It was sub-cultured to get the pure
104 culture isolated. Sample was appropriately labelled.

105 **Pour Plate**

106 This involves the preparation of the dilution of inoculums. The inoculums dilution is added to sterile
107 petric dish mixed with molten but cold media and allowed to solidify after incubation; some colonies
108 develop embedded and in addition to others on the surface of the plate but the plate were incubated in
109 closed container to avoid oxygen.

110 **Streak Plate**

111 The streak plate method appeared to be the most widely and routinely used method of plating in
112 laboratory. It involves the sterilization of wire loop by passing it over blue flame until it turns reddish.

113 The sterile, loop was used to pick a loopful of the organism and immediately used to make series of
114 parallel, non overlapping streak on the surface of the already solidified medium. The final streak will not
115 allow touching the source of the inoculums on the plates. The initial streak result to confluent growth
116 well isolate discrete colonies developed along the streaks on the plate. The discrete colonies on the
117 plate are clearly pure culture and further sub-culture was made from them for microbiological
118 investigation.

119 **Total Viable Counts**

120 This test was performed by pour plate techniques using nutrient Agar and fuel Agar (i.e. Kerosene,
121 diesel, and petrol).

122 The medium was prepared and sterilized; the sterile medium was allowed to cool. Duplicate of pour
123 plate of each sample were prepared by pipeting aseptically 0.1mls of sample into each sterile Petri dish
124 used for each sample and was discarded to avoid cross contamination.

125 The sterile and cool medium were poured aseptically and used to overlap the sample in the Petric dish.
126 The plate was rocked gently in single circular motion to allow proper mixing of medium and sample.
127 It was allowed to solidify and incubated in an upside position at 28C-37°C for 24 hour in a gallon
128 Kamp incubator. The plates were examine for microbial growth after 24 hour and colonies were
129 counted and recorded as colony forming units per millilitre.

130 **Gram Staining**

131 This test was performed by pour plate techniques using nutrient Agar and fuel Agar i.e (Kerosene,
132 Diesel, and petrol).

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135 each sterile Petric dish used from each sample and was discarded to avoid cross contamination. The
136 sterile and cooled mediums were poured aseptically and use to overlap the sample in the Petric dish.
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139 Kamp incubator. The plates were examined for microbial growth after 24 hour and colonies were
140 counted and recorded as colouring forming units per millilitre.

141 **Gram Staining**

142 The gram-staining was carried out as follows:- A loopful of water was dropped on a slide by sterile
143 inoculating loop. Inoculums from the young culture mixed with water in the film over the slide. The
144 smear was allowed to dry heated and fixed gently by passing it over the bursen flame. The smear on
145 the slide flooded with crystal violet solution of for 60 second, and rinsed with water. All the cells
146 turned purple, the smear will again flooded with fugal iodine for 1 minute and rinse 95% alcohol was
147 poured on the slide until the crystal violet was completely washed off. It will then counter strained
148 with safaranin and wash with water. At this point gram positive cells remained purple while gram
149 negative remain pink. This was viewed by means of oil immersion objective of the light microscope.

150 **Staining of Fungi**

151 Heat fixed smear was obtained with cotton blue lacto phenol for 1 minute, washed with distilled water
152 and dried. The slide was viewed under the microscope using objective x 100.

153 **Biological Test**

154 **a. Catalase Test**

155 Most of the aerobic microorganisms are capable of producing the enzymes catalane. A loopful of 48
156 hour culture of each isolate was placed on clean slide of which a drop of hydrogen peroxide had been
157 added. A formation of oxygen bubbles of effervescent caused by liberation of oxygen gas indicate the
158 presence of catalane.

159 **b. Sugar Fermentation**

160 Sugar fermentation was used to test for the ability of the organism to different sugar by breaking it
161 down to alcohol. Sugar such as sucrose, maltose, lactose, arabinos and mentol was used. 0.5% carbon
162 source was taken and emptied into McCartney bottle, containing peptone water and phenol red as an
163 indicator. The bottle was autoclaved for 121°C for 10 minutes. The autoclaving with tested organisms
164 was incubated with the inoculums. It was later used to compare the colour changes in tubes. Changing
165 in colour turned to yellow which indicates complete fermentation.

166 **c. Coaculate Test**

167 A small quantity of 24 hour old culture was taken with sterile inoculating loop; it then mixed with 1ms
168 of 10% V/V dilution of plasma on clean slide. Clothing of the mixture indicate positive test.

169 **d. Motility Test**

170 A 24 hour broth culture was taken with sterile inculpating loop into a cover slide, hanging drop
171 technique was used. It was viewed with the help of oil emission of X100 objective of light microscope.
172 Movement indicates motility positive while non-movement indicates motility negative.

173

174 **Biodegradation Test**

175 Kerosene agar was used after serial dilution to know the total number of organism that utilizes such
 176 organics solvent (i.e Kerosene), petrol and diesel. This was known through the increase in the number
 177 of cell per day.

178 4. RESULT

179 Organism isolated and identified were seven fungi and seven bacteria. The bacteria isolate are
 180 clostridium sparogermis. Aerococcus viridians, streptococcus Lactic, Micrococcus Lutes,
 181 Staphylococcus Lactic, Staphylococcus Epidermis, Streptococcus epidermis, Streptococcus Faecalis,
 182 Bacillus anthracis. The seven fungi isolated and identified are: *Articulospara Infalta*, *Dendospora*
 183 *erecta*, *Aspergillums Niger*, *Loidioderium Species*, *Geotichrum Albidum*, *Aspergillums funigatus* and
 184 *Streptothrix Atrax*. The result is shown on the table below.

185 **Table 1:** Biochemical/Morphological and microscopic identification of bacterial isolate.

Catalase	-	-	-	-	-	-	-	Z+
Motility	-	-	-	-	-	-	-	-
Hydrolysis	+	+	+	+	+	+	+	+
Glucose	A	A	A	A	A	A	A	A
Lactose	+	+	+	+	+	+	+	+
Sucrose	+	+	+	+	+	+	+	+
maltose	+	+	+	+	+	+	+	+
Arabinos	+	+	+	+	+	+	+	+
Coagulase	-	-	-	-	-	-	-	-
Shape	Circular	Sphere	sphere	Sphere	Round	Round	Dombel	Round
Edge	Dented	Enteric	Dented	Dented	Dented	Dented	Dented	Enteric
Elevation	Raised	Raised	Raised	Raised	Raised	Raised	Raised	Raised
Surface colour	Smooth	Smooth	Smooth	Smooth	Smooth	Rough	Smooth	Smooth
Pigmentation	Creamy	Creamy	Creamy	Creamy	Creamy	Pinkish	Pinkish	Creamy
G-stain	+ve	-ve	+ve	+ve	-ve	+ve	-ve	-ve
Probable organism	1 clostridium sprogenes	2 Aerococcs Viridams	3 Streptococcs lactic epidemis	4 Micrococcs luteus	5 Staphylococcus luteus	6 Streptococcs lactic faecalis	7 Streptococcs lactic faecalis	

186 KEY

187 + = Positive
 188 - = Negative
 189 A = Acid Production

190

191 **TABLE 2: Culture and Microscope Characterization of Fungi Isolate**

Cultural characteristics	Microscopic	Identification
White mycelia growth on PDA after 24 hours.	Cordidiophore hyaline slender upper part sparingly branch conidia.	Articlospara inflate
Submerge aquatic with branched septate mycelium, simple cordidiophore slender hyaline.	Whitish cotton like mycelia which turns red on PDA plate.	Dendropsora erecta
Black mycelia on culture media after 48 hours.	Chain of conidial bonne on phial ides with black glucose head supported by septet were observed.	Aspergillus Niger
Whitish mycelia which later turns grey on APDA plate.	Mycelia external conidiophores upright simple upper portion which increase in length as conidia formed.	Oidioderium species
White septate mycelia on PDA plate	Conidia arthrospore hyaline J. celled shut cylindrical with truncate end.	Geotrichum albidum
Gray mycelia on PDA Plate which were dusty.	Conidiophores upright simple terminating in a globule or elevate swelling bearing phralites at apex.	Aspergillus fumigates
Dark mycelia on PDA plate.	Loosely tall mycelia tall conidiophores branch spirally coiled.	Streptothric Atra

192

193 **TABLE 3: TOTAL COLONY COUNTS IN (KEROSENE FUEL, DIESEL) AGA R MEDIA.**

194 **Day 1**

	DIESEL (X10⁻⁴)	KEROSENE (X10⁻⁴)	PETROL (X10⁻⁴)
Clostridium Sprogenes	2	2	2
Aerococcus Viridams	2	3	4
Streptococcus lactic	-	1	2
Micrococcus luteus	1	3	2
Streptococcus epidemidis	-	2	4
Streptococcus faecalis	2	1	3
Bacillus anthracis	-	1	4

195

196 **Day 2**

	DIESEL (X10⁻⁴)	KEROSENE (X10⁻⁴)	PETROL (X10⁻⁴)
Clostridium Sprogenes	3	2	3
Aerococcus Viridams	5	1	5
Streptococcus lactic	2	2	4
Micrococcus luteus	4	2	3
Streptococcus epidemidis	6	3	5
Streptococcus faecalis	5	3	7
Bacillus anthracis	7	2	6

197

198 **Day 3**

	DIESEL (X10⁻⁴)	KEROSENE (X10⁻⁴)	PETROL (X10⁻⁴)
Clostridium Sprogenes	7	5	11
Aerococcus Viridams	9	10	16
Streptococcus lactic	4	8	14
Micrococcus luteus	6	4	11
Streptococcus epidemidis	8	9	4
Streptococcus faecalis	9	6	8
Bacillus anthracis	15	10	12

199

200 **Day 4**

	DIESEL (X10⁻⁴)	KEROSENE (X10⁻⁴)	PETROL (X10⁻⁴)
Clostridium Sprogenes	9	8	20
Aerococcus Viridams	15	17	30
Streptococcus lactic	8	7	22
Micrococcus luteus	11	14	7
Streptococcus epidemidis	15	9	16
Streptococcus faecalis	10	13	22
Bacillus anthracis	70	11	17

201

202 **5. DISCUSSION AND CONCLUSION**

203 The results obtained from this study revealed that arrays of microorganisms are associated with body
 204 of water. A total of seven fungi and seven bacteria were isolated as shown on table 1 and 2. Most of
 205 these colonies isolated are predominately microorganism that resides in water. The moisturization of
 206 the water body and its creaks create a conducive environment for the growth of microorganism and
 207 relationship exist between the enzymatic activity of fungi and amount of water in colonize substrate.
 208 The maximum relative humidity permitting the growth of fungi varies from 75% - 95% for different
 209 species of microorganisms.

210 The presence of microorganism in water which act to degrade the hydrocarbon components of
 211 contaminant, get the resident flora of certain contaminated water and play an important role in
 212 maintaining and bringing back the contaminated water to its uncontaminated state.

213 This study focused on bioremediation and treatment of high volume hydrocarbon wastes. Evidence
214 also provided that in land farming practices as in conventional bioremediation systems, a large fraction
215 of the volatile hydrocarbons is not biodegraded but is rather transferred to the atmosphere through
216 volatilization. An increasing focus on regulation and control of volatile organic carbon emissions calls
217 for hydrocarbon remediation and waste treatment system which contain or destroy the volatile organic
218 fraction.

219 The interest for exploiting petroleum degrading organism for environmental cleanup has become
220 central to petroleum geoscientist as this study has clearly shown that bacteria and fungi can be use to
221 degrade or detoxify pollutants or contaminant in a given environment.

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