

Original Research Article

PERFORMANCE OF *Cucurbita moschata* ON SOIL AND SOILLESS MEDIA

ABSTRACT

Cucurbita moschata is widely grown in both tropical and temperate region due to its structural adaptability. The study was carried out to assess the performance of *C. moschata* on soil (humus) and soilless media (NPK 15:15:15 and NPK 20:10:15 growth media). These treatments are designated as T_C, T_A and T_B, respectively. Standard procedures were followed in the assessment of mineral elements, nutritional composition, pigment compositions, and morphological characters (vein length, leaf area and number of leaves) of *C. moschata* in the three treatments. Among the treatments, *C. moschata* had the highest vein length, leaf area and number of leaves in T_A, while T_C recorded the least. Nutritional compositions of *C. moschata* were: moisture content (80.10%, 87.10% and 69.50%), carbohydrate (5.34%, 3.80% and 15.00%), ash content (3.61%, 1.20% and 4.10%), crude lipid (0.60%, 0.80% and 0.60%), crude protein (8.75%, 6.56% and 8.75%) and crude fibre (1.60%, 0.34% and 2.05%) for the treatments (T_A, T_B and T_C). The mineral composition of *C. moschata* grown in T_A, T_B and T_C growth media respectively were Mg (138.15 mg/kg, 43.90 mg/kg and 109.15 mg/kg), Mn (73.35 mg/kg, 0.25 mg/kg and 123.30 mg/kg), K (2,892.30 mg/kg, 3,338.80 mg/kg and 1,950.80 mg/kg), Zn (47.60 mg/kg, 10.55 mg/kg, 34.00 mg/kg), Ca (2,731.50 mg/kg, 337.95 mg/kg and 426.30 mg/kg), Na (89.65 mg/kg, 108.15 mg/kg and 66.60 mg/kg) and Fe (211.25 mg/kg, 0.00 mg/kg and 137.55 mg/kg) while copper was not detected. The pigment contents indicated the presence of chlorophyll *a* (0.32 mg/g, 0.39 mg/g and 0.24 mg/g), chlorophyll *b* (0.46 mg/g, 0.64 mg/g and 0.40 mg/g), carotenoid (0.33 mg/g, 0.42 mg/g and 0.30 mg/g), and xanthophyll (0.05 mg/g, 0.10 mg/g and 0.00 mg/g) for the three treatments, respectively.

Key Words: *Cucurbita moschata*, growth, development, minerals

1. INTRODUCTION

The production of vegetables and food for human consumption by many subsistent farmers has largely depended on soil as the growth medium. This means of production accounts for majority of vegetables such as *Telfairia occidentalis*, *Cucurbita moschata*, and *Talinum triangulare* found in our local markets. The volumes of production of vegetables have declined in recent years in rural and urban areas due to anthropogenic activities and reduction in soil fertility. However, the advent of scientific research led to the cultivation of plants in a soilless medium like hydroponics. According to Kumar and Cho, hydroponic is a technology which aids plant growth in nutrient solution involving or excluding the application of external source for provision of mechanical support [1]. It was earlier reported by Jensen that production of food in soilless medium is on the increase all over the world [2]. In addition, hydroponically grown vegetables and fruits have been recorded in literature as possessing more nutritional and desirable values as compared to soil grown food produce [3,4,5,6]. The

seedlings quality and vigour is dependent on the composition of media used [7, 8, 9, 10]. Most research carried out on hydroponic has been geared towards leafy greens, peppers and tomato fruit [6, 11, 12], while research on hydroponically grown *C. moschata* has been scarce.

Cucurbita moschata (Duschene *ex* Lam.) Duschene *ex* Poir belongs to the family Cucurbitaceae. Cucurbitaceae ranks amongst the highest of plant families used as human food, cultivated in tropics and temperate regions [13]. *C. moschata* possesses nutritional and therapeutic qualities and has gained the attention of food scientists in recent time [14]. The seeds of *C. moschata* are rich in minerals [15], useful source of nutrients and oils [16] and thus could be used as valuable food supplement [15, 17]. *C. moschata* as fruit vegetable is rich in carotenoids which have antioxidant activities and are easily converted to retinol, the active form of vitamin A [18, 19]. Beta carotene is the most predominant and active of the 5 or 6 provitamins present in commonly consumed foods [20]. It is locally consumed as freshly boiled and steamed or as processed food items in Thailand [21] and in cuisine or serve as desert in Malaysia [22]. There is also a wide variety carotenoid content of food from different races [23]. *C. moschata* is cultivated in Nigeria for both the fruits and leaves [24, 25].

In line with the challenges of population dynamics round the globe and the reduction in arable land for the cultivation of plants, the study is aimed at evaluating the growth and development of *C. moschata* on both soil and soilless media and proffers information in order to enhance its production for both human consumption and profit making.

64

2.0 MATERIALS AND METHODS

2.1 Source of materials used

Comment [P1]: What was the study design?

The seeds of *C. moschata* were collected from the Ecological Center of the University of Port Harcourt. The seeds were divided into two batches and planted in white-sand and humus soil, respectively. The medium of growth for the seedling were humus-soil (T_C) and two NPK solution formulations (15:15:15 and 20:10:15) designated as T_A and T_B , respectively. The two-week old seedlings raised with white-sand were transferred to hydroponic bowls containing different solutions of NPK formulation (T_A and T_B), which served as the soilless medium. The seedlings raised with humus soil served as the soil medium (T_C). The plants were allowed to stand for 8 weeks after planting. The morphological characters of *C. moschata* assessed were the vein length, number of leaves and leaf area. Minerals, pigment content and proximate composition of the leaves were determined following standard procedures.

78

2.2 Morphological characters

Vein length of *C. moschata* was measured with meter rule calibrated in centimeters while the number of leaves was obtained by direct counting. The leaf area of *C. moschata* was determined using the method of Akoroda [26]. Estimated leaf area (LA) = $0.9467 + 0.2475LW + 0.9724LWN$

Where N = Number of leaflets in a leaf; L = Length of the central length; W = Maximum width of the central leaflet.

86

87 2.3 Proximate composition

88 The proximate composition (crude protein, carbohydrate, crude fibre, crude lipid, ash and
89 moisture contents) of *C. moschata* was determined using method of Association of Official
90 Analytical Chemists [27].

91 2.4 Mineral content

92 The mineral contents (Mg, Cu, Mn, K, Zn, Ca, Na and Fe) of *C. moschata* were determined
93 using Atomic Absorption Spectrophotometer (AAS).

94 2.5 Pigment content

95 Sample (0.1 g) *C. moschata* was transferred into a test tube and acetone was added to make it
96 up to 10 ml. The test tube was then kept in the dark for 15 minutes with occasional shaking at
97 room temperature. The chlorophyll, carotenoid and xanthophyll contents were analyzed
98 spectrophotometrically by absorption measurement (A) at 350 nm to 700 nm with 1nm
99 interval and calculated according to the following equations:

$$\text{Chlorophyll } a \text{ (mg/g)} = \frac{13.7 \times A_{665} - 5.76 \times A_{649}}{\text{Mass} \times 200}$$

$$\text{Chlorophyll } b \text{ (mg/g)} = \frac{25.8 \times A_{649} - 7.6 \times A_{665}}{\text{Mass} \times 200}$$

$$\text{Carotenoid (mg/g)} = \frac{4.7 \times A_{440} - 0.263 \times \text{Chlorophyll (a + b)}}{\text{Mass} \times 200}$$

$$\text{Xanthophyll (mg/g)} = \frac{11.51 \times A_{480} - 20.61 \times A_{495}}{\text{Mass} \times 200}$$

100 The above pigments were extracted using acetone according to established methods [28, 29,
101 30].

102 2.6 Statistical analysis

103 The data obtained for the morphological characters and pigment contents of *C. moschata*
104 were subjected to statistical analysis.

Comment [P2]: Mention software you used and at what confidence level?

105

106 3.0 RESULTS AND DISCUSSION

107 3.1 Morphological characters:

108 3.1.1 Vein length:

109 The vein lengths of *C. moschata* grown in three different media are presented in Figure 1.
110 There was an increase in vein length from week 2 – 8 for the treatments. However, T_A
111 medium gave the highest vein length (13.25 cm) compared to other treatments (11.15 cm and
112 9.10 cm) at 8th week. The least vein length (9.10 cm) was recorded in the soil treatment at 8th
113 week.

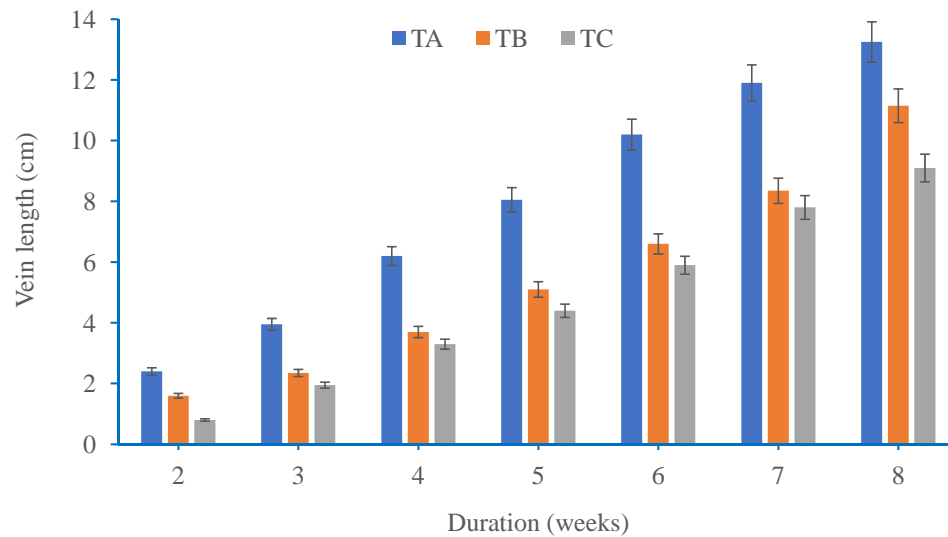


Figure 1: Vein length (cm) of *C. moschata* in three different growth media

3.1.2 Leaf area:

The leaf area of *C. moschata* grown in three different media are presented in Figure 2. There was an increase from week 2 – 8 for the treatments, T_A treatment had the highest leaf area (74.35 cm^2) compared to other treatments (67.05 cm^2 and 58.85 cm^2) at 8th week. The least leaf area (58.85 cm^2) was recorded in the T_C treatment at 8th week. This study has shown that the proportion of nitrogen, phosphorus and potassium available in the growth medium affects directly or indirectly the leaf area of plants.

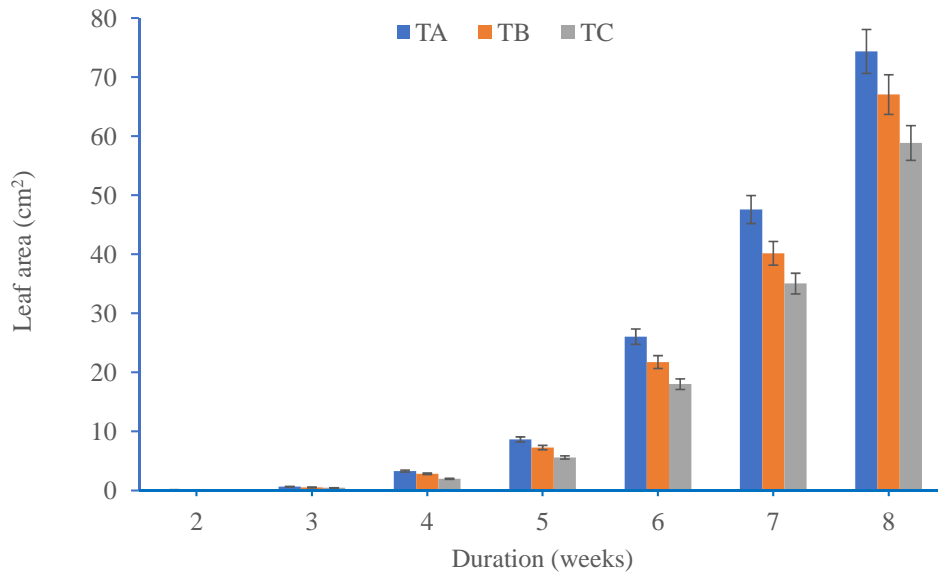


Figure 2: Leaf area (cm²) of *C. moschata* in different growth media

3.1.3 Number of leaves

The number of leaves of *C. moschata* grown in three different media are presented in Figure 3. From week 2 – 8, there was continuous increase in number of leaves among the three treatments (T_A , T_B and T_C). This observation is expected of growing plants. However, there was variation in the number of leaves of *C. moschata* in different treatments. At week 8, the highest increase in number of leaves was observed in T_A growth medium while the least recorded in T_C treatment. Apart from other factors that may interfere in plant growth, Nugawela *et al.* reported a correlation between CO₂ assimilation rate and planting conditions [31]. Plants experiences reduced dry biomass and this affects vegetative growth due to the reduction in CO₂ assimilation rate when planted under artificial shade such as green or shelter house. On the other hand, container and media interaction may affect fertility, pH, soluble salts, bulk density and root zone volume [32]. These may greatly influence plant growth on soilless substrate. However, the study has shown that the number of leaves of *C. moschata* was enhanced in soilless media containing varied proportion of NPK.

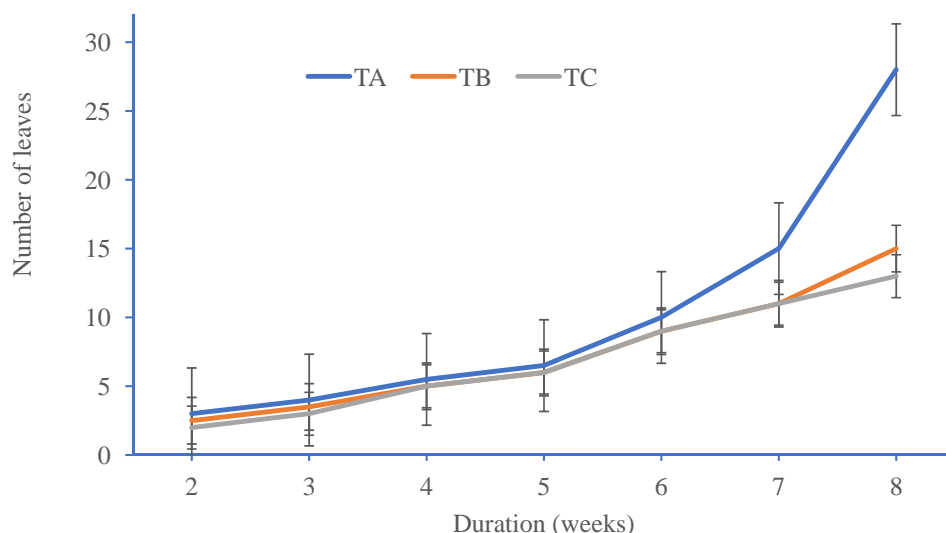


Figure 3: Number of leaves of *C. moschata* in three different growth media

3.2 Proximate compositions

The proximate composition of *C. moschata* leaves showed high amount of moisture content (80.10 %, 87.10 % and 69.50 %) for T_A, T_B and T_C treatments, respectively. The carbohydrate contents were 5.34 %, 3.80 % and 15.00 % in that order. Others were: ash (3.61%, 1.20 % and 4.10 %), protein (8.75%, 6.56% and 8.75%) and crude fibre (1.60%, 0.34% and 2.05%) were considerably low. The lipid contents (0.60 %, 0.80 % and 0.60%) were the lowest. The moisture and lipid contents were highest in *C. moschata* grown in T_B treatment. Ihenacho and Udebuani had earlier reported that high percentage moisture content provides for greater activity of water soluble enzymes and co-enzymes needed for metabolic activities [33]. Dietary fibre has some physiological effects in the gastro-intestinal tract such as: elimination of bile acids, fecal water [34]. It also serves as a source of human nutrition for diabetics in order to reduce glycaemic response to food and consequently the need for insulin [35]. Protein is an important part of catalytic activities, membrane build-up [36, 37]. The nutrient composition of plant materials varies with season, environment, age and cultural practice [38].

3.3 Mineral compositions

Cucurbita moschata leaves contain different minerals and their compositions ranging from lower concentrations of Zinc (Zn) : 47.60 mg/kg, 10.55 mg/kg, and 34.00 mg/kg; Manganese (Mn): 73.35 mg/kg, 0.25 mg/kg, and 123.30 mg/kg; Sodium (Na): 89.65 mg/kg, 108.15 mg/kg, and 66.60 mg/kg; Magnesium (Mg): 138.15 mg/kg, 43.90 mg/kg and 109.15 mg/kg; Iron (Fe): 211.25 mg/kg, 0.00 mg/kg, and 137.55 mg/kg for T_A, T_B and T_C treatments, respectively. Higher concentrations of mineral element were evident in Potassium (K): 2,892.30 mg/kg, 3,338.80 mg/kg, and 1,950.80 mg/kg respectively. In the three growth-

media, Calcium (Ca) was highest in T_A medium (2,731.50 mg/kg). Copper content was not detected in the three growth-media. The role of these elements in the well-being of humans has been previously documented by previous workers [39]. Mineral element plays diverse but essential role in plants, some of which include: catalytic, structural and electrochemical [37]. This implies that the consumption of *C. moschata* leaves will help to improve the nutritional status of human-beings.

3.4 Pigment content

The leaves of *C. moschata* had the highest composition of chlorophyll *a* and chlorophyll *b* in T_B medium, 0.39 mg and 0.64 mg respectively. The concentration of carotenoid and xanthophyll also had the highest concentration in T_B medium, 0.42 mg and 0.10 mg respectively, as shown in Figure 4 and higher than xanthophyll content in the growth media. Among the pigments, chlorophyll *b* content was in abundance than others in all the treatments. The carotenoids obtained in the leaves of *C. moschata* was in line with the work of Pritwani and Manthur, that reported the carotenoids value of 0.407 mg [40]. This trend could be associated with higher nitrogen content. Leaf growth, leaf area and photosynthetic rate may be influenced by the level of N in the soilless media. This ensures control of photosynthetic elements and production of carbohydrates. There may be probably a strong correlation and influence between chlorophyll and leaf area because the former indicates some level of N accumulation in leaves [41]. More so nitrogen use efficiency is said to be attributed to leaf area and other growth traits such as plant height [42]. Increase in N and P could increase leaf growth and chlorophyll content while its decrease may also be detrimental to crops [43]. Though, concentration of these elements sometimes may be advantageous or detrimental to the plants [44]. Other factors which may equally stimulate plant growth and development are better gaseous exchange; improved drainage and uniform extension of root systems sometimes are more advantageous than other growth factors [45].

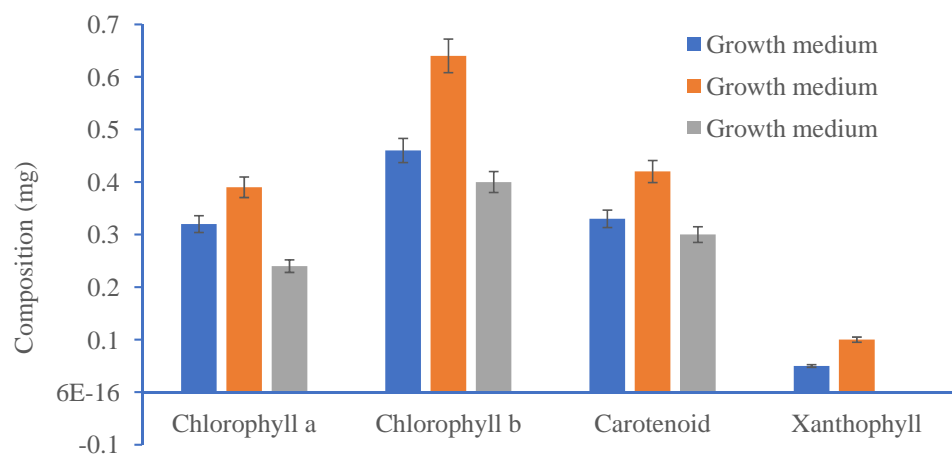


Figure 4: Pigment content (mg) of *C. moschata* in three different growth media

4.0 CONCLUSION

Cucurbita moschata is rich with nutrient and mineral composition. The mineral composition of any growth medium determines the growth and development of *C. moschata*. The study has shown that the variation in the macro-nutrients affects the vigour of *C. moschata*. T_A medium gave the highest vein length, leaf area, and number of leaves of *C. moschata* while the pigment compositions were slightly higher in T_B medium compared to other treatments. The study therefore recommends that *C. moschata* be grown in a moderate concentration of NPK solution with a view of tackling the problem of reduction in soil fertility and non-availability of arable land for the cultivation of *C. moschata*.

COMPETING INTEREST

Authors have declared that no competing interests exist.

REFERENCES

- Kumar RR, Cho JY. Reuse of hydroponic waste solution. *Environmental Science and Pollution Research*, 2014; 1-9.
- Jensen MH. Hydroponics worldwide. *Acta Horticulturae*, 1999; 481: 719-729.
- Gichuhi PN, Mortley D, Bromfield E, Bovell AC. Nutritional, physical and sensory evaluation of hydroponic Carrots (*Daucus carota* L.) from different nutrient delivery systems. *Journal of Food Science*, 2009; 74(9): 403-412.

- 220 4. Sgherri C, Cecconami S, Pinzino C, Navari IZ, Izzo R. Levels of anti-oxidants and
221 nutraceuticals in Basil grown in hydroponics and soil. *Food Chemistry*, 2010; 123(2):
222 416-422.
- 223 5. Selma MV, Luna MC, Martinez SA, Tudela JA, Beltran D, Baixauli C, Gil MI. Sensory
224 quality of green and red fresh-cut lettuces (*Lactuca sativa* L.) are influenced by soil and
225 soilless agricultural production systems. *Post-harvest Biology and Technology*, 2012;
226 63(1): 16-24.
- 227 6. Buchanan DN, Omaye ST. Comparative study of ascorbic acid and tocopherol
228 concentrations in hydroponic and soil-grown lettuce. *Food and Nutrition Sciences*,
229 2013; 4: 1047-1053.
- 230 7. Corti C, Crippa L, Genevini PL, Centemero M. Compost use in plant nurseries,
231 hydrological and physico-chemical characteristics. *Compost Science Utilization*, 1998;
232 6: 35-45.
- 233 8. Wilson SB, Stoffella PJ, Graetz DA. Use of compost as a media amendment for
234 containerized production of two subtropical perennials. *Journal of Environmental*
235 *Horticulture*, 2001; 19(1): 37-42.
- 236 9. Sahin U, Ors S, Ercisli S, Anapali O, Esitken A. Effect of pumice amendment on
237 physical soil properties and strawberry plant growth. *Journal of Central Europe*
238 *Agriculture*, 2005; 6(3): 361-366.
- 239 10. Baiyeri KP. Evaluation of nursery media for seedling emergence and early seedling
240 growth of two tropical tree species. *Journal of Agricultural Resources*, 2003; 4(1): 60-
241 65.
- 242 11. Arias R, Lee TC, Specca D, Janes H. Quality comparison of hydroponic tomatoes
243 (*Lycopersicon esculentum*) ripened on and off vine. *Journal of Food Science*, 2000;
244 65(3): 545-548.
- 245 12. Koyama M, Nakamura C, Kozo N. Changes in phenols contents from Buckwheat sprout
246 during growth stage. *Journal of Food Science and Technology*, 2013; 50(1): 86-91.
- 247 13. Christenhusz MJ, Byng JW. The number of known plants species in the world and it's
248 annual increase. *Phytotaxa*, 2010; 261(3): 201-217.
- 249 14. Fokou E, Achu M, Tchouanguep M. Preliminary nutritional evaluation of five species
250 of egusi seeds in Cameroon. *African Journal of Food and Agriculture Nutrition*
251 *Development*, 2004; 4: 1-11.
- 252 15. Nwofia GE, Nwogu N, Nwofia BK. Nutritional variation in fruits and seeds of
253 pumpkins (*Cucurbita* spp) accessions from Nigeria. *Pakistan Journal of Nutrition*,
254 2012; 11(10): 848-858.
- 255 16. Dhiman AK, Sharma KD, Attri S. Functional constituents and processing of pumpkin:
256 A review. *Journal of Food Science and Technology*, 2009; 46(5): 411-417.
- 257 17. Gorgonio CMS, Pumar M, Mothe CG. Macroscopic and physiochemical
258 characterization of a sugarless and gluten-free cake enriched with fibres made from
259 pumpkin seed (*Cucurbita maxima* L.) flour and corn starch. *Genc. Tecnol. Aliment.*
260 *Campinas*. 2011; 31(1): 109-118.

- 261 18. Dini I, Tenore GC, Dini A. (2003). Effects of industrial and domestic processing on
262 antioxidant properties of pumpkin pulp. *LWT- Food Science and Technology* 2003;
263 53(1):382-385.
- 264 19. Quiros ARB, Costa HS. Analysis of carotenoids in vegetable and plasma samples: a
265 review. *Journal of Food and Composition Analysis* 2006; 19(2-3) 97-111.
- 266 20. McLaren DSM. Chapter 5 Vitamin A: The Vitamins part 11 considering the individual
267 vitamins. Elsevier pp 2012; 93-138.
- 268 21. Pongjanta J, Naulbunrang A, Kawngdang S, Manon T, Thepjaikat T. Utilization of
269 pumpkin powder in bakery products Songklanakarin, *Journal of Science and*
270 *Technology*, 2006; 28(1): 71-79.
- 271 22. Norshazila S, Irwandi J, Othman R, Yumi HH. Carotenoid content in different locality
272 of pumpkin (*Cucurbita moschata*) in Malaysia. *International Journal of Pharmacy and*
273 *Pharmaceutical Sciences*, 2014; 6(3): 29-32.
- 274 23. Bhaskarachary K, Rao DSS, Deosthale YG, Reddy V. Carotene content of some
275 common and less familiar foods of plant origin. *Food Chemistry* 1995; 54: 189-193.
- 276 24. Okoli BE. Wild and cultivated cucurbits in Nigeria. *Economic Botany* 1984; 38: 350-
277 370.
- 278 25. Ndukwu BC, Okoli BE. Studies on Nigeria *Cucurbita moschata*. *Nigerian Journal of*
279 *Botany*, 1992; 5: 19-26.
- 280 26. Akoroda MO. Non-destructive estimation of area and variation in shape of leaf lamina
281 in the fluted pumpkin (*Telfairia occidentalis*). *Scientia Horticulturae*, 1993; 53(3): 261-
282 267.
- 283 27. AOAC. *Official methods of analysis*. Association of Official Analytical Chemicals,
284 (15th edition). Arlington, USA, 1990.
- 285 28. Kukric ZZ, Topalic LN, Kukavica BM, Matos SB, Pavicic SS, Boroja MM, Savic AV.
286 Characterization of anti-oxidant and microbial activities of nettle leaves (*Urticadioica*
287 L.). *Acta Periodica Technologica*, 2012; 43: 257-272.
- 288 29. Chang SK, Nagendra PK, Amin I. Carotenoid retention in leaf vegetables based on
289 cooking methods. *International Food Research Journal*, 2013; 20(1): 457-465.
- 290 30. Duma M, Alsina I, Zeipiria S, Lapse L, Dubova L. Leaf vegetables as source of
291 phytochemicals. *Foodbalt*, 2014; 20: 262-265.
- 292 31. Nugawela A, Ariyawansa P, Samarasekara RK. Physiological yield determinants of sun
293 and shade leaves of *Hevea brasiliensis*. *Journal Rubber Research Institute Sri Lanka*,
294 1995; 76: 1-10.
- 295 32. Hockenberry MM, Cunliffe BA. Effects of media porosity and container size on
296 overwintering and growth of ornamental grasses. *HortScience* 2004; 39(2): 248-250.
- 297 33. Ihenacho K, Udebuani AC. Nutritional composition of some leafy vegetable consumed
298 in Imo state, Nigeria. *Journal of Applied Science and Environmental Management*,
299 2009; 13(3): 35-38.

- 300 34. Akpabio UD, Akpan AE. Evaluation of nutritive and anti-nutritive composition of the
301 seeds of *Mondora myristica* (African nutmeg). *World Journal of Applied Science and*
302 *Technology*, 2012; 4: 49-55.
- 303 35. Onyije MO. *Elemental and proximate contents of Gnetum africanum and Telfairia*
304 *occidentalis* (B.Sc. project). Department of Plant Science and Biotechnology,
305 University of Port Harcourt, 2012; pp. 25-26.
- 306 36. Esenwo GJ. *Developmental biology and plant physiology*. Abeam Publishing Company,
307 Nigeria, 2004; pp.23-168.
- 308 37. Anoliefo CO. *Introductory Tropical Plant Biology*. Uniben Press, Nigeria, 2006; pp.
309 257-362.
- 310 38. Apoxi SO, Long RJ, Castro FB, Orakor ER. Chemical composition and nutritive value
311 of leaves and stems of tropical weeds. *Grass and Forage Science*, 2000; 55(1): 77-81.
- 312 39. Oluyemi EA, Akilua AA, Adenuya AA, Adebayo MB. Mineral contents of some
313 commonly consumed Nigerian foods. *Science Focus*, 2006; 11: 153-157.
- 314 40. Pritwani R, Manthur P. Beta carotene content of some commonly consumed vegetables
315 and fruits available in Delhi India. *Journal of Nutrition and Food Science* 2017; 7:625.
- 316 41. Nageswara RRC, Talwar HS, Wright GC. Rapid assessment of specific leaf area and
317 leaf nitrogen in peanut (*Arachis hypogaea* L.) using chlorophyll meter. *J. Agron. Crop*
318 *Sci.*, 2001; 189: 175-182.
- 319 42. Hirel B, Le Gouis J, Ney B, Gallais A. The challenge of improving nitrogen use
320 efficiency in crop plants: towards a more central role for genetic variability and
321 quantitative genetics within integrated approaches. *Journal of Experimental Botany*,
322 2007; 58(9): 2369–2387.
- 323 43. Sinclair TR, Vadez V. Physiological traits for crop yield improvement in low N and P
324 environments. *Plant and Soil* 2002; 245: 1-15.
- 325 44. Salisu M, Daud N, Ahmad I. Influence of fertilizer rates and soil series on growth
326 performance of natural rubber (*Hevea brasiliensis*) latex timber clones. *Australian*
327 *Journal Crop Science* 2013; 7: 1998 –2004.
- 328 45. Pinamonti F, Stringari G, Zorzi G. Use of compost in soilless cultivation. *Compost*
329 *Science & Utilization*, 1997; 5: 38-46.