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Original Research Article

Content of photosynthetic pigments and leaf gas exchanges of young coffee plants under light restriction and treated with paclobutrazol

ABSTRACT

The use of shading and paclobutrazol in coffee plants can be an important cultivation strategy to mitigate the negative effects of high solar radiation and atmospheric temperature. Therefore, the levels of photosynthetic pigments and foliar gas exchanges of young coffee plants submitted to doses of paclobutrazol were evaluated, in environments with artificial light restriction. Five experiments were performed: one in full sunlight and four in artificially shaded environments with black polyethylene meshes at 20%, 40%, 60% and 80% levels of light restriction. In each of these environments, an experiment was carried out, consisting of five treatments, defined by the application of paclobutrazol via substrate, at doses of 0, 10, 20, 30 and 40 mg of active ingredient per plant. Joint analysis of experiments and analysis of variance of the regression were made, for the study of levels of shading and doses of paclobutrazol. The light restriction optimized the photosynthetic apparatus of the plants, mainly at levels close to 60%, and considerably favored leaf gas exchanges of arabica coffee. The application of paclobutrazol in the studied dosages resulted in little or no effect on photosynthetic pigment contents and did not influence leaf gas exchanges of coffee plants.

Keywords: Coffea arabica L., shading, triazole, physiological changes.

1. INTRODUCTION

In several farming regions of Brazil, cultivated plants are constantly exposed to climatic adversities that limit their initial establishment in the field, negatively reflecting the yield potential. Among these, intense solar radiation, high temperatures, and low rainfall volume are the most limiting.

When plants are exposed to light energy higher than that required by photosynthesis, there may be energy imbalance that results in photoinhibition. This may promote the biosynthesis of reactive oxygen species and, consequently, cause oxidative stress. Photoinhibition may also be a result of photophysical parameters, which include response to light intensity or wavelength [1-2].

Coffee is a native species of understory regions and therefore is considered as a shade plant [3], with low point of light saturation. Thus, shading of coffee plantations may be an alternative cultivation method to mitigate negative effects of direct exposure to the sun, in order to favor the initial establishment of the crop and optimize its development in subsequent stages.

33 In general, coffee tree presents low rates of assimilation of CO₂ (A) when compared to other
34 tropical trees. Shading may favor certain environmental factors, such as temperature
35 attenuation and reduction of water vapor pressure deficit, in order to benefit the gas
36 exchange of coffee plants [4-5].

37 However, existing information on the effects of shading on gas exchange of coffee plants is
38 contrasting and depends on the level of light restriction [6-7].

39 In general, in comparison to sun leaves, leaves of shade present greater amount of
40 chlorophyll per reaction center, more developed antenna complexes, smaller ratio between
41 chlorophyll *a* and *b*, and lower content of carotenoids [8-9].

42 Plant growth regulators, especially inhibitors of biosynthesis of gibberellins, have been
43 applied to the traditional management of high technological standards of crops. These
44 products can influence various aspects of plant metabolism, both morphologically and
45 physiologically, reducing susceptibility to biotic and abiotic stresses [10]. This capacity of
46 modulation provided to the plants has substantial importance in face of the climatic
47 adversities verified in cropping environments.

48 Paclobutrazol (PBZ) [(2*RS*, 3*RS*) -1- (4-chlorophenyl) -4,4-dimethyl-2- (1,2,4-triazol-1-yl) -
49 pentan-3-ol] is triazole capable of inhibiting cytochrome P450 dependent mono-oxygenases
50 and, consequently, biosynthesis of gibberellins [11]. The changes in plant hormonal balance
51 caused by triazole, such as elevated levels of cytokinins and abscisic acid, can interfere with
52 foliar gas exchange and photosynthetic pigment content [12-15].

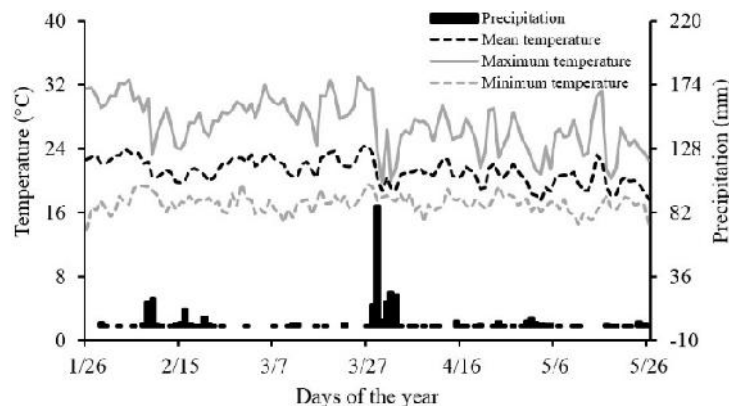
53 Several studies have demonstrated the ability of paclobutrazol to mitigate the damage
54 caused by abiotic stresses [16-21], including high temperature stress [11, 21].

55 The effects of this growth regulator have variations according to dosage, phenological stage,
56 and form of application [22]. For coffee plants, however, the knowledge about such
57 technology is still incipient, and there is a need for systematic and analytical studies on the
58 subject.

59 Therefore, the use of shading and the application of paclobutrazol in coffee plants is an
60 important strategy to minimize negative factors related to high solar radiation index and
61 elevation of atmospheric temperature. The objective of this study was to evaluate the levels
62 of photosynthetic pigments and foliar gas exchange of young coffee plants submitted to
63 doses of paclobutrazol in environments with artificial light restriction.
64

65 **2. MATERIAL AND METHODS**

66
67 The experiments were conducted at the Universidade Estadual do Sudoeste da Bahia,
68 Vitória da Conquista *Campus*, between January and May 2017. The experimental area is
69 located at 14° 53' 05" S and 40° 48' 00" W, at 852 meters of altitude. The climate of the
70 municipality, **Country name** according to Köppen-Geiger climatic classification, is of Cwa
71 (tropical of altitude) type, with mean annual temperature of 20.2°C and a mean precipitation
72 of 733.9 mm [23]. The meteorological data obtained during the period of tests can be
73 observed in Figure 1.



74

75 **Fig. 1. Meteorological data recorded in the automatic meteorological station of the**
 76 **Universidade Estadual do Sudoeste da Bahia, country name during the experimental**
 77 **period (INMET).**

78 *Coffea arabica* L. 'Catuaí Red IAC 144' seedlings were obtained in an accredited nursery.
 79 When they had four pairs of mature leaves (approximately five months old), they were
 80 individually transplanted to pots with a capacity of 20 dm³ (32.5 cm high x 34.5 cm higher
 81 diameter and 22 cm lower diameter).

82 Containers were filled with mix of soil (typical Eutrophic YELLOW LATOSOLO) and humus,
 83 in the ratio 9: 1, and homogenized through sieve of 5 mm. The chemical analysis of the soil
 84 used in the mixture showed the following results: pH (H₂O): 5.4; P: 2.0 mg dm⁻³; K⁺: 0.23
 85 cmol_c dm⁻³; Ca²⁺: 2.2 cmol dm⁻³; Mg²⁺: 0.8 cmol_c dm⁻³; Al³⁺: 0.1 cmol_c dm⁻³; H⁺: 2.7 cmol_c dm⁻³.
 86 Liming and fertilization of the substrate were carried out based on soil chemical analysis,
 87 and according to the technical recommendation of the Soil Fertility Commission of the State
 88 of Minas Gerais[24].

89 Immediately after transplanting, the pots were placed in different environments, with 0% (full
 90 sun), 20%, 40%, 60% and 80% of artificial light restriction. Shaded environments (4 meters
 91 wide x 8 meters long x 2 meters high) were obtained through black polyethylene meshes. In
 92 each environment (shaded and in full sun) an experiment was conducted, totaling five
 93 experiments.

94 Each experiment (0%, 20%, 40%, 60% and 80% of light restriction) consisted of five
 95 treatments, defined by the application of different doses of paclobutrazol via substrate (0, 10,
 96 20, 30 and 40 mg of active ingredient per plant). A completely randomized design was used,
 97 with four replications, totaling 20 plots. Each experimental unit consisted of a pot containing
 98 a coffee plant. For analysis of leaf gas exchanges, readings were made in blocks, with four
 99 replications, due to variations occurred during the evaluation period, from 8:00 a.m. to 12:00
 100 p.m.

101 Applications of paclobutrazol were carried out at 18 days after transplanting of seedlings,
 102 with the commercial product Cultar 250 SC® (250 g i.a. L⁻¹ of paclobutrazol), and volume of
 103 solution of 200 mL per plant, applied directly to the substrate.

104 Management of weeds and pests was performed according to the occurrence along the
 105 experiment conduction. All plants were irrigated every two days, with water volume
 106 determined by the gravimetric method (a control pot for each experiment), in which these
 107 containers were saturated with water, with subsequent gravimetric drainage until constant

108 weight. First, plant pot of each experiment was weighed to obtain the initial mass (IM). Every
109 two days, the control pots were weighed again, obtaining the final mass (FM). The volume of
110 water (V) to be applied at the date of each water replenishment, in liters, was determined by
111 the difference between the two masses, through the equation: $V = IM - FM$, with masses
112 being expressed in kilograms.

113 At 100 days after application (DAA) of paclobutrazol, SPAD (Soil Plant Analysis
114 Development) index and photosynthetic pigment content were evaluated. The intensity of
115 green color of leaf (SPAD index) was determined using a portable chlorophyllometer (SPAD
116 502, MINOLTA, Japan), with readings at three points of the first fully expanded leaf, from the
117 apex of the plant, and then the average.

118 The extraction of photosynthetic pigments was performed according to the modified
119 methodology of [25], by eliminating the stages of maceration and centrifugation of the discs,
120 described by [26]. The first fully expanded leaf of each plant was collected, from which 10
121 leaf discs of six millimeters of diameter were removed, with the aid of manual leaf disc
122 extractor. The material was immediately weighed on analytical balance and filled into
123 aluminum-coated test tubes containing 20 mL of 80% acetone (v/v). This procedure was
124 performed in an environment without direct incidence of light. The tubes were then capped,
125 sealed with plastic film, and kept in the dark for 48 hours to extract the pigments.

126 After this period, absorbance readings of the extracts were performed in spectrophotometer
127 (700 Plus, Femto, Brazil), at wavelengths of 663 nm, 646 nm and 470 nm. For the calibration
128 of the spectrophotometer, 80% acetone (v/v) was used as "blank". Concentrations ($\mu\text{g mL}^{-1}$
129 of extract) of *a*, *b*, and total chlorophyll, and carotenoids were calculated using specific
130 equations for each pigment [27]. Depending on the mass of each sample and the volume of
131 acetone used, the values were converted and the pigment content expressed as mg g^{-1} of
132 fresh leaf matter.

133 At 99 DAA of paclobutrazol, leaf gas exchanges were evaluated. These evaluations were
134 performed on the same leaf used for the other physiological analyzes, using an infrared gas
135 analyzer (IRGA), LCPro, ADC, UK coupled to an actinic light source of $1000 \mu\text{mol photons}$
136 $\text{m}^{-2} \text{s}^{-1}$ of photosynthetically active radiation.

137 Rate of CO_2 assimilation (A , $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), transpiration rate (E , $\text{mmol water vapor m}^{-2} \text{ s}^{-1}$),
138 stomatal conductance (g_s , $\text{mol m}^{-2} \text{ s}^{-1}$), and the internal CO_2 concentration in the leaf (C_i ,
139 $\mu\text{mol CO}_2 \text{ mol}^{-1} \text{ air}$). Carboxylation efficiency (A/C_i) was calculated by the ratio of CO_2
140 assimilation rate to internal CO_2 concentration in the leaf.

141 Data were submitted to normality tests (Lilliefors) and homogeneity of variances (Cochran).
142 After the analysis of variance of each experiment (each level of light restriction) was carried
143 out, joint analysis of experiments was performed, respecting for each variable relation
144 between mean squares of residue less than or equal to 1:7, according to [28]. When joint
145 analysis presented significance ($p < 0.05$), regression analysis was performed for the study
146 of paclobutrazol doses and levels of shading. The regression models were defined based on
147 the significance ($p < 0.05$), the highest coefficient of determination (R^2) and the biological
148 response for each characteristic studied. For statistical analysis, the program Statistical and
149 Genetic Analysis System (SAEG), version 9.1 was used.

150

151 **3. RESULTS AND DISCUSSION**

152

153 Light restriction was the factor with the greatest impact on the variables related to
154 photosynthetic pigments (content of chlorophyll *a*, *b*, and total, carotenoid content, and

155 chlorophyll *a:b* ratio), and intensity of green color in the leaf. For chlorophyll *a* and total
 156 content, there was interaction between the studied factors (levels of light restriction and
 157 doses of paclobutrazol). The environment with 60% of light restriction was not grouped for
 158 the analysis of the parameters chlorophyll *b* content and carotenoid content (Table 1).

159 For the unfolding of interaction between the factors, a cubic model for the relationship
 160 between chlorophyll *a* content and light restriction levels (LR) in coffee plants treated with 0
 161 and 30 mg of paclobutrazol (PBZ) was delineated. For the coffee plants submitted to 10, 20
 162 and 40 mg of the regulator, a linear model was established increasing as a function of levels
 163 of shading (Figure 2A).

164 Chlorophyll *a* content of plants not treated with PBZ (0 mg) was lower than treatment in full
 165 sun at levels below 33.6% of LR. From this level, the values were higher than the control,
 166 with an estimated maximum content of 2.34 mg g⁻¹ of chlorophyll *a* (65.7% LR). The
 167 maximum levels of chlorophyll *a* estimated for the treatments with 10, 20 and 40 mg of PBZ
 168 (2.45, 2.43 and 2.34 mg g⁻¹, respectively), remained close to the estimated maximum value
 169 for coffee plants without regulator application.

170 **Table 1. Analysis of variance summary and coefficients of variation (CV) of leaf**
 171 **greening (SPAD), chlorophyll *a* content (Chla), total chlorophyll content (Chla+b), ratio**
 172 **of chlorophyll *a* to *b* (Chla:b), chlorophyll *b* content (Chlb) and carotenoid content**
 173 **(Car) of *Coffea arabica* L. ‘Catuaí Vermelho IAC 144’ plants submitted to different**
 174 **light restriction levels (LR) and paclobutrazol doses (D), evaluated at 100 days after**
 175 **the application of the regulator. Vitória da Conquista - BA, 2017.**

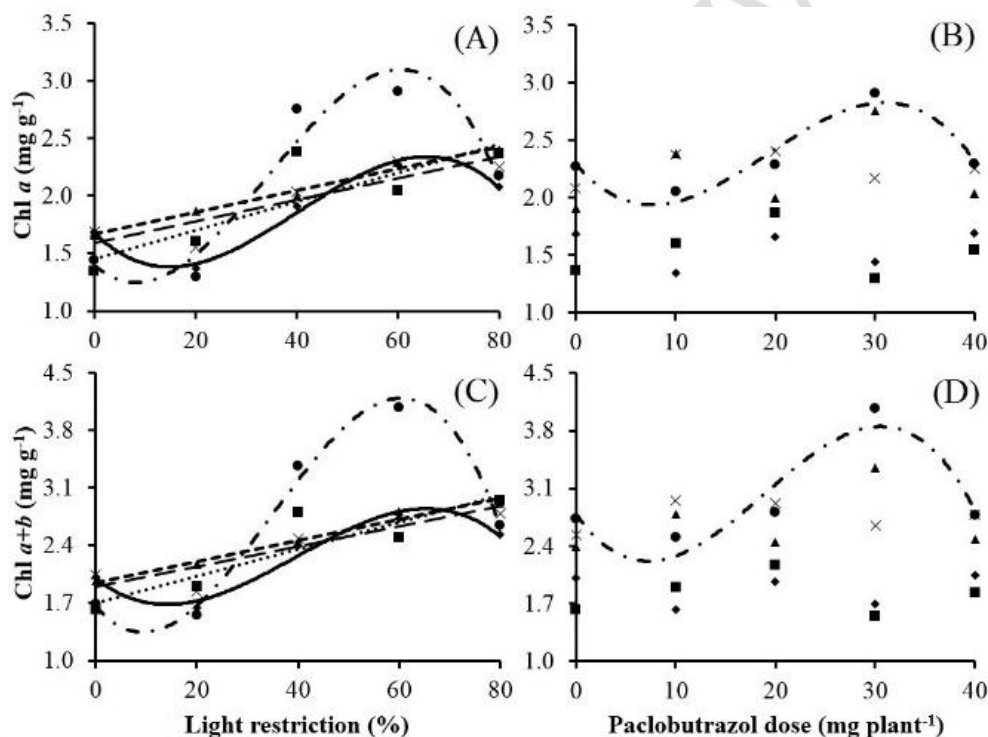
SV	df	MEAN SQUARES				df	Chlb	Car
		SPAD	Chla	Chla+b	Chla:b			
LR	4	391.1**	3.2**	5.9**	4.0*	3	0.2915**	0.042*
D	4	97.5 ^{ns}	0.2 ^{ns}	0.5 ^{ns}	0.3 ^{ns}	4	0.0008 ^{ns}	0.003 ^{ns}
LR*D	16	63.7 ^{ns}	0.3*	0.6**	1.0 ^{ns}	12	0.0137 ^{ns}	0.011 ^{ns}
Wn	75	66.1	0.1	0.3	0.7	60	0.0085	0.011
CV (%)		12.3	18.5	20.5	17.6		23.5	25.5

176 ^{ns}, * e **: non-significant, significative by “F” test at 5% and 1% of probability, respectively.

177 On the other hand, in coffee plants treated with 30 mg of PBZ, the effect of shading on
 178 increasing chlorophyll *a* content was potentiated. There was an expressive increase in the
 179 content of this pigment promoted by shading at levels above 17.2%, with an estimated
 180 maximum value of 3.09 mg g⁻¹ of chlorophyll *a* (121.75% higher than the full sun treatment),
 181 at the level of 60.4% of LR.

182 It was not possible to delineate a mathematical model to express the relationship between
 183 the chlorophyll a content and the PBZ doses of coffee plants conducted under levels of 0,
 184 20, 40 and 80% of LR. A cubic model was designed to express the effect of PBZ doses on
 185 the chlorophyll a content of coffee plants kept under 60% of light restriction. The values were
 186 higher than the control (without PBZ application) at doses higher than 17.7 mg of the
 187 regulator per plant, with an estimated maximum value for the dose of 31.0 mg PBZ (2.82 mg
 188 g⁻¹) (Figure 2B).

189 For the unfolding of interaction between LR levels and PBZ doses, in the evaluation of the
 190 total chlorophyll content, a similar trend was observed for chlorophyll a (Figure 2C and 2D).
 191 Maximum levels of total chlorophyll as a function of LR levels were estimated at 2.85, 3.00,
 192 2.97, 4.19 and 2.87 mg g⁻¹ of fresh matter for plants treated with 0, 10, 20, 30 and 40 mg of
 193 PBZ, respectively. As with chlorophyll a, coffee plants treated with 30 mg of PBZ via soil
 194 showed a more significant increase in total chlorophyll content as a function of shade levels
 195 compared to other doses (Figure 2C). For this treatment, maximum total chlorophyll content
 196 (4.19 mg g⁻¹), estimated at 60.4% of LR level, was approximately 2.5 times higher than the
 197 treatment in full sun.



198

199 **Fig. 2.** Chlorophyll a and total chlorophyll content in leaves of coffee plants (*Coffea*
 200 *arabica* L. 'Catuaí Vermelho IAC 144') in response to different light restriction levels
 201 (LR) and paclobutrazol doses (D), at 100 days after the application of the regulator. (A,
 202 B) chlorophyll a content (Chl a): (A) ♦0 mg – $\hat{Y}^* = 1.67811 - 0.0428795X + 0.00176607X^2 -$
 203 $0.0000146094X^3$ ($R^2 = 0.9874$); ■10 mg – $\hat{Y}^{**} = 1.451 + 0.0124625X$ ($R^2 = 0.7246$); ▲20
 204 mg – $\hat{Y}^{**} = 1.664 + 0.0095125X$ ($R^2 = 0.9839$); ●30 mg – $\hat{Y}^{**} = 1.39532 - 0.0378676X +$
 205 $0.00264821X^2 - 0.0000257552X^3$ ($R^2 = 0.9276$); x40 mg – $\hat{Y}^{**} = 1.592 + 0.0093125X$ ($R^2 =$
 206 0.7836). (B) ♦0%; ■20%; ▲40%; ●60% – $\hat{Y}^{**} = 2.29157 - 0.0999643X + 0.00811786X^2 -$
 207 $0.00014X^3$ ($R^2 = 0.9199$); x80%. (C, D) total chlorophyll content (Chl a+b): (C) ♦0 mg – \hat{Y}^*
 208 $= 1.99146 - 0.0472693X + 0.00204464X^2 - 0.0000171615X^3$ ($R^2 = 0.9584$); ■10 mg – $\hat{Y}^{**} =$

209 $1.706 + 0.016225X$ ($R^2 = 0.8051$); $\blacktriangle 20 \text{ mg} - \hat{Y}^{**} = 1.952 + 0.0127625X$ ($R^2 = 0.9797$); $\bullet 30$
210 $\text{mg} - \hat{Y}^{**} = 1.66129 - 0.0719911X + 0.00446161X^2 - 0.0000426562X^3$ ($R^2 = 0.9875$); $\times 40$
211 $\text{mg} - \hat{Y}^{**} = 1.8975 + 0.0122X$ ($R^2 = 0.7984$). (D) $\blacklozenge 0\%$; $\blacksquare 20\%$; $\blacktriangle 40\%$; $\bullet 60\%$ - $\hat{Y}^{**} = 2.78125$
212 $- 0.171187X + 0.0146375X^2 - 0.000258125X^3$ ($R^2 = 0.8583$); $\times 80\%$. * e **: significant by
213 regression analysis at 5% e 1% of probability, respectively.

214 As a strategy to increase the efficiency of light absorption processes, plants grown under
215 lower radiation levels tend to have higher density of light-picking complexes when compared
216 to plants kept in full sunlight [29]. In addition, the increase in chlorophyll content under low
217 light conditions may be associated, in part, with higher nitrogen allocation to photosystems
218 [30].

219 The total chlorophyll content of coffee plants conducted under 60% of LR, as a function of
220 doses of PBZ applied, was higher than the control at dosages above 16.5 mg of the inhibitor,
221 with a maximum point estimated for 30.6 mg of PBZ. Any mathematical model among those
222 studied expressed the effect of PBZ treatment on the total chlorophyll content of coffee
223 plants kept under 0, 20, 40 and 80% shading (Figure 2D).

224 Treatment with PBZ may result in increases cytokinin levels [14]. It is known that elevation in
225 cytokinin levels can accelerate chloroplast differentiation and chlorophyll biosynthesis, and
226 maintain the integrity of this molecule [31]. Working with *Solenostemon rotundifolius*, [32]
227 observed that PBZ treatment resulted in higher number of chloroplasts per cell unit in the
228 leaves when compared to the control treatment.

229 It was observed increase in SPAD index of arabica coffee plants, due to the increase in
230 shading levels, with maximum value (68.99) estimated for the level of 62.7% of LR (Figure
231 3A). There is positive correlation between SPAD index and chlorophyll content in leaves of
232 different plant species [33-36]. Therefore, the increase observed in the SPAD index in this
233 study was associated with higher chlorophyll content in leaves of the shaded plants (Figure
234 2C).

235 Generally, treatment with PBZ provides higher SPAD index in plants. This fact is commonly
236 associated with the increase of chlorophyll content, or the higher number of chloroplasts per
237 unit of leaf area, in response to increases of leaf thickness and decrease of leaf area [32, 13].
238 However, for the present study, there was no effect of the PBZ doses applied via soil, on the
239 coffee plants SPAD index (Figure 3B).

240 It should be noted that SPAD index is based on a unit of green light reflectance area, while
241 the chlorophyll content determined in the present work was defined based on the mass of
242 the leaf blade. Due to the existence of an impact intensity differential of PBZ on leaf
243 morphology (area reduction and thickness increase) and chloroplast metabolism (chlorophyll
244 synthesis, movement, distribution, and anatomy of these plastids), different associations
245 between these effects may interfere with the intensity of the green color of the leaf.

246 There was a tendency for linear increase of chlorophyll *b* content as a function of the
247 increase in LR levels (Figure 3C). [29] associated the decrease of the chlorophyll *b* content in
248 *Illicium floridanum* cultivated in full sun to the degradation of this pigment by the excess of
249 irradiation. In addition, the higher development of LHCI and LHCII antenna complexes in
250 shaded plants [8] may be associated with this response.

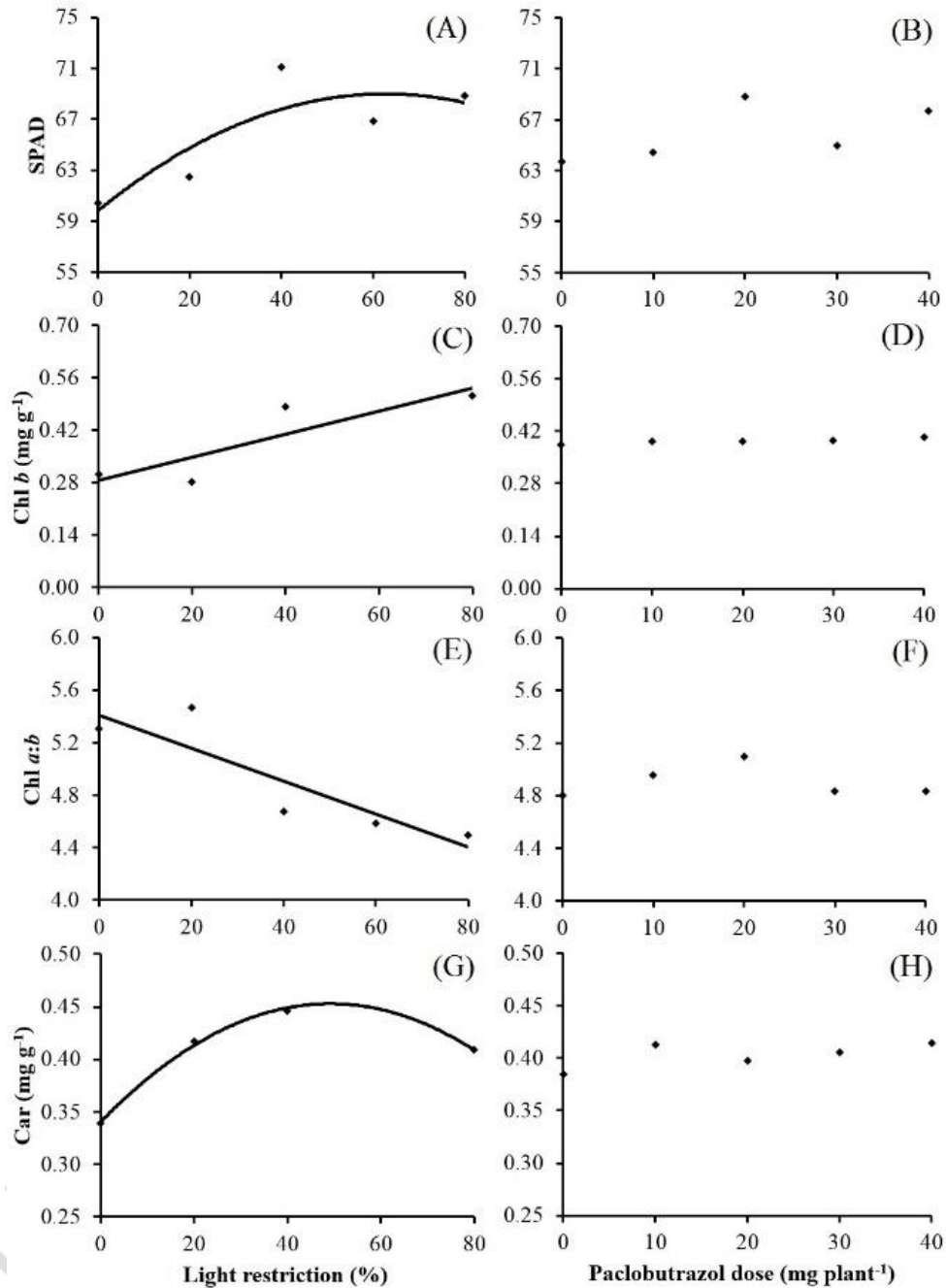
251 A linear decreasing effect was observed for the relationship between chlorophyll *a* and *b*
252 ratio (Chl*a*:*b*) and LR levels (Figure 3E). In general, the size of the antenna complexes
253 (LHCI and LHCII) of plants increases under low irradiation, while under high irradiation, it is

254 reduced to avoid overexcitation of the photosystems [8]. It is well-known that photosystems
255 only contain chlorophyll *a*, while antenna complexes present both chlorophyll *a* and *b*[9].
256 Thus, increase in LHCl and II complexes in shaded plants may result in lower chlorophyll *a:b*
257 ratio when compared to sun leaves.

258 Under shading conditions, the environment under the canopy of shading plants is enriched
259 with green light, as this is the predominant wavelength in the light transmitted and reflected
260 by leaves. Chlorophyll *b* shows the maximum absorption peak closest to green wavelength,
261 compared to chlorophyll *a*. Therefore, the reduction of chlorophyll *a:b* ratio is an important
262 strategy to increase the use of the predominant green light, which affects the leaves of
263 shaded plants [37].

264 A quadratic model for the relationship between carotenoid content of coffee plants and the
265 levels of shading was delineated. The conduction of coffee plants in shaded environments
266 resulted in higher levels of carotenoids, with maximum value estimated at 49.3% of LR
267 (Figure 3G). Generally, very intense solar radiation induces elevation of carotenoid levels,
268 since this pigment is involved in protecting the damage caused by excessive light [9], which
269 was not observed in the present study.

UNDER PEER REVIEW



270

271 Fig. 3.SPAD index, chlorophyll b content, ratio between chlorophyll a e b, and
 272 carotenoid content in leaves of coffee plants (*Coffea arabica* L. 'Catuai Vermelho IAC
 273 144') in response to differents light restriction levels (LR) and paclobutrazol doses
 274 (D), at 100 days after the application of the regulator. (A, B) SPAD index (SPAD): (A)
 275 $\hat{Y}^{**} = 59.8123 + 0.292836X - 0.00233598X^2$ ($R^2 = 0.7305$); (C, D) chlorophyll b
 276 content(Chl b): (C) $\hat{Y}^{**} = 0.2856 + 0.00307571X$ ($R^2 = 0.7583$); (E, F) ratio of chlorophyll
 277 a to b (Chl a:b): (E) $\hat{Y}^{**} = 5.4074 - 0.01257X$ ($R^2 = 0.7853$); (G, H) carotenoid content
 278 (Car): (G) $\hat{Y}^* = 0.339864 + 0.00457841X - 0.0000464205X^2$ ($R^2 = 0.9959$). * e **:
 279 significative by regression analysis at 5% e 1% of probability, respectively.

280 However, the effect verified in the present study corroborates with [3], who observed higher
281 carotenoid content in arabica coffee leaves conducted under 85% of light restriction, when
282 compared to those grown in full sun.

283 The higher carotenoid content in shaded coffee plants observed in this work may have
284 occurred due to the greater amount of light absorption complexes per unit of leaf area in
285 these plants [29], which has carotenoids as components of the complex antenna.

286 The content of chlorophyll *b*, chlorophyll *a:b* ratio and carotenoid content of coffee plants
287 were not altered by the application of paclobutrazol (Figure 3D, 3F, and 3H). According to
288 [38], the treatment with triazoles can increase abscisic acid and cytokinins, resulting in
289 increase in chlorophyll and carotenoid contents in leaves. However, for the present work, the
290 dosages of PBZ used were not effective in inducing such changes.

291 It should be emphasized again that the anatomical effect induced by PBZ in increasing
292 thickness and reducing leaf area may interfere with pigment contents when considering the
293 quantification based on the mass of the leaf blade.

294 Light restriction influenced all the characteristics related to leaf gas exchange, with the
295 exception of internal CO₂ concentration in the substomatic chamber. However, no
296 characteristics were affected by PBZ doses or the interaction between LR and PBZ (Table
297 2).

298 Often, limitations of leaf gas exchange in coffee plants are strictly associated with the
299 sensitivity of stomata to the increase in the vapor pressure deficit between leaf and
300 atmosphere [39, 5]. Air temperature attenuation is an important environmental change
301 promoted by shade cultivation [4], and may reduce the above limitations [40].

302 A quadratic model was designed to express stomatal conductance (g_s) response of coffee
303 plants as a function of the levels of light restriction. Initially, it is observed a slight decrease
304 of the values up to the level of 15.3% of shading (3.5% lower than the control). However, the
305 increase was more expressive from the 30.5% of LR level, with maximum g_s at 80% of
306 shading, 58.93% higher than the full sun treatment (Figure 4A).

307 Shading provides a modification in the microclimate of the growing environment, in order to
308 decrease wind speed and leaf temperature, and increase relative humidity of the air. This
309 results in a reduction in vapor pressure deficit and, therefore, reduces stomatal limitations of
310 coffee trees [40]. The highest values of g_s in coffee plants conducted under higher levels of
311 light restriction in this study were associated with this fact.

312 Direct relationship between the increases in light restriction levels and the potential net
313 assimilation rate of CO₂ (*A*) and transpiration rate (*E*) of the coffee plants were verified. The
314 elevation of *A* and *E* values in 73.04 and 43.27%, respectively, was observed for the highest
315 levels of shading (Figures 4C and 4E).

316 Stomatal conductance is the main limiting factor of the photosynthetic rate in plants grown in
317 full sun [40], a fact that may be associated to the increase in *A* values of shaded coffee
318 plants, since g_s was also elevated under these conditions.

319 It is worth mentioning that the increase of *A* under light restriction was similar to the increase
320 in the content of photosynthetic pigments under these conditions. Thus, the higher content of
321 chlorophylls and carotenoids (Figures 2A, 2C, 3C and 3G) may also have contributed to the
322 increase in photosynthetic rates.

323 **Table 2. Analysis of variance summary and coefficients of variation (CV) of stomatal**
 324 **conductance (g_s), net CO₂ assimilation rate (A), transpiration rate (E), internal CO₂**
 325 **concentration (C_i), and carboxylation efficiency (A/C_i) of *Coffea arabica* L. ‘Catuai**
 326 **Vermelho IAC 144’ plants submitted to different light restriction levels (LR) and**
 327 **paclobutrazol doses (D), evaluated at 99 days after the application of the regulator.**
 328 **Vitória da Conquista – BA, 2017.**

SV	df	MEAN SQUARES				
		g_s	A	E	C_i	A/C_i
LR	4	0,1488**	37,93**	2,38**	622,66 ^{ns}	0,000428**
D	4	0,0431 ^{ns}	5,07 ^{ns}	0,11 ^{ns}	254,59 ^{ns}	0,000061 ^{ns}
LR*D	16	0,0229 ^{ns}	3,89 ^{ns}	0,30 ^{ns}	399,76 ^{ns}	0,000049 ^{ns}
BL	3	0,5144**	11,86*	0,99*	3100,54**	0,000076 ^{ns}
Wn	72	0,0357	3,25	0,32	266,98	0,000036
CV (%)		60,76	27,84	24,49	5,31	28,34

329 ^{ns}, * e **: non-significant, significant by “F” test at 5% and 1% of probability, respectively.

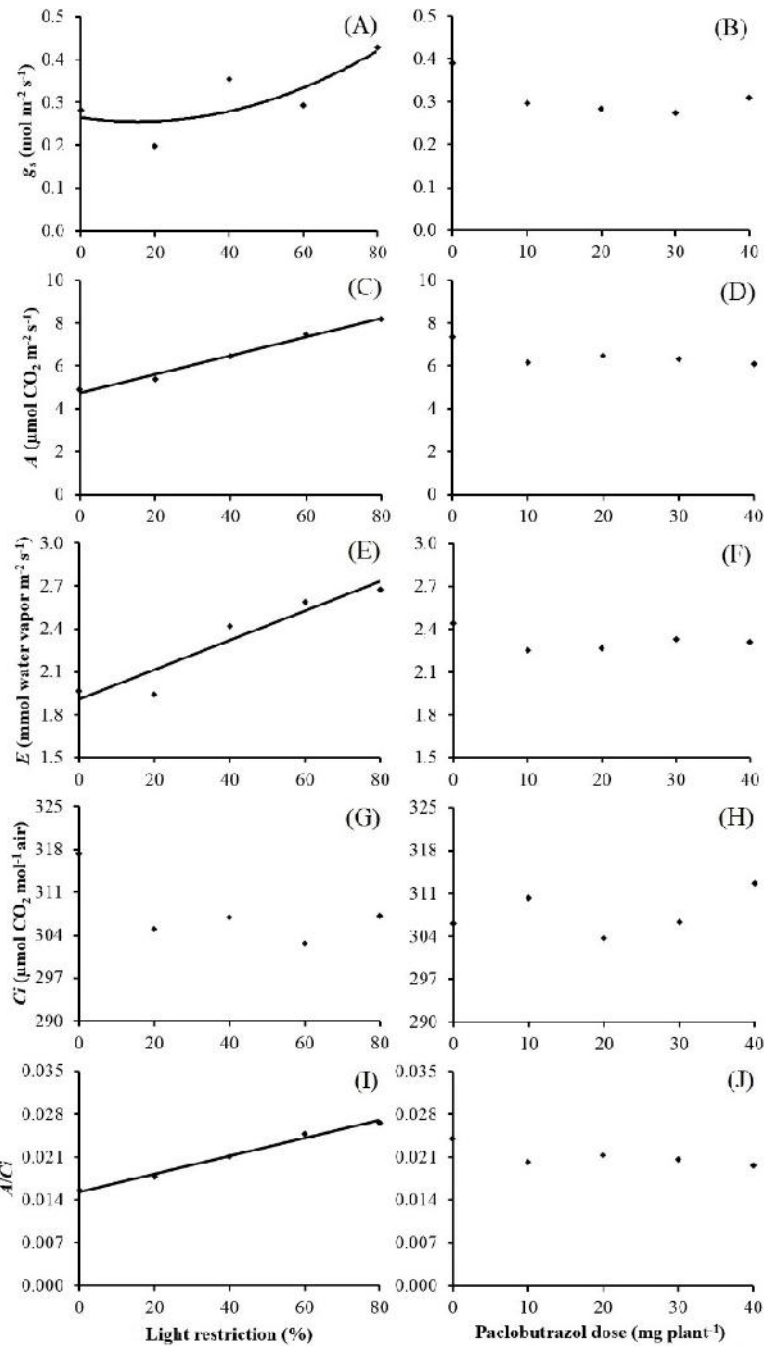
330 The increase of E observed in the shaded coffee plants was related to the higher values of
 331 g_s (less leaf stomatal resistance), since this process results mainly from the diffusion of
 332 water vapor through stomatal opening.

333 Higher values of g_s and E were verified in coffee plants conducted under artificial light
 334 restriction, compared to coffee plants grown in full sun [6]. Higher photosynthetic rates in
 335 shaded coffee plants were verified by [41], compared to plants conducted without light
 336 restriction.

337 In the present work, although the shaded coffee plants presented less resistance to gas
 338 diffusion, internal CO₂ concentration (C_i) was not altered by light restriction levels (Figure
 339 4G). [7] also observed no difference between the C_i of shaded coffee trees and full sun.

340 On the other hand, carboxylation efficiency (A/C_i) showed a tendency of linear increase as a
 341 function of the increase of shading levels (Figure 4I). This parameter was elevated up to
 342 75.9% at the level of 80% of shading, compared to the control treatment (full sun). This
 343 result was related, in part, to the temperature attenuation in shaded environments.

344 Ribulose-1,5-bisphosphate-carboxylase/oxygenase (rubisco) enzyme present in chloroplasts
 345 can catalyze both photosynthesis and photorespiration. The rates of each of these
 346 processes depend on the activity of rubisco as carboxylase or oxygenase, and they are
 347 modified by the environmental conditions [9].



348

349 **Fig. 4.** Leaf gas exchanges of coffee plants (*Coffea arabica* L. 'Catuaí Vermelho IAC
 350 144') in response to different light restriction levels (LR) and paclobutrazol doses
 351 (D), at 99 days after the application of the regulator. (A, B) stomatal conductance (g_s):
 352 (A) $\hat{Y}^* = 0,2644 - 0,0012025X + 0,000039375X^2$ ($R^2 = 0,6289$); (C, D) net CO₂
 353 assimilation rate (A): (C) $\hat{Y}^{**} = 4,7421 + 0,043295X$ ($R^2 = 0,9884$); (E, F) transpiration
 354 rate (E): (E) $\hat{Y}^{**} = 1,9066 + 0,0103125X$ ($R^2 = 0,8940$); (G, H) internal CO₂ concentration
 355 (C_i); (I, J) carboxylation efficiency (A/C_i): (I) $\hat{Y}^{**} = 0,015343 + 0,000145525X$ ($R^2 =$
 356 $0,9892$). * e **: significative by regression analysis at 5% e 1% of probability,
 357 respectively.

358 Although the activity of the enzyme as carboxylase increases with temperature, the affinity of
359 rubisco by CO₂, as well as the solubility of CO₂, decrease. This results in increases in
360 photorespiratory activity at higher temperatures and, consequently, lower carboxylation
361 efficiency [42].

362 On this way, increases in the carbon assimilation rate due to the light restriction, associated
363 to a constant *C_i* between the treatments, resulted in higher carboxylation efficiency in
364 shaded coffee plants.

365 PBZ treatment can alter several aspects of leaf gas exchange in many species [12,43-44]. In
366 the present study, however, PBZ application via soil did not influence any of the parameters
367 related to gas exchange of coffee plants (Figures 4B, 4D, 4F, 4H and 4J).

368 According to [12], the increase in abscisic acid contents resulting from triazole application
369 may result in partial stomatal closure and reduction in the transpiration rate of treated plants.
370 On the other hand, PBZ application did not alter stomatal conductance in coffee plants [45].

371 The effect of PBZ on increasing [44] or reducing [46] the rate of CO₂ assimilation is
372 modulated by dosage and form of application. In coffee plants, [45] found that application of
373 lower concentrations of PBZ via leaf yielded higher photosynthetic rates and carboxylation
374 efficiency, while higher concentrations restricted both processes.

375 The absence of the effect of PBZ on leaf gas exchanges of coffee plants, in this work, was
376 possibly due to the fact that the dosages studied were too low to alter these parameters.
377

378 **4. CONCLUSION**

379
380 Light restriction optimized the photosynthetic apparatus of the plants, mainly at levels
381 (explain) close to 60%, and favored the leaf gas exchanges of arabica coffee in initial
382 growth. The application of paclobutrazol in the dosages studied resulted in little or no effect
383 (explain doses) on the levels of photosynthetic pigments, and did not influence the leaf gas
384 changes of young arabica coffee plants.

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392
393

394 **REFERENCES-CAN INCLUDE RECENT YEAR REFERENCES**

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