

1 **EFFECTS OF BLACK SEED (*NIGELLA SATIVA*) AND UZIZA LEAF (*PIPER***
2 ***GUINEENSE*) ON ELECTROLYTES, UREA AND CREATININE OF WISTAR ALBINO**
3 **RATS**

4 **ABSTRACT**

5 **Aim:** For centuries, plant and plant products have played a pivotal role in medication. This study
6 evaluated the effect of aqueous extract of black seed (*Nigella sativa*) and uziza leaf (*Piper*
7 *guineense*) on electrolytes, urea and creatinine of Wistar rats.

8 **Materials and method:** Twenty-five Wistar rats were used for the study, the rats were arranged
9 into five groups with five rats in each of the groups. The rats had access to their normal feed but
10 sucrose and margarine were used to induce hyperglycemia and hyperlipidemia respectively on
11 the rats with the exception of the rats in the positive control group. The rats in the negative
12 control were induced using the sucrose and margarine but were not treated using the aqueous
13 extracts. The rats in the uziza group were treated with 2ml of uziza aqueous leaf extract, while
14 the rats in the black seed group were treated with 2ml of black seed aqueous extract. The rats in
15 the black seed **and** uziza leaf group were treated with 2ml of the combined aqueous extract.

16 **Results:** The results showed that the extracts had a decreasing effect which was time dependent
17 on the electrolytes. The highest decrease was obtained on the third week of feeding when
18 compared to the control ($p=0.05$). The sodium levels (mmol/L) showed for the negative control
19 (198.23 ± 1.96), positive control (108.15 ± 1.60), uziza leaf (98.28 ± 4.17), black seed ($101.67 \pm$
20 4.24), black seed **and** uziza (90.83 ± 2.14). The decrease for potassium levels (mEq/L) showed
21 for the negative control (0.90 ± 0.06), positive control (0.05 ± 0.10), uziza leaf (0.07 ± 0.18),
22 black seed (0.06 ± 0.19), black seed **and** Uziza (0.05 ± 0.10). Furthermore, the extracts also had a
23 reducing effect on urea and creatinine levels with the highest reduction obtained on the third
24 week ($p=0.05$). The urea levels (mmol/L) showed for the negative control (26.84 ± 0.05),
25 positive control (15.15 ± 1.20), uziza leaf (12.83 ± 0.98), black seed (12.16 ± 2.01), black seed
26 **and** uziza (11.48 ± 1.78). The extracts also decreased creatinine levels (mmol/L) with the
27 negative control (284.58 ± 0.33), positive control (182.73 ± 3.67), uziza leaf (194.16 ± 18.30),
28 black seed (167.34 ± 14.66), black seed **and** uziza (174.46 ± 10.66).

29 **Conclusion:** The extracts significantly decreased the elevated electrolytes levels and therefore
30 uziza leaf and black seed can be used to restore kidney function.

31 **Keywords:** Creatinine, Electrolytes, *Nigella sativa*, *Piper guineense*, Potassium, Sodium, Urea,

32 1. INTRODUCTION

33 Naturally existing plants have been found to contain varieties of chemical substances which are
34 of paramount importance to the medical world [1]. The use of natural products with therapeutic
35 properties is as ancient as human civilization and for a long time, mineral, plant, and animal
36 products were the main sources of drugs for therapeutic purpose [2]. Plants have always been a
37 major source of nutrition and health care for both humans and animals. In recent years, there has
38 been a growing interest in alternative therapies and the therapeutic use of natural products,
39 especially those derived from plants [3].

40 *Nigella sativa* is commonly known as Black cumin in traditional medicine. *Nigella sativa* is a
41 spice plant that is widely used for prevention and treatment of many ailments in many countries
42 worldwide. It has been shown that the biological activity of the black cumin is related to the
43 composition of its essential oil, which contains 30 to 48% of thymoquinone, 7 to 15% P-cymene,
44 6 to 12% carvacrol, 2 to 7% 4-terpineol, 1 to 4% Tanethole and 1 to 8% Sesquiterpene [4, 5].
45 Pharmacologically, thymoquinone and its derivatives are the most important components of
46 Uziza or black pepper (*Piper guineese*) is a flowering plant in the family Piperaceae. The fruit,
47 known as a peppercorn when dried, is a small drupe, five millimeters in diameter, dark red when
48 fully matured, containing a single seed. It is a native to India and long been considered the
49 world's most important spice. It is cultivated for its fruit, which is usually dried and used as a
50 spice and seasoning [1] and also as preservative [6]. It is one of the most common spice in the
51 European cuisine and has been known and prized since antiquity for both its flavor and its
52 medicine [7]. Black pepper has been used to flavor foods for over 3000 years. The same fruit is
53 also used to produce white pepper and green pepper [8]. The plant has a fruit which contains
54 angular black seeds, and the seeds are considered to be the most valuable part contributing
55 beneficial health effects. *Nigella sativa* as a natural remedy has been documented to possess
56 numerous therapeutic values, including diabetes, tumour, hypercholesterolemia, hypertension,
57 inflammation, and gastrointestinal disorders [9, 10].

58

59 In recent years, there has been a growing interest and demand in using medicinal plants for
60 treating and preventing various diseases including cardiovascular diseases. Traditional medicines

61 of plants origin have received much attention due to several factors such as easy availability,
62 affordable cost, safety, and efficacy as well as cultural acceptability. Uziza leaf (*Piper guineense*)
63 is an African plant with its leaves having a peppery taste and green when fresh and darker green
64 when dried. *Piper guineense* contains over 700 species all over the world. It is a local spice
65 mainly used in Nigerian dishes and it known to provide nutritional, culinary, insecticidal and
66 medicinal benefits. It contains proteins, carbohydrates, alkaloids, steroids, glycosides, saponins,
67 flavonoids, tannins and phenolic compounds; also vitamins, minerals and fat. The
68 pharmacological properties of *N. sativa* is attributed to several component including proteins,
69 amino acids, carbohydrates, fibers, oils (combination of fatty acids, especially polyunsaturated
70 fatty acids), volatile oil (frequently thymoquinone), mineral, alkaloids, flavonoids, saponins, and
71 others [11, 12, 13].

72 Uziza leaves have a peppery taste, are pale greenish color when fresh and darker green when
73 frozen or dried. The inflorescence is a pedicel flower spike between 3 and 6 cm long and the
74 peduncle 5 mm long. Flowers are greenish yellow and arranged in a spiral along the spine [14].
75 The fruits of *P. guineense* occur in clusters, small, reddish or reddish brown when ripe and black
76 when dry. The fruit is a **drupe, oval**, 5mm in diameter [15, 16, 17].

77 *P. guineense* have been characterized and their chemical composition determined. They are used
78 as therapeutic agents in minor ailments [18]. Phytochemicals are not vitamins or minerals but are
79 bioactive compound found in plant foods that work with nutrient and dietary fibers to protect
80 against disease [19]. The presence of phytochemicals like alkaloids in both the leaves and seed
81 extracts of *P. guineense* signified the possession of medicinal properties within the plant. The
82 flavonoids possess antioxidant, anti-inflammatory, anti-tumor, anti-allergic and antiplatelet
83 properties [20]. They are also found to have cholesterol lowering ability. Alkaloids which are
84 natural products present in *P. guineense* are made up of heterocyclic nitrogen that has anti-
85 malarial, antihypertensive, antiarrhythmic and anticancer properties. Alkaloids are being used as
86 central nervous system stimulant, powerful pain relievers, topical anesthetic in ophthalmology
87 among others [21]. Tannins are compounds with proline-rich proteins that help to inhibit the
88 absorption of iron when present in the gastrointestinal lumen thus reducing the bioavailability of
89 iron due to the presence of compounds that help in the treatment of diseases like enteritis,
90 gastritis, and esophagitis. Plants that contain tannins as their primary component are astringent,

91 thus very beneficial for the management of diarrhea, dysentery, inflammation of the mucous
92 membrane [22]. Saponins have anti-carcinogenic properties and may also play an important role
93 in antimalarial activity of plants. *P.guineense* also contains cardiac glycosides in a significant
94 amount and cardiac glycosides are useful in the management of diseases associated with the
95 heart. *P.guineense* also contains dillapiol, 5-8% of piperine, elemicine, 10% of myristicine and
96 safrole and these chemicals exhibit bactericidal and antimicrobial effects on some micro-
97 organisms [23]. *P. guineense* like other members of the piper family contains 5-8% of the
98 chemical “piperine” which gives them their “heat”. They also contain large amounts of beta-
99 carophyllene which is being investigated as an anti-inflammatory agent [24].

100

101 The kidneys are two bean-shaped organs in the renal system. They help the body pass waste as
102 urine. They also help filter blood before sending it back to the heart. The kidneys perform many
103 crucial functions, including: maintaining overall fluid balance, regulating and filtering minerals
104 from blood, filtering waste materials from food, medications, and toxic substances, creating
105 hormones that help produce red blood cells, promote bone health, and regulate blood pressure.

106 Blood urea nitrogen (BUN) provides a rough measurement of the glomerular filtration rate, the
107 rate at which blood is filtered in the kidneys. Urea is formed in the liver as an end product of
108 protein metabolism and is carried to the kidneys for excretion. Nearly all kidney diseases cause
109 inadequate excretion of urea, elevating BUN levels in the blood. (Other causes of high BUN
110 levels include gastrointestinal bleeding and steroid treatment). It can be done to determine the
111 amount of urea nitrogen in the blood [25].

112 Creatinine is a breakdown product of creatine, an important component of muscle. The
113 production of creatinine depends on muscle mass, which varies very little. Creatinine is excreted
114 exclusively by the kidneys, and its level in the blood is proportional to the glomerular filtration
115 rate. The serum creatinine level (serum is the clear liquid that remains after whole blood has
116 clotted) provides a more sensitive test of kidney function than BUN because kidney impairment
117 is almost the only cause of elevated creatinine. It can also be measured with a urine test.
118 Creatinine clearance rate determines how efficiently the kidneys are clearing creatinine from the
119 blood and serves as an estimate of kidney function. For renal function test, urine and serum

120 levels of creatinine are measured, as well as the volume of urine excreted over a 24-hour period.
121 The creatinine clearance rate is then calculated and expressed as the volume of blood, in
122 milliliters, that can be cleared of creatinine in 1 minute. A low creatinine clearance value
123 indicates abnormal kidney function. It requires both a urine and blood sample [25].

124 Sodium is one of the body's electrolytes, which are minerals that the body needs in relatively
125 large amounts. Electrolytes carry an electric charge when dissolved in body fluids such as blood.
126 Most of the body's sodium is located in blood and in the fluid around cells. Sodium helps the
127 body keep fluids in a normal balance (see About Body Water). Sodium plays a key role in
128 normal nerve and muscle function. The body obtains sodium through food and drink and loses it
129 primarily in sweat and urine. Healthy kidneys maintain a consistent level of sodium in the body
130 by adjusting the amount excreted in the urine. When sodium consumption and loss are not in
131 balance, the total amount of sodium in the body is affected. The concentration of sodium in the
132 blood may be too low (hyponatremia) or too high (hypernatremia) [25]

133 Potassium is one of the body's electrolytes, which are minerals that carry an electric charge when
134 dissolved in body fluids such as blood. Most of the body's potassium is located inside the
135 cells. Potassium is necessary for the normal functioning of cells, nerves, and muscles. The body
136 must maintain the potassium level in blood within a narrow range. A blood potassium level that
137 is too high (hyperkalemia) or too low (hypokalemia) can have serious consequences, such as an
138 abnormal heart rhythm or even stopping of the heart (cardiac arrest). The body can use the large
139 reservoir of potassium stored within cells to help maintain a constant level of potassium in blood.

140 The body maintains the right level of potassium by matching the amount of potassium consumed
141 with the amount lost. Potassium is consumed in food and drinks that contain electrolytes
142 (including potassium) and lost primarily in urine. Some potassium is also lost through the
143 digestive tract and in sweat. Healthy kidneys can adjust the excretion of potassium to match
144 changes in consumption. Some drugs and certain conditions affect the movement of potassium
145 into and out of cells, which greatly influences the potassium level in blood [25].

146 The main aim of this study is to determine the effect of black seed (*Nigella sativa*) and uziza leaf
147 (*Piper guineense*) on kidney profile (urea and creatinine) and electrolyte (sodium and potassium)
148 of sucrose induced hyperglycaemia and margarine induced hyperlipidemia on Wistar albino rats.

149

150 2. MATERIALS AND METHODS

151 Reagent kits were bought from Randox Laboratories Ltd. Ardmore, Diamond Road, Crumlin,
152 Co. Antrim, United Kingdom BT29 4QY.

153 2.1. Experimental Animal and Design

154

155 Twenty five Wistar rats were purchased from the Biochemistry animal house in Choba
156 University of Port Harcourt. The mean weight was 150 ± 10 g. The experimental animals were
157 grouped into 5 groups with 5 rats in each group and the method of feed was by gavaging. The
158 animals were acclimatized for one week.

159 **GROUP 1:** this group served as the positive control. This group had access to normal feed (ad
160 libitum). They were not induced with sucrose and margarine. Furthermore, they were not treated
161 with Uziza leaf and black seed extracts.

162 **GROUP 2:** this group served as negative control, it had 5 rats fed with normal feed (ad libitum)
163 and distilled water but was induced with sucrose and margarine without treatment with either
164 black seed or uziza leaf extract.

165 **GROUP 3:** this group contained 5 rats fed with normal feed (ad libitum) and distilled water, was
166 induced with sucrose and margarine but treated with aqueous extract of black seed.

167 **GROUP 4:** this group contained 5 rats fed with normal feed (ad libitum) and distilled water was
168 induced with sucrose and margarine but treated with aqueous extract of uziza leaf.

169 **GROUP 5:** this group contained 5 rats fed with normal feed (ad libitum) and distilled water was
170 induced with sucrose and margarine but treated with equal proportion of the uziza leaf and black
171 seed aqueous extracts.

172 .

173 **2.2. Sample Preparation**

174

175 The black seed (*Nigella sativa*) was bought from a local market in Kaduna State, Nigeria, while
176 the uziza leaf (*Piper guineense*) was obtained from a compound in Choba, Obio-Akpor Local
177 Government area, Rivers State, Nigeria. The plants were identified as *Nigella sativa* and *Piper*
178 *guineense* by a staff of the Department of Plant Science and Biotechnology, Faculty of Sciences
179 University of Port Harcourt

180 50g of each of the samples; Uziza leaf (*Piper guineense*) and black seed (*Nigella sativa*), was
181 soaked in 500ml of distilled water. After the stock preparation using a syringe, 2ml of the
182 aqueous extract solution was collected and administered to the animals once daily. Also the feed
183 used was formulated thus;

184

185

186 **Table 1: Feed formulation table**

	COMPOSITION BY WEIGHT (g)	COMPOSITION IN PERCENTAGE (%)
Normal feed	500	50
Margarine	200	20
Sucrose	200	20
Vitamin	100	10

187

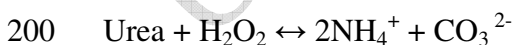
188 **2.3. Blood Collection**

189 The animals after inducement with sucrose and margarine for one month were treated and
190 sacrificed on a weekly bases. A desiccator with chloroform soaked cotton wool was used to
191 weaken each of the animal put inside after some minutes, when properly anaesthetized it was
192 brought out of the desiccator and dissected, some of the blood was put into a heparin bottle,
193 fluoride oxalate and ethylene diamine tetra acetic acid (EDTA) bottle according to the
194 parameters in consideration, the organs were also taken and put in a sterile bottle and all taken to
195 the laboratory for analysis.

196 **2.4. Determination of blood urea**

197 Urease-glutamate dehydrogenase -UV method according to Berthelot's method [26] was used to
198 determine the level of Urea in the samples. Mindray test kits was used for the analysis.

199 Reaction Principle



202 Urea is hydrolyzed by urease, and one of the products, ammonia, oxidises NADH to NAD⁺
203 catalysed by glutamate dehydrogenase (GLDH). The absorbance decrease is directly
204 proportional to the concentration of urea.

205 Procedure

206 Two test tubes labeled T1 (reagent blank) and T2 (test sample) were set up. T1 contained 1000
207 μL of reagent (R1) and 10 μL of distilled water, while T2 contained 1000 μL of reagent (R1) and
208 10 μL of test sample. The contents of each tube were mixed and incubated at 37°C for 2 min.
209 After incubating, 250 μL of the second reagent (R2) was added to both test tubes. The contents
210 of each tube was incubated again for 30 seconds at 37°C, the absorbance was read after 2 minutes
211 at a wavelength of 546 nm.

212 Calculation

213 $\Delta A = [\Delta A \text{ sample}] - [\Delta A \text{ blank}]$

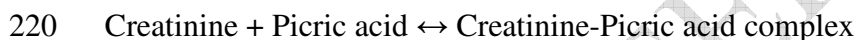
214 $\text{Conc. of urea} = [\text{change in absorbance of sample}] - [\text{change in absorbance of blank}]$.

215 The result is expressed in mmol/L.

216 **2.5. Determination of blood creatinine**

217 Modified Jaffé method according to Bartels and Bohmer [27] was used to determine the level of
218 creatinine in the samples. Mindray test kits was used for the analysis.

219 Reaction Principle



221 At an alkaline solution, creatinine combines with picric acid to form an orange-red colored
222 complex. The absorbance increase is directly proportional to the concentration of creatinine.

223 Procedure

224 Two test tubes labeled T1 (reagent blank) and T2 (test sample) were set up. T1 contained 180 μL
225 of reagent (R1) and 18 μL of distilled water, while T2 contained 180 μL of reagent (R1) and 18
226 μL of test sample. The contents of each tube were mixed thoroughly at 37°C for 1 min. After
227 incubating, 180 μL of the second reagent (R 2) was added to both test tubes. The content of the
228 tube was mixed thoroughly, incubated at 37°C for 30 seconds and the absorbance read at 492 nm
229 wavelength 2 minutes later.

230 Calculation

231 $\Delta A = [\Delta A \text{ sample}] - [\Delta A \text{ blank}]$

232 $\text{Conc. of creatinine} = [\text{change in absorbance of sample}] - [\text{change in absorbance of blank}]$.

233 The result is expressed in mmol/L.

234

235

236 **2.6. Determination of blood sodium**

237 Sodium levels were determined by colorimetric test. Magnesium-uranyl acetate method. The
238 Principle of this method is that after the precipitation of sodium magnesium-uranyl acetate, in
239 the supernatant form with uranyl ions in solution with thioglycolic acid a yellow-brown
240 coloured complex is formed. The optical density difference between the reagent blank
241 (without precipitation of sodium) and the result of the analysis is proportional to the sodium
242 concentration [28]. Reagent A kit contained uranylacetate (19mM) and magnesium acetate
243 (140mM) while reagent B kit contained ammonium thioglycolate (550mM), ammonia
244 (550mM) and the standard aqueous solution of sodium equivalent 150mmol. 2.00ml of reagent
245 A was mixed with 0.02 ml of the sample. For the standard, 2.00 ml of reagent A and 0.02 ml
246 of the standard were mixed. The mixtures were let to stand for 5 minutes, they were then
247 shaken thoroughly for 30 seconds. The mixtures were allowed to stand for 30 minutes. They
248 were centrifuged at 2,000rpm for 5 minutes. The supernatant was then separated. 0.05ml of
249 the clear supernatant was mixed with 2.00ml of reagent B. For the blank, 0.05 ml of reagent
250 A and 2.00 ml of reagent B were mixed, while the standard tube contained 0.05 ml of
251 supernatant and 2.00ml of reagent B. The absorbance of the mixtures was read after 10
252 minutes at 405nm with spectronic – 20 spectrophotometer.

253 Calculations: $\frac{\text{Blank O.D} - \text{Sample O.D}}{\text{Blank O.D} - \text{Standard O.D}} \times 150 = \text{mmol/L}$

254 $\text{Blank O.D} - \text{Standard O.D}$

255

256 **2.7. Determination of potassium**

257 Potassium levels were determined by colorimetric endpoint method [29]. One millilitre of
258 reagent was mixed with 0.1ml of sample except for the controls, which had no samples. The
259 blank tube contained 1.0ml of reagent while the standard tube contained 1.0ml of reagent and
260 0.1ml of standard. The mixtures were incubated at 25°C for 3mins. The absorbance was read
261 against reagent blank at 500nm with Spectronic -20 spectrophotometer.

262

263 Calculations: $\frac{\Delta A_{\text{unknown}}}{\Delta A_{\text{standard}}} \times C_{\text{standard}} = \text{potassium concentration mEq/L}$

264 $\Delta A_{\text{standard}}$

265

266

267

268 **2.8. Statistical Analysis**

269 Data analysis was performed using the Statistical package for the Social Sciences software
270 (SPSS, version 11.0). Data is displayed in mean \pm SD. The statistical method of one way analysis
271 of variance (ANOVA) was used to compare the mean values obtained among different groups.
272 Differences were considered significant whenever the p-value is $p=0.05$.

273

274 **3. RESULTS**

275
276 **TABLE 2: WEIGHT OF THE RATS BEFORE AND AFTER ADMINISTRATION OF**
277 **THE EXTRACTS**

Groups	Weight before administration (g)	Weight after administration (g)	Body weight change (g)
Negative control	66.33±13.22	117.18±20.79	50.85
Positive control	129.92±2.02	141.62±5.39	11.70
Uziza leaf	121.25±2.00	112.33±2.79	8.92
Black seed	105.53±0.19	81.54±4.14	23.94
Black seed & Uziza leaf	100.92±3.09	87.77±8.19	13.15

278 Results are expressed as mean ± standard deviation

279

280

281 **Table 3: Effect of first, second & third week oral administration of Uziza leaf and black**
 282 **seed on sodium levels (Na) of Wistar rat.**

(mmol/L)			
Treatment	Week 1	Week 2	Week 3
Negative control	194.43 ± 3.15^a	195.95 ± 2.76^b	198.23 ± 1.96^c
Positive control	108.20 ± 3.08	108.60 ± 0.97	108.15 ± 1.60
Uziza leaf	131.49 ± 8.95^a	120.73 ± 6.65^b	98.28 ± 4.17^c
Black seed	130.28 ± 7.87^a	122.95 ± 5.75^b	101.67 ± 4.24^c
Uziza & black seed	118.64 ± 7.16^a	100.17 ± 1.08^b	90.83 ± 2.14^c

283 Results are means of three determinations \pm standard deviation.

284 ^{abc} Different letters in a given row denote significant difference, $p=0.05$.

285

286

287 **Table 4: Effect of first, second & third week oral administration of Uziza leaf and black**
 288 **seed on potassium levels (K) of Wistar rat.**

(mEq/L)			
Treatment	Week 1	Week 2	Week 3
Negative control	0.07 ± 0.08^a	0.08 ± 0.06^b	0.09 ± 0.06^c
Positive control	0.49 ± 0.05	0.05 ± 0.05	0.05 ± 0.10
Uziza leaf	0.05 ± 0.47^a	0.06 ± 0.50^b	0.07 ± 0.18^c

Black seed	0.04 ± 0.33 ^a	0.05 ± 0.23 ^b	0.06 ± 0.19 ^c
Uziza & black seed	0.52 ± 0.18 ^a	0.51 ± 0.21 ^b	0.05 ± 0.10 ^c

289 Results are means of three determinations ± standard deviation.

290 ^{abc} Different letters in a given row denote significant difference, p=0.05.

291

292

293 **Table 5: Effect of first, second and third week oral administration of Uziza leaf and black**
 294 **seed on Urea concentration on Wistar rat.**

UREA CONCENTRATION (mmol/L)			
Treatment	Week 1	Week 2	Week 3
Negative control	26.18 ± 0.21 ^a	26.80 ± 0.09 ^b	26.84 ± 0.05 ^c
Positive control	15.22 ± 0.60	15.27 ± 0.20	15.15 ± 1.20
Uziza leaf	18.43 ± 1.83 ^a	16.24 ± 0.57 ^b	12.83 ± 0.98 ^c
Black seed	18.37 ± 3.16 ^a	16.18 ± 0.88 ^b	12.16 ± 2.01 ^c
Uziza & black seed	16.14 ± 2.44 ^a	15.00 ± 0.78 ^b	11.48 ± 1.78 ^c

295 Results are means of three determinations ± standard deviation.

296 ^{abc} Different letters in a given row denote significant difference, p=0.05.

297

298

299 **Table 6: Effect of first, second and third week oral administration of Uziza leaf and black**
 300 **seed on Creatinine concentration of Wistar rat.**

(mmol/L)			
Treatment	Week 1	Week 2	Week 3
Negative control	285.27 ± 0.45	285.39 ± 0.23	284.58 ± 0.33
Positive control	194.44 ± 4.22	198.57 ± 2.53	182.73 ± 3.67
Uziza leaf	235.36 ± 23.18 ^a	196.16 ± 10.80 ^b	194.16 ± 18.30 ^c
Black seed	210.53 ± 22.24 ^a	198.26 ± 11.88 ^b	167.34 ± 14.66 ^c
Uziza & black seed	195.80 ± 16.38 ^a	163.76 ± 12.14 ^b	174.46 ± 10.66 ^c

301 Results are means of three determinations ± standard deviation.

302 ^{abc} Different letters in a given row denote significant difference, p=0.05.

303

304

305

306 **4. RESULTS AND DISCUSSION**

307 Table 3 shows the result of the effect of aqueous extract of uziza leaf and black seed on sodium
308 level of Wistar rat. After three weeks of inducing the negative control group with sucrose and
309 margarine without treatment, the value of 198.23 ± 1.96 mmol/L when compared with the
310 positive control which was not induced 108.15 ± 1.60 mmol/L was obtained. The results showed
311 that there was a significant effect on the kidney that led to the increase in the concentration of the
312 plasma sodium ($P < 0.05$). This agrees with the research by **Igile et al.** [30] stating that electrolyte
313 abnormalities are common in diabetic patients and may be associated with increased morbidity
314 and mortality. The disturbances of electrolyte homeostasis are also frequently observed in
315 community subjects. Community-acquired electrolyte disorders, even chronic and mild, are
316 related to poor prognosis [31]. Electrolyte disorders are usually multifactorial in nature. Various
317 pathophysiological factors, such as nutritional status, gastrointestinal absorption capacity,
318 coexistent acid-base abnormalities, pharmacological agents, other comorbid diseases (mainly
319 renal disease) or acute illnesses, alone or in combination, play a key role.

320 Uziza group with value of 98.28 ± 4.17 mmol/L showed that uziza significantly decreased the
321 concentration of the plasma sodium when compared to the negative control group at ($P < 0.05$).
322 The traditional and scientific relevance of *P. guineense* are numerous. It is endowed with
323 therapeutic phytochemicals and nutrients which confer therapeutic effects on it and nutritional
324 relevance as well [32]. Research has shown that *P. guineense* contains aromatic substances,
325 alkaloids, salt and substitutes, another earlier report has shown that the leaf of *P. guineense* is
326 rich in flavonoids and phenolic compounds and this compounds has been reported as being
327 beneficial to the kidney electrolytes [33].

328 Black seed group with value of 101.67 ± 4.24 mmol/L also decreased the sodium levels. *Nigella*
329 *sativa* have been used for thousands of years as a spice and food preservative, as well as a
330 protective and curative remedy for several disorders [34]. Black seed extract, seed oil and the
331 isolated bioactive compound thymoquinone possess significant non-toxic phytochemicals
332 beneficial to health [34]. According to the previous and recent scientific researches carried out
333 in various parts of the world, black seed is found effective in providing healing for 129 types of

334 human ailments including 16 different types of cancer, diabetes, asthma, cold, hypertension,
335 Alzheimer's disease, Parkinson's syndrome safety [35,36]. The black seed and uziza group with
336 the value of 90.83 ± 2.14 mmol/L showed that the extract significantly decreased the elevated
337 serum sodium concentration ($P < 0.05$).

338 The extracts significantly reduced serum potassium levels ($p < 0.05$) with uziza leaf (0.07 ± 0.18
339 mEq/L), black seed (0.06 ± 0.19 mEq/L), black seed and Uziza leaf (0.05 ± 0.10 mEq/L) when
340 compared to the negative control group.

341 The extracts also showed reduction in serum urea levels with uziza leaf (12.83 ± 0.98) mmol/L,
342 black seed (12.16 ± 2.01) mmol/L and the combination of black seed and Uziza leaf ($11.48 \pm$
343 1.78) mmol/L ($p < 0.05$). Also the extracts reduced the serum creatinine levels ($p < 0.05$) with uziza
344 leaf (197.16 ± 18.30) mmol/L, black seed (167.34 ± 14.66) mmol/L, black seed and Uziza leaf
345 (174.46 ± 10.66) mmol/L. In a previous study it was shown that oral administration of aqueous
346 extract of *N. sativa* seeds showed no significant changes in kidney function [37]. Another study
347 also failed to show any toxicity for *N. sativa* fixed oil in mice [12, 38]. This study showed that
348 oral administration of *N. sativa* has no toxicity by the concentration doses used. These results is
349 in agreement with previous data reporting that *N. sativa* has a wide margin of safety [35, 36]. It
350 also suggests that there are no toxic effect on kidney function of *N. sativa* at different doses a
351 short period.

352 5. CONCLUSION

353 In conclusion, the extracts significantly decreased the elevated urea, creatinine and electrolytes
354 levels and therefore uziza leaf and black seed can be used to restore kidney function. The results
355 of the present study showed the absence of toxic effect of black seed and uziza leaf on rat kidney.
356 Black seed and Uziza leaf are safe and effective herb that can be used by almost anyone. In
357 general, the aqueous extract is not associated with serious side effects. No irritations or side
358 effects are caused when the right dose is correctly applied.

359 Competing Interests

360 Authors have declared that no competing interests exist.

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363 **6. ETHICAL APPROVAL:**

364 This research work was carried out with the approval of the University of Port Harcourt research
365 ethics committee.

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