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2 **AGE AND SEX RELATED PREVALENCE AND DISTRIBUTION OF**
3 **HOOKWORM INFECTION AMONG PUPILS OF UNIVERSITY OF CALABAR**
4 **STAFF SCHOOL, CALABAR, NIGERIA**

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6

7 **ABSTRACT**

8 For decades, hookworm infections have been known to be a major problem of public
9 health, affecting mostly children and causing significant impact on their health. The aim
10 of this study was to investigate the prevalence rates between age and sex of school
11 children affected using University of Calabar Staff School, Calabar, Nigeria. Stool
12 samples were collected from the pupils and examined using direct smear, formal-ether
13 concentration and brine floatation techniques. The frequency of hookworm infection was
14 more in males, 11(11.3%) than in females, 7(6.8%) consisting the 18 positive children
15 sampled from the 200 pupils and the highest prevalence was found among the 13 – 15
16 years age group (33.3%). The statistical analysis showed no significant difference for
17 both age and sex ($p>0.05$). Hookworm infections have decreased in recent years due to
18 the increasing development of the country and other personal hygiene observed thus,
19 continuous deworming program should be conducted in primary schools to lower the rate
20 of the infection.

21

22 **KEYWORDS:** Public health, prevalence, hookworm, smear and infection

23 **INTRODUCTION**

24 Hookworm infection caused by *Ancylostoma duodenale* and *Neatorsmmericanus*
25 affects about 740 million people worldwide with 80 million people severely infected.
26 Developing countries are the most affected and within these, the major cases occur
27 among school aged children (Bethony *et al.*, 2006). The distribution of hookworm
28 infection depends on many factors like socio-demographic variables associated with
29 poverty such as reduced access to adequate sanitation, potable water, and healthcare, as

30 well as the prevailing climatic and environmental conditions (Monstessor *et al.*, 1998).

31 The economic burden caused by hookworm infection is high. Recently, it has been
32 estimated to cost 1.8 million Disability-Adjusted-Life-Years (DALYs). Young children
33 are reported to be disproportionately affected by hookworm infection compared to adults
34 due to increased nutritional requirements and less developed immune system. Hookworm
35 infection in this age group has been linked with significant reduced growth and increased
36 risk for protein-energy malnutrition (Stephenson *et al.*, 2000) including growth stunting,
37 iron-deficiency, anemia intellectual retardation, cognitive and educational deficit.

38 The precise impact of hookworm infection on child nutrition, growth and
39 development appears to depend on the species, burden and its impact on host nutrition
40 tends to be long-termed. Despite recent advances, a number of important questions still
41 remain unanswered regarding hookworm infection risk and its consequences in child
42 population. For example, it is still not clear whether certain age, gender and even ethnic
43 groups are most likely to become infected.

44

45 **Life Cycle and Classification**

46 Humans are infected with hookworm's third stage filarial-form larvae. The larva in
47 soil penetrates through the skin particularly into area such as unprotected feet. Once
48 infected, the filarial-form larva migrates into blood circulation. They break out of the
49 pulmonary blood vessels into alveoli, then crawl up the trachea and are swallowed with
50 saliva to re-enter the intestinal tract. They attach themselves to the mucous membrane of
51 the small intestine to mature into adults. The female adult releases eggs (*N. americanus*
52 about 9000 – 10,000 eggs per day and *A. duodenale* 25000 – 34000 eggs per day) which

53 are passed in the faeces of the human host. These eggs hatch in the environment within
54 several days and cycle starts anew (Hawdow and Hotez, 1996).

55

56 **Epidemiology**

57 No international surveillance mechanisms are in place to determine the prevalence
58 and global distribution of hookworm infection. However, based on extensive research,
59 hookworm is estimated to be in 740 million people (de Silva *et al.*, 2003), highest
60 prevalence occurring in sub-Saharan Africa and Eastern Asia. Other area of rural poverty
61 in the tropics including Southern China (Hotez, 2003). Indian subcontinent (Yadla *et al.*,
62 2003) and America (HJotez, 2003) also have high transmission rates. In all regions, there
63 is striking relationship between hookworm prevalence and low socioeconomic status.

64 Compared to other soil transmitted helminthes (STH) infections and schistosomiasis,
65 hookworm infection exhibits a unique age-intensity profile. Whereas, the intensity of the
66 former peaks in childhood and adolescence, hookworm intensity usually either steadily
67 rises in intensity with age or plateau in adulthood. (Bethony *et al.*, 2002). According to
68 Hotez (2005), sub-Sahara and East Asia have about 198 and 149 million infected people
69 respectively. Others are South Asia – 59 million, Latin America and Caribbean – 50
70 million, North Africa and Middle East – 10 million, India – 71 million, China – 39
71 million infected people. Other studies in Nigeria shows that in Niger-Delta 34.9% of
72 4990 people were infected (Agi and Awi-Waadu, 2008) and in Vom, Plateau State, 3.2%
73 of total 463 samples were positive (Odebunmi *et al.*, 2007).

74 Recent technological development are now used by researchers like Geographical
75 Information System (GIS) and Remote Sensing (RS) to examine helminth ecology and

76 epidemiology, other focus on the development of DNA-based tools that can be used for
77 diagnosis of infection, specific identification and analysis of genetic variability in
78 hookworm populations (Brooker *et al.*, 2007, Gassen *et al.*, 2009).

79

80 **Pathogenesis and Clinical Presentations**

81 Hookworm larva get to the host either orally or by penetrating the skin to reach the
82 small intestine. Hookworm's haematophagous habits cause pathogenesis of anemia and
83 malnutrition. The worms attach using their mouth parts, each female worm is estimated
84 to ingest a minimal of 0.1ml of blood per day. However, actual blood loss can be
85 significantly greater, the worms change, their feeding sites several times a day, and the
86 secretion of anticoagulants or proteins means that the vacated sites continues to bleed,
87 contributing greatly to blood loss (Bungiro and Capello, 2004, Devaney, 2005)

88 **METHODS**

89 **Study Location**

90 The study was carried out at the University of Calabar Staff School, Calabar, Cross River
91 State, Nigeria, between the periods of October – December 2013. The age range of the
92 pupils sampled was from 2 – 13 years.

93 **Study Population**

94 Stool samples were collected from 200 pupils aged 2-13 years. The faecal samples were
95 collected in October – December, 2013. School was selected by convenience sampling.
96 To augment the sample size due to a lower than expected number of children, a younger
97 category was added.

98

99 **Ethical Approval**

100 Informed consent was taken from parents and caretakers and the school authorities after
101 explanation about the objectives and aims of the study.

102 **Sample Collection**

103 Pupils were provided with wide mouthed specimen bottles with screw caps and spoons
104 with specific instructions to collect samples in the morning. The sample bottles were
105 properly labeled with date, number, age and sex of the pupil. Total of 200 samples were
106 collected, from 97 males and 103 females. They were given instructions to avoid
107 contamination of samples. The samples were transported to the Microbiology laboratory,
108 University of Calabar for analysis.

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110 **Questionnaire**

111 Simple structured closed –ended questionnaires were given to pupils to gain information
112 from the parents or guardians of subjects regarding child and household socio-
113 demographic, housing, water and sanitation characteristics. These items included subject
114 age, sex, ethnicity, family size, house construction, water, sanitation and garbage
115 disposal, characteristics and the presence versus absence, type and density of domestic
116 animals living in and around the home, shoe wearing habit, personal hygiene practices
117 and related symptoms like headache, nausea, abdominal pains etc.

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122 **MACROSCOPY AND MICROSCOPY**

123 **Macroscopy**

124 Before carrying out microscopic examination, macro-scopy was carried out to check the
125 consistency of the stool, presence of blood or mucus, presence of proglottids, colour and
126 smell of the stool, then recorded against the appropriate sample number.

127 **Microscopy**

128 Wet mounts (saline and iodine)

129 **Principle:** - It is a simple preliminary microscopic method that detects motility in
130 organisms, eggs, cysts, larvae, trophozoites of the organism and gives clear images of
131 fresh specimen under the microscope.

132 **Procedure:** - Place a drop of saline on left half of the slide and one drop of iodine on the
133 right half. With an applicator stick, pick up a small portion of the specimen and mix with
134 saline drop, do same with iodine drop. Put the cover slip separately on both and examine
135 under the microscope using 10X objectives to focus, the 40X objective to get a clearer
136 view.

137 **Results:** - Ova, cysts and trophozoites of organisms were found and identified mostly in
138 the saline wet mount. Iodine wet mount revealed mostly amoebic and flagella cysts.

139 **Concentration techniques**

140 **Principle:** - This technique is used mostly when the number of parasites in the stool
141 specimen is low. Eggs, cysts and larvae are covered but trophozoites get destroyed during
142 this procedure.

143 Concentration procedures can be grouped under 2 categories:-

144 i. sedimentation procedure

145 ii. Flotation procedure

146 **Formal-ether sedimentation technique**

147 **Procedure:** - Transfer half teaspoon of faeces in 10ml of water in a glass container and
148 mix thoroughly. Place two layers of gauze in a funnel and strain the contents into a 15ml
149 centrifuge tube. Centrifuge for two minutes at about 500g. Discard the supernatant and
150 re-suspend the sediment in 10ml of physiological saline. Centrifuge at 500g and discard
151 supernatant. Re-suspend the sediment in 7ml of 10% formaldehyde (one part of 40%
152 formalin in three parts of saline) and 3ml of ether (or ethyl acetate). Close the tube with a
153 stopper and shake vigorously or mix. Remove the stopper and centrifuge, rest the tube in
154 a stand, four layers become visible. Pour off the liquid leaving a small amount of
155 formalin for suspension of the sediment. With a pipette, remove the sediment and mix it
156 with a drop of iodine, then examine under the microscope. (Cheessbrough, 2002).

157 **Result:** - Ova or cysts can be detected, since this method is very sensitive. Size and shape
158 of the parasitic structure is maintained.

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161 **Brine flotation technique**

162 **Procedure:-** Place about one milliliter of faeces in a flat bottomed container with
163 capacity of 15-20ml, add few drops of brine and stir it to make a fine emulsion, then
164 more brine so that the container is nearly full, stirring the solution thoroughly. Remove
165 floating coarse materials, fill the container with a dropper till a meniscus is formed. Place
166 a glass slide carefully on top, allow for 20 minutes, lift slide quickly and overturn
167 smoothly, after putting a coverslip, examine under the microscope. (Koneman, 1997).

168 **Result:** - Ova or cysts can be detected, but unfertilized eggs of *A. lumbricoides* cannot be
169 seen because they do not float. If suspension is left for more than 20 minutes, protozoan
170 cysts and thin walled nematodes eggs will collapse and become distorted in appearance.

171 **Statistical Analysis**

172 Frequency distribution tables, percentage prevalence and intensity of infection are
173 estimated using standard formulae. Simple percentage was used to compare differences in
174 prevalence between age groups and sex, Fisher's Exact (SISA) was used to find odd
175 ratios (O.R), confidence interval (C.I) and probability (P)-values. Significant level was
176 set at confidence limit 95% and $p < 0.05$.

177

178 **RESULTS**

179 Two hundred (200) samples were from the pupils consisting of 97 males and 103
180 females. The mean age group fell under 7-9 years, the age range of the pupil was from 1-
181 13 years. Hookworm was recovered from a total of 18 samples revealing a prevalence
182 rate of 9%. The highest prevalence for hookworm infection was found in age group 13-15
183 years (33.3%) and lowest in <4 age group, which had no positive sample. Frequency
184 distribution was highest in males than in females (11.3% and 6.8% respectively).
185 Statistical analysis of the data showed no significant difference for both age and sex
186 ($p > 0.05$).

187

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Table 1

189

Prevalence of Hookworm infection in sample population (n=200)

Category	No. of subject	Percentage frequency (%)
Hookworm +	18	9
Hookworm -	182	91
Total	200	100

190

191 This table shows that 9% of the samples examined were hookworm positive, 18 samples
 192 out of the 200 samples had hookworm.

193

Table 2

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Distribution of hookworm infection by age of study subjects

Age (Years)	No. involved	No. +	% +	C.I	O.R	p-value
<4	2	-	-	-	-	-
4 – 6	42	5	11.90	0.62-3.04	1.50	0.17
7 – 9	103	9	8.70	0.60-1.57	0.94	0.19
10 – 12	50	3	6.00	0.22-1.87	0.57	0.17
13 – 15	3	1	33.30	0.48-53.07	5.29	0.23

195

196 This table reveals that the highest prevalence is found among age group 13 – 15 years
 197 (33.3%), followed by 4 – 6 years (11.9%). This does not show a significant difference
 198 ($p>0.05$).

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Table 3

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Distribution of Hookworm Infection by Sex of Study Subjects

Sex	No. examination	Positive	% age	C.I	O.R	p-value
Male	97	11	11.3	0.87-1.93	1.75	0.11
Female	103	7	6.8			

203

204 This table shows distribution of hookworm infection by sex, males had highest
 205 prevalence (11.3%) compared to females (6.8%), but this shows no significant difference
 206 ($p > 0.05$).

207

Table 4

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Prevalence of hookworm infection according to demographic characteristics of study subjects

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Residential Area	No. (%)	No. + (%)
Unical Staff Quarters	30 (15.00)	2 (6.67)
Calabar South	97 (48.50)	11 (11.34)
Calabar Municipality	73 (36.50)	5 (6.85)

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211 **KEY:** Unical = University of Calabar

212

No. = Number

213

% = Percentage

214

+ = Positive

215 This table shows the prevalence of hookworm infection based on the residential area, it
216 reveals that those who stay in Calabar South had highest frequency of infection (11.34%)
217 and University of Calabar Staff Quarters had the lowest frequency (6.67%).

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Table 5

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Distribution of Hookworm Infection by Age and Sex

Age (years)	Sex	No. Examined	No. Infected	% Infected
>4	M	-	-	-
	F	2	-	-
4 -6	M	19	2	11.9
	F	23	3	
7 – 9	M	47	5	8.7
	F	56	4	
10 – 12	M	29	3	6
	F	21	-	
13 – 15	M	2	1	33.3
	F	1	-	

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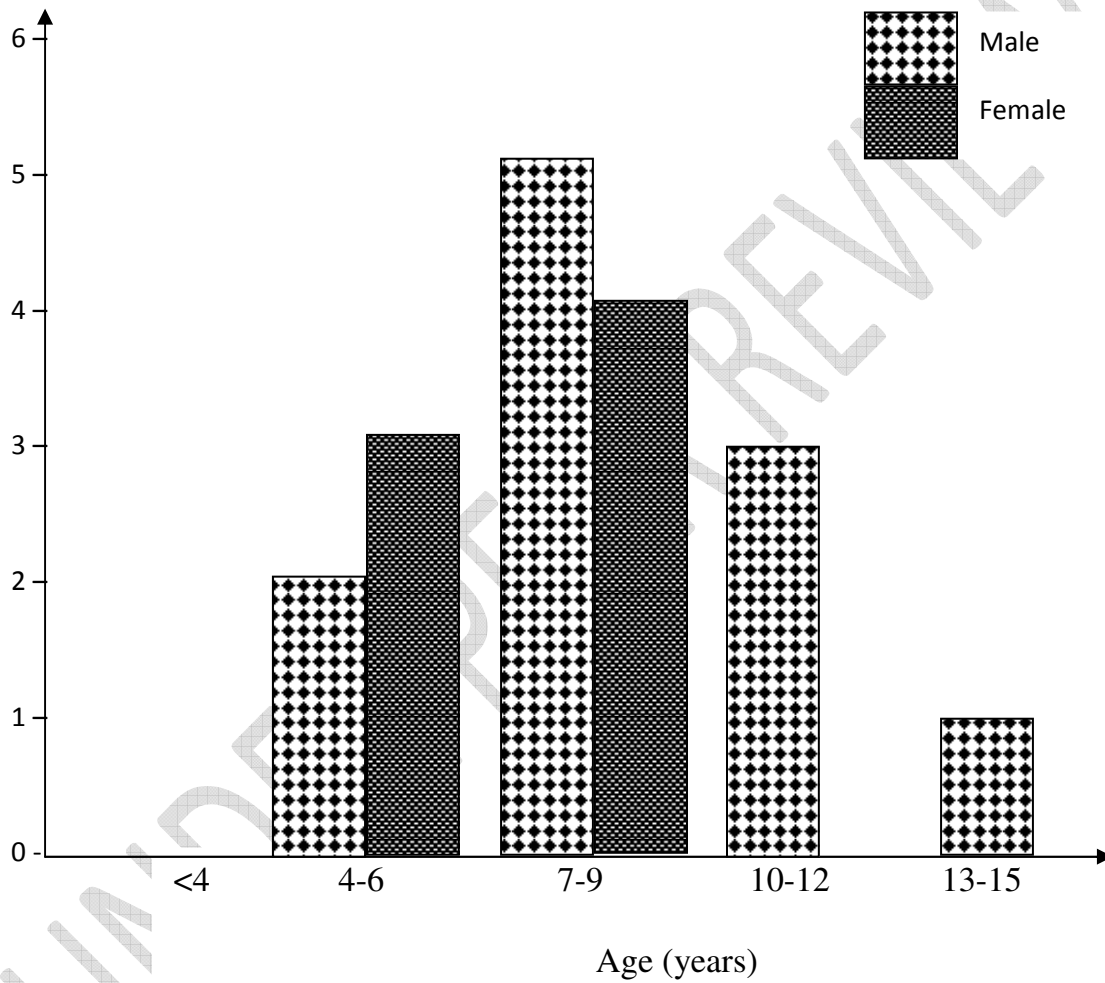


Figure 1: Distribution of hookworm infection by age and sex.

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