

Reduction of some Food-Borne Pathogens in Chicken Fillets Using Aluminum and Silicon Nanocomposite

An assessment of Aluminium and Silicon Nanocomposite on some Food-Borne Pathogens associated with Chicken Fillets

Running head: Nanoparticles and improving quality of chicken fillets

ABSTRACT

Nanotechnology is an innovative technology for improving food quality and safety.

Aims: The aim of this study was to evaluate the efficacy of hydroxy propyl methyl cellulose (HPMC) films containing nanoparticles against 3 foodborne pathogens.

Study design: Data collection study.

Place and Duration: All experiments were done in Food Technology Department, Benha University, Egypt; Nanomaterial Laboratory, Beni-Suef University, Egypt and Agricultural Research Center, Egypt and were done within take 3 months.

Methodology: All methods were collected by different references such as preparation edible film, antimicrobial activity, mode of action, challenge study and the scanning electron microscope (SEM) and mechanical properties of HPMC films were evaluated.

Results: The results obtained from this study showed that in initial experiments, the nanoparticles (~80 nm) at 80 ppm were active against *Bacillus cereus*, *Staphylococcus aureus*, and *Salmonella* Typhimurium compared with 20 and 40 ppm. The HPMC films including Al₂O₃-NPs were active against *B. cereus* than *S. aureus* and *S. Typhimurium*, while the SiO₂-NPs were more effective against *S. Typhimurium* and *B. cereus* compared with *S. aureus*. In challenge studies, HPMC films including Al₂O₃-NPs and SiO₂-NPs at 80 ppm decreased the viability of the three were highly decreasing the 3 foodborne pathogens growth associated with chicken fillets stored at 4±1 °C for up to 15 days, as compared with the control sample. HPMC films incorporated with nanoparticles inhibited the microbial population ~ 2-3 log₁₀ CFU/cm² over the chicken fillet during storage period.

Conclusion: This work indicated that the results conducted that HPMC films incorporated with nanoparticles (~ 80 nm) at 80 ppm could be enhanced the safety of refrigerated chicken fillets.

Keywords: antimicrobial activity, HPMC edible film, nanoparticles, chicken fillets, cold storage.

INTRODUCTION

Foodborne pathogen are one of the important biological hazards which caused a lot of disease, harmful in food products, and lose much money (17). According to Center for Disease Control and

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38 Prevention report, food-borne diseases account for approximately 48 million illnesses, 128000
39 hospitalizations and 3000 deaths cases, as well costed 15.6 billion \$ each year in the United States
40 (10). Five foodborne pathogens record about (88%) of food poisons: *Norovirus* (26%), *Salmonella*
41 *nontyphoidal* (35%), *Campylobacter* (15%), *E. coli* (STEC) O157 (4%), and *Toxoplasma gondii* (8%).
42 Moreover, twenty food products recalled in which exposure occurred in one state such as apple cider,
43 bread, chicken, drink mix, ground beef, muffins, pork, raw tuna, and roast beef. (10).

44 Recently, nanotechnology have many applications in food sector particularly food industry, quality and
45 safety (3). These applications used to improve food safety and extend shelf-life of food products (6).
46 Nanoparticles one of the most shape utilized in food safety as antimicrobial and supplementation. As
47 well, inorganic nanoparticles as antimicrobial have taken more attention against food-borne
48 pathogens i.e. aluminum oxide nanoparticles (Al_2O_3 -NPs) and silica oxide nanoparticles (SiO_2 -NPs)
49 (16).

50 Al_2O_3 -NPs food grad are non-toxic, active against food-borne pathogens and permitted by FDA.
51 Al_2O_3 NPs at 1000 mg ml⁻¹ significantly affected against the *Escherichia coli* growth in ready to eat
52 foods (22). One study demonstrated Al_2O_3 -NPs incorporated with polyvinylidene fluoride films
53 reduced the *E. coli* growth (33). A study conducted by the author (29) reported that Other study
54 found aluminum oxide nanoparticles were active against *Salmonella* Typhimurium, *Listeria*
55 *monocytogenes*, *Fusarium oxysporum*, *Chromobacterium violaceum*, and *Aspergillus flavus* the result
56 obtained by (29).

57 Food grade SiO_2 -NPs are non- toxic, anticaking, has been used as food additive and permitted by
58 FDA (7). Oregano silane containing SiO_2 -NPs has been reported to prevented biofilm formation of
59 food-borne pathogens (14). SiO_2 -NPs reduce food-borne pathogens growth and make significate
60 changes in cell morphology such as *Salmonella enterica* (32).

61 Hydroxy propyl methyl cellulose (HPMC) edible film is approved by the by FDA for food packaging (21
62 CFR 172.8741). It has a good characters such as tasteless and odorless, transparent, and barrier
63 (31). As well, HPMC films including poly lactic acid and incorporated with green tea extract
64 nanoparticles improved shelf-life of fatty foods (34). Additionally, HPMC films contained TiO_2
65 nanoparticles was reported to inhibited *E. coli* and *S. aureus* growth (26).

66 In Egypt, the chicken products consumption is growing up nowadays. That is revert to high nutritional
67 value, available un expensive, and easy cooked, however, spoiled rapidly. The aim of this work was
68 to evaluate nanoparticles i.e. Al_2O_3 -NPs and SiO_2 -NPs antimicrobials against food-borne pathogens in
69 chicken fillets.

70 All over Goals

71 Improve the quality and safety of chicken fillets.

72 Development the packaging systems.

73 Extending the shelf-life of products.

74 Discovering a new antimicrobial.

75 2. MATERIALS AND METHODS

76 2.1. Bacterial strains

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77 Three bacterial strains utilized in this work were *Bacillus cereus* (ATCC 10876), *Staphylococcus*
78 *aureus* (ATCC 11988), and *S. Typhimurium* (ATCC 14028). The strains activated at Food Technology
79 Department, Benha University, Egypt. All strains were cultivated twice on Tryptic Soy Agar (TSB; Bio-
80 life company, Italy) at 37 °C for 24 h, and kept at 4 °C till using (18).

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81 2.2. Antimicrobials agents

82 Food-grade aluminum oxide nanoparticles (Al_2O_3 -NPs), and silica oxide nanoparticles (SiO_2 -NPs) at
83 (~80 nm) were obtained from Nanomaterial Laboratory, Beni-Suef University, Egypt.

84 2.3. Preparation of Hydroxy Propyl Methyl Cellulose (HPMC) films

85 Hydroxy propyl methyl cellulose films (HPMC) were prepared according to follow. Briefly, 4 % of
86 HPMC was dissolved in 100 mL distilled water at 70 °C with stirring at 1000 rpm/min for 2 h. A 1 mL
87 of glycerol 30% was added with stirring at 1000 rpm for 30 min. The nanoparticles were added and
88 stirred at 1000 rpm/min for 15 min. The solution was sterilized at (121°C/15 min). Then, casted and
89 dried, as well kept under cold storage till utilized (26).

90 2.4. Antimicrobial activity of nanoparticles against food-borne pathogens

91 Antimicrobial activity of nanoparticles was evaluated by disk diffusion method. In briefly, different
92 concentration of nanoparticles i.e. 20, 40 and 80 ppm against food-borne pathogens. Add 10µl from
93 bacterial strains. Then, 100µl from nanoparticles agent were added. Afterward, the dishes put in
94 incubator at 37°C for 48 h. At the end of incubation time clear zones were appeared and measured by
95 ruler (24).

Comment [RJW26]: Disk diffusion technique by Kirby Bawer. Indicate the medium used

96 2.5. Mode of nanoparticles action against bacterial strains

97 The mode of action was done according to (15) with slightly modification. Briefly, 2 ml of sterilized
98 Tryptic Soy Broth (TSB) were added. 1 ml of bacterial strain and 1 mL of antimicrobial were added.
99 After that, the tube was incubated the tubes overnight at 37°C for 24 h. Then, the pellets were
100 collected by centrifuge at 2500 rpm for 10 min. Then, examined by scanning electron microscope
101 after spread the cells onto a glass slices pre-washed with ethanol and acetone, and drying at 37 °C
102 for 15 min.

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103 2.6. Challenge study

104 Raw chicken fillets were purchased from local Cairo, Egypt. The fillets were transferred in ice box to
105 laboratory, and freshly used. The fillets were cut down (5 × 5 cm) sections under sterilized conditions.
106 Then, the samples treated with ultraviolet light (UV) for 15 min to decrease bacterial population.
107 Chicken fillets were inoculated overnight by aseptically diluted cultures of *S. Typhimurium*, *S. aureus*
108 and *B. cereus* approximately $5 \log_{10}$ CFU/cm² on the surface. After impregnation, the samples were
109 kept at room temperature for 20 min to allow cell attachment. Then, raw chicken fillets were coated
110 with HPMC films (5 × 5 cm) incorporated with nanoparticles. Control samples covered by control
111 HPMC films. After 0, 3, 6, 9, 12 and 15 days, the samples were tested to determine remain microbial
112 colonies. 1mL was spread plated in duplicate onto brilliant green agar for *S. Typhimurium*, paired
113 parker (M043) for *S. aureus*, *Bacillus cereus* agar base (M833) for *B. cereus* to demonstrate microbial
114 growth. Resulting colonies were counted after 24:48 h incubation at 37°C, populations measured by
115 \log_{10} , and expressed as \log_{10} CFU/cm² (21).

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Comment [RJW30]: Use 24 h

Comment [RJW31]: substrict

Comment [RJW32]: indicate temperature 27 °C

116 2.7. Scanning electron microscope (SEM) of HPMC films

117 Hitachi S-4700 scanning electron microscope (Hitachi, Toronto, Ontario, Canada) was used to study
118 the morphology of nanoparticles and films. The samples were deposited onto aluminum specimen
119 stubs using double-stick carbon tabs (Ted Pella Inc., Redding, CA, USA) and coated with
120 gold/palladium on an ion sputter coated (Denton Vacuum Inc., Moorestown, NJ, USA) for 45 s at 20
121 mA. All samples were examined using an accelerating beam at a voltage of 1.5 kV. Magnifications of
122 40,000x and 60,000x were used (11).

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123 2.8. Film solubility and thickness characterization

124 The solubility of films in water were studied. Thickness was determined by using digital micrometer
125 model 7326 (Mitutoyo Manufacturing, Tokyo, Japan) at 6 different positions on the film according to
126 (30).

127 2.9. Tensile of HPMC films determination

128 The tensile of films were determined by Texture Analyzer TA.XT2 (Stable Micro System, Surrey, UK),
129 according to the ASTM Standard Method D 88283 (initial grip separation = 50 mm and cross head
130 speed = 100 mm/min) according to (11).

131 2.10. Water vapor permeability

132 Water vapor permeability was evaluated by ASTM E96-92 gravimetric method with some
133 modifications to measure the relative humidity (RH) of HPMC films according to (12). Water vapor
134 permeability was calculated according to follow relation: $WVP = \frac{WVTR}{(P_2 - P_3) \cdot y}$

135 Where WVTR was obtained from the slope of the weight loss rate through the film surface and p^2 was
136 the water vapor partial pressure on the film underside. p^3 was water vapor partial pressure at the film
137 underside, y the average film thickness. Water vapor permeability of each film was measured as the
138 mean and standard deviations of 5 replications.

139 2.11. Gases vapor permeability (O₂ and CO₂)

140 The gas vapor permeability was determined at 30°C in a designed stainless cell by gas testing
141 instrument, model Witt Oxybaby headspace gas analyzer (O₂/CO₂) following the method described by
142 following equation: $P = \frac{Q \cdot X}{A \cdot t \cdot \Delta p}$

143 The gas permeability (P) was calculated according to (12).

144 Where, P is the permeability of gas, (m³/m. day. mmHg), Q is the quantity of gas diffused m³, X is the
145 thickness of film, A an area of the film, m², t is the time, day and Δp is the pressure difference across
146 the film.

147

148 2.12. Statistical analysis

149 The challenge study, statistical analyses for bacterial growth were carried out utilizing one-way
150 ANOVA with a significant value of $P \leq 0.05$ by using SPSS software, var. 18 (IBM; Armonk, N.Y.,
151 U.S.A.). Results were analyzed as a completely randomized design according to (28). All challenge

152 experiments were performed in triplicate, using 3 samples per treatment. Multiple comparisons were
153 carried out applying least significant difference and Tukey's test.

154 3. RESULTS AND DISCUSSION

155 3.1 Antimicrobial activity of nanoparticles against food-borne pathogens

156 As shown in **Table 1 and 2**. The antibacterial activity of inorganic nanoparticles i.e. aluminum oxide
157 nanoparticles (Al_2O_3 -NPs) and silica oxide nanoparticles (SiO_2 -NPs) against food-borne pathogens
158 such as *Bacillus cereus*, *Salmonella* Typhimurium and *Staphylococcus aureus* were evaluated. **The**
159 **result showed that Results-conducted-that** Al_2O_3 -NPs and SiO_2 -NPs (~80 nm) at 80 ppm were
160 effective against food-borne pathogens i.e. *B. cereus*, *S. Typhimurium* and *S. aureus*, than 20 and 40
161 ppm respectively, as reported by (9). Moreover, Al_2O_3 -NPs were more active against *B. cereus* and
162 *S. aureus* than *S. Typhimurium* that is agreement with (13). In addition, SiO_2 -NPs were more active
163 against *B. cereus*, and *S. Typhimurium* compared *S. aureus* that is partially agreement with (14).
164 The results found that the Al_2O_3 -NPs were more active against spores and gram positive than gram
165 negative bacteria, while SiO_2 -NPs more effective against gram negative and spores compared with
166 gram positive bacteria. The results are agreement with data reported by (4).

167 Furthermore, according to **Table 3**, the effect of hydroxy propyl methyl cellulose (HPMC) edible films
168 incorporated with nanoparticles were reduced *B. cereus*, *S. aureus* and *S. Typhimurium* population
169 growth. The results showed that Al_2O_3 -NPs were inhibited *B. cereus* and *S. aureus* growth than *S.*
170 *Typhimurium*. Although, SiO_2 -NPs less effective against *S. aureus* than *B. cereus*, and *S.*
171 *Typhimurium*. The results agreement with data reported by (5).

172 3.2 Mode of action nanoparticles against foodborne pathogens

173 Based on the results of nanoparticles activity against food-borne pathogens, the mode of action it
174 seems necessary. **Fig. 1**, illustrated that Al_2O_3 -NPs were highly effective against gram positive than
175 gram negative bacteria, this is reverting to the Al_2O_3 -NPs action as follows, Al_2O_3 -NPs interact with
176 bacteria membrane and made changes in cell morphology such as (a) the formation of 'pits' in their
177 cell wall. Moreover, made disruption and drastic in cell wall. (b) As well, it produces reactive oxygen
178 species (ROS) which allow to penetrate the cell membrane and led the cell to death. (c) Moreover,
179 causes cell oxidative stress and formed free-radical scavenging that is led the bacteria to die that is
180 reported by (19).

181 In addition to, SiO_2 -NPs more effective against gram negative and spores than gram positive bacteria.
182 That is due to (a) the ability of SiO_2 -NPs to make morphological changes, lose the cell to preform it in
183 function role. (b) As well, reactive oxygen spices (ROS) generation, and lose the DNA function and
184 led to damage. (c) Additionally, cause the oxidative stress regulation in gens according to (16).

185 3.3 Challenge study

186 Based on the results of antimicrobial activity of HPMC films incorporated with nanoparticles, the films
187 were utilized to cover raw chicken fillets at $4\pm 1^\circ\text{C}$ up to 15 days. **Fig. 2, 3, and 4**, demonstrated that
188 the bacterial population was gradually increase during the storage period over 15 days, when used
189 control films compared with the nanoparticles films. HPMC films including nanoparticles reduced the
190 food-borne pathogens growth approximately $2:3 \log_{10}$ during the challenge study.

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191 HPMC films include SiO₂-NPs were stronger antimicrobial against *B. cereus*, *S. Typhimurium* and *S.*
192 *aureus* than Al₂O₃-NPs on raw chicken fillets, these results are agreement with (19), (29) and (25).

193 3.4 Scanning electron microscope of HPMC films including nanoparticles agent

194 **Fig. 5** showed that, the cross sections and surface appearance of the control film, which appear to be
195 homogeneous, smooth, colorless and free of any dimples or crevices. The HPMC films incorporated
196 with nanoparticles were completely dispersion. Al₂O₃-NPs and SiO₂-NPs loaded films show no pores
197 with smooth surface. The presence of these pores is likely due to the flocculation and coalescence of
198 small drops during film preparation. Also, the nanoparticles distribution were found to be
199 homogeneous in all films according to (1).

200 3.5 Mechanical properties of films

201 As shown in **Table. 4**, the tensile, water vapor permeability oxygen vapor permeability and carbon
202 dioxide vapor permeability were evaluated, HPMC films containing SiO₂-NPs were the highest values
203 compared with HPMC films control and Al₂O₃-NPs films in mechanical properties. Additionally, SiO₂-
204 NPs increased the films water vapor permeability, carbon dioxide vapor permeability, tensile, oxygen
205 vapor permeability and formed strong structure of films. That is due to (a) the ability of SiO₂-NPs to fill
206 the pores between the HPMC films structure (b) HPMC diffusion with SiO₂-NPs and form
207 homogenized structure (c) the ration of glycerol and it is ability to prevent water evaporation. As well,
208 Al₂O₃-NPs were the lowest values and formed a weak structure, that is revert to the Al₂O₃-NPs can
209 not interference with HPMC films and there is heterogenous distribution. In the control HPMC films,
210 the transparence and thickness, was the lowest values than Al₂O₃-NPs and SiO₂-NPs films. That is
211 refer to the color of nanoparticles and nanoparticles doses in films solution. Regarding solubility, there
212 are non-significant results between HPMC films control and HPMC films including nanoparticles,
213 these data agreement (2), (23) and (27).

214 4. CONCLUSION

215 The results of this investigation ~~were~~ demonstrated that HPMC films including Al₂O₃-NPs and SiO₂-
216 NPs were active against food-borne pathogens such as *S. Typhimurium*, *B. cereus* and *S. aureus* in
217 chicken fillets. Additionally, nanoparticles (~80 nm) at 80 ppm showed a significant inhibition
218 compared with 20 and 40 ppm respectively. Moreover, SiO₂-NPs ~~has~~ are stronger antimicrobial
219 ~~activity~~ against food-borne pathogens than Al₂O₃-NPs. However. HPMC films incorporated with SiO₂-
220 NPs had a better mechanical property than HPMC films included Al₂O₃-NPs. HPMC films containing
221 nanoparticles ~~have~~ ~~ia~~ longer ~~recreasing the~~ shelf-life ~~proprerty~~ and ~~improve~~ ~~the~~ chicken fillets safety
222 and quality.

223

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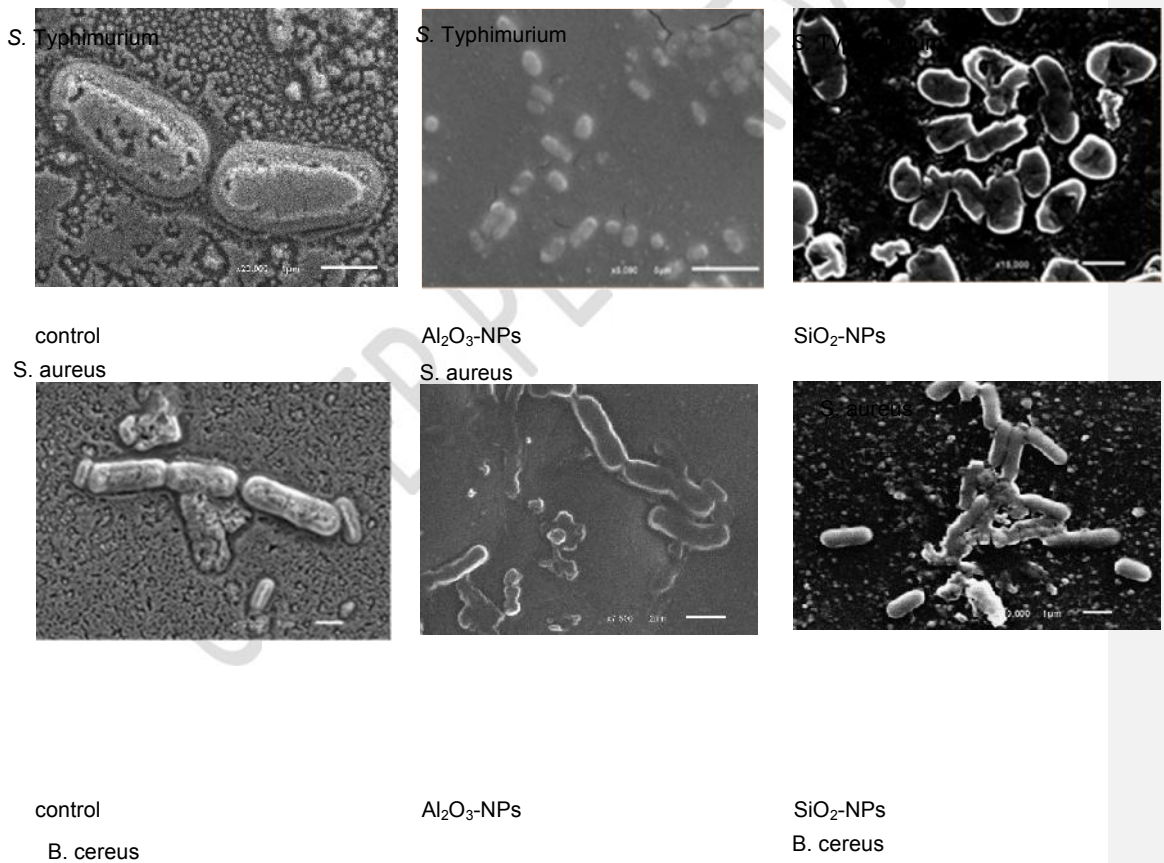
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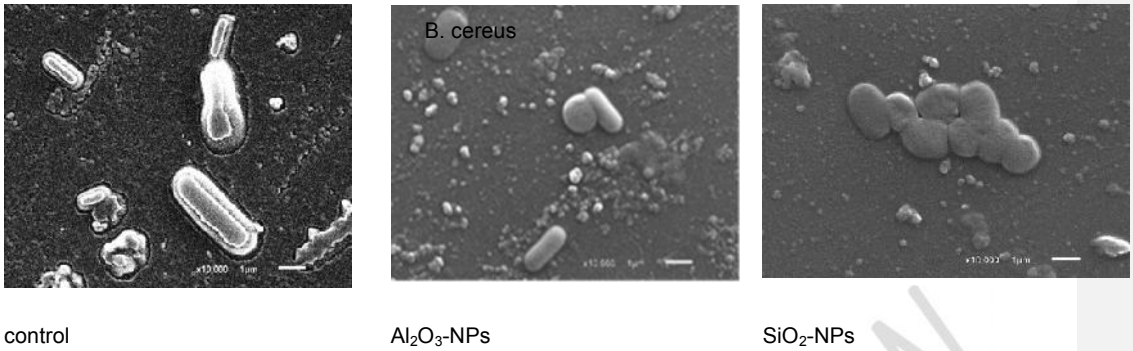
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 347 Fig.1 The mode of action of nanoparticles against foodborne pathogens using SEM.

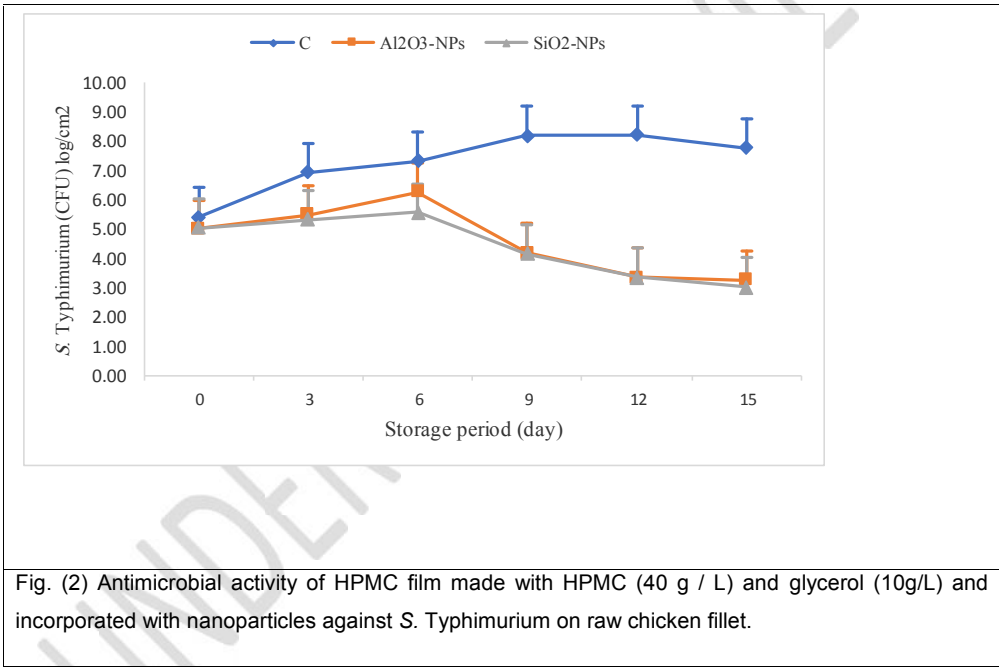


Fig. (2) Antimicrobial activity of HPMC film made with HPMC (40 g / L) and glycerol (10g/L) and incorporated with nanoparticles against S. Typhimurium on raw chicken fillet.

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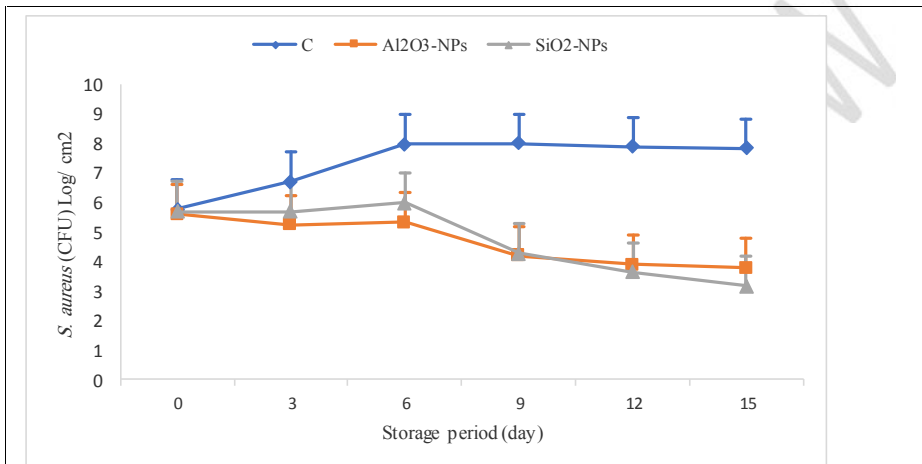


Fig. (3) Antimicrobial activity of HPMC film made with HPMC (40 g / L) and glycerol (10g/L) and incorporated with nanoparticles against *S. aureus* on raw chicken fillet.

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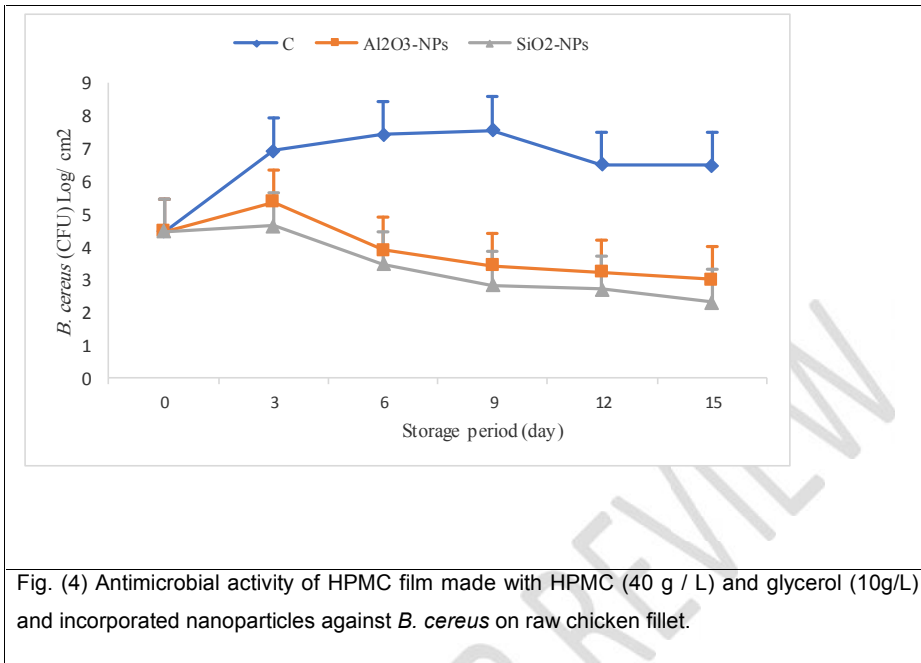
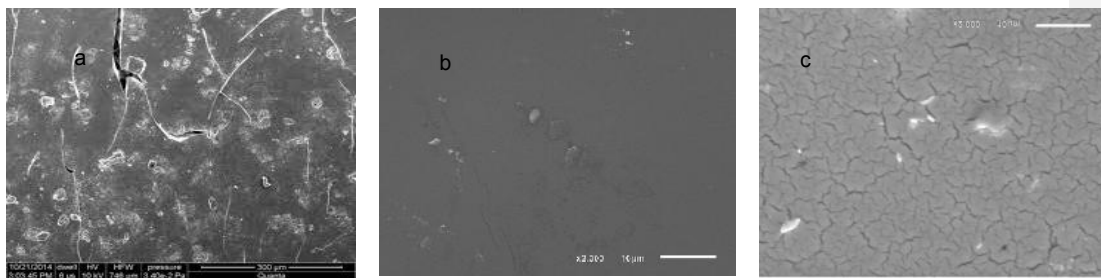


Fig. (4) Antimicrobial activity of HPMC film made with HPMC (40 g / L) and glycerol (10g/L) and incorporated nanoparticles against *B. cereus* on raw chicken fillet.

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control

Al₂O₃-NPs

SiO₂-NPs

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394 Fig. (5) The SEM of (a) HPMC films incorporation (b) Al₂O₃-NPs and (c) SiO₂-NPs.

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412 Table (1). Antibacterial activity of Al₂O₃-NPs and SiO₂-NPs nanoparticles (~80 nm) at different

413 concentration against foodborne pathogens.

| Bacterial strains | Al ₂ O ₃ -NPs | | | SiO ₂ -NPs | | |
|-----------------------|-------------------------------------|--------|--------|-----------------------|--------|--------|
| | 20 ppm | 40 ppm | 80 ppm | 20 ppm | 40 ppm | 80 ppm |
| <i>S. Typhimurium</i> | 9±0.3 | 11±0.3 | 13±0.2 | 11±0.3 | 15±0.2 | 18±0.3 |
| <i>S. aureus</i> | 8±0.3 | 12±0.3 | 14±0.3 | 12±0.3 | 13±0.3 | 16±0.3 |
| <i>B. cereus</i> | ND | 12±0.3 | 15±0.3 | 13±0.3 | 15±0.3 | 18±0.3 |

414

415 ND: Not Detect

416 Al₂O₃-NPs: Aluminum oxide nanoparticles

417 SiO₂-NPs: Silica oxide nanoparticles

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419

420 Table (2). Antibacterial activity of Al₂O₃-NPs and SiO₂-NPs nanoparticles (~80 nm) at 80 ppm against

421 foodborne pathogens.

| Bacterial strains | Nanoparticles agents | |
|-------------------|-------------------------------------|-----------------------|
| | Al ₂ O ₃ -NPs | SiO ₂ -NPs |
| S. Typhimurium | 13±0.2 | 18±0.3 |
| S. aureus | 14±0.3 | 16±0.3 |
| B. cereus | 15±0.3 | 18±0.3 |

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423 Al₂O₃-NPs: Aluminum oxide nanoparticles

424 SiO₂-NPs: Silica oxide nanoparticles

425

426 Table (3). Antibacterial activity of HPMC film incorporation with nanoparticles (~80 nm) at 80 ppm

427 against foodborne pathogens.

| Bacterial strains | HPMC films incorporation nanoparticles | |
|-------------------|--|-----------------------|
| | Al ₂ O ₃ -NPs | SiO ₂ -NPs |
| S. Typhimurium | 16±0.2 | 22±0.4 |
| S. aureus | 17±0.3 | 20±0.3 |
| B. cereus | 18±0.3 | 22±0.4 |

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429 HPMC: Hydroxy propyl methyl cellulose

430 Al₂O₃-NPs: Aluminum oxide nanoparticles

431 SiO₂-NPs: Silica oxide nanoparticles

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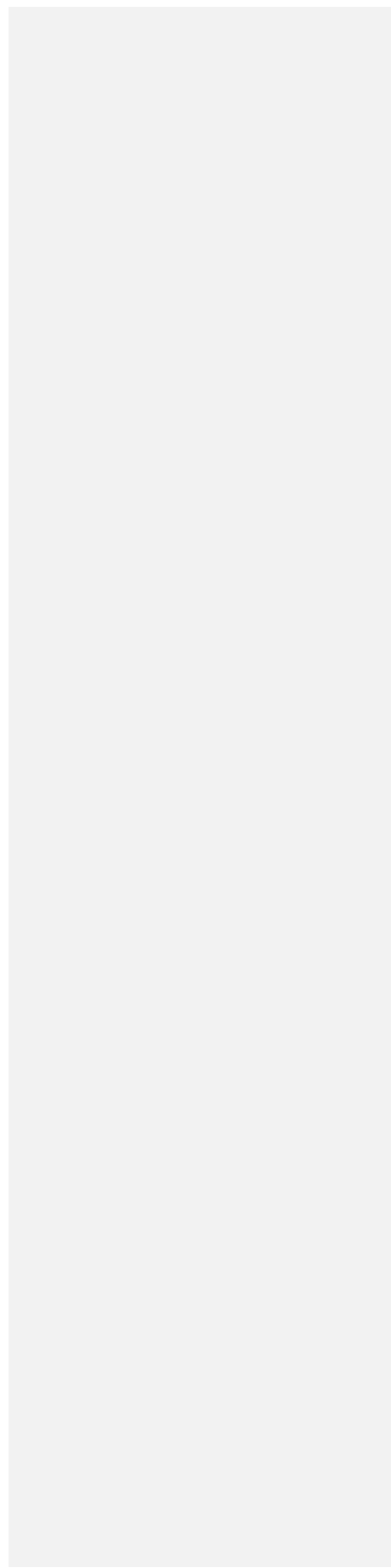
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UNDER PEER REVIEW



444 Table (4). Physical and mechanical properties of HPMC films incorporated with Al₂O₃-NPs and SiO₂-NPs

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| samples | Properties (tests results) | | | | | | |
|---|----------------------------|---|---|--|--------------|-----------|------------|
| | Tensile (MPa) | Water vapor permeability (g mm K ⁻¹ Pa ⁻¹ h ⁻¹ m ⁻²) | O ₂ vapor permeability P (ml mm cm ⁻² s ⁻¹ cm Hg ⁻¹) | Co ₂ vapor permeability P (ml mm cm ⁻² s ⁻¹ cm Hg ⁻¹) | Transparence | Thickness | Solubility |
| control | 38.1 | 0.108 | 0.188×10 ⁻⁸ | 2.25×10 ⁻⁹ | 0.065 | 0.5 mm | 100% |
| HPMC- Al ₂ O ₃ -NPs | 31.6 | 0.056 | 1.074×10 ⁻⁸ | 1.44×10 ⁻⁹ | 0.079 | 0.5mm | 100% |
| HPM -SiO ₂ -NPs | 43.17 | 0.541 | 2.17×10 ⁻⁸ | 14.4×10 ⁻⁹ | 0.082 | 0.51 mm | 100% |

446 HPMC: Hydroxy propyl methyl cellulose

447 Al₂O₃-NPs: Aluminum oxide nanoparticles

448 SiO₂-NPs: Silica oxide nanoparticles

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