

Combining Ability Analysis of Yield Components and Late Leaf Spot Resistance Trait of Nine Groundnut Genotypes.

Abstract

Four susceptible genotypes (SAMNUT 23, 24, 25 and 26) were used as females and five resistant genotypes (ICGV 12991, ICGV 7878, FDR-F7 82, FDR-F7 67 and FDR-F7 61) were used as males using line x tester mating design with three replications. The parents and progenies were evaluated for late leaf spot resistance. Highly significant negative GCA effects were recorded for disease incidence in SAMNUT 25 (-0.05), indicating the genotype is a good general combiner for LLS disease incidence and SAMNUT 26 (-5.90) is a good general combiner for LLS disease severity. ICGV 12991 (-12.00), FDR-F7 67 (-.68) and FDR-F7 61 (-2.23) genotypes are good general combiner for LLS disease tolerance (disease incidence and severity). Among the crosses, significant and negative SCA effects were obtained for most of the crosses i.e., SAMNUT 24 x FDR-F7 67 and SAMNUT 24 x FDR-F7 61 for LLS tolerance (disease incidence and severity), indicating that they are good specific combiners for LLS tolerance. The ratio of the GCA and SCA variances indicated the preponderance of SCA variance over GCA variance for disease incidence and severity, indicating the role of non-additive gene effect and it may be due to difference in genotypes used as parents.

Introduction

Late leaf spot is the most devastating fungal disease accounting for yield loss of over 60% (Okello *et al.*, 2010). The disease is caused by *Phaeoisariopsis personata* with symptoms that are seen as small necrotic flecks that enlarge and become light to dark brown. Efforts have been made to control late leaf spot disease using a combination of cultural and chemical measures with limited success (Page *et al.*, 2002). Use of fungicides to control leaf spots usually increases production costs by 10% (Coffelt and Porter, 1986). Effective chemical control is heavily reliant upon multiple fungicide applications (Jordan *et al.*, 2012), which are costly for resource poor farmers in Nigeria, and as well raises environmental and health concerns with significant decrease in crude protein and fiber contents with increasing disease severity (Coffelt and Porter, 1986). These factors coupled with health hazards associated with the use of insecticides suggested the use of host plant resistance as the most effective and environmentally friendly control measure for the management of LLS. The use of resistant genotypes and genetic information on inheritance of LLS will help in the development and utilization of LLS resistant cultivars which will reduce production costs; and boost groundnut production in Nigeria. It is also necessary to know about the nature and magnitude of gene action responsible for controlling the inheritance of various yield attributes along with combining ability of the parents and their cross combinations in order to make use of them in further crop improvement program. The line x tester analysis is one of the efficient methods of evaluating large number of inbred as well as providing information on the relative importance of GCA and SCA effects for interpreting the genetic basis of important plant traits (Singh and Chaudhury, 1985). The most commonly used designs for combining ability studies are line x tester (L x T) and diallel analysis. Combining ability analysis following line x tester given by Kempthorne (1957) and Arunachalam (1974) is frequently used for testing the performance of lines in hybrid combinations. It is also useful in characterizing the nature and magnitude of gene action involved in controlling the quantitative

traits. The general and specific combining ability effects and variances obtained from a set of F1s would enable a breeder to select desirable parents and crosses for each of the quantitative components separately. From their results Tatum and Spargue (1942) concluded that the general combining ability was mainly the results of additive gene action while the specific combining ability due to dominance, epistasis and genotypic environment interaction. Baker's ratio close to unity indicates additive.

Due to different genetic control of LLS and yield associated traits in various genetic materials, the objectives of the present study were therefore to identify general and specific combining abilities and narrow-sense heritability estimates for yield component and LLS resistance traits in nine groundnut genotypes.

Materials and Methods

The experimental materials for this study comprised of nine early maturing groundnut genotypes representing a range of resistance levels to Late Leaf Spot (LLS) obtained from IAR Samaru. The resistant genotypes were validated in the 2013/2014 growing season IAR at the farm, Samaru. Five of the groundnut genotypes are tolerant to the Late Leaf Spot (LLS), these are ICGV 12991, ICGV 7878, FDR-F7 82, FDR-F7 67 and FDR-F7 61 which were used as male. The other four genotypes are the improved and released material by IAR, which are all susceptible to the Late Leaf Spot. These are SAMNUT 23, SAMNUT 24, SAMNUT 25 and SAMNUT 26 which were used as females. The 20 F1 along with 9 parents were evaluated for LLS in the screen house using line x tester design in a randomized complete block design (RCBD) with three replications. Two pots were allocated for each genotype and two seed were sown per pot. All the plant protection and agronomic measures were adopted. Thirty-five days old plants were inoculated with LLS at 106 conidia/ml inoculum concentrations. Hand held sprayer was used for the inoculation; 0.1 ml spore suspension was dropped on the leaves. High relative humidity around the plants was maintained by covering the plants with wet polythene bags 24 hours before inoculation. Inoculated plants were covered for another 24 hours to maintain high humidity. Plants were observed weekly for development of disease after inoculation and disease score was recorded using 1-9 scale describes by Subrahmanyam *et al.*, (1995). Data were recorded on four randomly selected plants of each entry of each replication for plant height, days to 50% flowering, days to maturity, number of matured pods per plant, number of seeds per pod, 100-seed weight, LLS disease severity and LLS disease incidence. Data collected on disease severity and incidences were transformed using log10 and were all subjected to analysis of variance using General Linear Model procedure of Statistical Analysis System (SAS) package (SAS, 2002). The combining abilities (GCA and SCA) were carried out as per Singh and Chaudhury (1985).

Table 3.2: Description of Leaf Spot scale (1-9)

Leaf spot score	Description	Disease Severity (%)
1	No disease	0
2	Lesion larger on lower leaves, no defoliation	1 – 5
3	Lesion larger on lower leaves, very few lesion on middle leaves, defoliation of some leaflets evident	6 – 10
4	Lesion on the lower middle leaves, but severe on lower leaves, defoliation of some leaflets evident on lower leaves	11 – 20
5	Lesion on all lower and middle leaves, over 50% defoliation of lower leaves	21 – 30
6	Lesion severe on lower and middle leaves, defoliation of some leaflets evident on middle leaves	31 – 40
7	Lesion on all leaves but less severe on top leaves, defoliation of all lower and some middle leaves	41 – 60
8	Defoliation of almost all middle leaves, lesion severe on top leaves and some defoliation of top leaves evident	61 – 80
9	Defoliation of almost all leaves having bare stems, some leaflets may be present, but with severe leaf spot	81– 100

Results and Discussion

The analysis of variance for male parents revealed the presence of significant variation for all studied traits except days to 50% flowering and days to maturity **traits**. Further partitioning of variance indicated lack of variability among females for important characters *viz* disease incidence and disease severity. The crosses showed a significant difference among all **investigated** traits except plant height, days to 50% flowering and days to maturity **traits**.

The results of **general combining ability (GCA)** are presented in Table 4.2. Negative GCA effect observed for days to maturity **trait** is required for the development of early maturing genotype as reported by Vishnuvardhan, *et al.* (2012). The negative and significant GCA effect exhibited by SAMNUT 25 indicated that it is a good general combiner for disease incidence. SAMNUT 26 is a good general combiner with a negative and significant GCA effect for disease severity. ICGV 12991, FDR-F7 67 and FDR-F7 61 are good general combiners with negative GCA effects on LLS tolerance (disease severity and incidence).

The progenies with negative SCA effects for disease incidence and disease severity SAMNUT 24 x FDR-F7 61 and SAMNUT 25 x FDR-F7 67 **crosses** were identified as the most promising genotypes in breeding program for LLS resistance traits. These progenies originated from

parents with negative and positive GCA values in negative direction. This suggest the difficulty in predicting the *P. personata* tolerance level of the progenies based on GCA alone and should necessitate testing of specific male x female combinations (add the reference). Arunga *et al.* (2010) reported that the SCA effect alone has limited value for parental choice in breeding programs. They, therefore suggested that SCA effects should be used in combination with other parameters, such as hybrid means and the GCA of the respective parents such that hybrid combination with both high mean and favorable SCA estimates and involving at least one of the parents with high GCA, would tend to increase the concentration of favorable alleles; which is desired by any breeder (add the reference). Furthermore, it was observed that crosses involving one good combiner and one average or poor combiner showed negative SCA effects. For instance, SAMNUT 25 and FDR-F7 67 genotypes have a poor GCA values for disease incidence and severity resistance, while their cross shown a negative and desirable SCA effects. This manifestation of progenies having reactions not related to the parent's attributes introduced a different dimension in the inheritance of groundnuts resistance to LLS. John *et al.* (2012) and Ayo-Vaughan *et al.* (2013) observe similar phenomena in groundnuts and cowpeas, respectively and attributed it to genetic interaction between favorable alleles and between unfavorable alleles contributed by both parents. This suggests that inheritance of this trait is not simple and that inter-allelic interactions due to epistasis could be responsible. However, SCA effects are not very important for crops like groundnuts that are highly self-pollinated and difficult to produce commercial hybrids, a point advanced by Kimani and Derera (2009) while working on beans; self-pollinated counterpart of groundnut. (Table 4.3).

The relative importance of GCA to SCA variance was judged from the ratio of the GCA to SCA variance which helps to indicate the predominance of either additive or non-additive action (add the reference). The predominance of non-additive genetic variance over additive genetic variance indicated by the values with magnitude less than unity for all the traits measured, it may be due to differences in the genotypes used as parents as stated by Abul-Kalam, *et al.* (2014), Table 4.4.

Table 4.1: Mean Squares for some Agronomic Traits and Late Leaf Spot of Groundnut Genotypes Generated using North Carolina Design II Evaluated at Samaru in 2016.

Sources of variation	Df	PLHT(cm)	50%F	DM	MPPP	SPP	SWGT(g)	DI (%)	DS (%)
Rep	2	16.65	19.72	18.15	0.82	0.02	0.16	0.05	0.07
Male	4	59.85**	2.28	10.03	35.79**	0.20**	1.63**	0.02**	0.03**
Female	3	10.27	1.93	33.91**	28.06**	0.26**	65.38**	0.01	0.01
Female x Male	12	12.85	4.72	4.37	24.88**	0.12**	4.44**	0.01**	0.01**
Error	38	12.70	6.97	5.63	0.97	0.01	0.14	0.01	0.01

** = 0.01 probability levels respectively. PLHT (cm) = plant height, 50%F=days to 50% flowering, DM=days to maturity, MPPP= number of mature pod per plant, SPP= number of seed per pod, SWGT (g), = 100 seed weight, DS (%) =disease severity in percentage, DI (%) =disease incidence in percentage.

135 Table 4.2: Estimates of General Combining Ability effects of Groundnut Genotypes Evaluated for some
136 Agronomic Traits and Late Leaf Spot at Samaru in 2016

GENOTYPE	PLHT(cm)	50%F	DM	MPPP	SPP	SWGT(g)	DI (%)	DS (%)
<i>Males</i>								
ICGV12991	-3.18**	0.37	-20.87**	-0.50	-0.13**	3.28**	-0.01	-12.00**
ICGV 7878	-0.93	-0.35	23.72**	-1.12**	0.10**	6.72**	0.01	2.08**
FDR-F7 82	0.07	0.45	27.05**	1.22**	0.02	-3.02**	-0.02	0.83**
FDR-F7 67	1.07	0.12	23.63**	-1.25**	-0.19**	-2.02**	0.04*	-0.68**
FDR-F7 61	-0.20	-0.22	26.80**	1.15**	0.08**	-1.68	-0.03	-2.23**
SE±	0.92	0.68	0.61	0.25	0.02	1.85	0.02	0.02
<i>Females</i>								
SAMNUT 23	2.40**	-0.63	-20.80**	0.33	0.13**	1.03	0.06**	11.93**
SAMNUT 24	-0.85	0.45	-20.07**	2.25**	0.13**	0.53	0.01	6.00**
SAMNUT 25	1.82*	-0.13	-20.40**	-1.75**	-0.02	-1.80	-0.05**	-0.02
SAMNUT 26	-0.18	-0.05	-19.07**	-0.33	-0.12*	-3.05	-0.01	-5.90**
SE±	1.03	0.76	0.69	0.28	0.03	2.07	0.02	0.02

137 * and ** = significant at 0.05 and 0.01 probability levels respectively. PLHT (cm)= plant height, 50%F=days to 50%
138 flowering, DM=days to maturity, MPPP= number of mature pod per plant, SPP= number of seed per pod,
139 SWGT(g),= 100 seed weight, DS (%)=disease severity in percentage, DI (%)=disease incidence in percentage.

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143 Table 4.3: Estimates of Specific Combining Ability effects of Groundnut Genotypes Evaluated for some
144 Agronomic Traits and Late Leaf Spot at Samaru in 2016.

CROSSES	PLHT(cm)	50%F	DM	MPPP	SPP	SWGT(g)	DI(%)	DS (%)
SAMNUT 23 x ICGV12991	-4.07	-1.23	-0.23	-3.80**	-0.25**	-11.30*	-0.04	-0.74**
SAMNUT 23 x ICGV7878	2.85	2.35	-0.48	0.95	-0.02	1.87	-0.05	0.22**
SAMNUT 23 x FDR F782	0.85	-0.40	0.27	1.28*	0.00	6.20	0.06	0.13**
SAMNUT 23 x FDR F767	-0.15	-0.82	-1.40	3.53**	0.16**	-0.88	0.00	0.19**
SAMNUT 23 x FDR F761	0.52	0.10	1.85	-1.97**	0.11*	4.12	0.04	0.20**
SAMNUT 24 x ICGV12991	0.93	-0.03	0.10	2.53**	0.02	-1.57	0.06	0.34**
SAMNUT 24 x ICGV7878	-0.15	-0.78	0.85	3.62**	0.05	6.27	0.04	-0.01

SAMNUT 24 x FDR F782	0.52	-0.20	-0.40	-0.72	-0.10*	-3.40	0.00	-0.12**
SAMNUT 24 x FDR F767	0.52	1.05	0.93	-4.47**	0.14*	6.52	0.00	-0.09*
SAMNUT 24 x FDR F761	-1.82	-0.03	-1.48	-0.97	-0.11*	-7.82	-0.10*	-0.11*
SAMNUT 25 x ICGV12991	1.60	0.63	1.50	1.33*	0.23**	11.77*	-0.03	0.22**
SAMNUT25 x ICGV7878	-2.15	-1.78	-0.42	-0.92	-0.10*	-1.40	0.06	-0.08*
SAMNUT 25 x FDR F782	0.52	1.47	-1.00	-0.25	0.28**	0.60	-0.04	-0.06
SAMNUT 25 x FDR F767	0.85	-0.95	-0.33	-0.67	-0.22**	-2.15	-0.08*	-0.08*
SAMNUT 25 x FDR F761	-0.82	0.63	0.25	0.50	-0.20**	-8.82*	0.09*	-0.01
SAMNUT 26 x ICGV12991	1.53	0.63	-1.37	-0.07	0.00	1.10	0.01	0.18**
SAMNUT 26 x ICGV7878	-0.55	0.22	0.05	-3.65**	0.07	-6.73	-0.04	-0.13**
SAMNUT 26 x FDR F782	-1.88	-0.87	1.13	-0.32	-0.18**	-3.40	-0.01	0.05
SAMNUT 26 x FDR F767	-1.22	0.72	0.80	1.60*	-0.08	-3.48	0.08	-0.02
SAMNUT 26 x FDR F761	2.12	-0.70	-0.62	2.43**	0.20**	12.52**	-0.03	-0.08*
SE±	2.06	1.52	1.37	0.57	0.05	4.13	0.04	0.04

* and ** = significant at 0.05 and 0.01 probability levels respectively. PLHT (cm)= plant height, 50%F=days to 50% flowering, DM=days to maturity, MPPP= number of mature pod per plant, SPP= number of seed per pod, SWGT(g)= 100 seed weight, DS (%)=disease severity in percentage, DI (%)=disease incidence in percentage.

Table 4.4: Component GCA and SCA Variance of Groundnut Genotypes Evaluated for some Agronomic Traits and Late Leaf Spot at Samaru in 2016.

Estimates	PLHT(cm)	50%F	DM	MPPP	SPP	SWGT(g)	DI(%)	DS (%)
σ_{GCA}^2	8.57	3.15	2.91	16.59	0.08	2.96	0.01	0.01
σ_{SCA}^2	40.99	-9.59	30.09	63.46	0.44	63.38	0.02	0.03
$\frac{\sigma_{GCA}^2}{\sigma_{SCA}^2}$	0.21	-0.33	0.10	0.26	0.18	0.05	0.40	0.27

PLHT(cm)= plant height, 50%F=days to 50% flowering, DM=days to maturity, MPPP= number of mature pod per plant, SPP= number of seed per pod, SWGT(g)= 100 seed weight, DS (%)=disease severity in percentage, DI (%)=disease incidence in percentage.

References

- Abul-Kalam A., Shah-E-Alam., Abdul-Hamid., Mohd Y.R., and Malek, M. A. (2014). Combining Ability of Pod Yield and Related Traits of Groundnut (*Arachis hypogaea* L) under Salinity Stress. *The Scientific World Journal*. Doi.org/10.1155/2014/589586.
- Arunachalam, V., Srivastava, P.S.L. (1977). Heterosis as a function of genetic divergene in triticale, *Z Pflanzenzuchtg*, **79**: 269-275

160 Arunga, E.E, Van-Rheenem, H.A. and Owuoche, J.O. (2010). Diallel analysis of Snap bean
 161 (*Phaseolus vulgaris* L.) varieties for important traits. *African Journal of Agricultural Research*
 162 5(15):1951-1957.

163 Ayo-Vaughan, M. A., Ariyo, O. J., and Alake, C. O. (2013). Combining ability and genetic
 164 components for pod and seed traits in cowpeas. *Italian Journal of Agronomy*; Vol.8:10.

165 Coffelt, T. A. and Porter, D. M. (1986). Field screening of reciprocal Chico x Florigiant peanut
 166 population for resistance to leafspot in Virginia, *Peanut Science*. 13: 57-60.

167 John, K., Reddy. R. P., Reddy, K.H., Sudhakar. P. and Reddy, N.P.E. (2012). Identification of best
 168 heterotic crosses for yield and water use efficiency traits in groundnut (*Arachis hypogaea* L.).
 169 *Journal of Plant Breeding and Crop Science*. 4 :17-24.

170 Jordan.D.L., Brandenburg, R.L., Brown, A.B., Bullen, G.S., Roberson, G.T., Shew, B. and Spears,
 171 F.J. (2012). Peanut Information. North Carolina Cooperative Extension Service, 54 College
 172 of Agriculture and Life Sciences North Carolina State University. Pages, 100- 127.

173 Kempthorne, O., (1957). An Introduction to Genetics Statistics. John Wiley and Sons Inc., New
 174 York, pp. 176-178.

175 Kimani, J. M. and Derera, J. (2009). Combining ability analysis across environments for some traits
 176 in dry bean (*Phaseolus vulgaris* L.) under low and high soil phosphorus conditions.

177 Okello D. K., Biruma, M. and Deom, C. M. (2010). Overview of groundnuts research in Uganda:
 178 Past, present and future. *African Journal of Biotechnology* 9:6448-6459.

179 Page, W. W., Busolo-Bulafu, C.M., vander Merwe, P.J.A. and Chancellor, T.C.B . (2002).
 180 Groundnut manual for Uganda: Recommended groundnut production practices for
 181 smallholder farmers in Uganda. Chatham, UK: Natural Resources Institute. Pages 1-12.

182 SAS Institute (2002) SAS/STAT. User's guide version 9.6th edition. SAS Institute Carry NC.

183 Singh, R. K. and Chaudhary, B. D. (1985). *Biometrical Methods in Quantitative Genetic Analysis*.
 184 Rev. ed. Kalyani Publishers, New Delhi 110 002, India.pp318.

185 Subrahmanyam, P., McDonald, D., Waliyar, F., Reddy, L.J., Nigam, S.N., Gibbons, R.W., Rao, V.
 186 R., Singh, A.K., Pande, S., Reddy, P.M. and SubbaRao, P.V.S. (1995). Screening methods
 187 and sources of resistance to rust and late leaf spot of groundnut. *Information Bulletin* No: 47
 188 ICRISAT. Page 24.

189 Tatum, L. A., and Sprague, G.F.(1942). General vs specific combining ability in single cross of corn.
 190 *Journal of American Society of Agronomy*, **34**: 923-932.

191 Vishnuvardhan. K. M., Vasanthi. R. P., Reddy K. H. P., and Reddy, B.V. (2012). Genetic variability
 192 studies for yield attributes and resistance to foliar diseases in groundnut (*Arachis hypogaea*
 193 l.). *International Journal of Applied Biology and Pharmaceutical Technology*.3: 390-394.