

Original Research Article

POTENTIAL PROTECTIVE EFFECT OF *ANACARDIUM OCCIDENTALE* AGAINST PARACETAMOL-INDUCED HEPATOTOXICITY

ABSTRACT

This study investigates the protective effects of the ethanolic extract of *Anacardium occidentale* on paracetamol induced liver toxicity in Wistar rats. Twenty five (25) female Wistar rats were randomly divided into five (5) groups of five (5) rats each. The groups were designated as follows; A, B, C, D and E. Group A served as the normal control, group B served as the negative control and received 1000 mg/kg of paracetamol only, group C received 1000 mg/kg of paracetamol then treated with 250 mg/kg of *A. occidentale* extract, group D received 1000 mg/kg of paracetamol then treated with 500 mg/kg of *A. occidentale* extract and group E received 1000 mg/kg of paracetamol together with 500 mg/kg of the extract for 4 weeks respectively. After the last day of administration, the rats were anaesthetized using ketamine and the liver tissues were harvested and sent for histopathological examination. The result revealed significant ($P < 0.05$) decrease in body weight of testing groups compared to their controls after paracetamol administration. The histopathological findings revealed severe focal aggregation of inflammatory cells (SFAIC), severe hepatocellular necrosis in the negative control and showed regeneration of hepatic tissues, which was more significant in the group that received high doses of the extract (500 mg/kg). It can be deduced that the ethanolic extract of *A. occidentale* has protective effect against liver toxicity and should be recommended as an alternative to traditional treatment.

Comment [U1]: Reduce the abstract

Keywords: *Anacardium occidentale*, paracetamol, liver, acetaminophen, Wistar rats.

1.0. INTRODUCTION

Paracetamol, also known as acetaminophen, is the most widely used over the counter medication to treat pain and fever [1]. It is a common analgesic used worldwide and is the first step on the World Health Organization (WHO) pain ladder [2]. Its mechanism of action is unknown; however, recent studies demonstrate that paracetamol inhibits the production of prostaglandin within the central

27 nervous system and peripheral tissues [3]. The use of paracetamol has been generally considered
28 safe when consumed to the maximum recommended dosage of 4 g/day than other commonly used
29 analgesics like the opiates and non-steroidal anti-inflammatory drugs (NSAIDs); but its safety and
30 efficacy has been questioned [4]. Paracetamol toxicity is one of the leading causes of intentional and
31 unintentional poisoning in the United State which could be attributed to its availability as both a single
32 ingredient and in combination with other medications in various concentrations [5]. Its toxicity could
33 also result from either acute or chronic overdose. Hepatotoxicity is a consequence of over
34 consumption of paracetamol which could result to abnormalities in liver function, acute liver failure
35 and sometimes death [6]. Paracetamol could also cause skin reactions such as Steven-Johnson
36 syndrome, kidney cancer [7] and increased risk of childhood asthma during pregnancy [8].

37 *A. occidentale* (commonly called cashew) is a well-known member of the *Anacardiaceae* family.
38 Although, poisonous plants are ubiquitous, herbal medicine is used by up to 80% of the population in
39 the developing countries [9]. It is widely cultivated in the subtropical and tropical countries of the world
40 and is used in the tropics for the treatment of diarrhoea and *E.coli*, the extract of the leaf has been
41 used to lower blood pressure and sugar [10]. In the traditional Nigerian and Brazilian pharmacopoeia,
42 stem bark of *A. occidentale* is known for its anti-inflammatory effects [11,12]. The leaves of *A.*
43 *occidentale* have been reported to possess phytoconstituents such as saponins, tannins and
44 flavonoids, which exerts antioxidant activities [13]. The seeds are consumed orally in Columbia as
45 aphrodisiac and to cure impotency and also used as anti-venom for snake bites [14]. It has also been
46 found to have pharmacological effect such as anti-ulcerogenic [15], antihyperglycaemic [16] and anti-
47 diabetic properties [17].

48 The existing management of liver toxicity using orthodox and various methods of medications are very
49 expensive, which require prolonged use and sometimes are accompanied by serious side effects.
50 However, medicinal plants seem inexpensive and very assessable with little or no side effects. Thus,
51 this study aimed at evaluating the hepatoprotective activity of ethanolic extract of *A. occidentale*
52 leaves on paracetamol induced toxicity.

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55 2.0. Materials and Methods

56 2.1. Preparation of the plants extract

57 Fresh leaves of cashew were collected from Uli, Anambra State, Nigeria and washed in a basin of
58 water to remove dirt and debris. The leaves were dried under ambient room temperature after which it
59 was grounded using laboratory mill into coarse form. 50 g of the coarse powder was macerated in 250
60 ml of ethanol for 72 hrs and then filtered using a filter paper. The filtrate was concentrated using a
61 rotary evaporator and further dried using thermostat oven into a gel-like form.

62 2.2. Chemical reagents

63 The chemicals of this study were purchased from Emmy enterprise, Onitsha, Anambra State.
64 Paracetamol manufactured by Emzor was procured from the pharmaceutical unit of Chukwuemeka
65 Odumegwu Ojukwu University while 10% formal saline was from the histopathology section of the
66 college of medicine, Chukwuemeka Odumegwu Ojukwu University.

67 2.3 Ethical approval

68 Ethical clearance was obtained from the research and ethics committee of the Faculty of Basic
69 Medical Sciences, Chukwuemeka Odumegwu Ojukwu University, Nigeria. The animals were treated
70 in line with the National Institute for Health Guide on the Care and Use of Laboratory Animals (1985).

71 2.4. Animals

72 Twenty five (25) healthy female Wistar rats weighing about 100 to 190 g were purchased from the
73 Chris Animal farm, Nwagpadolu estate, G.R.A Awka, Anambra State and were transferred to the
74 animal house of the Department of Anatomy, Faculty of Basic Medical Science, Chukwuemeka
75 Odumegwu Ojukwu University, Uli, Anambra State. They were then housed in standard cages under
76 normal temperature (27-30°C) and fed with guinea feed (produced by Grand Cereals Ltd.) and water
77 *ad libitum*. The animals were allowed to acclimatize for a period of two weeks before administration.

78 2.5. Experimental procedure

79 The animals were weighed using digital balance before commencement of administration and after
80 the last day of administration.

81 2.5.1. Induction of toxicity

82 Twenty rats were induced with toxicity. 1000 mg of paracetamol was dissolved in 100 ml of distilled
83 water to give a concentration 10 mg/ml.

84 2.5.2. Experimental design

85 The rats were randomly divided into five (5) groups of five rats each (n=5) designated as groups A, B,
86 C, D and E. The extracts were given orally with the use of an oral intubation tube between the hours
87 of 11 am-1 pm. The breakdown of the treatment of each group is as follows;

88 GROUP A (Normal control): Fed with rat feed and water

89 GROUP B (Negative control): Administered 1000 mg/kg body weight of paracetamol only for three
90 days, with normal feed and water.

91 GROUP C: Administered 1000 mg/kg body weight of paracetamol only for three days and then
92 treated with 250 mg/kg body weight of the extract.

93 GROUP D: Administered 1000 mg/kg body weight of paracetamol only for three days and then
94 treated with 500 mg/kg body weight of the extract.

95 GROUP E: Administered 1000 mg/kg body weight of paracetamol and 500 mg/kg body weight of the
96 extract concurrently.

97 The administration was given orally, once daily between the hours of 11 am and 1 pm for a period of
98 28 days. On the 29th day, the animals were anesthetized by the injection of ketamine and dissected.

99 The liver was harvested and fixed on 10% formal saline for histological examination.

100 2.5.3. Histological examination

101 The liver was harvested from the animals were fixed on 10% formal saline. The tissues were
102 processed by passing them through ascending grades of alcohol and then cleared in xylene after
103 which embedding in paraffin wax was carried out. Sections of about 3-5 µm was obtained using
104 rotatory microtome which were later deparaffinised, hydrated and stained using haematoxylin and
105 eosin (H&E) dye. The tissues were then mounted using neutral dibutylphthalate xylene (DPX) medium
106 for microscopic examination at x 600 magnification.

2.5.4. Statistical analysis

Data were analyzed using students'-test, with values expressed as Mean \pm SEM (Standard Error of Mean). It was done with the use of Statistical Package for Social Sciences (SPSS) software (Version 20). Differences between means were considered at $P < 0.05$.

3.0. RESULTS

3.1 Body weight change

Table 1. Effect of ethanolic extract of *A. occidentale* leaves on body weight of paracetamol induced toxicity rats

		MEAN	\pm SEM	P-VALUE	T-VALUE
Group A	Initial	116.66	\pm 8.81		
	Final	170.00	\pm 10.00	0.004**	-16.000
Group B	Initial	150.00	\pm 5.77		
	Final	120.00	\pm 5.77	0.035*	5.196
Group C	Initial	140.00	\pm 5.77		
	Final	113.33	\pm 3.33	0.015*	8.000
Group D	Initial	153.33	\pm 8.81		
	Final	120.00	\pm 5.77	0.038*	5.000
Group E	Initial	190.00	\pm 5.77		
	Final	156.66	\pm 3.33	0.038*	5.000

Values are expressed as Mean \pm SEM, * $P < 0.05$, ** $P < 0.01$, final body weight compared to initial body weight.

3.2 Histopathological findings

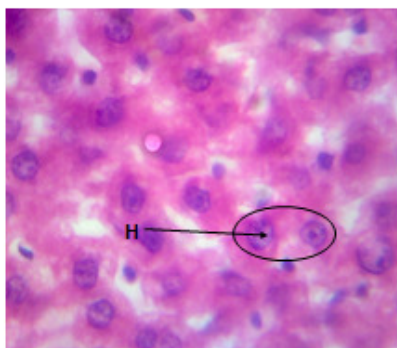


Plate A: Photomicrograph of the normal control section of the liver of rats administered with distilled water only showing normal Hepatocyte (H) and cytoarchitecture.

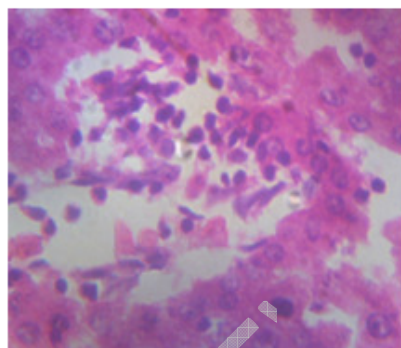


Plate B: Photomicrograph of liver tissue of rats administered with 1000 mg/kg of paracetamol only group showing severe focal aggregate of inflammation cells (SFAIC), cytoplasmic ground glass appearance (CGGA) and severe hepatocellular necrosis (HCN).

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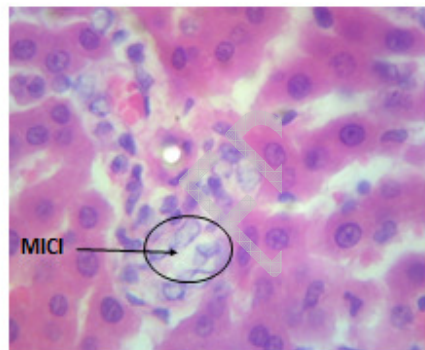


Plate C: Photomicrograph of liver tissue of rats administered with 1000 mg/kg of paracetamol and treated with 250 mg/kg of the extract showing moderate regenerated hepatic tissue with moderate inflammatory cell infiltration (MICI) and mild intrahepatic hemorrhage (MIHH).

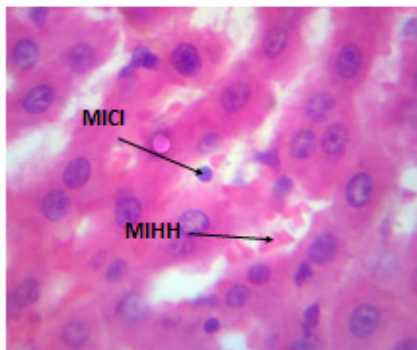


Plate D: Photomicrograph of liver tissue of rats administered with 1000 mg/kg of paracetamol and treated with 500 mg/kg of the extract showing well regenerated hepatic tissue with mild inflammatory cell infiltration (MICI) and mild intrahepatic hemorrhage (MIHH).

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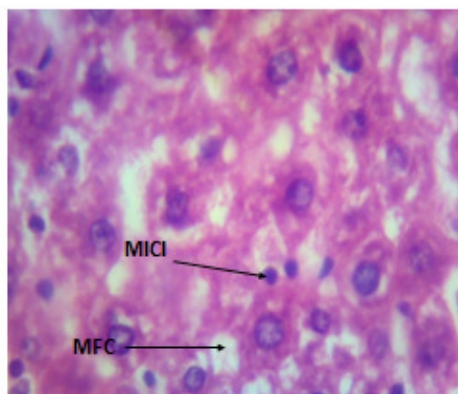


Plate E: Photomicrograph of liver tissue of rats administered with 1000 mg/kg of paracetamol and 500 mg/kg of the extract showing moderate cytoarchitectural regeneration with moderate inflammatory cell infiltration (MICI) and mild fatty changes (MFC).

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121 4.0 DISCUSSION

122 The use of plants has shown promising effect in the treatment and management of diseases and
 123 toxicity. This is due to the presence of potent bioactive compounds in the plant which have anti-
 124 hepatotoxic properties [18].

125 Findings from this study revealed a significant increase in body weight of the group A animals.
 126 However, there was a significant loss of body weight in groups B, C, D and E. The increase in body
 127 weight could be physiological as the animals were only exposed to water and feed throughout the
 128 study. The reduction in body weight could be as a result of exposure to toxicity or due to loss of
 129 appetite by the animals. This is in line with that of Jaouad, 2004 who reported a significant decrease
 130 in body weight of rats when administered 225 mg/kg of paracetamol. The groups treated with the
 131 extract also showed reduction in weight and could be as a result of the action of anacardiac acid,
 132 which is a component of the extract that has been found to reduce the deposition of fat by its
 133 uncoupling action [19]. This corresponds with the study of Dare et al., [20].

134 Histopathological study of toxicity induced untreated rats showed severe histopathological changes
135 on the hepatic tissue, which was due to the paracetamol used in the study. This corresponds with
136 previous reports by [21]. The histological study of the treated groups (C, D and E) showed the
137 regenerative effect of the liver tissues. *A. occidentale* may possess antioxidant properties that are
138 related to their ability to inhibit peroxidative damage caused by environmental toxicants [22,23].
139 Higher doses of the extract showed the greater restorative effect of the liver tissue than lower dose of
140 the extract. This protective effect is in agreement with previous reports [24].

141 5.0. CONCLUSION

142 Findings from this study indicate that consumption of ethanolic extract of *A. occidentale* leaf
143 possesses hepatoprotective effects on the liver against paracetamol induced toxicity. It should
144 therefore be recommended in the management of liver disorders.

145 ACKNOWLEDGMENT

146 None

147 COMPETING INTERESTS

148 Authors have declared that no competing interests exist

149 ETHICAL APPROVAL

150 All authors hereby declare that principles of laboratory animal care (NIH publication No. 85-23,
151 revised 1985) were followed as well as specific national laws where applicable. All experiments have
152 been examined and approved by the appropriate ethics committee.

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