

Haematinic Effects of Aqueous Extract of *Lophira lanceolata* Leaves in Phenylhydrazine- induced Haemato-toxicity in Wistar Rats

Abstract

Aim: The haematinic activity of the aqueous extract of *Lophira lanceolata* leaves was investigated using rat model of phenylhydrazine- induced anaemia.

Methods and Results: Red Blood Cell (RBC) count, Haemoglobin (Hb) concentration and Packed Cell Volume (PCV) were analysed as indices of anaemia. Following phenylhydrazine administration to rats at a dose of 10mg/kg for 8 days, a significant decrease ($P<0.05$) in the haematological parameters was observed indicating anaemia. However, treatment with graded doses (200, 400 and 800 mg/kg) of the aqueous extract of *Lophira lanceolata* leaves produced a significant ($P<0.05$) increase in the RBC count, Hb concentration and PCV time- and dose-dependently.

Conclusion: It was concluded that *Lophira lanceolata* leaves possess haematinic activity, making it useful in the management of anaemia.

Key words: Haematinic, *Lophira lanceolata*, Phenylhydrazine, Haemato- toxicity, Wistar rats

1.0 INTRODUCTION

Anaemia is the most prevalent nutritional deficiency disorder in the world. WHO defines anaemia as the condition in which the haemoglobin content of blood is lower than normal as a result of deficiency of one or more essential nutrients ^[1]. According to WHO, more than 2 billion people worldwide suffer from anaemia with iron deficiency responsible for 50% of the cases ^[2]. Anaemia occurs in women of reproductive age anaemia due to menorrhagia while in pregnancy the excess need of iron usually results to anaemia ^[3]. Iron is important for formation of haemoglobin, myoglobin, cytochrome oxidase, peroxidase and catalase. The total quantity of iron in the body averages 4 to 5 grams, about 65 percent of which is in the form of haemoglobin ^[4]. A man excretes about 1 mg of iron each day, mainly into the faeces. For a woman, the menstrual lost of blood brings the iron loss to a value of about 2mg/day ^[5].

Through the ages man has learnt to take advantage of the many resources placed at his disposal by nature to meet his essential needs in all fields. As important reserves and sources of abundance, natural resources are indispensable for socio-economic development. According to Gbile ^[6], the diversity of the flora in Africa partly explains the strength of traditional medicine.

38 Some Africans resort to orthodox methods of treatment due to mitigating circumstances such as high cost
39 of drugs, poverty and poor nutrition ^[7]. Many herbs have been used locally for the management of
40 anaemia and one of such plants is *Lophira lanceolata*. Therefore, this study was undertaken to
41 determine the haematinic potentials of the aqueous leaf extract of *Lophira lanceolata* in rats to
42 provide a scientific basis justifying the use of the plant in traditional medicine for the acclaimed
43 treatment of anaemia.

44 *Lophira lanceolata* is a tree of the tropical and sub-tropical regions. It is a common tree in
45 Cameroun, Nigeria and Sudan. It often grows gregariously on fallow land at the edge of forests.
46 It is a tree of 8 to 10 m tall, straight or twisted, with leaves alternate, clustered at the end of short
47 straight branches, glabrous, bright and blade oblong-lanceolate. The bark surface is corky grey
48 ^[8]. *Lophira lanceolata* is used in traditional medicine to treat several illnesses. The decoction of
49 the fresh leaves is administered orally against headaches, dysentery, diarrhoea, cough, abdominal
50 pains and cardiovascular diseases. It is also used on skin to cure wounds ^[8].

51
52

53 **2.0 MATERIALS AND METHODS**

54 **2.1 Materials**

55 **2.1.1 Chemicals and drugs**

56 All chemicals/drugs used were purchased locally.

57 **2.1.2 Animals**

58 Adult Wistar rats of either sex weighing 180–220g were used for this study. They were kept in
59 stainless steel cages under standard laboratory conditions. They were maintained on clean water
60 and standard rodent feed.

61 **2.2 Methods**

62 **2.2.1 Plant Collection and Identification**

63 The leaves of *Lophira lanceolata* were collected from a natural habitat in Okpella Area of Edo
64 State, Nigeria. The plants were identified at the herbarium unit of the Department of Biology,
65 University of Benin, Benin-City, Nigeria and voucher specimens were deposited for future
66 references.

67 **2.2.2 Preparation of Extracts**

68 The leaves of *Lophira lanceolata* were shade- dried for seven (7) days and pulverized using an
69 electric blender. Two thousand (2000) gram of the pulverized leaves was soaked in distilled
70 water for 72- hours. The resulting mixture was filtered using **Whatman** filter paper (Size No1)
71 and the extract (referred to as LLAE henceforth) was concentrated using a free- dryer.

72 **2.2.3 Acute Toxicity Study**

73 The oral median lethal dose (LD₅₀) of the extract was determined in rats according to the method of
74 Lorke ^[9].

75 **2.2.4 Experimental Design**

76 A total of 25 adult wistar rats were weighed and divided into 5 groups of 5 animals each and
77 treated as follows:

78 Group 1: received distilled water (1 ml) daily (normal control),

79 Group 2: Anaemic and received distilled water (1 ml) daily (anaemic control),

80 Group 3: Anaemic and received LLAE at 200 mg/kg body weight/day

81 Group 4: Anaemic and received LLAE at 400 mg/kg body weight/day

82 Group 5: Anaemic and received LLAE at 800 mg/kg body weight/day

83 Rats were made anaemic by oral intubations of phenylhydrazine (10 mg/kg body weight) daily
84 for 8 days. Rats that developed anaemia with haemoglobin concentration lower than 13 g/dl were
85 selected for the study.

86 **2.2.4.1 Haematological investigation**

87 Blood was collected by ocular puncture after overnight fast. The blood was collected before
88 induction of anaemia, after induction of anaemia with PHZ and during **1, 2, and 3 weeks** of
89 treatments. The haemoglobin concentration (Hb), red blood cell count and pack cell volume
90 (PCV) were assessed.

91 **2.2.5 Statistical Analysis**

92 Statistical analysis was carried out using SPSS version 20.0. All the data were expressed as mean
93 ± SEM and the statistical differences between the means were determined by one way analysis of

94 variance (ANOVA) which was followed by Duncan test and difference between means at $P >$
95 0.05 were considered significant.

96

97 **3.0 RESULTS**

98 **3.1 Acute Toxicity**

99 The results of acute toxicity studies showed no mortality or physical changes in skin and fur,
100 eyes and mucus membrane, respiratory rate, circulatory signs, autonomic and central nervous
101 system effects up to a dose of 5000 mg/kg of aqueous extract of *Lophira lanceolata*. The oral
102 LD₅₀ of the extract was then taken to be > 5000 mg/kg.

103 **3.2 Effect of the Administration of Aqueous Extract of *Lophira lanceolata* Leaves on** 104 **Haematological Parameters of Wistar rats**

105 Tables 1, 2, 3 and 4 shows the changes in haematological parameters of rats treated with
106 phenylhydrazine and the extract. Following the administration of phenylhydrazine (PHZ) for 8
107 days, the RBC, Hb, and PCV of rats decreased significantly ($P < 0.05$) (Table 1) giving rise to
108 macrocytic anaemia. After one week of treatment of the anaemic rats in groups 3, 4, and 5 with
109 *Lophira lanceolata* extract, there was no significant ($P > 0.05$) changes in RBC, Hb, and PVC
110 compared to the anaemic control (group 2) (Table 2). However, at 2 week- post treatment, The
111 Hb, RBC and PCV significantly ($P < 0.05$) and dose- dependently increased with 800mg/ kg of
112 the extract taking the values to near normal range (Table 3) Similarly, at 3 week- post treatment,
113 the Hb, RBC and PCV significantly ($P < 0.05$) and dose- dependently increased but with 800mg/
114 kg of the extract this time taking the values to the normal range (Table 4). Figures 1-3 shows the
115 changes in Hb, PCV and RBC per group during the 3- week treatment regimen. The Hb of the
116 treated rats did not show significant changes within the first week of the experiment but there
117 was a steady increase up to 3 week- post treatment (Figure 1). Similar observations were made
118 for RBC and PCV (Figures 2 and 3 respectively).

119

120

121

122

123

124 **Table 1: Effect of Phenylhydrazine on some Haematological Parameters after 8- days of**
 125 **Administration**
 126

Treatment	Group 1	Group 2	Group 3	Group 4	Group 5
Hb (g/ dl)	16.31±1.11 ^b	11.23±1.11 ^a	12.06±1.13 ^a	11.17±1.23 ^a	11.33±1.25 ^a
RBC (x10 ⁶ µg /l)	6.29±0.80 ^b	3.29±0.81 ^a	3.07±0.33 ^a	3.05±0.44 ^a	3.07±0.53 ^a
PCV (%)	51.11±2.33 ^b	43.23±3.17 ^a	42.11±1.13 ^a	42.13±2.26 ^a	43.12±3.01 ^a

127 Data are presented as mean ± SD. Data was analyzed by one- way ANOVA followed by
 128 Duncan post- hoc test for multiple comparisons, (n=5). Mean values having different lower
 129 case alphabets as superscripts are considered significant (p< 0.05) within the rows. **Group**
 130 **1: Control received 1ml distilled water. Groups 2-5: received 10mg/ kg phenylhydrazine for**
 131 **8 days**

132
 133
 134 **Table 2: Effect of the Aqueous Extract of *Lophira lanceolata* on the Haematological**
 135 **Parameters of Wistar rats one week- post treatment**
 136

Treatment	Group 1	Group 2	Group 3	Group 4	Group 5
Hb (g/ dl)	16.58±2.01 ^b	11.73±1.01 ^a	12.16±1.13 ^a	12.22±1.78 ^a	12.05±1.33 ^a
RBC (x10 ⁶ µg /l)	6.67±1.02 ^b	3.03±0.72 ^a	3.67±0.42 ^a	3.53±0.79 ^a	3.77±0.30 ^a
PCV (%)	51.55±2.13 ^b	43.43±2.42 ^a	44.21±1.32 ^a	45.23±3.21 ^a	45.22±2.99 ^a

137 Data are presented as mean ± SD. Data was analysed by one- way ANOVA followed by
 138 Duncan post- hoc test for multiple comparisons, (n=5). Mean values having different lower
 139 case alphabets as superscripts are considered significant (p< 0.05) within the rows. **Group**
 140 **1: Normal control and received 1ml distilled water. Group 2: Anaemic control and received**
 141 **1ml distilled water. Group 3: Anaemic and received 200 mg/ kg LLAE, Group 4: Anaemic**
 142 **and received 400 mg/ kg LLAE, Group 5: Anaemic and received 800 mg/ kg LLAE**

143
 144
 145

146 **Table 3: Effect of the Aqueous Extract of *Lophira lanceolata* on the Haematological**
 147 **Parameters of Wistar rats two week- post treatment**

Treatment	Group 1	Group 2	Group 3	Group 4	Group 5
Hb (g/ dl)	16.98±1.45 ^b	11.43±1.25 ^a	12.12±1.32 ^a	14.28±1.23 ^{ab}	16.12±2.21 ^b
RBC (x10 ⁶ µg /l)	6.29±0.80 ^b	3.13±0.52 ^a	3.68±0.48 ^a	4.31±0.99 ^{ab}	5.43±0.98 ^{ab}
PCV (%)	51.11±2.33 ^b	42.08±2.44 ^a	44.23±2.32 ^a	48.15±2.73 ^{ab}	50.45±2.91 ^b

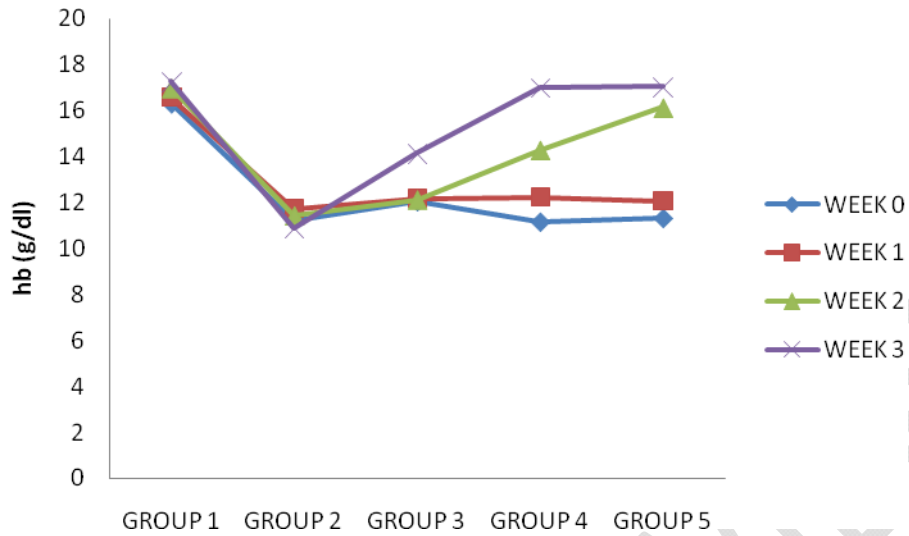
148 Data are presented as mean ± SD. Data was analysed by one- way ANOVA followed by
 149 Duncan post- hoc test for multiple comparisons, (n=5). Mean values having different lower
 150 case alphabets as superscripts are considered significant (p< 0.05) within the rows. **Group**
 151 **1: Normal control and received 1ml distilled water. Group 2: Anaemic control and received**
 152 **1ml distilled water. Group 3: Anaemic and received 200 mg/ kg LLAE, Group 4: Anaemic**
 153 **and received 400 mg/ kg LLAE, Group 5: Anaemic and received 800 mg/ kg LLAE**

154
 155 **Table 4: Effect of the Aqueous Extract of *Lophira lanceolata* on the Haematological**
 156 **Parameters of Wistar rats three week- post treatment**
 157

Treatment	Group 1	Group 2	Group 3	Group 4	Group 5
Hb (g/ dl)	17.26±1.05 ^b	10.86±1.42 ^a	14.12±1.18 ^{ab}	16.99±1.42 ^b	17.01±2.06 ^b
RBC (x10 ⁶ µg /l)	6.97±1.01 ^b	3.25±0.63 ^a	5.31±0.41 ^{ab}	5.81±0.78 ^{ab}	6.93±1.01 ^b
PCV (%)	51.85±3.26 ^b	41.19±1.32 ^a	46.25±2.43 ^{ab}	47.31±1.23 ^{ab}	50.96±2.08 ^b

158 Data are presented as mean ± SD. Data was analysed by one- way ANOVA followed by
 159 Duncan post- hoc test for multiple comparisons, (n=5). Mean values having different lower
 160 case alphabets as superscripts are considered significant (p< 0.05) within the rows. **Group**
 161 **1: Normal control and received 1ml distilled water. Group 2: Anaemic control and received**
 162 **1ml distilled water. Group 3: Anaemic and received 200 mg/ kg LLAE, Group 4: Anaemic**
 163 **and received 400 mg/ kg LLAE, Group 5: Anaemic and received 800 mg/ kg LLAE**

164
 165
 166
 167
 168



169

170 **Figure 1: Changes in Haemoglobin concentration (g/dl) across the groups within the 3-**
 171 **week treatment period. Group 1: Normal control and received 1ml distilled**
 172 **water. Group 2: Anaemic control and received 1ml distilled water. Group 3:**
 173 **Anaemic and received 200 mg/ kg LLAE, Group 4: Anaemic and received 400**
 174 **mg/ kg LLAE, Group 5: Anaemic and received 800 mg/ kg LLAE**

175

176

177

178

179

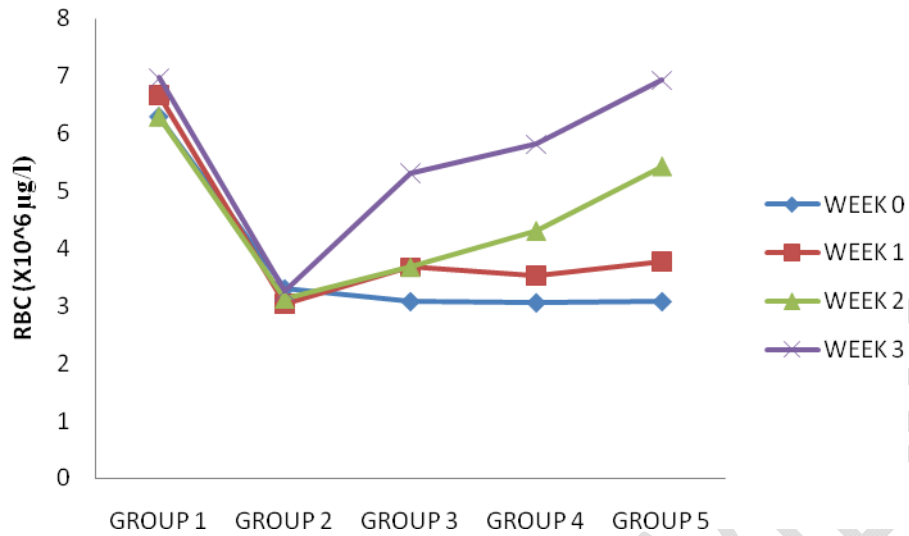
180

181

182

183

184



185
186
187
188
189
190
191
192
193
194
195
196
197
198
199
200
201
202
203
204
205
206
207

Figure 2: Changes in RBC concentration ($\times 10^6 \mu\text{g/l}$) across the groups within the 3- week treatment period. Group 1: Normal control and received 1ml distilled water. Group 2: Anaemic control and received 1ml distilled water. Group 3: Anaemic and received 200 mg/ kg LLAE, Group 4: Anaemic and received 400 mg/ kg LLAE, Group 5: Anaemic and received 800 mg/ kg LLAE

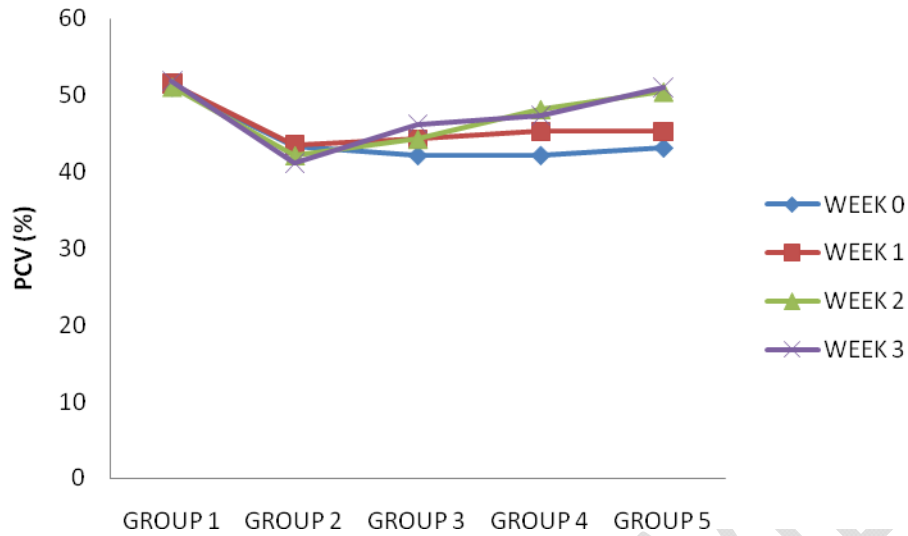


Figure 3: Changes in PVC (%) across the groups within the 3- week treatment period.
Group 1: Normal control and received 1ml distilled water. Group 2: Anaemic control and received 1ml distilled water. Group 3: Anaemic and received 200 mg/ kg LLAE, Group 4: Anaemic and received 400 mg/ kg LLAE, Group 5: Anaemic and received 800 mg/ kg LLAE

208
 209
 210
 211
 212
 213
 214
 215
 216
 217
 218
 219
 220
 221
 222
 223
 224
 225
 226
 227
 228
 229
 230

231 4.0 DISCUSSION

232 Phenylhydrazine produces both aryl and hydroxyl radicals when incubated with rat liver
233 microsomes ^[10] and oxidized by hydrogen peroxide at pH 7.4 and 37°C ^[11]. The radicals induce
234 oxidative stress in the red cell membrane resulting in haemolysis, which may have resulted via
235 lipid peroxidation ^{[12], [13], [14], [15]}. Sub-chronic intoxication of rats with PHZ (10 mg/kg/day for 8
236 days) results in marked haemolytic anaemia characterised by decreased RBC, Hb and PCV ^[16].
237 Similar results were obtained in our study when experimental rats were administered PHZ (Table
238 1).

239 The main function of the RBC is the transportation of oxygen to tissues in the body. As such, any
240 pathological or physiological condition that affects the RBC alters its function and this may be
241 detrimental to the body. In this study PHZ altered the function of RBC by haemolysis
242 characterised by decreased levels of RBC, Hb and PCV. However, our results indicated that the
243 aqueous leaf extract of *Lophira lanceolata* markedly increased the concentration of
244 haemoglobin, red blood cell count and the packed cell volume beginning from 2 week after
245 treatment. It was also observed that the recovery of the treated groups was dose related with the
246 highest dose of 800 mg/kg effecting the highest change. At the third week of the experiment,
247 treatment of anaemic rats with *Lophira lanceolata* increased the RBC, Hb and PCV to near
248 normal values. *Lophira lanceolata* is the well known source of minerals, Sterols, Proteins and
249 other vitamins. These chemical constituents of the plant might be responsible for the haematonic
250 activity.

251 5.0 Conclusion

252 The oral administration of the aqueous leaf- extract of *Lophira lanceolata* significantly improved
253 the haematological parameters affected by phenylhydrazine administration. From this study, it
254 was inferred that *Lophira lanceolata* leaves possess haematonic potential, therefore it could be
255 useful in the management of anaemia.

256

257

258

259

260

261

262 **REFERENCES**

- 263 [1]. Kawaljit kaur, B.D. (2014). Arya girls college,jalandar cantt,Punjab, *European journal of*
264 *zoological research*,3(1):32-36.
- 265 [2]. Ebtesen mehdi and hany sady (2014). Prevalence and risk factors of iron deficiency anaemia,
266 *American journal of health research*,2(5):319-326
- 267 [3]. Ramesh chellan, lopamudra paul (2010); prevalence of iron deficiency anaemia in india,
268 *journal of population and social studies*,19.
- 269 [4]. Sembulingam, K. and Prema Sembulingam, Iron and Haemoglobin metabolism, *Essentials*
270 *Of Medical Physiology*, fourth edition 2007, p.no.72.
- 271 [5]. Arthur. C. (2014). Guyton and John.E.Hall, *Blood, Iron Metabolism, Text book of medical*
272 *physiology*, 9th edition, p 430.
- 273 [6]. Gbile, R. (1986). Haematology of conventionally maintained Lac P Wistar rats during the
274 first year of life. *Lab. Anim.* 16: 198-200.
- 275 [7]. Brown D. (2001). The Herb Society of America New Encyclopedia of Herbs and their
276 uses.New York: (D.K), 31
- 277 [8]. Arbonier M. (2000) Arbres, arbustes et lianes des zones sèches d’Afrique de l’ouest. *CIRAD,*
278 *MNHN, UICN*, 425-427.
- 279 [9]. Lorke, D. 1983. "A new Approach to Practical Acute Toxicity Testing." *Archives of*
280 *Toxicology* 54: 275-287.
- 281 [10]. Gannett PM, Lawson TS, Kolar C, Toth B (1997). Aryl radical formation during the
282 metabolism of arylhydrazines by microsomes. *Chem. Res. Toxicol.* 10 (12): 1372-1377.
- 283 [12]. McMillan DC, Jensen CB, Jollow DJ (1998). Role of lipid peroxidation in dapsone induced
284 haemolytic anaemia. *J. Pharmacol. Exp. Ther.* 287 (3): 868-876.
- 285 [13]. Cighetti G, Debiasis S, Paroni R, Allevi (1999). Free and total malondialdehyde assessment
286 in biological matrices by GC/MS. What is needed for an accurate detection? *Analytical*
287 *Chemistry.* 266 (2): 222-229.
- 288 [14]. Zimmermann L, Antebi H, Morvan BC, Alcindor G (1997). In vitro peroxidation of plasma
289 and erythrocyte lipids during WR 1339 induced hyperlipidemia in Wistar rats. *Ann.*
290 *Pharm. Fr.* 55 (6): 246- 253.
- 291 [15]. Nelson C, Erikson K, Pinero DJ, Beard JL (1997). In Vivo dopamine metabolism is altered
292 in iron deficient anaemic rats. *J. Nutr.* 127(12): 2282-2288.
- 293 [16]. Unami A, Nishina N, Terai T, Sato S, Tamura T, Noda K, Mine Y (1996). Effect of
294 cisplatin on erythropoietin production in rats. *J. Toxicol. Sci.* 21(3): 157-65.

295