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Bacteriological Quality of Kunu Drink Sold In Bayelsa State Nigeria and the Pathogenic Potential of Some Isolates.

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9 10 ABSTRACT

Introduction: Kunu or kununzaki is a beverage drink made from grains such as millet, sorghum and maize or other combinations. It is a non-alcoholic beverage marketed in public places such as offices, markets, schools, motor parks and used in festivities such as weddings, birthday celebration, naming ceremonies etc. The high bacterial content of kunu calls for investigation.

Aim: The aim of this research is to isolate, identify bacterial contaminants in kunu and determine enterotoxin producing abilities of some isolates.

Methodology: A total of 150 bottles of kunu were purchased, 50 each from Yenagoa, Sagbama and Ogbia respectvely. Each bottle of kunu was properly mixed by gentle inversion several times and 1mL of the kunu was pipetted and added to 9mL sterile peptone water. Subsequent serial dilution was made to 10^5 . Then 0.1mL was placed on agar media in duplicate. The plates were incubated at 37°C for 18-24 hours and examined for growth.

Results: The bacteria isolated from Kunu were *S. aureus* 150 (27.8%), *E. coli* 150 (27.8%), *Bacillus sp.* 150 (27.8%) and *Staphylococci sp.* 90 (16.7%) respectively. Out of the *S. aureus* isolated, 25 (16.6%) produced enterotoxin and *E. coli* isolated, 19 (13%) produced enterotoxin respectively.

Conclusions: The contamination of kunu occurs during processing, packaging and by vendors. Improved personal hygiene of the producers, environment and proper preservation methods will reduces microbial proliferation and spoilage of kunu. The consumption of kunu is of public health interest.

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Keywords: Bacteria, Contamination, Kunu, Enterotoxin, Strains

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1. INTRODUCTION

Kunu or kununzaki is a beverage drink made from grains such as millet, sorghum and maize 17 or their combinations. It is a popular drink in northern parts of Nigeria. Kunu made from 18 19 sorghum is milky light-brown in colour, while that made from maize is whitish in colour [1.2]. 20 The grain seeds used for the production of kunu drink were allowed to germinate while 21 steeped in water for few days and after which blended with sweet potatoes and ginger or 22 pepper to form a smooth paste. The paste is divided into two, one part is placed in a vessel 23 and boiled water is added to it to form a thick mixture. The unheated half is added to the 24 previous and stirred to give a thick mixture. The mixture is left for a day or two for the grain 25 husk to settle. The husk and other sediments are filtered out of the mixture and the filtrate is 26 boiled for consumption.

Kunu is a non-alcoholic beverage marketed in several public places such as offices,
 markets, schools, motor parks and a very common consumed beverage in ocasions such as
 weddings, naming ceremonies, birthday celebrations, burials etc.[3] Kunu is an appetizer,

Comment [WU1]: Lines 17-18: Authors should provide information from literature about other similar fermented cereal based beverages from Europe.

Citation: <u>Pasqualone A, Summo</u> <u>C, Laddomada B, Mudura E, Coldea TE,</u> 2018. Effect of processing variables on the physico-chemical characteristics and aroma

of bors, a traditional beverage derived from wheat bran., Food Chemistry, 265:242-252. doi: 10.1016/j.foodchem.2018.05.095

Comment [WU2]: Line 28: Authors should check English grammar. English proofing is needed in entire document.

food complement and refresher to quench thirst [4,5,6] The proximate analysis of kunu was
 determined and the content includes; protein 2.31 – 3.63%, fats 3.35 – 3.65%, ash content

32 1.16 - 1.21% and carbohydrates 82.92 - 83.55% [2]

33 There are varieties of Kunu depending on the feed stock used for processing, they are; 34 kunuzaki, kunugyada, kunusamiya, kunubaule, kununjiko and kunugayamba [1,7] Out of 35 these, kununzaki is most widely produced and consumed [8,9] Some of the microorganisms involved in the fermentation of kunu were Lactobacilli, Lactococcus, Enterococcus, 36 37 Streptococcus, Penicillium and Sacharomyces sp.[10] The high bacterial content of kunu 38 may be an indicator of poor hygiene, poor quality cereals and water used in preparation and packaging processes [11] The bacteria isolated from kunu were E. coli 33.3%, S. aureus 39 40 26.7%, Streptococcus sp. 23.3%, Pseudomonas sp. 10% and Bacillus sp. 6.7% [11]. In a study to determine the microbiological quality of kunu in Yenagoa, Bayelsa State, the 41 42 bacteria isolated were E. coli, Enterobacter sp, Bacillus sp, Salmonella sp, Micrococcus and 43 Streptococcus sp. It was noted that most of the bacteria isolated were of public health 44 importance and they were introduced during processing and handling due to poor hygiene [12] I n another research investigating microbial quality of locally produced kunu in Calabar 45 46 the bacteria isolated were Bacillus sp. 15%, E. coli 15%, Salmonella sp. 12.5%, Streptococcus sp.10%, Pseudomonas sp. 7.5%, Proteus sp. 7.5%, Lacobacillus 22.5% and 47 S. aureus 10% respectively [13]. The microbiological qualities of kunu sold in Calaber were 48 49 below acceptable standard and unfit for human consumption soughtlt was noted that the contamination of kunu could come from different source, the bacteria isolated may present 50 51 health risk to consumers [14]. Bacteria isolated from kunu might be associated with food 52 spoilage, food infections and poisons [15]. Similar organisms were isolated by other researches and they attributed the contamination of kunu to processing and handling. The 53 54 processing and handling of kunu should be improved for consumers wellbeing [16]. 55 Kunuzaki contamination with pathogenic bacteria is of public health importance and might 56 cause diverse food related illnesses and infection to consumers [17]. The aim of this work is 57 to determine the bacteriological quality of kunu sold in Bayelsa, identy isloted bacteria and 58 the enterotoxin producing strains of some isolates. 59

60 2. MATERIAL AND METHODS

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2 2.1 Study Area

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The study was conducted in Bayelsa State, Nigeria. The samples of Zobo drink were purchased from the three (3) senatorial district headquarters of Bayelsa, namely; Yenagoa (the capital), Sagbama and Ogbia town. Bayelsa state was carved out of River State in 1996. Bayelsa is located in latitude 4° [[15]]^/ North, latitude $5^{\circ}23'$ South and longitude $5^{\circ}22'$ west and longitude $6^{\circ}45'$ east. It is bound by Delta State on the North, River State on the East and Atlantic Ocean on the West and South. Bayelsa has the largest wetland in West African sub-region. It has a population of about 1.7 million people.

72 2.2 Collection of Samples

The organism used for the experiment is *P. aeruginosa*. It was molecularly identified at
Lahor Research Laboratories, Benin, Edo State, Nigeria. The organism was stored in 50%
glycerol and kept at -20°C.

78 2.3 Bacteriological Examination of Samples79

Each sample of kunu was gently mixed by inversion several times and 1mL of the sample
 (neat) was added to 9ml of sterile peptone water (sterilized by autoclaving at 121° C for 15
 minutes). Subsequent serial dilutions were made up to 105 and 0.mL of the last dilution

Comment [WU3]: The collection of samples is not clearly described. Authors should specify if beverage samples were provided from local stores, markets, storage temperatures is the store, their shelf life, storage conditions until the moment of analyze. Volume of collect samples and how were they collected? (105) was dropped on already prepared and dried agar plates in duplicates (nutrient, MacC
 onkey and salmonella/shigella). These was spread evenly on agar media with aid of sterile
 glass rod (sterilized by dipping in absolute alcohol and flaming in bunsen flame).

The inoculated plates were allowed to dry and incubated at 37° C for 18 - 24 hours before
 examining for growth.

88 2.4 Test for Bacterial Load in stored Kunu

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A set of freshly prepared kunu were kept at room temperature and another in the refrigerator at about 4° C after the initial determination of the bacteria counts in CFU/mL. The counts from the preserved kunu at room and refrigeration temperature were determined on the second and third day respectively.

2.5 Identification of Isolated Bacteria

The bacteria isolated were identified using morphology, cultural, Gram's stain reaction,
chemical and biochemical reactions such as citrate, VP, Methyl red, indole, catalase,
coagulase and carbohydrate fermentation etc.

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101 **2.6 Detection of Enterotoxin Producing** *E. coli* from Kunu

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PROTM 0157 KIT detects enterotoxin producing E. coli. The Hardy Diagnostics PROTM 0157 KIT provides a rapid latex agglutination method to detect E. coli serogroup 0157 antigen from colonies isolated in the laboratory. These were E. coli producing verotoxin (VTproducing pathogen). Hardy diagnostic E. coli PROTM 0157 Kit contains due latex particles coated with an antiserum against E. coli 0157 antigen. When the coated latex particle is mixed with fresh colonies of E. coli serotype 0157, the bacteria will bind to the antiserum, causing the latex particles to visibly agglutinate, which is indicative of positive result.

111 <u>2.6.1 Procedure</u>

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113 The reagents were allowed to attain room temperature for about 20 minutes prior to use. 114 Then a drop of sterile saline (Cat. no.K59) was dropped in the circle on the test card and 115 overnight cultures of E. coli were emulsified by mixing it with the saline on the test card. The 116 Latex Reagents were mixed by inverting the tubes several times, prior to use. One (1) drop of E. coli PRO[™] O157 Latex Reagent were dispensed onto the test circle on the test card. 117 The Latex Reagent and the organism suspension were then mixed with the wooden 118 119 applicator provided, using the complete area of the circle. A new stick was used for each 120 reagent. Then the entire card was gently hand-rocked, allowing the mixture to flow slowly 121 over the ring area for up to 2 minutes. Under normal lighting conditions, agglutination (strong clumping) of the latex particles were examined. All organisms yielding a positive 122 123 agglutination reaction were retested with the Negative Control Latex Reagent. 124

125 2.7 Enterotoxicity Testing For S. auerus

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The Prolex[™] Staph Latex Kit provides a rapid platform for the identification of
Staphylococcal isolates particularly S. aureus that produce enterotoxin. The Prolex[™] Staph
Latex Kit utilizes blue polystyrene latex particles that have been sensitized with fibrinogen
and IgG

- 131132 2.7.1 Procedure
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134 The test kit was removed from the refrigerator 20 minutes prior to use and the latex reagents 135 were allowed to attain room temperature. The latex reagent was re-suspended by inverting the dropper bottle several times. This was followed by dispensing 1 drop of Staph Test Latex 136 137 Reagent into a circle on the test card. A sterile loop was used to transfer two colonies of the test isolate into the circle. The reagent and colonies were mixed and spread to cover the 138 139 entire area of the circle and this was rocked gently on the card allowing the mixture to flow 140 slowly over the entire test ring area. Agglutination was observed for 20 seconds. Negative 141 Control Latex Reagent was included as quality control. Strong agglutination within 20 142 seconds with the Staph Test Latex Reagent indicates positive result.

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145 3. RESULTS

146 147 **3.1**

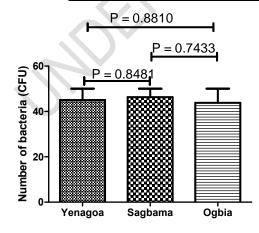
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A total of 150 samples of kunu were examined for the presence of bacteria. The result 149 150 obtained showed that from Yenaguoa S. aureus was 50 (27.8%), E. coli 50 (27.8%), Bacillus sp. 50 (27.8%), and Streptococcus sp. 30 (16.7%) respectively. From kunu bought from 151 Sagbama, S. aureus were 50 (27.0%), E. coli 50 (28.6%), Bacillus sp. 50 (27.0%) and 152 153 Streptococcus sp. 35 (18.9%) respectively. Kunu purchased from Ogbia town had S. aureus 50 (28.6%), E. coli 50 (28.6%), Bacillus sp. 50 (28.6%) and Streptococcus sp. 25 (14.3%) 154 155 respectively. The overall percentage occurrences of isolated bacteria were S. aureus 150 (27.8%), E. coli 150 (27.8%), Bacillus sp. 150 (27.8%) and Streptococcus sp. 90 (16.9%) 156 157 respectively shown of Table 1.

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Table 1. Percentage Occurrence of Bacteria Isolated from Kunu Drink

Location	S. aureus	E. coli	Bacillus sp.	Streptococcus sp.	Total
Yenegoa	50 (27.8)	50 (27.8)	50 (27.8)	30 (16.7)	185 (33.3)
Sagbama	50 (27.8)	50 (27.8)	50 (27.8)	35 (27.8)	185 (34.6)
Ogbia	50 (27.8)	50 (27.8)	50 (27.8)	25 (14.3)	175 (32.4)
Total	150 (27.8)	150 (27.8)	150 (27.8)	90 (16.7)	540



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161 Fig Comparison of the bacteria isolated from different locations

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spelling.

163 4.2 Enterotoxin Producing S. aureus and E. coli from Kunu

A total of 50 S. aureus were isolated from kunu in Yenegoa out of which 7 (14%) were
enterotoxin producing strains and 50 S. aureus were isolated from Sagbama, 10 (20%)
produced enterotoxin, while in Ogbia, Town 50 S. aureus were isolated, 8 (16%) were
positive for enterotoxin production respectively. Overall total of S. aureus that prodused
enterotoxin were 25 (16.6%) as shown in Table 2.

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170 Table 2. Percentage Occurrence of Enterotoxin Producing S. aureus and E. coli from Kunu

Location	S. aureus	Number positive	E. coli	Number positive
Yenegoa	50	7 (14)	50	7 (14)
Sagbama	50	10 (20)	50	5 (10)
Ogbia	50	8 (16)	50	7 (14)
Total	150	25 (16.6)	150	19 (13)

171 Numbesr in parentheses =percentages

172 Numbers in parentheses = percentages

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 Table3. Bacterial Counts from Preserved Refrigerated and Non-Refrigerated Kunu

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 (CFU/mL)

Temperature	0 Hr	24 Hrs	48 Hrs
Refrigeration	7.48×10^{5}	8.40×10^{5}	8.72×10^{5}
Room	7.48×10^{5}	11.96×10^{5}	13×10^{5}

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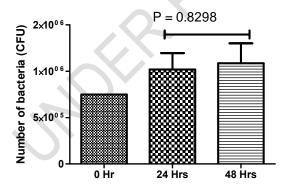


Fig. Comparison of bacterial counts from 0 Hr, 24Hrs and 48Hrs.

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192 The bacteria isolated from kunu drinks in this study were Staphylococcus sp, E. coli, Bacillus 193 sp. and Streptococcus sp. The contamination after the boiling process (post CCP 194 contamination) might be responsible for the presence bacteria in kunu. Similar bacteria were 195 isolated from kunu by other researchers in other parts of Nigeria such as Bacillus sp. Salmonella sp. Micrococcus, Staphylococcus aureus and Streptococcus sp.[13], E. coli, S. 196 197 aureus, Salmonella sp and Shigella sp in Maiduguri [18], In Kaduna, Lactobacillus, Bacillus sp. and E. coli [19], while some reported E. coli, S. aureus, Streptococcus pyogenes [20]. 198 199 Other researchers had S. aureus 4(10%), Lactobacillus 9(22.5%), Proteus sp. 3(7.5%), Streptococcus sp. 4(10%), Pseudmonas sp. 3(7.5%), Bacillus sp. 6(15%) and E. coli 6(15%) 200 respectively [13]. Most researchers had comparable bacteria in Port Harcourt [7], Oyo and 201 Lagos [1,2], Kano [21], Maiduguri [18], Jalingo [3] in Calabar cross River State [5]. The poor 202 microbial quality of kunu produced locally were because of poor hygiene and poor 203 environmental conditions under which kunu were produced. Health hazard may be 204 205 associated with consumer of locally produced kunu.

206 Among the bacteria isolated, Staphylococci were the most prevalent organism. 207 Staphylococcus is normal inhabitant of the human body which can be found on the skin, 208 mouth, nostril, hands, various surfaces etc. these were possible sources from where Staphylococci sp. can contaminate kunu during processing and packaging. Bacteria might 209 be present in storage containers, sieve used to filter the finished product and contamination 210 211 from handlers. The percentage of S. aureus was 21.7% [11], while in this study S. aureus 212 were 26.7%. The prevalence of E. coli was 27.8% from this study but others E. coli as 33.3% 213 [11] and E. coli as 5.0% [5]. E. coli is the most prevalent aerobic bacteria in human and mammal faeces. Contamination by E. coli might be by faecal contamination, contaminated 214 215 water, handlers, processing and packing. Bacillus sp. were 27.8% occurrence. Bacillus sp. are geophilic and the spores are found in the soil, dust etc. The contamination of grains and 216 spices by Bacillus sp. and their spores from soil and dust are likely. The percentage of 217 Bacillus sp. isolated other workers were 23.3% [5], and 7.6% [12]. Spores may survive 218 during boiling at about 100°C (the only Critical Control Point (CCP) in the processes of kunu 219 220 production) and germinate to re-contaminate kunu. Kunu should be preserved under refrigeration temperature at 4° C and or pasteurized to reduce the microbial load and to 221 increase the shelf life [21,5]. 222

223 It was noted that local drinks such as kunu may act as vehicle for the transmission of 224 zoonotic and bacterial infections such as staphylococosis, salmonellosis, shigellosis, 225 tuberculosis, listerosis etc. (Umaru et al., 2014)[3]. Kunu as a beverage sold in public places 226 such as markets, schools, offices etc. is patronized because of the cheap price compared to 227 other soft drinks and is served in occasions such as weddings, naming ceremonies, birthday celebrations for economic reasons and public acceptability. The consumption of 228 229 contaminated kunu drink may result in outbreak of food borne illness. Preparation of kunu in 230 environment with poor sanitary conditions predisposes the preparation and packaging 231 processes to contamination and exposes the consumers to health risk.

The isolation of 13% of *E. coli* and 17% of *S. aureus* capable of producing enterotoxin indicates that the consumption of kunu is of public health concern. The production of enterotoxin in bacteria is mostly associated with gastrointestinal disturbances and/ or food borne illnesses.

236 Conclusions: The production kunu as non alcoholic beverage for public consumption 237 should be regulated by appropriate regulatory agencies to reduce the risk of consumer 238 infection. Producers should be made to have fair knowledge of food preservation and food 239 sanitation. The isolation of 13% of *E. coli* and 17% of *S. aureus* capable of producing 240 enterotoxin indicates that the consumption of locally produced kunu is a public health 241 concern.

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UNDERPETRATION