

Original Research Article

Genetic Variability, Correlation and Path Coefficient Analysis in advanced Generation of *Brassica napus* L.

ABSTRACT

The present study was conducted involving 62 F₃ genotypes of *Brassica napus* L. at the experimental farm of Sher-e-Bangla Agricultural University, Dhaka to ponder the genetic variability, phenotypic, genotypic and environmental coefficient of variation, heritability and genetic advance, correlation, path coefficient and genetic diversity analysis in a RCBD with three replications during November 2014 to February 2015. The investigations aimed to select the best segregating genotypes for the yield improvement of rapeseed. Analysis of variance indicated that the genotypes were found significantly different for all the characters considered. The relatively phenotypic coefficient of variation (PCV) was higher than the genotypic coefficient of variation (GCV) for all the traits inspected. The high GCV value was observed for the NSP. PH, NSP, SL, NSS, and SYP indicated high broad sense heritability. The significant positive correlation with SYP was found in PH (0.368**), the NPB (0.332**), NSB (0.382**), NSP (0.549**), and SL (0.037**). The results of path coefficient analysis uncovered that PH (0.582), 50F (0.390), DM (0.575), NPB (0.678), NSB (0.182), and TSW (0.289) had a positive direct impact on SYP and thus it was concluded that these traits could be exploited for the enhancement of yield potential of rapeseed. Based on the agronomic execution genotypes G8, G14, G19, G21, G47, and G55 might be proposed for future hybridization program on Bangladesh and helps rapeseed breeders to amend their breeding activities.

24 **Key words:** *Brassica juncea*, *Brassica rapa*, Genetic advance; Genetic advance percentage
25 of mean; Heritability.

26 INTRODUCTION

27 *Brassica* oil is the world's most important edible vegetable oils. In Bangladesh,
28 different types of *Brassica* species are developed through the breeding program. The genomic
29 constitutions of the three diploid elemental species of *Brassica* are AA for *B. rapa*, BB for *B.*
30 *nigra* and CC for *B. oleracea* having the diploid chromosome number of 20, 16, and 18
31 respectively. On the other hand, the species *B. juncea* (AABB), *B. carinata* (BBCC) and *B.*
32 *napus* L. (AACC) is the amphidiploids [1]. Approximately, 70% of the total cultivated
33 mustard in Bangladesh is occupied of either *B. rapa* or *B. napus* L [2]. *Brassica* oil crops are
34 the most critical group of species that supply essential edible oil in Bangladesh [3]. Mustard
35 and rapeseed seeds contain 40%–45% oil and 25% protein [3].

36 The per capita consumption of consumable oil in our country is 8 g/day when
37 contrasted with a need of 40 g/day [4]. The shortage of edible oil has turned into a constant
38 issue for the country [5]. The average per hectare yield of oilseed crops in Bangladesh was
39 740 kg, and average world production was 2400 kg [6]. The productivity of oilseed crops in
40 Bangladesh is comparatively lower than the oilseed growing countries of the world. The
41 logical reason behind such poor yield in Bangladesh might be credited because of the lack of
42 improved varieties and poor management practices [7]. Besides, the cultivated area of
43 mustard is comparatively lower than other crops as a consequence of rice-dependent cropping
44 pattern, and as such, it is strenuous to change [8].

45 A plant breeding program may be divided into three main steps viz. developing
46 germplasms with various genetic resources, selection of the best individual from the
47 expanded resources and utilized the best selected individual to develop a suitable and

48 superior variety. There is plenty of scopes to increase yield per unit of area through breeding
49 unrivaled varieties. The knowledge on genetic variability [9], heritability and genetic advance
50 [10] and character association is a prerequisite for starting a fruitful breeding program
51 expecting to develop high yielding varieties [8]. High heritability value indicates the strategy
52 for selection of suitable character by the phenotypic performance of the respective genotype
53 and genetic advance showed the progress for the choice of the best individual [11]

54 Determination of correlation coefficient between the characters has considerable
55 importance in selecting breeding materials. Path coefficient technique splits the correlation
56 coefficient into direct and indirect effects [12] via alternative aspects or pathways and in this
57 way allows an essential examination of components that influence a given correlation and can
58 be useful in detailing an efficient selection strategy [13]. Therefore, the path coefficient
59 analysis has been found to provide more particular data on the direct and indirect impact of
60 each of the segment characters upon seed yield [14]. Inter-varietal and inter-specific
61 hybridization are essential for creating the variation or transfer gene of interest from wild
62 species in rapeseed improvement programme [15]. Genetic variability is one of the criteria
63 for parent choice [16]. Consideration of more diverse parents (inside the farthest point) in
64 hybridization should build the possibility of acquiring the most extreme heterosis [17] and
65 give the full range of variability in segregating generations. This present investigation was
66 undertaken to evaluate the variability, character association and the selection criteria for the
67 best genotypes among the advanced generations of *B. napus*.

68 **MATERIALS AND METHODS**

69 The present research was led at the experimental farm of Sher-e-Bangla Agricultural
70 University, Dhaka during November 2014 to February 2015. The area of the trial site is 23⁰
71 74' N latitude and 90⁰ 35' E longitudes with 8.2 meters above from the ocean level. The soil

72 of the experimental site in Agro-ecological region of “Madhupur Tract” (AEZ No. 28). The
 73 land was clay loam in texture and olive gray with common fine to medium distinct dark
 74 yellowish brown mottles. The pH range is 5.47–5.63 and organic carbon content is 0.82%.

75 The healthy seeds of sixty-two F_3 of *B. napus* L. were collected from the Dept. of
 76 Genetics and Plant Breeding, Sher-e-Bangla Agricultural University, which were utilized as
 77 test materials (Table 1). Randomized complete block design (RCBD) was used with three
 78 replications. The total area of the plot was $56 \text{ m} \times 14 \text{ m} = 784 \text{ m}^2$ along with $56 \text{ m} \times 3.5 \text{ m}$
 79 replication⁻¹ plot and the distance between replication to replication was 1 m. 30 cm spacing
 80 was used between the line to line.

81 The data were recorded on ten randomly selected plants for each cross and each
 82 parent on 50F, days to 80% maturity, PH (cm), NPB, NSB, NSP, SL (cm), NSS, 1000 seed
 83 weight (g) and SYP (g).

84 **Table 1.** List of sixty two genotypes of advanced generation *B. napus* used for the experiment
 85

Genotypes	F₃ Populations	Genotypes	F₃ Populations
G1	BS-7 × Nap-206	G32	Nap-2012 × Nap-2013
G2	BS-7 × Nap-2012	G33	Nap-2012 × Nap-2022
G3	BS-7 × Nap-2013	G34	Nap-2037 × Bs-13
G4	BS-7 × Nap-2057	G35	Nap-2037 × Nap-206
G5	BS-13 × Nap-179	G36	Nap-2037 × Nap-248
G6	BS-13 × Nap-206	G37	Nap-2037 × Nap-2001
G7	BS-13 × Nap-2001	G38	Nap-2037 × Nap-2012
G8	BS-13 × Nap-2013	G39	Nap-2037 × Nap-2013
G9	BS-13 × Nap-2022	G40	Nap-2037 × Nap-2022
G10	Bs-13 × Nap-2057	G41	Nap-2037 × Nap-2057
G11	Nap-179 × Nap-206	G42	Nap-2057 × Nap-248
G12	Nap-179 × Nap-2001	G43	Nap-2057 × Nap-2012
G13	Nap-179 × Nap-2012	G44	Nap-2057 × Nap-2022
G14	Nap-179 × Nap-2013	G45	Nap-9908 × Bs-13
G15	Nap-179 × Nap-2022	G46	Nap-9908 × Nap-206
G16	Nap-179 × Nap-2057	G47	Nap-9908 × Nap-248
G17	Nap-206 × Nap-2012	G48	Nap-9908 × Nap-2001
G18	Nap-206 × Nap-2013	G49	Nap-9908 × Nap-2012
G19	Nap-206 × Nap-2022	G50	Nap-9908 × Nap-2013

G20	Nap-206 × Nap-2057	G51	Nap-9908 × Nap-2022
G21	Nap-248 × Nap-159	G52	Nap-9908 × Nap-2037
G22	Nap-248 × Nap-206	G53	Nap-9908 × Nap-94006
G23	Nap-248 × Nap-2012	G54	Nap-94006 × Bs-13
G24	Nap-248 × Nap-2013	G55	Nap-94006 × Bs-7
G25	Nap-248 × Nap-2022	G56	Nap-94006 × Nap-179
G26	Nap-248 × Nap-2057	G57	Nap-94006 × Nap-206
G27	Nap-2001 × Nap-179	G58	Nap-94006 × Nap-2001
G28	Nap-2001 × Nap-206	G59	Nap-94006 × Nap-2012
G29	Nap-2001 × Nap-248	G60	Nap-94006 × Nap-2013
G30	Nap-2001 × Nap-2013	G61	Nap-94006 × Nap-2022
G31	Nap-2001 × Nap-2022	G62	Nap-94006 × Nap-2057

86 Analysis of variance was calculated using MS Excel software using MSTAT-C software.
87 The phenotypic and genotypic variance was evaluated by [18]. The genotypic (GCV) and
88 phenotypic (PCV) coefficient of variation was computed by [19]. Heritability and genetic
89 advance were determined as described by [20, 21]. The simple correlation coefficient was
90 obtained by the method of [20, 22] and path coefficient analysis was carried out by [23].

91 RESULTS AND DISCUSSION

92 Variability, Heritability and Genetic advance

93 Significant variations were observed for most of the characters among sixty two F₃
94 materials of *B. napus* L. The values of mean, range CV%, phenotypic variances, genotypic
95 variances, PCV and GCV for different yield related characters were shown in Table 2a and
96 2b.

97 **Table 2a.** Estimation of range and genetic parameters in ten characters of 62 genotypes in *B. napus*
98 L.
99

Parameters	Range	Mean	MS	CV (%)	σ^2_p	σ^2_g	σ^2_e
50F	32.50-44.00	38.12	9.30**	5.02	6.49	2.82	3.67
DM	79.50-91.50	86.98	17.22**	3.44	13.08	4.14	8.93
PH	90.73-136.00	111.81	156.52**	4.26	89.59	66.92	22.67
NPB	2.30-3.94	3.09	0.27**	13.32	0.22	0.05	0.17
NSB	0.88-3.38	2.14	0.59**	15.44	0.45	0.15	0.30
NSP	85.35-223.80	129.17	1706.54**	9.94	935.67	770.88	164.79

SL	6.86-14.26	7.97	2.05**	5.67	1.13	0.92	0.20
NSS	16.73-29.20	22.66	13.09**	6.36	7.59	5.51	2.08
TSW	3.14-3.87	3.46	0.05**	5.71	0.05	0.01	0.04
SYP	45.62-76.72	60.45	126.52**	3.61	65.64	60.89	4.75

100

101 **, * Correlation is significant at the 0.01 and 0.05 level, respectively.

102 Here, 50F= Days to 50% flowering, DM= Days to maturity, PH=Plant height, NPB= Number of
 103 primary branches per plant, NSB= Number secondary branches per plant, NSP= Number of siliqua
 104 per plant, NSS= Number of seed per siliqua, SL= Siliqua length (cm), TSW=Thousand seed weight
 105 (g), SYP=Seed yield per plant, MS= Mean sum of square, CV(%)= Coefficient of variation, σ^2_p =
 106 Phenotypic variance, σ^2_g = Genotypic variance, σ^2_e =Environmental variance.

107

108

109 **Table 2b.** Estimation of range and genetic parameters in ten characters of 62 genotypes in *B. napus*
 110 L.

111

Parameters	PCV	GCV	ECV	Heritability	Genetic advance (5%)	Genetic advance (% mean)
50F	6.68	4.40	5.02	43.46	2.28	5.98
DM	4.16	2.34	3.44	31.70	2.36	2.71
PH	8.47	7.32	4.26	74.70	14.57	13.03
NPB	15.26	7.43	13.32	43.73	0.23	7.46
NSB	31.23	18.12	25.44	63.66	0.46	16.62
NSP	23.68	21.49	9.94	82.39	51.91	40.19
SL	13.32	12.06	5.67	81.90	1.79	22.49
NSS	12.15	10.35	6.36	72.60	4.12	18.18
TSW	6.19	2.39	5.71	14.89	0.07	1.90
SYP	13.40	12.91	3.61	92.76	15.48	25.61

112

113 Here, PCV = Phenotypic coefficient of variation, GCV = Genotypic coefficient of variation, ECV =
 114 Environmental coefficient of variation.

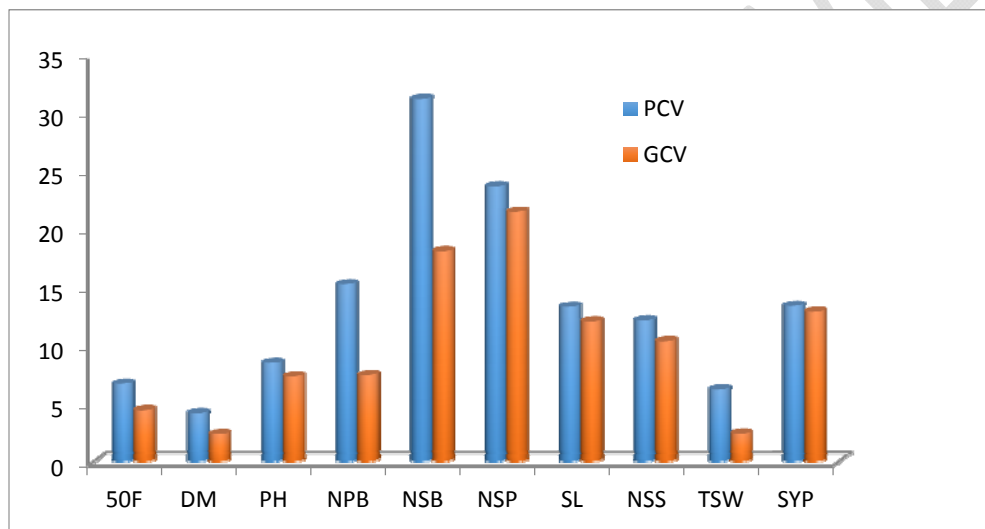
115

116 **Days to 50% flowering (50F)**

117 The days to 50% flowering were observed the lowest (32.50 days) in G19 and most
 118 noteworthy (44 days) was observed in G48 (Table 2a). The PCV (6.68) was slightly higher
 119 than the GCV (4.40) (Table 2b). Days to 50% flowering exhibited low heritability (43.46%)
 120 with low genetic advance (2.28), and genetic advance in the percentage of the mean (5.98)
 121 demonstrated that this attribute was controlled by non-additive gene (Table 2b).

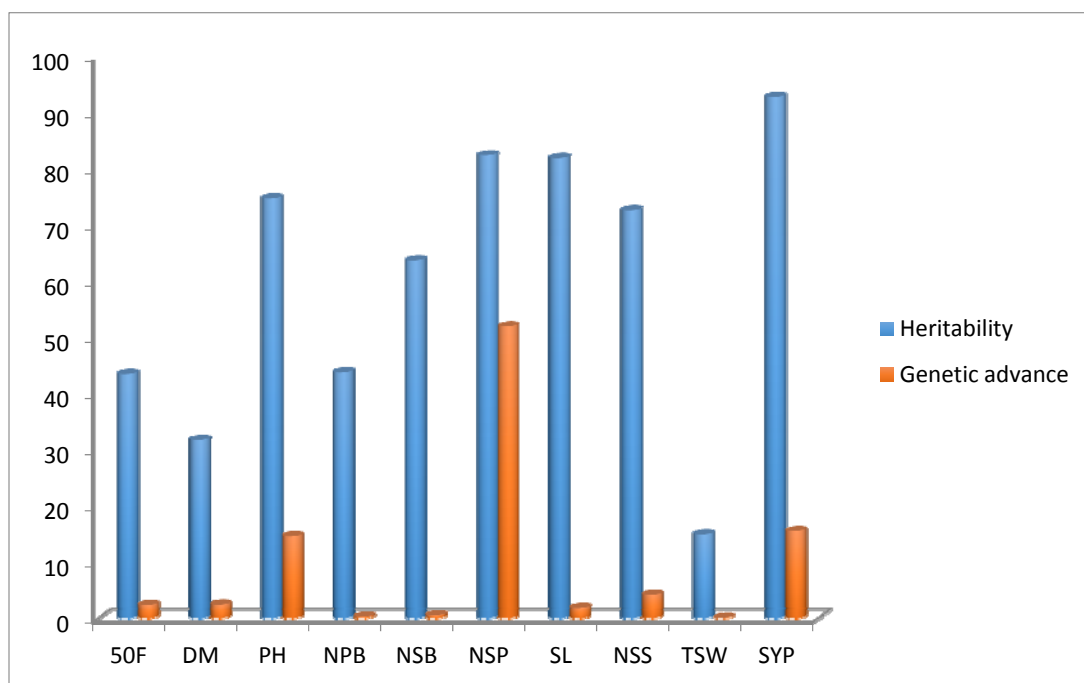
122 **Days to maturity (DM)**

123 Maturity delayed the maximum in G32 (91.50 days), and the earliest maturity was
124 observed in G19 (79.50 days) (Table 2a). The PCV (4.16) was higher than the GCV (2.34)
125 (Table 2b), which proposed that the environment has a significant role in the expression of
126 this trait. Days to maturity demonstrates low heritability (31.70) with low genetic advance
127 (2.36), and the genetic advance in the percentage of the mean indicated that this trait was
128 controlled by the non-additive gene and medium probability of choosing genotypes that
129 would mature earlier (2.71) (Table 2b). The frequency of the segregating plants showing
130 reduced maturity was comparatively higher than the other crosses.



131

132 **Figure 1.** Genotypic and phenotypic coefficient of variation in *Brassica napus* L.



133

134 **Figure 2.** Heritability and genetic advance over mean in *Brassica napus* L.

135 **Plant height (PH, c)**

136 In this investigation, the highest PH was observed in G19 (136c) whereas the
 137 minimum PH was observed in G13 (90.73c) (Table 2a). The PCV (8.47) value is slightly
 138 higher than GCV (7.32) value (Table 2b). The most noteworthy variation in PH among
 139 parents and their hybrid was observed by [24, 25]. Plant height showed high heritability
 140 (74.70%) with the moderately high genetic advance (14.57) and the genetic advance in the
 141 percentage of mean (13.03) (Table 2b), uncovered the likelihood of the prevalence of the
 142 additive gene action in the inheritance of this trait and indicating that this trait could be
 143 improved through the selection process [26]. High variability in PH for *B. juncea*, *B. rapa*,
 144 and *B. napus* L. was likewise seen by [27].

145 **Number of primary branches per plant (NPB)**

146 The highest NPB was observed in G47 (3.94) whereas the minimum was in G26
 147 (2.30) (Table 2a). PCV (15.26) value is comparatively higher than the corresponding GCV
 148 (7.43) value indicating the apparent variation not only due to genotypes but also due to the

149 considerable influence of the environment (Table 2b). [28] also found significant differences
150 in the NPB. The NPB displayed low heritability (43.73%) with the low genetic advance
151 (0.23) and the genetic advance in the percentage of the mean (7.46), which uncovered that the
152 non-additive gene controlled this trait. As a whole, the low heritability and the consequent
153 low genetic advance indicated the lower plausibility of choosing genotypes for this attribute
154 [29]. However, some of the individual plants showed quite a reasonable lower primary
155 branches which were selected for further study in the next generation. Low heritability
156 coupled with low genetic advance was also found by [30].

157 **Number of secondary branches per plant (NSB)**

158 The highest NSB was observed in G55 (3.38) whereas the minimum number was in
159 G18 (0.88) (Table 2a). The PCV value (31.23) is higher than the corresponding GCV value
160 (18.12) (Table 2b). It indicated the presence of considerable variability among the genotypes
161 for this trait. [31] found the highest GCV for the NSB while working on 24 genotypes of *B.*
162 *napus* L. [28] found significant differences for the NSB. Moderately high heritability (63.66)
163 along with the low genetic advance (0.46) and the genetic advance in the percentage of the
164 mean (16.62) (Table 2b) revealed that the non-additive gene controlled this trait [16, 32]. As
165 a whole, the moderately high heritability and the consequent low genetic advance indicated
166 the lower possibility of selecting genotypes. Moderately high heritability coupled with low
167 genetic advance was also found by [33].

168 **Number of siliqua per plant (NSP)**

169 The NSP was observed the highest in G14 (223.80) and the lowest in G24 (85.35)
170 (Table 2a). PCV (23.68) had a similar trend as GCV (21.49) (Table 2b). The difference
171 between the PCV (23.68) and GCV (21.49) indicated the existence of adequate variation
172 among the genotype. The high heritability (82.39) with the high genetic advance (51.91) and

173 the genetic advance in the percentage of the mean (40.19) was observed for this trait revealed
174 the possibility of the predominance of additive gene action in the inheritance of this trait. This
175 trait possessed high variation; it is the high potential for active selection for further genetic
176 improvement of this character [32]. [34] also observed high heritability coupled with the high
177 genetic advance for this trait. [34] reported that the NSP were highly heritable coupled with
178 high genetic advance.

179 **Length of silique (SL)**

180 Length of silique was observed the highest in G21 (14.26c), and the minimum length
181 of the pod was observed in G55 (6.86c) (Table 2a). Relatively medium PCV (13.32) and
182 GCV (12.06) was found for this trait (Table 2b). Silique length showed the high heritability
183 (81.90) with the low genetic advance (1.79), and the low genetic advance in the percentage of
184 the mean (22.49) indicated that this trait was controlled by non-additive gene (Table 2b).

185 **Number of seeds per silique (NSS)**

186 The NSS was observed highest in G21 (29.20), and the lowest was in G55 (16.73)
187 (Table 2a). The value of PCV and GCV were 12.15 and 10.35 respectively for the number of
188 seeds per silique (Table 2b) which indicating that medium variation exists among the
189 different genotypes [35]. Similar variability was also recorded by [36]. Number of seeds per
190 silique showed high heritability (72.60%) coupled with the high genetic advance (4.12) and
191 the high genetic advance in the percentage of the mean (18.18) (Table 2b) indicated that this
192 trait was controlled by additive gene and choice for this character would be helpful [15, 17,
193 32]. High heritability coupled with the high genetic advance for this trait was likewise seen
194 by [37].

195 **Thousand seed weight (TSW, g)**

196 Thousand seed weight was found the maximum in G15 (3.87g) whereas the minimum
197 was found in G50 (3.14g) (Table 2a). The PCV (6.19) and GCV (2.39) were close to each
198 other (Table 2b). This trait had low heritability (14.89%), low genetic advance (0.07) and
199 genetic advance in the percentage of the mean (1.90) revealed that this trait was controlled by
200 non-additive gene. High heritability for this trait was also observed by [38].

201 **Seed yield per plant (SYP, g)**

202 Seed yield per plant was found the maximum in G8 (76.72 g), and the minimum was
203 in G24 (45.62 g) (Table 2a). The values of PCV and GCV were 13.40 and 12.91. Similar
204 variability was also found by [39, 40]. Seed yield per plant showed the high heritability
205 (92.76%) with the high genetic advance (15.48) and moderately the high genetic advance in
206 the percentage of the mean (25.61) (Table 2b) indicated that this trait was controlled by the
207 additive gene and selection for this character would be effective [32]. High heritability
208 coupled with the high genetic advance for this trait was also observed by [33]. High
209 heritability and genetic advance for SYP were reported by [37] while working with 22
210 genotypes of *B. napus* L.

211 The knowledge of variability is prerequisite for simultaneous selection and significant
212 improvement of rapeseed genotypes. The NSB and number of silique per plant demonstrates
213 the presence of broader variability suggesting that these traits could be the excellent
214 candidate for developing new high yielding rapeseed varieties [41]. GCV is a good indicator
215 that imparts information on the existence of genetic variability present in various quantitative
216 traits, but it is lack of providing the clear picture about the heritable variation contributed to
217 GCV [42]. In the current investigation, we found a cross-link between GCV and PCV for PH,
218 NSP, SL, NSS, and SYP indicating that environment influenced less for the expression of
219 these traits. The highest GCV and PCV value observed for traits- NPB, NSB, and NSP

220 suggesting that selection for these traits would be rewarding to isolate more promising lines.
221 Moderate GCV and PCV value were found for SL, NSS, and SYP indicating that vigorous
222 selection method is utilized for the improvement of these parameters. The descriptors such as
223 50F, DM, PH, and TSW displayed low GCV and PCV value recommended breeder to find
224 out the high variability source for these traits for the future improvement.

225 GCV along with the heritable components estimation would render the outcome for
226 proper selection for utilizing them in the future breeding program [43]. Genetic and
227 environmental factors are the contributors to the observed variation in a population. Genetic
228 factors are the only heritable portion from generation after generation. We cannot solely
229 confirm the expected genetic gain in the next generation unless we consider heritability in
230 conjunction with the genetic advance [44] because it provides reliability for the selection of
231 the trait of interest from the variable entries [44]. Characters have high heritability and high
232 genetic advance as a percentage of mean is considered as a powerful genetic tool in the
233 selection round of the best genotype. These characters are governed by the additive gene
234 action and have a less chance to influence by the environment [45]. We found high
235 heritability coupled with high genetic advance as a percentage of the mean for the trait NSB
236 and NSP. Thus these traits have a less tendency to guide by the environment. Three types of
237 heritability was found in corn- low (0-20%), medium (20-60%) and high (above 60%) [46].

238 **Correlation coefficient**

239 Seed yield is a complex product being influenced by several quantitative traits. Some
240 of these traits are highly associated with seed yield. The analysis of the relationship among
241 those traits and their association with seed yield is very much essential to establish selection
242 criteria. The correlation co-efficient between pairs of the attribute for F₃ materials of *B. napus*
243 L. is shown in (Table 3).

244 Days to 50% flowering showed a highly significant and positive correlation with DM
 245 (G= 0.533, P= 0.182) indicated that if 50F increased then DM also increased. It also exhibited
 246 interaction with NSP (G= 0.458, P= 0.316), SL (G= 0.051, P= -0.056) and SYP (G= 0.201, P
 247 = 0.132). However, it had negative interaction with NSP (G= -0.282, P=-0.206) (Table 3).
 248 Insignificant association of these traits indicated that environmental factors largely influenced
 249 the associations between these traits.

250 **Table 3.** Genotypic and phenotypic correlation coefficients among different pairs of yield and yield
 251 contributing characters for different genotype of *Brassica napus* L.

Parameters		DM	PH	NPB	NSB	NSP	SL	NSS	TSW	SYP
50F	G	0.533**	0.320**	-0.184	-0.176	-0.282	0.051	0.458	-0.341**	0.201
	P	0.182*	0.169**	-0.083	-0.02	-0.206	-0.056	0.316	-0.314**	0.132
DM	G		0.330	-	-0.284	0.063**	0.091**	0.018**	-0.586	-0.065
	P		0.074	0.032	-0.016	0.132**	0.037**	0.074**	-0.109	-0.04
PH	G			0.055**	0.194*	0.396**	0.038**	0.038**	-0.597	0.368**
	P			0.078**	0.187*	0.375**	0.039**	0.041**	-0.234	0.317**
NPB	G				0.576**	0.397**	0.398	0.581	-0.165	0.332**
	P				0.626**	0.276**	0.160	0.163	-0.164	0.167*
NSB	G					0.507**	0.381	-0.284	-0.188*	0.382**
	P					0.414**	0.180	0.188	-0.190*	0.236**
NSP	G						-0.159	0.039	0.200	0.549**
	P						0.136**	0.013	0.071	0.531**
SL	G							0.489**	-0.018	0.037**
	P							0.341**	-0.009	0.048**
NSS	G								0.849	0.074
	P								0.230*	0.047
TSW	G									0.663
	P									0.304

252 ** = Significant at 1%. , * = Significant at 5%.

253 Days to maturity showed significant and positive correlation with NSS (G= 0.018, P=
 254 0.074), SL (G= 0.091, P= 0.037) and NSP (G= 0.063, P= 0.132) (Table 3). It had negative
 255 correlation with SYP (G= -0.065, P= -0.04), TSW (G= -0.586, P= -0.109) (Table 3).
 256 Insignificant association of these traits indicated that environmental factors largely influenced
 257 the associations between these traits. [47] also revealed that DM had an insignificant and
 258 positive interaction with SYP.

259 Plant height showed highly significant and positive interaction with NPB ($G= 0.055$,
260 $P = 0.078$), NSB ($G= 0.194$, $P= 0.187$), NSP ($G= 0.396$, $P= 0.375$), SL ($G= 0.038$, $P= 0.039$),
261 NSS ($G= 0.038$, $P= 0.041$) and SYP ($G= 0.368$, $P= 0.317$) (Table 3). Highly significant
262 positive associations between PH and other characters indicate that the same gene governed
263 the traits and simultaneous improvement would be effective. It had insignificant and negative
264 interaction with TSW ($G= -0.597$, $P= -0.234$) (Table 3). Insignificant association of these
265 traits indicated that environmental factors largely influence the association between these
266 traits. These findings are showed resemblance to the reports of [47]. The significant positive
267 correlation between PH and SYP was found by [48].

268 Number of primary branches per plant showed positive and significant interaction
269 with NSB ($G= 0.575$, $P= 0.626$), NSP ($G= 0.397$, $P= 0.276$) and SYP ($G= 0.332$, $P= 0.167$)
270 (Table 3). These were suggesting if the NPB increases then SYP also increases. It had
271 insignificant and positive correlation with SL ($G= 0.398$, $P= 0.160$) and NSS ($G= 0.581$, $P=$
272 0.163). However, insignificant and negative interaction was found in TSW ($G= -0.165$, $P= -$
273 0.164) (Table 3). Insignificant association of these traits indicated that environmental factors
274 largely influence the association between these traits. Number of secondary branch showed
275 highly significant and positive interaction with NSP ($G= 0.507$, $P= 0.414$) and SYP ($G=$
276 0.382 , $P= 0.236$) indicated that the same gene governed the traits, and simultaneous
277 improvement would be effective, and branching was an important contributor to yield,
278 independent of its association with plant canopy size. It had insignificant correlation with SL
279 ($G= 0.381$, $P= 0.180$) and NSS ($G= -0.284$, $P= 0.188$). However, it had a significant and
280 negative interaction with a TSW ($G = -0.188$, $P= -0.190$) (Table 3). Insignificant association
281 of these traits indicated that environmental factors largely influence the association between
282 these traits. These findings are showing similar to the reports of [28].

283 The NSP showed a significant and positive correlation with SYP ($G= 0.549$, $P=$
284 0.531) (Table 3) whereas the insignificant and positive interaction was found in NSS ($G=$
285 0.039 , $P= 0.013$), TSW ($G= 0.200$, $P= 0.071$) (Table 3). Insignificant association of these
286 traits indicated that environmental factors largely influence the association between these
287 traits. [49] reported that NSS had a positive and insignificant effect on SYP.

288 Siliqua length showed a highly significant and positive correlation with SYP ($G=$
289 0.037 , $P= 0.048$) and NSS ($G= 0.489$, $P= 0.341$) (Table 3) indicated that the traits were
290 governed by same gene and simultaneous improvement would be effective. [50] reported that
291 if SL increased then SYP will increase. Insignificant and negative correlation found with
292 TSW ($G= -0.018$, $P= -0.009$). Number of seeds per silique showed positive interaction with
293 TSW ($G= 0.849$, $P= 0.230$) and SYP ($G= 0.074$, $P= 0.047$) (Table 3).

294 Thousand seed weight showed insignificant and positive interaction with SYP ($G=$
295 0.663 , $P= 0.304$) (Table 3). Insignificant association of these traits indicated that
296 environmental factors largely influence the association between these traits. [51] found
297 positive associations which support the results.

298 **Path Coefficient analysis**

299 Association of character determined by correlation coefficient may not provide an
300 exact picture of the relative importance of the direct and indirect influence of each of yield
301 components on seed yield per hectore. To find out a clear view of the inter-relationship
302 between SYP and other yield attributes, direct and indirect effects were worked out using
303 path analysis at the phenotypic level which also measured the relative importance of each
304 component. Seed yield per plant was considered as a resultant (dependent) variable, and all

305 other characters were causal (independent) variables. Estimation of the direct and indirect
306 effect of path coefficient analysis for *B. napus* L. is presented in Table 4.

307 Path coefficient analysis revealed that 50F had a positive direct effect (0.390) on SYP.
308 [52] explained that 50F had a positive direct effect on SYP. 50F showed a positive indirect
309 effect on DM (0.036), PH (0.125), NSP (0.027) and negative effect on NPB (-0.072), NSB (-
310 0.069), SL (-0.006), NSS (-0.195) and TSW (-0.388) (Table 4).

311 Days to maturity had a positive direct effect (0.575) on SYP. This trait had a positive
312 indirect effect through NPB (0.172), NSB (0.163), NSP (0.036), SL (0.052), NSS (0.010) and
313 TSW (0.169) (Table 4). [53] revealed that DM had a positive direct effect on yield. On the
314 other hand, DM had negative indirect effect via 50F (-0.208) and PH (-0.306).

315

316 **Table 4.** Path coefficient analysis showing direct and indirect effects of different characters on yield
 317 of mustard
 318

Parameter s	Direct (Bold) and Indirect effect									Genotypic correlation with yield
	50F	DM	PH	NPB	NSB	NSP	SL	NSS	TSW	
50F	0.390	0.036	0.125	- 0.072	- 0.069	0.027	- 0.006	- 0.195	- 0.388	-0.201*
DM	- 0.208	0.575	- 0.306	0.172	0.163	0.036	0.052	0.010	0.169	0.065
PH	- 0.186	- 0.192	0.582	- 0.037	- 0.113	0.037	0.005	- 0.016	- 0.173	0.368**
NPB	- 0.125	0.203	0.032	0.678	0.105	0.037	- 0.049	- 0.247	- 0.048	0.332**
NSB	- 0.032	0.052	0.035	0.390	0.182	0.048	- 0.047	- 0.166	- 0.054	0.382**
NSP	0.110	0.006	0.230	0.269	0.092	- 0.094	- 0.020	- 0.017	- 0.058	0.549**
SL	0.020	- 0.011	- 0.022	0.270	0.069	0.015	- 0.124	- 0.061	- 0.005	0.037
NSS	0.179	- 0.008	- 0.022	0.394	- 0.071	0.004	- 0.208	- 0.425	- 0.245	-0.074
TSW	0.523	- 0.337	0.347	0.112	- 0.034	- 0.019	- 0.002	0.361	0.289	0.663**

319 Residual effect: **0.3123**

320 **, * Correlation is significant at the 0.01 and 0.05 level, respectively

321
 322 Plant height had a positive direct effect (0.582) on yield per plant. Also had a positive
 323 indirect effect via the NSP (0.037) and SL (0.005) (Table 4). Plant height showed negative
 324 indirect effect on 50F (-0.186), DM (-0.192), NPB (-0.037), NSS (-0.113), NSS (-0.016) and
 325 TSW (-0.173).
 326

327 The NPB had a positive direct effect on SYP (0.678) (Table 4). This trait had a
 328 positive indirect effect via DM (0.203), PH (0.032), NSB (0.105) and NSP (0.037). On the
 329 other hand, the negative indirect effect was found on 50F (-0.125), SL (-0.049), NSS (-0.247)
 330 and TSW (-0.048). [54 and 55] reported that the NPB had the direct positive effect on seed
 331 yield.

332 The number of secondary branches had a positive direct effect (0.182) on SYP. It had
 333 a positive indirect effect via DM (0.052), PH (0.035), NPB (0.390), NSP (0.048), NSS

334 (0.166) and TSW (0.054) (Table 4). [53] observed that the NSB had a direct effect on SYP.
335 On the other hand 50F (-0.032) and SL (-0.047) had negative indirect effect on the NSB.

336 The NSP had a negative direct effect (-0.094) on seed yield. This trait had a positive
337 indirect effect on 50F (0.110), DM (0.006), PH (0.230), NPB (0.296) and NSB (0.092) (Table
338 4). This trait had a negative indirect effect via SL (-0.020), NSS (-0.017) and TSW (-0.058).

339 Siliqua length had a direct negative effect (-0.124) on SYP. [56] reported that SL had
340 a negative direct effect on SYP. This trait had indirect positive effect 50F (0.020), on NPB
341 (0.270), NSB (0.069) and NSP (0.015). On the other hand, SL showed indirect negative effect
342 via DM (-0.011) PH (-0.022), NSS (-0.061) and TSW (-0.005).

343 The NSS had a direct negative effect (-0.425) on SYP. This trait had an indirect
344 positive effect on 50F (0.179), NPB (0.394) and NSP (0.004) (Table 4). On the other hand,
345 this trait showed indirect negative effect via DM (-0.008), PH (-0.022), NSB (-0.071), SL (-
346 0.208) and TSW (-0.245).

347 Path coefficient analysis revealed that TSW had positive direct effect on yield per
348 plant (0.289) followed by positive indirect effect via 50F (0.523), PH (0.347), NPB (0.112)
349 and NSS (0.361) (Table 4). [33] reported that TSW had a positive direct effect on SYP. On
350 the other hand, this trait showed negative indirect effect on DM (-0.337), NSB (-0.034), NSP
351 (-0.019) and SL (-0.002).

352 **Selection of parents for future hybridization**

353 Selection of genetically diverse parents is the prime task for any plant breeding
354 activities. Therefore, considering the magnitude agronomic performance the genotypes G8
355 (Nap BS-13 X Nap-2013) for higher SYP, and G47 (Nap-9908 X Nap-248) higher NPB, G19

356 (Nap-206 X Nap-2022) for tallness, lowest 50F and the lowest DM, G21(Nap-248 X Nap-
357 159) for the highest SL and highest NSS, G14(Nap-179 X Nap-2013) highest NSP and G55
358 (Nap-94006 X BS-7) for highest NSB.

359

360 REFERENCES

361

- 362 1. Nath UK, Naz S, Rahman MM. Genetic divergence of *Brassica campestris*, *Brassica*
363 *juncea* parents and their hybrids. Pakistan J. Biol. Sci.2003; 6(10): 936-938.
- 364 2. Rashid MH, Bhuiyan MSR, Akbar MA, Parveen S. Diversity analysis of the germplasm
365 oleiferous *Brassica* species. J. Sher-e-Bangla Agric. Univ. 2009; 3 (1): 30-34.
- 366 3. BBS. Statistical Yearbook of Bangladesh. Bangladesh Bureau of Statistics. Statistics
367 Division, Ministry of Planning. Govt. of the People's Republic of Bangladesh, Dhaka;
368 2011.
- 369 4. Kaul AK, Das ML. Oil seeds in Bangladesh Canada Agric. Sector team. Ministry of
370 Agric. Govt. of the Peoples Republic of Bangladesh; 1978
- 371 5. Parveen S, Bhuiyan MSR, Hossain MS, Rashid MH. Variability study in F₂ progenies of
372 the inter-variety Crosses of *Brassica rapa*. J. Sher-e-Bangla Agric. Univ.2008; 2(2): 34-
373 40.
- 374 6. FAO. FAOSTAT Database of Agriculture (Crops); 2011. Available:<http://www.fao.org>
- 375 7. Rashid MH, Parveen S, Bhuiya MSR. Genetic variability, correlation and path coefficient
376 analysis in nineteen *Brassica rapa* germplasm. J. Sher-e-Bangla Agric. Univ. 2010; 4(1):
377 84-89.
- 378 8. Parveen S, Rashid MH, Bhuiyan MSR. Genetic variation and selection criteria for seed
379 yield related traits in rape seed (*Brassica napus* L.). Bangladesh J. Pl. Breed. Genet. 2013;
380 26(2): 15-22.
- 381 9. Rahman MM, Chowdhury MAZ, Hossain MG, Amin MN, Muktedir MA, Rashid MH.
382 Gene action for seed yield and yield contributing characters in turnip rape (*Brassica rapa*
383 L.). J. Expt. Biosci. 2011; 2(2):67-76.
- 384 10. Sarwar G, Hossain MS, Rashid MH, Parveen S. Assessment of genetic variability for
385 agro-morphological important traits in aman rice (*Oryza sativa* L.). Int. J. Appl. Sci.
386 Biotechnol. 2015; 3 (1): 73-79.

- 387 11. Sarwar G, Rashid MH, Parveen S, Hossain MS. Evaluation of genetic diversity in
388 agromorphological traits of forty two aman rice genotypes (*Oryza sativa* L.) using D²
389 analysis. American J. Experimental Agri. 2015;8(5): 280-288
- 390 12. Sarwar G, Rashid MH, Parveen S, Hossain MS. Correlation and path coefficient analysis
391 for agro-morphological important traits in aman rice genotypes (*Oryza sativa* L.). Adv.
392 Biores. 2015; 6 (4): 40-47.
- 393 13. Sabaghnia N, Dehghani H, Alizadeh B, Mohghaddam M. Interrelationships between seed
394 yield and 20 related traits of 49 canola genotypes in non-stressed and water stressed
395 environments. Spanish J. Agri. Res. 2010; 8: 356-370.
- 396 14. Behl RK, Chowdhury BD, Shing RP, Shing DP. Morphophysiological determinates of oil
397 yield in *Brassica juncea* under dryland conditions. Indian J. Genet. Pl. Breed.1992; 52(3):
398 280-284.
- 399 15. Parveen S, Rashid MH, Bhuiyan MSR. Assessment of breeding potential of rapeseed
400 germplasm using D² analysis. J. Expt. Biosci. 2015; 6(1): 59-64.
- 401 16. Bhatt GM. Comparison of various methods of selecting parent for hybridization in
402 common bread wheat (*Triticum aestivum*). Aus. J. Agric. Res. 1973; 24: 457-464.
- 403 17. Taiana T, Rashid MH, Parveen S, Hossain MS, Haque MA. Selection strategies to choose
404 better parents in tomato using genetic parameters. 2015; 4(1): 33-39.
- 405 18. Johnson HW, Robinson HF, Comstock RE. Estimation of genetic and environmental
406 variability in soybean. Agron. J. 1955; 47: 314-318.
- 407 19. Burton GW. Quantitative inheritance in grass pea. Proc. 6thGrassl. Cong. 1952; 1: 277-
408 283.
- 409 20. Singh RK, Chaudhary BD. Biometrical methods in quantitative genetic analysis. Kalyani
410 Publishers, New Delhi, India; 1985.
- 411 21. Allard RW. Principles of plant breeding. John Willey and Sons. Inc. New York;1960
- 412 22. Clarke GM. Statistics and experimental design. Edward Arnold. London;1973
- 413 23. Dewey DR, Lu KH. A correlation and path coefficient analysis of components of crested
414 wheat grass seed production. Agron. J.1995; 51: 515-518.
- 415 24. Malik V, Singh H, Singh D. Gene action of seed yield and other desirable characters in
416 rapeseed. Analysis Biol (Ludhiana). 1995; 11(1/2): 94-97.
- 417 25. Rao TS. Genetics of yield components in brown sarson. Geneticalberica.1977; 29(3/4):
418 219-227.
- 419 26. Tyagi MK, Chauhan JS, Kumar PR, Singh KH. Estimation of heterosis in indian mustard
420 [*Brassica juncea* (L.) Czern and Coss.]. Annals Agric. Bio. Res.2001; 69(2):193-200.

- 421 27. Afrin T, Bhuiyan MSR, Rashid MH, Parveen S. Variability and comparative analysis
422 among advanced generations of *Brassica rapa* L. Plant Knowledge J. 2016; 5(1): 18-26.
- 423 28. Varshney SK, Rai B, Singh B. Component analysis of harvest index in *Brassica* oilseeds.
424 Indian J. Agric. Rev. 1986; 20(3): 129-134.
- 425 29. Chowdhury BD, Thakural SK, Singh DP, Singh P. Genetics of yield and its components
426 in Indian mustard. Narendra Deva J. Agril. Res. 1987; 3(1): 37-43.
- 427 30. Ali MA, Md. Bhuiyan MSR, Rashid MH, Parveen S, Robbani MG, Sonom M. Breeding
428 for an ideal plant type in *Brassica rapa* L. Plant Knowledge J. 2016; 5(1): 36-43.
- 429 31. Singh RP, Malik BPS, Singh DP. Variation for morphological characters in genotypes of
430 Indian mustard. Indian J. Agric. Sci. 1987; 57(4): 225-230.
- 431 32. Lekh R, Hari S, Singh VP, Raj L, Singh H. Variability studies in rapeseed and mustard.
432 Ann. Agril. Res. 1998; 19(1): 87-88.
- 433 33. Rashid MH, Parveen S, Bhuiyan MSR. Morphological attributes species identification of
434 oleiferous *Brassica* species and better parents selection criteria for *Brassica juncea*. Int. J.
435 Current Res. 2015; 7 (9): 19847-19854.
- 436 34. Singh RS, Singh P, Dixit RK. Combining ability analysis of yield and developmental
437 traits in Indian canola (*Brassica campestris* L. var. yellow sarson). Farm Sci. 1987; 12(2):
438 170-174.
- 439 35. Mahmud F, Rasul MG, Rahim MA. Genetic diversity analysis in some advanced lines of
440 *Brassica napus*. Sci. Asia. 2008; 34: 432-434.
- 441 36. Kwon BS, Lee JI, Chae YA. Genetic studies on some agronomic characters in rapeseed.
442 Korean J. Pl. Breed. 1989; 21(1): 22-27.
- 443 37. Kumar V, Singh D. Genetics of yield and its components in Indian mustard (*Brassica*
444 *juncea* L. Czern and Coss). Crop Res. 1994; 7(2): 243-246.
- 445 38. Singh H. Genetic variability, heritability and drought indices analysis in *Brassica* species.
446 J. oilseeds Res. 1986; 3(2): 170-177.
- 447 39. Yadava YP, Singh H, Singh D. Gene action for seed yield and its attributes under
448 research. Indian J. Genet. Pl. Breed. 1993; 6(1): 168-172.
- 449 40. Khera MK, Singh P. Sensitivity and performance of some *Brassica napus* genotypes in
450 stress and non-stress environments. Crop improve. 1988; 15(2): 209-211.
- 451 41. Ejaz-ul-hasan, Mustafa HSB, Bibi T, Mahmood T. Genetic variability, correlation and
452 path analysis in advanced lines of rapeseed (*Brassica napus* L.) for yield components.
453 Cercetări agronomice în moldova. 2014; 157(1): 71-79.

- 454 42. Awas G, Mekbib F, Ayana A. Variability, heritability and genetic advance for some yield
455 and yield related traits and oil content in ethiopian coriander (*Coriandrum sativum* L.)
456 Genotypes. Int. J. Plant Breed. Genet. 2015; 9:116-125.
- 457 43. Burton GW, DeVane EH. Estimations of heritability in tall festca (*Festuca arundinacea*)
458 from replicated clonal materials. Agron. J. 1953; 45: 478-481.
- 459 44. Ahsan MZ, Majidano MS, Bhutto H, Soomro AW, Panhwar FH, Channa AR, Sial KB.
460 Genetic variability, coefficient of variance, heritability and genetic advance of some
461 *Gossypium hirsutum* L. accessions. J. Agric. Sci. 2015; 7(2): 147-151.
- 462 45. Panes VG, Sukhatme PV. Statistical methods for agricultural workers.3rd Edn. ICAR.
463 New Delhi; 1995.
- 464 46. Robinson HF, Comstock RE, Harvey PH. Estimates of heritability and the degree of
465 dominance in corn. Agron. J. 1949; 41: 353-359.
- 466 47. Parveen S. Variability study in F₂ progenies of the inter-varietal crosses of *Brassica rapa*.
467 MS thesis, Department of Genetics and Plant Breeding, Shere-e-Bangla Agricultural
468 University, Dhaka; 2007
- 469 48. Khan and Khan. Evaluation of genetic potential of some Brassica germplasm collections.
470 Int. J. Agric. Biol. 2003; 6(4): 30-31.
- 471 49. Tyagi PK, Singh K, Rao V, Kumar A. Correlation and path coefficient analysis in Indian
472 mustard (*Brassica juncea* L.). Crop Res. Hisar.1996; 11(3): 319-322.
- 473 50. Nasim M, Rahman L, Quddus MA, Shah-E-Alam M. Correlation and path analysis in
474 *Brassica campestris* L. Bangladesh J. Agril. Sci. 1994; 21(10): 15-23.
- 475 51. Saini HC, Kumar RP. Model plant architecture through association and path co-efficient
476 analysis in Indian Colza. Indian J. Agric. Res.1995; 29(3): 109-115.
- 477 52. Chauhan J, Singh P. Association of some morpho-physiological determinants with seed
478 yield in toria (*Brassica campestris* L var. toria). Thesis Abst. 1995; XI(1): 42-43.
- 479 53. Rashid MH. Characterization and diversity analysis of the oleiferous *Brassica* species.
480 MS thesis, Department of Genetics and Plant Breeding, Sher-e-Bangla Agricultural
481 University, Dhaka; 2007.
- 482 54. Mahla HR, Jambhulkar SJ, Yadav DK, Sharma R. Genetic variability, correlation and
483 path analysis in Indian Mustard. [*Brassica juncea* (L.) Czern and Coss.]. Indian J. Genet.
484 Pl. Breed. 2003; 63(2):171-172.
- 485 55. Singh RP, Khera MK, Gupta VP. Variability and correlation studies for oil and seed yield
486 in gobhi sarson. Crop improv.1991; 18(2): 99-102.

487 56. Han JX. Genetic analysis of oil content in rape *Brassica napus*. Oil Crops of China.1990;
488 2: 1-6.

UNDER PEER REVIEW