# Study the effect of watermelon white rind extract as decreased the detrimental of soybean oil in vivo.

4

\_5

## **ABSTRACT**

Dried watermelon white rind was subjected to mycotoxin determination to assure its safe usage. Results ensured that the rind was free from aflatoxin and ochratoxin. Watermelon white rind aqueous extract contained iron, copper and potassium at concentration range of 3.4, 0.53 and 45.51 ppm, respectively. While, the amounted of Chrome and Selenium were 14.2 and 98.51 µg/Kg, respectively. Soybean oil had free fatty acid, peroxide value, iodine number and anisidine value of 0.43%, 13.62 meq O<sub>2</sub>/Kg, 132 and 0.7, respectively. GC-MS analysis of soy oil ascertained the presence of twenty-four compounds: linoleic acid, methyl ester (25.27%), monensin (15.75%), elaidic acid (9.24%), nonadecanoic acid, methyl ester (7.04%), cis-13-eicosenoic acid (4.92%), cis-vaccenic acid (4.68%), linoleic acid (4.67%), palmitoleic acid (4.46%), 9-tetradecenal (4.42%) and cysteine (4.18%)were the most predominant. Fatty acid profile of the oil showed that the ratio of saturated fatty acid to unsaturated fatty acids was 1:5. Biological experiment was designed to evaluate safety usage and impact of soybean oil on different blood parameters of rats for two months' interval period. Rats fed diet prepared by soybean oil had a decreased calcium level in comparison with negative control (p<0.05). Supplementation with watermelon white rind aqueous extract rendered calcium level to normal status as negative control. Phosphorus level wasn't affected by soya oil.

24

*Keywords: watermelon; GC-MS; blood parameters; calcium level.* 

25

26

27

### 1. INTRODUCTION

Moatermelon (*Citrullus lanatus* var. *lanatus*, family Cucurbitaceae) is a flowering plant plant from southern Africa. The white rind is thrown as unused-agro waste. Rind constitutes 30% of the weight of whole watermelon fruit.

[331] cited that ethanolic and aqueous extracts of watermelon white rind possessed antibacterial activity against *E. coli and Salmonella sp.*, Gas Chromatography-Mass Spe35 metry analysis revealed the existence of methionine, L-Aspartic acid, Glycyl-Dasp36 gine, 9-Cis-Retinoic acid, Stearic acid allyl ester and Ascorbic acid permethyl that 37 ontributed to its antibacterial activity.

33e rind had total antioxidant activity of 297 mg AAE/100g, total phenols content of 13939 mg GAE/100g and total flavonoids of 40.4 mg QE/100g. FRAP assay indicated the 40gh reducing ability of the rind. Crude protein content amounted to 13.3%, crude fibe41(14.7%) and fat (2.11%). The rind is a source of iron (30.4 mg/kg), potassium (6.942%), copper (9.4 mg/kg), chromium (85μg/100g) and selenium (542μg/100g). Uns43urated fatty acid amounted to 81.2%. Vitamins A and E valued 383.44 μg/100g and 43.72 mg/100g, respectively [10]. Wastes are source of sugars, minerals, organic acid45 dietary fiber, and bioactive compounds [6].

Soy **b6** an oil affected negatively bone structure as reported by [5].

A7 study investigated the adverse effect of soybean oil in rat found that oil induced sign48 cant fatty liver [13].

40

## 2. MATERIAL AND METHODS

51

\$2\text{atermelon white rind was cut into small pieces, dried at 40°C and pulverized into fine 53\text{owder.}

# Preparation of white rind aqueous extract:

65 gram of dried powder was mixed with one liter of hot water, stirred, filtered and use 65 the sole source of fluid.

### Determination of Aflatoxin and Okratoxin

**58** tal Aflatoxin and Ochratoxin were determined according to AOAC [3].

# Elemental analysis of rind aqueous extract

bon, copper, potassium, chrome and selenium were determined according to AOAC [2].61

# Chemical analysis of soybean oil

Quality of oil was assessed by determining anisidine value, iodine number, peroxide value4 and free fatty acid according to AOAC [4]. Fatty acid composition was determined according to AOAC [4].

66C-MS analysis of soybean oil was carried out using GC (Agilent Technologies 7896A). The components were verified by matching their mass spectra and retention times with the database of National Institute of Standard and Technology (NIST) library.

# Biodogical experiment

Eighten rats were distributed into three groups:

Grozp (1) served as negative control and fed normal diet [9] and supplied with drinking wat 22

Group (2) served as positive control fed normal diet to which 150 ml soybean oil was add 74 per kilo and supplied with drinking water.

Gro**75** (3) fed diet as group (2) supplied with aqueous watermelon white rind extract. Die**76**nd fluids were supplied *ad-libtum* for all groups.

At the end of the experiment, blood samples were collected, centrifuged at 4000 rpm and Rerum was subjected to the analysis of calcium and phosphorus.

91

## 3. RESULTS AND DISCUSSION

93 lemental analysis of watermelon white rind aqueous extract (Table 1) ensured the pressance of iron (3.4 ppm), cupper (0.53 ppm), potassium (45.5 ppm), chrome (14.2 μg/(Σ)) and selenium (98.5 μg/(Kg)).

Data in Table (2) revealed that soybean oil had anisidine value of 0.7, iodine number 13197, free fatty acid 0.43% and peroxide value of 13.62 meq O<sub>2</sub>/Kg.

**98**venty-four compounds were detected in the GC-MS chromatogram of soybean oil. Lin**99**cic acid (25.27%) was the most predominant in the tested oil, followed by mon**160**sin (15.75%), elaidic acid (9.24%), nonadecanoic acid (7%), cis-vaccenic acid (4.6**201**), linoleic acid (4.67%), palmitoleic acid (4.46%), 9-tetradecenol (4.42%) and cyste**02**c (4.18%) and accounted for 59.34% of oil constituent (Table 3).

**A03**in Table (4), fatty acid profile of soybean oil showed the existence of linoleic acid (0.21%), oleic acid (22.85%), linoleic acid (6.2%) and gadolic acid (0.21%) as unsatos ated fatty acids accounting for 83.54% of total oil content. Saturated fatty acids contos palmitic acid (10.99%), stearic acid (4.82%), arachidic acid (0.36%) and behator acid (0.29%) representing 16.46% of soybean oil content. These results are in accountance with [7] who stated that soybean had low level of saturated fat and high conton of linoleic acid [1].

**∆10**shown in Table (5), a significant difference (p<0.05) existed between negative con**1101** (G1) and rats group fed diet with soybean oil (G2). A decrease in calcium level was 1112 served indicating that soybean oil affected calcium blood level.

Soybean had high phytate level [12]. Phytates can block the uptake of essential minerals as calcium, copper, iron, zinc and magnesium in intestinal tract that may contribute to mineral deficiencies [8].

**The**re was non-significant difference between negative control (G1) and Group 3 fed soy **ha**? noil and drunk rind extract, nor between G2 and G3.

Data 18 vealed that phosphorus blood level was not affected by any treatment and non-sign 16 ant differences existed between G1 and both groups G2 and G3.

The 120 traction of plant material and isolation of biologically active compounds are essential to understand their role in disease prevention and treatment.

122

123

## Table (1): Elemental analysis of watermelon rind aqueous extract

Element	Result
Copper (ppm)	0.5
Iron (ppm)	3.4
Potassium (ppm)	45.5
Chrome (µg/Kg)	14.2
Selenium (μg/Kg)	98.5

125

Table (2): Chemical evaluation of soy oil

Tested parameters	Result
Free fatty acid (%)	0.43
Peroxide number (meq O <sub>2</sub> /Kg)	13.62
Iodine number	131.8
Anisidine value	0.7

127

Table (3): GC-MS analysis of soy oil

RT	Compound name	Area sum (%)
3.88	Chicoric acid	0.29
5.7	Phytanic acid	0.59
6.187	3,2',4',5'-Tetramethoxyflavone	0.27
8.04	Gardenin	0.49
8.96	Isovitexin	0.59
11.7	Lutein	1.33
12.03	Stevioside	0.57
13.23	Hexadecanoic acid, methyl ester	2.63
13.43	Pentadecanoic acid	0.73
13.5	Monensin	15.75
13.9	Zearalenone	1.59
14.17	Oleic acid	2.83
14.35	Cis-vaccenic acid	4.68

Linoleic acid, methyl ester	25.27
Elaidic acid	9.24
Cis-13-eicosenoic acid	4.92
Nonadecanoic acid, methyl ester	7.0
Linoleic acid	4.67
Quinine	0.5
3-(3,4-dimethoxyphenyl)-4,6-dimethylcoumarin	0.98
Di-γ-linolenin	1.97
Palmitoleic acid	4.46
Cystine	4.18
9-tetradecenal, (Z)-	4.42
	Elaidic acid  Cis-13-eicosenoic acid  Nonadecanoic acid, methyl ester  Linoleic acid  Quinine  3-(3,4-dimethoxyphenyl)-4,6-dimethylcoumarin  Di-γ-linolenin  Palmitoleic acid  Cystine

Table (4): Fatty acid analysis of soybean oil

Fatty acid	Classification	Relative distribution			
Palmitic acid C16:0	Saturated fatty acid	10.99%			
Stearic acid C18:0	Saturated fatty acid	4.82%			
Arachidic acid C20:0	Saturated fatty acid	0.36%			
Behenic acid C22:0	Saturated fatty acid	0.29%			
Oleic acid C18:1n9	Unsaturated fatty acid	22.85%			
Linoleic acid C18:2n6	Unsaturated fatty acid	54.28%			
Linolenic acid C18:3n3	Unsaturated fatty acid	6.2%			
Gadolic acid C20:1ω9	Unsaturated fatty acid	0.21%			

Table (5): Serum calcium and phosphorus levels in treated rat groups

Groups Parameters	Group 1 (n=6)	Group 2 (n=6)	Group 3 (n=6)
Calcium (mg/dl)	13.2±0.64	11.3±0.48 *	12.8±0.62
Phosphorus (mg/dl)	10.5±0.66	10.38±0.76	11.96±0.44

\*Significant difference (p<0.05) in comparison with negative control

## 4. CONCLUSION

1\$8 ybean oil decreased blood calcium level, while phosphorus was stable in all treated groups. Supplementation with watermelon white rind aqueous extract rendered calcium level to normal status as negative control.

141

## 5. REFERENCES

- [1] Anderson, J.W., Smith, B.M. and Washnock, C.S. 1999. Cardiovascular and renal latenefits of dry soybean intake. The American Journal of Clinical Nutrition, Vol. 1705(3): 464-474.
- [2] A46AC (2006): 968.22. "Aflatoxin in Peanuts and Peanut Product" CB Method. 147 Washington, D.C.
- [3] **A48**AC (2012): Official method of analysis, 18<sup>th</sup> ed, *Washington D.C. USA*. Volume **1(49)** Chapter (49): No. 991.31p21–23 for Aflatoxins and No. 2000.03 p65–66 for **150**hratoxins. AOAC- IUPAC Method Codex- Adopted- AOAC Method.
- [4] **A52**AC Official Methods of Analysis No. 969.33, Chapter 41, P. 19-20, 19<sup>th</sup> ed. 201652
- [5] (53) los, A.S. da Costa, Aline de Sousa dos Santos and Cellycristina Alves do 154 scimento Saba. 2015. Impact of a high-fat diet containing canola or soybean oil 155 body development and bone. Nutr. Hosp., 31: 2147-2153.
- [6] 156 ilas, S., Canadanovic-Brunet, J. and Cetkovic, G. 2009. By-products of fruits 1570 cessin g as a source of phytochemicals. Chem. Ind. Chem. Eng. 15: 191-202.
- [7] **158**edman, M. and Brandon, D.L. 2001. Nutrition and health benefits of soy **159**teins. J. of Agri and Food Chemistry, 49(3): 1069-1086.
- [8] **M60**land, B.F., Smith, S.A. and Smith, J.C. Jr. 1988. Nutritional status and phytate. **161**urnal of the American Dietetic Association, 88(12): 1562-1566. J. Agric. Chem. **162**d Biotechn., Mansoura Univ. Vol. 6(5): 117-125.
- [9] Maional Research Council (NRC). Nutrient Requirements of Laboratory Animals, 16th ed. Nati onal Academy Press, Washinngton, DC, 1995.
- [10**169**la A. Wahdan, Neamat I. Bassuony, Zeinab M. Abd El-Ghany and Ghadir A. El-166 haghaby. 2017. Watermelon white rind as a natural valuable source of 16phytochemicals and multinutrients. Egyptian Nutrition Society- Special Issue for 168 he First International Conference of Nutrition, Hurghada city, April 2017, p. 89-16904.

- [1170] a Aly Wahdan, Neamat I. Bassuony, Zeinab M. Abd El-Ghany and Amal 17Mustafa Ahmed. 2015. Evaluation of antibacterial activity and gas 172hromatography-mass spectrometry analysis of watermelon white rind extracts.
- [127]3Ologhobo, A.D. and Fetuga, B.L. 1984. Distribution of phosphorus and phytate 17th some Nigerian varieties of legumes. J. of Food Sci., 49(1): 199-201.
- [ 187\$ Yang F., Zhang Y., Xu Q. and Xue C. 2013. Effects of oils on lipid metabolism 176n obese mice induced by a high fat diet. Wei Sheng Yan Jui, 42(6): 17901-914.

178

179

180